

MIT Open Access Articles

Cellular and synaptic network defects in autism

The MIT Faculty has made this article openly available. *Please share* how this access benefits you. Your story matters.

Citation: Peca, Joao, and Guoping Feng. "Cellular and Synaptic Network Defects in Autism." Current Opinion in Neurobiology 22, no. 5 (October 2012): 866–872.

As Published: http://dx.doi.org/10.1016/j.conb.2012.02.015

Publisher: Elsevier

Persistent URL: http://hdl.handle.net/1721.1/102179

Version: Author's final manuscript: final author's manuscript post peer review, without publisher's formatting or copy editing

Terms of use: Creative Commons Attribution-Noncommercial-NoDerivatives





NIH Public Access

Author Manuscript

Curr Opin Neurobiol. Author manuscript; available in PMC 2013 October 01.

Published in final edited form as:

Curr Opin Neurobiol. 2012 October; 22(5): 866–872. doi:10.1016/j.conb.2012.02.015.

Cellular and synaptic network defects in autism

João Peça¹ and Guoping Feng^{1,2}

¹McGovern Institute for Brain Research, Department of Brain and Cognitive Sciences, Massachusetts Institute of Technology, Cambridge, MA 02139, USA

²Stanley Center for Psychiatric Research, Broad Institute, Cambridge, MA 02142, USA

Abstract

Many candidate genes are now thought to confer susceptibility to autism spectrum disorder (ASD). Here we review four interrelated complexes, each composed of multiple families of genes that functionally coalesce on common cellular pathways. We illustrate a common thread in the organization of glutamatergic synapses and suggest a link between genes involved in Tuberous Sclerosis Complex, Fragile X syndrome, Angelman syndrome and several synaptic ASD candidate genes. When viewed in this context, progress in deciphering the molecular architecture of cellular protein-protein interactions together with the unraveling of synaptic dysfunction in neural networks may prove pivotal to advancing our understanding of ASDs.

Introduction

Autistic behaviors were independently identified as recognizable syndromes in the early 20th century by Kaner [1], Asperger [2] and Heller [3]. In the DSM-IV, the autism diagnosis spans a broad continuum of what are collectively known as ASDs or Pervasive Developmental Disorders. These encompass Rett syndrome, Asperger syndrome, Childhood Disintegrative Disorder (Heller syndrome), idiopathic autism and atypical autism (Pervasive Developmental Disorder Not Otherwise Specified) [4]. Although heterogeneous, ASDs are united by a combination of three core behavioral symptoms: a) impairments in language and communication; b) abnormal sociability or deficits in social interaction; and c) restricted interested and repetitive or stereotyped behaviors. In addition to these, other common pathological disturbances include gait and motor disturbances, anxiety, epilepsy, sensorial abnormalities, sleep disturbances and comorbidity with psychiatric disorders such as attention deficit hyperactivity disorder, obsessive-compulsive disorder (OCD) and mood disorders [5][•]. ASDs are considered to be highly heritable and a genetic predisposition is thought to play an important role in the etiology of these disorders [5]. For Rett syndrome in particular, mutations in the MECP2 gene are known to be responsible for the majority of cases and only a small percentage remain idiopathic. Moreover, autism is also highly prevalent in patients afflicted with other genetic syndromes such as Tuberous Sclerosis, Cowden, Fragile-X, Angelman, 16p11.2 deletion, Cohen, Smith-Magenis, and Phelan-McDermid syndromes among others [6]. Therefore, the prominence of ASD symptomatology associated with genetic conditions again further suggests that genetic factors may play a key role in the etiological origins of the idiopathic autism phenotype.

^{© 2012} Elsevier Ltd. All rights reserved.

Publisher's Disclaimer: This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

Page 2

As our understanding and sophistication for genetic diagnosis grows, so do the number of discrete genes and chromosomal regions that may afford susceptibility to ASDs. Currently, more than 100 genes are susceptibility candidates, indicating ASDs represent a collection of conditions with heterogeneous causation [7][•]. Nevertheless, recent evidences have unveiled a remarkable convergence of several of these genes on common cellular pathways that intersect at neuronal glutamatergic synapses. Functional overlap such as this may provide a framework that reconciles the presence of a wide array of genes triggering a set of conditions with a high degree of phenotypic similarity in the core behavioral manifestations. Therefore, it is expected that the identification of common pathways, the better understating of the underlying neurobiological disease substrates, and the analysis of neural circuitry dysfunctional in ASD may open the door to new and broadly effective therapies. Here, we highlight some of the intersecting pathways overlapping at glutamatergic synapses and explore how these pathways interconnect. Our focus is directed to the Shank3 and synaptic protein network, and their relations with the genes involved in Tuberous Sclerosis Complex, Fragile X syndrome and Angelman syndrome.

Shank3 and Postsynaptic Density Proteins in ASD

The interdependence among proteins working as a biochemical complex, suggests that even single gene disorders reflect the perturbation of complex intracellular pathways and cellular networks. Therefore, an emerging notion is that if a particular gene or protein plays a role in pathogenesis, its direct and close network interactors are associated with an increased likelihood of precipitating similar endophenotypical manifestations when they are dysfunctional [8,9]. Thus, the analysis of protein networks and their association at the cellular level may shed light on the etiological landscape of interrelated conditions. For ASDs, the transsynaptic complex composed of Neurexin/Neuroligin/PSD-95/SAPAP/Shank pathway is a good example of a set of genes converging on both function, location and associated disorders (Figure 1). From these, Neurexin-1 [10-13], Neuroligin-3 and -4 [14], PSD-95 [15], SAP97 [16], SAPAP2 [17], Shank2 and Shank3 [17–19]** have all been implicated in ASDs and validated across multiple studies. Shank3 in particular is present in the terminal q13 region of chromosome 22 that is deleted/rearranged in patients afflicted with Phelan-McDermid syndrome [20-22]. Interestingly, while even minimal deletions in Shank3 have been shown to induce the complete symptomatology of PMS, a ring chromosome aberration that leaves the Shank3 gene intact does not [23,24]. Moreover, point mutations in the Shank3 gene have been identified in patients diagnosed with ASDs [18,25]. Together, evidence from mutations and chromosomal abnormalities affecting Shank3 in ASDs and PMS patients suggest that deficits in this gene can trigger major neurobehavioral complications and autism-spectrum related behaviors.

At the cellular level, all three genes in the Shank family (Shank1-3) give rise to proteins containing multiple protein-protein interaction domains. Shank proteins prominently concentrate in the postsynaptic density (PSD) of dendritic spines [26]. The PSD is key for organizing and maintaining proper synaptic communication, and within this structure Shanks have been hypothesized to function as master scaffolds. Among other roles, Shank can act by bridging together multiple synaptic pathways and provide a link between the PSD and the actin cytoskeleton. Together, Shank and Homer multimers [27,28]* form a common platform (with the SAPAP and PSD-95 family of proteins) for the anchoring of ionotropic (AMPAR and NMDAR) and metabotropic glutamate receptors (mGluR) at synapses (Figure 1) [29]. To understand the function of the Shank proteins, overexpression and knockdown strategies have been used to show a clear role in the genesis, maturation and stabilization of neuronal spines [30,31]**. Shank3 overexpression in particular has been demonstrated to induce the formation of functional spines in cultured aspiny cerebellar granule cells [31]**. More recently, targeted manipulations in the mouse genome have provided further insight

into the *in vivo* functions of Shank1 and Shank3. An analysis of these mutant animals showed that spine density was reduced in hippocampal neurons of Shank1 knockout mice and in medium spiny neurons of Shank3 knockout mice [32,33]^{••}. Moreover, the four different lines of Shank3 mutant mice characterized so far have unveiled neuronal deficits concentrating in decreases of glutamatergic signalizing, loss of synaptic strength, and at the behavioral level, in deficiencies pertaining to social interaction and other ASD-relevant behaviors [33–36]^{••}. Interestingly, the expression of Shank3 is enriched in the striatum [37] and Shank3 mutant mice have cortico-striatal circuitry defects [33]^{••}.

Tuberous Sclerosis Complex

Tuberous Sclerosis Complex (TSC) originates from two genetic disorders caused by mutation in either the TSC1/hamartin or TSC2/tuberin genes [38,39]. TSC is commonly recognized by the appearance of hamartomas (tumor-like nodules) in the skin and the central nervous system. Common neurological and behavioral symptoms include seizures, developmental delays, intellectual disability [40] and up to 45% of patients also fulfill the diagnostic criteria for ASD [41]. Hamartin and tuberin form a complex that participates in the inhibition of the mammalian target of rapamycin (mTOR) [42]. mTOR is a biochemical energy sensor and a regulator of cell growth that plays an important role in stimulating protein synthesis at the synapse [43]. In line with these roles, Ehninger and colleagues observed a basal increase in activity of the mTOR signaling-pathway in Tsc2 heterozygous mutant mice [44]. This in turn leads to abnormal neuronal plasticity and memory defects that are rescued by treating the Tsc2 mutant mice with rapamycin—an inhibitor for mTOR activity [44][•]. Additionally, post-mitotic perturbation of Tsc1 levels leads to neuronal morphology alterations, such as an increase in soma and spine size, but a decrease in dendritic spine density [45]. Interestingly, recent work from Zoghbi and colleagues has provided evidence for the existence of shared binding partners (e.g. Homer3 and Actinin) between the Tsc1/Tsc2 protein complex and Shank3, indicating an intermolecular overlap between these two ASD relevant pathways [46]". Furthermore, the Tsc1/Tsc2 complex also overlaps with the Fragile X mental retardation 1 (FMR1) gene network at synaptic sites. Specifically, FMR1 was shown to interact with TSC2 transcripts and other autism associated genes [47][•]. Nevertheless, recent results alert to the notion that the precise molecular changes occurring from the disruption of these pathways may not be trivial to predict. Specifically, Tsc2 mutant mice display defects that oppose those caused by FMR1 mutants in regards to mGluR-dependent synaptic plasticity [48]. Moreover, the genetic cross of both mutant mice for Tsc2+/ and Fmr1/y rescued synaptic and behavioral impairments, further corroborating an apposition in molecular roles [48]. This could suggest that Tsc2 and FMRP proteins/transcripts may be regulated through complex feedback and compensatory mechanism, or alternatively, that Tsc2/mTOR and FMR1 prime the regulation of distinct pools of synaptic proteins necessary for mGluR-dependent synaptic plasticity.

Fragile X Syndrome

Trinucleotide repeats and mutations leading to the silencing of fragile X mental retardation protein (FMRP) production gives rise to the Fragile X syndrome (FXS) [49,50]. This disorder is the single most common cause for heritable intellectual disability, and due to the high prevalence in the general population; it is also the most widely diagnosed cause for ASD [51]. The fragile X mental retardation protein (FMRP) binds and transports mRNA transcripts to spines where it associates with polyribosomes [52] to exert control over protein translation and regulate several families of synaptic and other autism-relevant proteins [53,54]. Recent evidence suggests that the neuronal deficits in FXS may occur in part due to the loss of translational constraint imposed by FMRP on transcripts for Neuroligin-3, Neurexin-1, Shank3, PTEN (Cowden syndrome), TSC2 and NF1

(Neurofibromatosis type 1) [47][•]. In hippocampal synapses, long-term depression (LTD) induced by group 1 mGluR requires translation of synaptic transcripts [55]. In FMR1 mutant mice, mGluR-dependent LTD is enhanced [56]; and FMRP mutant mice crossed to harbor a mGluR5 (a group 1 mGluR) heterozygous deletion display an amelioration of several abnormalities present in the FMRP mutants [57]. This has led to the hypothesis that the manipulation of mGluR signaling is a plausible new target for treating symptoms in FXS patients [58]. Furthermore, the FMRP/mGluR pathway also intersects with that of Shank proteins. Besides associating with FMRP [47][•], Shank3 mRNA is one of rare transcripts strongly enriched in dendritic spines [37], and protein levels of Shank1 and Shank3 are increased in the PSD of FMR1 knockout mice [59]. Therefore, the interaction between mGluRs and Shank3 through Homer, the interrelatedness in FMRP/mGluR synaptic pathways and the regulation of Shank3 transcripts by FMRP again points to a molecular and functional convergence at the PSD of glutamatergic synapses.

Ubiquitin-pathway Genes

While inducing or inhibiting protein synthesis can accomplish control over synaptic protein function, another prominent cellular mechanism for the rapid regulation of protein abundance is the ubiquitination and proteasomal system. Perhaps the correlation between the ubiquitin system and autism is best illustrated by UBE3A (Ubiquitin protein ligase 3A), a gene which when silenced or mutated can cause Angelman syndrome, a disorder with high comorbidity with ASDs [60,61]. In addition to UBE3A, information collected from ASD patients revealed copy number variations (CNV) in other ubiquitin genes, including – PARK2, RFWD2 and FBXO40 [62]*. In vivo, Ube3a localizes to synaptic sites and plays a role in dendritic spine development [63]. Ube3a deficient mice display abnormal spine morphology and decreases in spine density [63]. However, even while the full gamut of Ube3a targets may not be entirely understood, the in vivo importance of this gene is clear for synaptic plasticity in and in the experience-dependent maturation of neocortical circuits in Ube3a mutant mice [64,65]. Additionally an Ube3a mouse model engineered to carry an increased gene dosage was also reported to display the core symptomatology of ASD [66]. Furthermore at the synaptic level the increase in Ube3a level promoted the suppression of excitatory glutamatergic signaling [66], suggesting a critical role for this protein at excitatory synapses [66]. Interestingly, changing neuronal activity levels produces a rapid bidirectional change in the composition of PSD proteins, an effect that is mediated by the proteasome system [67][•]. Additionally, from a set of PSD proteins investigated, Shanks were among the few found to display obvious variations in levels of ubiquitination in response to changes in neuronal activity [67]. More recently, a mutant mouse line that produces a truncated Shank3 protein was found increased ubiquitination and abnormal protein turnover of other Shank3 isoforms and also of NMDA receptors [34][•]. Together, these results establish the importance of ubiquitin ligases to the regulation of excitatory synapses, in the etiology of ASDs and also hint at overlap with the network surrounding Shank proteins.

Concluding remarks

Though recent years have seen important strides in the discovery of genes that may confer susceptibility to ASDs, it is increasingly important to further our grasp on the cellular and molecular consequences of putative candidate gene mutations. Additionally, some of the more auspicious results to date have come from studies showing that postnatal genetic rescue of certain genetic lesions can ameliorate phenotypical defects, as demonstrated for genetic mouse models of Rett syndrome, Fragile X syndrome and OCD-like behaviors [68–71][•]. These results establish the prospect of reversibility of symptoms in some neurodevelopmental disorders, encompassing also reversibility at the level of cellular defects underlying aspects of these disorders. Thus, the continued accrual of information

across multiple animal models and the delineation of which nodes may be more amenable to intervention will be essential. This will require further experimental data confirming the relationship between these pathways and the identification of targets for therapeutical intervention.

Yet, one concern that arises as a corollary to the molecular interconnectedness of ASD gene pathways is as follows: mutations affecting ubiquitously expressed gene products may result in ASD specific phenotypes in an indirect manner, due to perturbation of functional interactions with affiliated gene products that are enriched or expressed only in specific brain regions. This could lead to a focal disruption of discrete brain circuits. As a putative example, FMRP mutations, while broadly affecting protein translation of many target transcripts, would only precipitate aberrant Shank3 mRNA translation in neurons and brain regions that normally express Shank3. Understanding this potential cellular asymmetry in brain disorders will require continued exploration of cell autonomous function but with additional consideration to the unique expression patterns of functionally linked genes, as well as higher order neural networks between brain regions. Thus, even if genetic mutations form the etiology of many ASDs, it is essential to understand the neuropathology in the context of the specific brain circuits underpinning the hallmark neurological and behavioral manifestations.

Acknowledgments

We thank Dr. Jonathan Ting for critical comments on this manuscript. G.F. is supported by grants from the National Institutes of Health (NIMH R01MH081201), The Hartwell Foundation, Simons Foundation Autism Research Initiative (SFARI), Stanley Center for Psychiatric Research, and the SPARC program from Broad Institute of MIT and Harvard. J.P. is funded by an Autism Speaks Translational Postdoctoral Fellowship (#7649).

References and Recommended Readings

- 1. Kanner L. Autistic disturbances of affective contact. Nervous Child. 1943:217-250.
- Asperger H. Die autistischen Psychopathen im Kindesalter. Archiv f
 ür Psychiatrie und Nervenkrankheiten. 1944:76–136.
- 3. Heller T. Ueber Dementia infantilis. Zeitschrift fuer die Erforschung und Behandlung des Jugendlichen Schwachsinns. 1908; 2:17–28.
- 4. American Psychiatric Association., American Psychiatric Association. Task Force on DSM-IV.: Diagnostic and statistical manual of mental disorders : DSM-IV-TR. 4. Washington, DC: American Psychiatric Association; 2000.
- 5•. Geschwind DH. Advances in autism. Annu Rev Med. 2009; 60:367–380. This review focuses on recent advances in the genetics of autism and emphasizes some of the etiological heterogeneity presented by patients. The author also discusses the data on the co-morbidity of ASDs with psychiatric disorders and suggests that the three autism endophenotypes may provide a referencing diagnosis to increase the ability to detect common genetic risk variants associated with ASDs. [PubMed: 19630577]
- Cohen D, Pichard N, Tordjman S, Baumann C, Burglen L, Excoffier E, Lazar G, Mazet P, Pinquier C, Verloes A, et al. Specific genetic disorders and autism: clinical contribution towards their identification. Journal of autism and developmental disorders. 2005; 35:103–116. [PubMed: 15796126]
- 7•. Betancur C. Etiological heterogeneity in autism spectrum disorders: more than 100 genetic and genomic disorders and still counting. Brain research. 2011; 1380:42–77. This review provides an exhaustive compilation of the genes described in the literature (100+) that may afford susceptibility to ASDs. The author focuses and highlights the etiological genetic heterogeneity of the broad gamut of ASDs. [PubMed: 21129364]
- 8••. Barabasi AL, Gulbahce N, Loscalzo J. Network medicine: a network-based approach to human disease. Nat Rev Genet. 2011; 12:56–68. The authors discuss how diseases often reflect the perturbations of complex intracellular network and intercellular networks. The authors also

discuss the concept of interactome, prediction of disease genes and disease modules and their relevance to human medicine. [PubMed: 21164525]

- Goh KI, Cusick ME, Valle D, Childs B, Vidal M, Barabasi AL. The human disease network. Proc Natl Acad Sci U S A. 2007; 104:8685–8690. [PubMed: 17502601]
- Kim HG, Kishikawa S, Higgins AW, Seong IS, Donovan DJ, Shen Y, Lally E, Weiss LA, Najm J, Kutsche K, et al. Disruption of neurexin 1 associated with autism spectrum disorder. Am J Hum Genet. 2008; 82:199–207. [PubMed: 18179900]
- Yan J, Noltner K, Feng J, Li W, Schroer R, Skinner C, Zeng W, Schwartz CE, Sommer SS. Neurexin 1alpha structural variants associated with autism. Neuroscience letters. 2008; 438:368– 370. [PubMed: 18490107]
- Wisniowiecka-Kowalnik B, Nesteruk M, Peters SU, Xia Z, Cooper ML, Savage S, Amato RS, Bader P, Browning MF, Haun CL, et al. Intragenic rearrangements in NRXN1 in three families with autism spectrum disorder, developmental delay, and speech delay. Am J Med Genet B Neuropsychiatr Genet. 2010; 153B:983–993. [PubMed: 20162629]
- Feng J, Schroer R, Yan J, Song W, Yang C, Bockholt A, Cook EH Jr, Skinner C, Schwartz CE, Sommer SS. High frequency of neurexin 1beta signal peptide structural variants in patients with autism. Neurosci Lett. 2006; 409:10–13. [PubMed: 17034946]
- 14. Jamain S, Quach H, Betancur C, Rastam M, Colineaux C, Gillberg IC, Soderstrom H, Giros B, Leboyer M, Gillberg C, et al. Mutations of the X-linked genes encoding neuroligins NLGN3 and NLGN4 are associated with autism. Nat Genet. 2003; 34:27–29. [PubMed: 12669065]
- 15. Feyder M, Karlsson RM, Mathur P, Lyman M, Bock R, Momenan R, Munasinghe J, Scattoni ML, Ihne J, Camp M, et al. Association of mouse Dlg4 (PSD-95) gene deletion and human DLG4 gene variation with phenotypes relevant to autism spectrum disorders and Williams' syndrome. The American journal of psychiatry. 2010; 167:1508–1517. [PubMed: 20952458]
- 16. Willatt L, Cox J, Barber J, Cabanas ED, Collins A, Donnai D, FitzPatrick DR, Maher E, Martin H, Parnau J, et al. 3q29 microdeletion syndrome: clinical and molecular characterization of a new syndrome. American journal of human genetics. 2005; 77:154–160. [PubMed: 15918153]
- 17•. Pinto D, Pagnamenta AT, Klei L, Anney R, Merico D, Regan R, Conroy J, Magalhaes TR, Correia C, Abrahams BS, et al. Functional impact of global rare copy number variation in autism spectrum disorders. Nature. 2010; 466:368–372. In this article, the authors identify copy number variations in several synaptic genes suggesting that rare variations may significantly contribute to ASDs. They identify variants in SHANK2, DLGAP2, SYNGAP1, and the DDX53–PTCHD1 locus. [PubMed: 20531469]
- 18••. Durand CM, Betancur C, Boeckers TM, Bockmann J, Chaste P, Fauchereau F, Nygren G, Rastam M, Gillberg IC, Anckarsater H, et al. Mutations in the gene encoding the synaptic scaffolding protein SHANK3 are associated with autism spectrum disorders. Nat Genet. 2007; 39:25–27. The authors identify various rare point mutations in the SHANK3 gene in patients affected with ASDs outside of a diagnosed Phelan-McDermid syndrome. [PubMed: 17173049]
- 19•. Berkel S, Marshall CR, Weiss B, Howe J, Roeth R, Moog U, Endris V, Roberts W, Szatmari P, Pinto D, et al. Mutations in the SHANK2 synaptic scaffolding gene in autism spectrum disorder and mental retardation. Nat Genet. 2010; 42:489–491. Using techniques to identify copy number variations and DNA sequencing, the authors find mutations in the SHANK2 gene in patients with intellectual disability and ASDs. [PubMed: 20473310]
- 20. Wilson HL, Wong AC, Shaw SR, Tse WY, Stapleton GA, Phelan MC, Hu S, Marshall J, McDermid HE. Molecular characterisation of the 22q13 deletion syndrome supports the role of haploinsufficiency of SHANK3/PROSAP2 in the major neurological symptoms. Journal of medical genetics. 2003; 40:575–584. [PubMed: 12920066]
- 21. Bonaglia MC, Giorda R, Mani E, Aceti G, Anderlid BM, Baroncini A, Pramparo T, Zuffardi O. Identification of a recurrent breakpoint within the SHANK3 gene in the 22q13. 3 deletion syndrome. Journal of medical genetics. 2006; 43:822–828. [PubMed: 16284256]
- 22. Phelan K, McDermid HE. The 22q13.3 Deletion Syndrome (Phelan-McDermid Syndrome). Molecular Syndromology. 2011
- Jeffries AR, Curran S, Elmslie F, Sharma A, Wenger S, Hummel M, Powell J. Molecular and phenotypic characterization of ring chromosome 22. American journal of medical genetics Part A. 2005; 137:139–147. [PubMed: 16059935]

- 24. Misceo D, Rodningen OK, Baroy T, Sorte H, Mellembakken JR, Stromme P, Fannemel M, Frengen E. A translocation between Xq21.33 and 22q13.33 causes an intragenic SHANK3 deletion in a woman with Phelan-McDermid syndrome and hypergonadotropic hypogonadism. American journal of medical genetics Part A. 2011; 155A:403–408. [PubMed: 21271662]
- Moessner R, Marshall CR, Sutcliffe JS, Skaug J, Pinto D, Vincent J, Zwaigenbaum L, Fernandez B, Roberts W, Szatmari P, et al. Contribution of SHANK3 mutations to autism spectrum disorder. Am J Hum Genet. 2007; 81:1289–1297. [PubMed: 17999366]
- 26. Sheng M, Kim E. The Shank family of scaffold proteins. Journal of cell science. 2000; 113 (Pt 11):1851–1856. [PubMed: 10806096]
- 27•. Hayashi MK, Tang C, Verpelli C, Narayanan R, Stearns MH, Xu RM, Li H, Sala C, Hayashi Y. The postsynaptic density proteins Homer and Shank form a polymeric network structure. Cell. 2009; 137:159–171. Biochemical characterization and protein crystallography of Shank3 and Homer provide structural findings as to how these proteins may associate in a higher order molecular platform for the broader PSD. [PubMed: 19345194]
- 28•. Baron MK, Boeckers TM, Vaida B, Faham S, Gingery M, Sawaya MR, Salyer D, Gundelfinger ED, Bowie JU. An architectural framework that may lie at the core of the postsynaptic density. Science. 2006; 311:531–535. Biochemical characterization and protein crystallography of Shank3 and Homer provide structural findings as to how these proteins may associate in a higher order molecular platform for the broader PSD. [PubMed: 16439662]
- Tu JC, Xiao B, Naisbitt S, Yuan JP, Petralia RS, Brakeman P, Doan A, Aakalu VK, Lanahan AA, Sheng M, et al. Coupling of mGluR/Homer and PSD-95 complexes by the Shank family of postsynaptic density proteins. Neuron. 1999; 23:583–592. [PubMed: 10433269]
- Sala C, Piech V, Wilson NR, Passafaro M, Liu G, Sheng M. Regulation of dendritic spine morphology and synaptic function by Shank and Homer. Neuron. 2001; 31:115–130. [PubMed: 11498055]
- 31••. Roussignol G, Ango F, Romorini S, Tu JC, Sala C, Worley PF, Bockaert J, Fagni L. Shank expression is sufficient to induce functional dendritic spine synapses in aspiny neurons. J Neurosci. 2005; 25:3560–3570. This article describes how Shank3 expression can induce functional dendrite spines, while conversely a Shank3 knockdown leads to a reduction in spine density. The authors also propose a specific role for discrete Shank3 protein domains in spine morphology and cellular localization. [PubMed: 15814786]
- 32. Hung AY, Futai K, Sala C, Valtschanoff JG, Ryu J, Woodworth MA, Kidd FL, Sung CC, Miyakawa T, Bear MF, et al. Smaller dendritic spines, weaker synaptic transmission, but enhanced spatial learning in mice lacking Shank1. J Neurosci. 2008; 28:1697–1708. [PubMed: 18272690]
- 33•. Peca J, Feliciano C, Ting JT, Wang W, Wells MF, Venkatraman TN, Lascola CD, Fu Z, Feng G. Shank3 mutant mice display autistic-like behaviours and striatal dysfunction. Nature. 2011; 472:437–442. A total of four Shank3 mutant mouse lines described by 3 independent laboratories. Despite the variety of mutations and methodologies, a striking convergence of ASD-like phenotypes is observed across Shank3 mouse lines. A perturbation in the glutamatergic system is also consistently observed. Bozdagi et al, offered the first analysis of heterozygous mutant Shank3 mice; Wang et al. described in detail the multiple different protein isoforms arising from the Shank3 gene; Peca et al. describe the enrichment of Shank3 in the striatum and observe defects in cortico-striatal circuits; Bozdagi et al. suggest that part of the cellular defects in the Shank3 mutants are due to defects in the ubiquitination system. [PubMed: 21423165]
- 34•. Bangash MA, Park JM, Melnikova T, Wang D, Jeon SK, Lee D, Syeda S, Kim J, Kouser M, Schwartz J, et al. Enhanced polyubiquitination of Shank3 and NMDA receptor in a mouse model of autism. Cell. 2011; 145:758–772. A total of four Shank3 mutant mouse lines described by 3 independent laboratories. Despite the variety of mutations and methodologies, a striking convergence of ASD-like phenotypes is observed across Shank3 mouse lines. A perturbation in the glutamatergic system is also consistently observed. Bozdagi et al, offered the first analysis of heterozygous mutant Shank3 mice; Wang et al. described in detail the multiple different protein isoforms arising from the Shank3 gene; Peca et al. describe the enrichment of Shank3 in the striatum and observe defects in cortico-striatal circuits; Bozdagi et al. suggest that part of the cellular defects in the Shank3 mutants are due to defects in the ubiquitination system. [PubMed: 21565394]

- 35•. Wang X, McCoy PA, Rodriguiz RM, Pan Y, Je HS, Roberts AC, Kim CJ, Berrios J, Colvin JS, Bousquet-Moore D, et al. Synaptic dysfunction and abnormal behaviors in mice lacking major isoforms of Shank3. Human molecular genetics. 2011; 20:3093–3108. A total of four Shank3 mutant mouse lines described by 3 independent laboratories. Despite the variety of mutations and methodologies, a striking convergence of ASD-like phenotypes is observed across Shank3 mouse lines. A perturbation in the glutamatergic system is also consistently observed. Bozdagi et al, offered the first analysis of heterozygous mutant Shank3 mice; Wang et al. described in detail the multiple different protein isoforms arising from the Shank3 gene; Peca et al. describe the enrichment of Shank3 in the striatum and observe defects in cortico-striatal circuits; Bozdagi et al. suggest that part of the cellular defects in the Shank3 mutants are due to defects in the ubiquitination system. [PubMed: 21558424]
- 36•. Bozdagi O, Sakurai T, Papapetrou D, Wang X, Dickstein DL, Takahashi N, Kajiwara Y, Yang M, Katz AM, Scattoni ML, et al. Haploinsufficiency of the autism-associated Shank3 gene leads to deficits in synaptic function, social interaction, and social communication. Molecular autism. 2010; 1:15. A total of four Shank3 mutant mouse lines described by 3 independent laboratories. Despite the variety of mutations and methodologies, a striking convergence of ASD-like phenotypes is observed across Shank3 mouse lines. A perturbation in the glutamatergic system is also consistently observed. Bozdagi et al, offered the first analysis of heterozygous mutant Shank3 gene; Peca et al. described in detail the multiple different protein isoforms arising from the Shank3 gene; Peca et al. describe the enrichment of Shank3 in the striatum and observe defects in cortico-striatal circuits; Bozdagi et al. suggest that part of the cellular defects in the Shank3 mutants are due to defects in the ubiquitination system. [PubMed: 21167025]
- 37. Bockers TM, Segger-Junius M, Iglauer P, Bockmann J, Gundelfinger ED, Kreutz MR, Richter D, Kindler S, Kreienkamp HJ. Differential expression and dendritic transcript localization of Shank family members: identification of a dendritic targeting element in the 3['] untranslated region of Shank1 mRNA. Mol Cell Neurosci. 2004; 26:182–190. [PubMed: 15121189]
- 38. Consortium ECTS. Identification and characterization of the tuberous sclerosis gene on chromosome 16. Cell. 1993; 75:1305–1315. [PubMed: 8269512]
- 39. van Slegtenhorst M, de Hoogt R, Hermans C, Nellist M, Janssen B, Verhoef S, Lindhout D, van den Ouweland A, Halley D, Young J, et al. Identification of the tuberous sclerosis gene TSC1 on chromosome 9q34. Science. 1997; 277:805–808. [PubMed: 9242607]
- 40. Crino PB, Nathanson KL, Henske EP. The tuberous sclerosis complex. The New England journal of medicine. 2006; 355:1345–1356. [PubMed: 17005952]
- Smalley SL. Autism and tuberous sclerosis. Journal of autism and developmental disorders. 1998; 28:407–414. [PubMed: 9813776]
- 42. Inoki K, Li Y, Zhu T, Wu J, Guan KL. TSC2 is phosphorylated and inhibited by Akt and suppresses mTOR signalling. Nature cell biology. 2002; 4:648–657.
- 43. Tang SJ, Reis G, Kang H, Gingras AC, Sonenberg N, Schuman EM. A rapamycin-sensitive signaling pathway contributes to long-term synaptic plasticity in the hippocampus. Proceedings of the National Academy of Sciences of the United States of America. 2002; 99:467–472. [PubMed: 11756682]
- 44•. Ehninger D, Han S, Shilyansky C, Zhou Y, Li W, Kwiatkowski DJ, Ramesh V, Silva AJ. Reversal of learning deficits in a Tsc2+/- mouse model of tuberous sclerosis. Nature Medicine. 2008; 14:843–848. The authors describe a reversal of deficits in the Tsc2 mutant mice suggesting that postnatal intervention may alleviate certain defects in complex genetic disorders.
- 45. Tavazoie SF, Alvarez VA, Ridenour DA, Kwiatkowski DJ, Sabatini BL. Regulation of neuronal morphology and function by the tumor suppressors Tsc1 and Tsc2. Nature neuroscience. 2005; 8:1727–1734.
- 46••. Sakai Y, Shaw CA, Dawson BC, Dugas DV, Al-Mohtaseb Z, Hill DE, Zoghbi HY. Protein interactome reveals converging molecular pathways among autism disorders. Science translational medicine. 2011; 3:86ra49. Using proteomic approaches, a range of converging molecular pathways are found to congregate on susceptibility to ASD. Here the authors provide clues on how, Tsc1 and Shank3 share multiple binding partners. Furthermore, the authors also highlight how interaction networks provide a framework to identify novel genetic targets for syndromic and non-syndromic autism.

- 47•. Darnell JC, Van Driesche SJ, Zhang C, Hung KY, Mele A, Fraser CE, Stone EF, Chen C, Fak JJ, Chi SW, et al. FMRP stalls ribosomal translocation on mRNAs linked to synaptic function and autism. Cell. 2011; 146:247–261. The mechanism of action of FMRP is explored and a robust system to assess FMRP target-mRNAs is presented by the authors. The loss of translational brake of synaptic proteins is suggested to contribute to FXS. [PubMed: 21784246]
- 48•. Auerbach BD, Osterweil EK, Bear MF. Mutations causing syndromic autism define an axis of synaptic pathophysiology. Nature. 2011; 480:63–68. Mutations in Tsc1/2 are hypothesized to drive abnormal synaptic function by leading to excessive protein production. Nevertheless, this study challenges the assumption that a simplistic view can be taken when considering mutation involving Tsc2 and the downstream effects on mGluR-dependent synaptic plasticity. The authors suggest that Tsc2 and FMRP mutations may precipitate opposing consequences on certain cellular mechanisms. [PubMed: 22113615]
- Verkerk AJ, Pieretti M, Sutcliffe JS, Fu YH, Kuhl DP, Pizzuti A, Reiner O, Richards S, Victoria MF, Zhang FP, et al. Identification of a gene (FMR-1) containing a CGG repeat coincident with a breakpoint cluster region exhibiting length variation in fragile X syndrome. Cell. 1991; 65:905– 914. [PubMed: 1710175]
- 50. De Boulle K, Verkerk AJ, Reyniers E, Vits L, Hendrickx J, Van Roy B, Van den Bos F, de Graaff E, Oostra BA, Willems PJ. A point mutation in the FMR-1 gene associated with fragile X mental retardation. Nature genetics. 1993; 3:31–35. [PubMed: 8490650]
- Hatton DD, Sideris J, Skinner M, Mankowski J, Bailey DB Jr, Roberts J, Mirrett P. Autistic behavior in children with fragile X syndrome: prevalence, stability, and the impact of FMRP. American journal of medical genetics Part A. 2006; 140A:1804–1813. [PubMed: 16700053]
- 52. Feng Y, Gutekunst CA, Eberhart DE, Yi H, Warren ST, Hersch SM. Fragile X mental retardation protein: nucleocytoplasmic shuttling and association with somatodendritic ribosomes. The Journal of neuroscience : the official journal of the Society for Neuroscience. 1997; 17:1539–1547. [PubMed: 9030614]
- 53. Brown V, Jin P, Ceman S, Darnell JC, O'Donnell WT, Tenenbaum SA, Jin X, Feng Y, Wilkinson KD, Keene JD, et al. Microarray identification of FMRP-associated brain mRNAs and altered mRNA translational profiles in fragile X syndrome. Cell. 2001; 107:477–487. [PubMed: 11719188]
- 54. Kindler S, Kreienkamp HJ. The role of the postsynaptic density in the pathology of the fragile X syndrome. Results and problems in cell differentiation. 2012; 54:61–80. [PubMed: 22009348]
- Huber KM, Kayser MS, Bear MF. Role for rapid dendritic protein synthesis in hippocampal mGluR-dependent long-term depression. Science. 2000; 288:1254–1257. [PubMed: 10818003]
- Huber KM, Gallagher SM, Warren ST, Bear MF. Altered synaptic plasticity in a mouse model of fragile X mental retardation. Proc Natl Acad Sci U S A. 2002; 99:7746–7750. [PubMed: 12032354]
- Dolen G, Osterweil E, Rao BS, Smith GB, Auerbach BD, Chattarji S, Bear MF. Correction of fragile X syndrome in mice. Neuron. 2007; 56:955–962. [PubMed: 18093519]
- 58. Krueger DD, Bear MF. Toward fulfilling the promise of molecular medicine in fragile X syndrome. Annual review of medicine. 2011; 62:411–429.
- Schutt J, Falley K, Richter D, Kreienkamp HJ, Kindler S. Fragile X mental retardation protein regulates the levels of scaffold proteins and glutamate receptors in postsynaptic densities. The Journal of biological chemistry. 2009; 284:25479–25487. [PubMed: 19640847]
- Kishino T, Lalande M, Wagstaff J. UBE3A/E6-AP mutations cause Angelman syndrome. Nature genetics. 1997; 15:70–73. [PubMed: 8988171]
- 61. Matsuura T, Sutcliffe JS, Fang P, Galjaard RJ, Jiang YH, Benton CS, Rommens JM, Beaudet AL. De novo truncating mutations in E6-AP ubiquitin-protein ligase gene (UBE3A) in Angelman syndrome. Nature genetics. 1997; 15:74–77. [PubMed: 8988172]
- 62••. Glessner JT, Wang K, Cai G, Korvatska O, Kim CE, Wood S, Zhang H, Estes A, Brune CW, Bradfield JP, et al. Autism genome-wide copy number variation reveals ubiquitin and neuronal genes. Nature. 2009; 459:569–573. In this study the authors find a link between copy number variations in ubiquitin genes and other neuronal genes. This is one of the first studies to show a robust link between ubiquitin ligases and ASDs apart from the already characterized UBE3A gene in Angelman syndrome. [PubMed: 19404257]

- 63. Dindot SV, Antalffy BA, Bhattacharjee MB, Beaudet AL. The Angelman syndrome ubiquitin ligase localizes to the synapse and nucleus, and maternal deficiency results in abnormal dendritic spine morphology. Human molecular genetics. 2008; 17:111–118. [PubMed: 17940072]
- 64. Yashiro K, Riday TT, Condon KH, Roberts AC, Bernardo DR, Prakash R, Weinberg RJ, Ehlers MD, Philpot BD. Ube3a is required for experience-dependent maturation of the neocortex. Nature neuroscience. 2009; 12:777–783.
- Mabb AM, Judson MC, Zylka MJ, Philpot BD. Angelman syndrome: insights into genomic imprinting and neurodevelopmental phenotypes. Trends in neurosciences. 2011; 34:293–303. [PubMed: 21592595]
- 66. Smith SE, Zhou YD, Zhang G, Jin Z, Stoppel DC, Anderson MP. Increased gene dosage of Ube3a results in autism traits and decreased glutamate synaptic transmission in mice. Science translational medicine. 2011; 3:103ra197.
- 67•. Ehlers MD. Activity level controls postsynaptic composition and signaling via the ubiquitinproteasome system. Nat Neurosci. 2003; 6:231–242. This study examines the consequences on the composition of the postsynaptic density as a consequence of changing the activity levels in neuronal cultures. The author identifies ensembles of proteins that vary bidirectionally with activity and further examines the role of ubiquitination in several PSD target proteins with increased or decreased synaptic activity. [PubMed: 12577062]
- 68•. Giacometti E, Luikenhuis S, Beard C, Jaenisch R. Partial rescue of MeCP2 deficiency by postnatal activation of MeCP2. Proc Natl Acad Sci U S A. 2007; 104:1931–1936. In these studies the authors describe how a postnatal rescue of MECP2 expression can rescue behavioral deficits in mutant mice. These results underscore the possibility to intervene and ameliorate symptoms in the case of early intervention with appropriate therapies. [PubMed: 17267601]
- 69•. Guy J, Gan J, Selfridge J, Cobb S, Bird A. Reversal of neurological defects in a mouse model of Rett syndrome. Science. 2007; 315:1143–1147. In these studies the authors describe how a postnatal rescue of MECP2 expression can rescue behavioral deficits in mutant mice. These results underscore the possibility to intervene and ameliorate symptoms in the case of early intervention with appropriate therapies. [PubMed: 17289941]
- 70•. Welch JM, Lu J, Rodriguiz RM, Trotta NC, Peca J, Ding JD, Feliciano C, Chen M, Adams JP, Luo J, et al. Cortico-striatal synaptic defects and OCD-like behaviours in Sapap3-mutant mice. Nature. 2007; 448:894–900. In this study the authors identify behavioral abnormalities reminiscent of OCD-like behaviors in SAPAP3 mutant mice. SAPAP3 is enriched in the striatum and the perturbation of this gene is found to produce cortico-striatal synaptic dysfunction. A postnatal, viral-mediated reintroduction of SAPAP3 in the striatum rescued both synaptic deficits and behavioral abnormalities. [PubMed: 17713528]
- 71•. Zeier Z, Kumar A, Bodhinathan K, Feller JA, Foster TC, Bloom DC. Fragile X mental retardation protein replacement restores hippocampal synaptic function in a mouse model of fragile X syndrome. Gene Ther. 2009; 16:1122–1129. In this study the authors use viral mediated gene therapy to reintroduce FMRP expression in the hippocampus of FMRP deficient mice. The reintroduction of FMRP rescued synaptic plasticity in the hippocampus of mutant mice again suggesting that postnatal interventions may alleviate part of the neuronal deficits in mouse models of genetic disorders. [PubMed: 19571888]
- 72. Zuchner S, Wendland JR, Ashley-Koch AE, Collins AL, Tran-Viet KN, Quinn K, Timpano KC, Cuccaro ML, Pericak-Vance MA, Steffens DC, et al. Multiple rare SAPAP3 missense variants in trichotillomania and OCD. Mol Psychiatry. 2009; 14:6–9. [PubMed: 19096451]

Research Highlights

- Mutations in several genes encoding the core transsynaptic complex of Neurexin/Neuroligin/PSD-95/SAPAP/Shank confer susceptibility to ASDs
- Tuberous Sclerosis Complex, Fragile X syndrome, Angelman syndrome and synaptic ASD candidate genes have overlapping interaction networks
- Mouse models for ASDs have helped identify commonalities in glutamatergic dysfunction
- Identifying important nodes in interaction networks may facilitate therapeutical intervention
- ASD endophenotypes may be triggered by common neural circuit defects

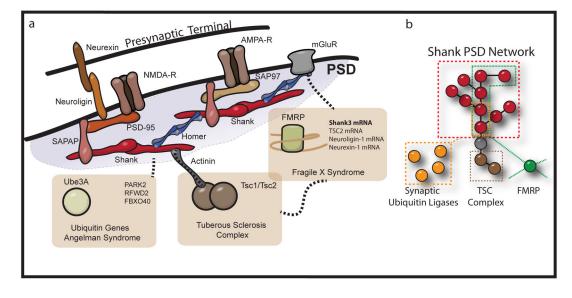


Figure 1.

Shank proteins at the center of an ASD disease-module. A model for the overlap between synaptic proteins involved in susceptibility to syndromic and non-syndromic autism. (a) Neurexin and Neuroligins are transsynaptic partners and candidate genes for susceptibility to autism [10–14], in the postsynaptic density these bind to the SAPAP family of proteins which have been linked to ASDs and OCD [17,72], PSD-95 and SAP97 which are involved in intellectual disability and autism [15,16] and Shank2 and Shank3 which are also candidate genes for autism [18,19]. Shank dimers are thought to organize a molecular platform in concert with Homer tetramers to stabilize the larger PSD, connecting AMPAR, NMDAR and mGluR into one protein hub. In the deeper synaptic compartment the control of PSD protein levels may be tightly controlled by independent complexes such as TSC1/2 through mTOR, or via FMRP regulation of synaptic transcripts, and most likely also through synaptic ubiquitin ligases. (b) Simplified putative disease-module network centering on Shank proteins at the PSD (red) displaying the overlap between synaptic ubiquitin ligases and ubiquitination of SAPAP/Shank (see text – orange), the Tuberous sclerosis genes TSC1/ TSC2 (brown) and FMRP regulation of several PSD protein transcripts (green).