Measurement of the nonlinear elasticity of red blood cell membranes

The MIT Faculty has made this article openly available. Please share how this access benefits you. Your story matters.

Citation

As Published
http://dx.doi.org/10.1103/PhysRevE.83.051925

Publisher
American Physical Society

Version
Final published version

Citable link
http://hdl.handle.net/1721.1/65332

Terms of Use
Article is made available in accordance with the publisher’s policy and may be subject to US copyright law. Please refer to the publisher’s site for terms of use.
Measurement of the nonlinear elasticity of red blood cell membranes

YongKeun Park,1,2,4 Catherine A. Best,3 Tatiana Kuriabova,4 Mark L. Henle,5 Michael S. Feld,1,† Alex J. Levine,6 and Gabriel Popescu7,‡

1G. R. Harrison Spectroscopy Laboratory, Massachusetts Institute of Technology, Cambridge, Massachusetts 02139, USA
2Harvard-MIT Division of Health Science and Technology, Massachusetts Institute of Technology, Cambridge, Massachusetts 02139, USA
3College of Medicine, University of Illinois at Urbana-Champaign, Urbana, Illinois 61801, USA
4Department of Physics, University of Colorado, Boulder, Colorado 80309, USA
5School of Engineering and Applied Sciences, Harvard University, Cambridge, Massachusetts 02138, USA
6Department of Chemistry & Biochemistry and Department of Physics & Astronomy, UCLA, Los Angeles, California 90095, USA
7Quantitative Light Imaging Laboratory, Department of Electrical and Computer Engineering, Beckman Institute for Advanced Science & Technology, University of Illinois at Urbana-Champaign, Urbana, Illinois 61801, USA

(Received 28 February 2010; published 27 May 2011)

The membranes of human red blood cells (RBCs) are a composite of a fluid lipid bilayer and a triangular network of semiflexible filaments (spectrin). We perform cellular microrheology using the dynamic membrane fluctuations of the RBCs to extract the elastic moduli of this composite membrane. By applying known osmotic stresses, we measure the changes in the elastic constants under imposed strain and thereby determine the nonlinear elastic properties of the membrane. We find that the elastic nonlinearities of the shear modulus in tensed RBC membranes can be well understood in terms of a simple wormlike chain model. Our results show that the elasticity of the spectrin network can mostly account for the area compression modulus at physiological osmolality, suggesting that the lipid bilayer has significant excess area. As the cell swells, the elastic contribution from the now tensed lipid membrane becomes dominant.

DOI: 10.1103/PhysRevE.83.051925 PACS number(s): 87.16.dj, 83.60.Df, 87.16.dm

I. INTRODUCTION

Blood serum is typically maintained at fixed osmolality to control the flux of water to the circulating RBCs. When placed in hypotonic solutions (i.e., solutions with a lower concentration of solutes), the hemoglobin (Hb)-rich interior of the RBCs draws water into the cells, causing swelling and, in extreme cases, bursting. Alternatively, RBCs shrink when placed in hypertonic solutions, as the osmotic pressure difference forces water out of the cytosol of the RBCs.

In addition to changing the cells’ morphology, these volume changes driven by osmotic pressure variations modify the mechanics of the cells. Measurements of the mechanical properties at various fixed osmotic pressures thus allow one access to the nonlinear elasticity of the cells. Since RBCs do not have an internal cytoskeleton or other complex subcellular organelles, their mechanics is determined entirely by their membrane, which is a lipid bilayer coupled to a triangular lattice of semiflexible polymer filaments mainly composed of spectrin. Any observed change in the mechanics of the cells under osmotic stress must reflect the elastic nonlinearity of this composite membrane as it is tensed or relaxed by the influx or efflux, respectively, of water. Thus, the ability to measure the linear response properties of the cell membrane at varying states of osmotic stress allows one to experimentally probe the nonlinear elastic response of this tethered membrane.

In this paper we report on a series of microrheological measurements of RBC membrane mechanics obtained at varying levels of osmotic stress. We simultaneously measure the cellular volume so that we may quantitatively measure the change in the elastic properties of the composite membrane as a function of applied strain. Our results provide insight into the general problem of the nonlinear mechanics of tethered membranes, but also have biological implications for understanding the effects of blood serum osmolality on RBC mechanics. Since these cells undergo large deformations and significant changes in osmotic stress in the microvasculature, understanding the nonlinear mechanics of RBCs under osmotic pressure may have direct physiological implications.

II. MODEL AND METHODS

The RBC plasma membrane is a 4–5-nm-thick fluid lipid bilayer mechanically characterized by a bending modulus \( \kappa \) and an area compression modulus \( K_A \). On the cytoplasmic or interior side of the membrane, there is the triangular spectrin network with mesh size of 90 nm that is anchored to the bilayer via transmembrane proteins at the nodes of this crosslinked network [1]. On length scales that are long compared to the lattice constant, the mechanics of this network can be described as a two-dimensional elastic continuum having two equal Lamé constants, \( \mu = \lambda \). Given the viscosity of the fluid lipid bilayer, viscous stresses in the membrane are subdominant up to frequencies of \( \sim 10^3 \) Hz [2]. The cytosol of the RBC consists mainly of a spatially homogenous Hb solution of viscosity \( \eta_c \); that, due to the high Hb concentration, is significantly more viscous than that of the surrounding solvent, \( \eta_s < \eta_c \). Thus, the mechanical description of the RBC contains four parameters: \( \kappa, K_A, \mu, \eta_c \). Of these, \( \kappa \) is thought to depend solely on the lipid composition of the bilayer, \( K_A \) depends on a combination of the bilayer’s surface tension and the area modulus of the underlying spectrin network, and \( \mu \) depends solely on the...
shear modulus of that network. Finally, while the viscosity of the surrounding solvent is known, the viscosity of the highly concentrated cytosol solution $\eta_c$ will vary strongly with the Hb concentration and thus with osmotic pressure.

A number of techniques has been used to obtain quantitative information about the mechanics of live RBCs, including pipette aspiration [3], electric field deformation [4], magnetic beads cytometry [5], and optical tweezers [6,7]. However, none of these methods can probe all of the mechanical parameters of the RBC membrane simultaneously. Moreover, these approaches require contact by an external probe and operate with a large force (or deformation) that may invalidate the assumption of a linear response. Fluctuations in RBC membranes are intrinsic reporters of the linear mechanical response; they offer a nonperturbing window into the structure, dynamics, and function of the intact cell [8–13]. In order to measure the nanoscale membrane fluctuations with high accuracy and low noise, we employed diffraction phase microscopy (DPM) [14], which has been used successfully for imaging cell structures and dynamics [15–19]. The analysis of the observed fluctuation spectrum of a soft elastic object to determine its (visco-)elastic properties is commonly referred to as microrheology [20,21]. The analysis of the fluctuation data requires (i) a model of the frequency-dependent linear response to applied forces and (ii) an assumption of thermal equilibrium. By using the latter assumption and the fluctuation-dissipation theorem [22], one may fit a model of the response function containing the appropriate elastic moduli to the observed fluctuations. For a case in question, we use cellular microrheology to probe the effective linear elastic response of the membrane at varying states of stress, controlled by an external osmotic pressure.

To modulate the osmolality of the medium, we prepared RBC suspensions (10⁶ cell/μl) with 11 different osmolalities ranging from 100 to 600 mOsm/kg H₂O. We followed the standard protocols for RBC preparation [11]. Fresh blood samples were collected and diluted 1:5 in Hank’s buffer saline solution (HBSS) and then immediately centrifuged at 2000 g at 5 °C for 10 minutes to separate RBCs from plasma. The RBCs were washed three times and then resuspended in the given NaCl solutions. The NaCl solutions contained increasing concentrations of NaCl (0.3%–1.8%), which correspond to suspension osmolalities ranging from 100 to 600 mOsm/kg. Using DPM, we extracted quantitative optical phase shifts $\phi(x, y, t)$ associated with the cells at spatial and temporal resolutions of nm and ms, respectively [14,16]. The cell thickness profile was obtained from the optical phase shift as $h(x, y, t) = \lambda / 2\pi (\Delta n)_{\phi}(x, y, t)$. Since the refractive index difference $\Delta n$ is mainly contributed from the homogeneous Hb solution in cytoplasm, the integration of optical phase shifts over the cell area, i.e., the dry mass [24], is related to the volume of RBCs as $\text{Volume} = \int (h(x, y)) \, dA$. The $\Delta n$ was calculated using the RBC volume data in the literature [25].

III. RESULTS AND DISCUSSIONS

A. Topography and membrane fluctuations of RBCs at different osmotic pressure

Thickness profiles and horizontal cross sections of RBCs in hypotonic, isotonic, and hypertonic media are shown in Fig. 1. It is clear that different osmolalities of the extracellular medium result in significant changes in RBC shape. In a hypotonic medium [100 mOsm/kg; Fig. 1(a)], RBCs are swollen due to water influx, but still maintain the dimpled region in the center. At osmotic pressure of less than 100 mOsm/kg, most of the RBCs are lysed. In the hypertonic case [600 mOsm/kg; Fig. 1(c)], RBCs shrink due to water efflux. In a hypertonic medium, the projected area of the membrane is lower than that of normal RBCs, which is indicative of an increase in the membrane tension caused by cell swelling;
see Fig. 2(a). In contrast, the projected area in a hypertonic medium does not change significantly. We extracted the mean corpuscular hemoglobin (MCH), i.e., the total amount of Hb in the cell, as the product of the Hb concentration and cytoplasmic volume. The Hb concentration was retrieved using its known relationship with the refractive index [26]. MCH maintains constant values at different osmolalities [Fig. 2(c)], which is consistent with the impermeability of the RBC membrane to large Hb proteins.

The membrane fluctuations were obtained by subtracting the time-averaged cell profile from each instantaneous topography map in the series, \( \Delta h(x,y,t) = h(x,y,t) - \langle h(x,y) \rangle \). The root mean squared (RMS) displacement of membrane fluctuations, \( \sqrt{\langle \Delta h^2 \rangle} \), shows that the maximum membrane fluctuations occur around 300 mOsm/kg, which is within the normal physiological blood osmolality [Fig. 2(d)]. The decreased deformability of RBCs in both hypo- and hypertonic conditions is consistent with a variety of experimental techniques, including laser scattering, cell elongation measurements, and blood filtration experiments [27,28].

### B. Mechanical properties of RBCs at different osmotic pressure

In order to investigate the mechanical properties of RBCs, we analyze the spatial correlations of the out-of-plane membrane fluctuations using a continuum model of the composite spectrin-network/lipid membrane [29]. Our recent theoretical description incorporates the coupling between the bending and compression modes of the curved membrane [29] and allows us to determine quantitatively the cell’s mechanical parameters: \( \kappa, K_A, \mu, \eta_c \). The details of the model have been discussed elsewhere [23,29,30]. The two-point correlation function of the membrane fluctuations was calculated as \( C(d,t) = \langle \Delta h(d,t) \Delta h(0,0) \rangle \), where the angular brackets denote both spatial and temporal averaging. As shown in Fig. 3, the theoretical model provides a very good description of the experimental data, which allows us to extract the cell material properties. In addition, we find that there is generically a single best fit, since the bending modulus controls the high wave-number features of the correlation function, while the other elastic constants dominate the lower wave-number features. To obtain these fits to \( \tilde{C}(d,\omega) \), we adjust the following parameters: the shear \( \mu \) and area compression \( K_A \) moduli of the spectrin network and lipid bilayer, the bending modulus \( \kappa \) of the lipid bilayer, the viscosity of the cytosol \( \eta_c \), and the radius of the sphere \( R \). We constrain our fits by setting \( R \) to the average radius of curvature of the RBCs obtained directly from the data and fixing the viscosity for extracellular medium to be \( \eta_s = 1.2 \) mPa s [31]. Furthermore, we assume that the elastic properties of the composite membrane are dominated by reactive (i.e., nondissipative) terms, so that we may treat the elastic constants as real and frequency independent. This assumption is supported \textit{a posteriori} by our making fits to correlation functions at two frequencies separated by a factor of \( \sim 8 \) with the same elastic parameters. This leaves a four-dimensional space of fitting parameters spanned by \( \kappa \),
$K_A$, $\mu$, and $\eta$. For a triangular elastic network, we expect $\mu = \lambda$ [32], so positive values of dimensionless parameter $\delta = K_A/\mu = 2$ measure the importance of the contribution of the lipid bilayer’s surface tension to the effective area compression modulus of the composite membrane.

From these fits, we determined the three mechanical properties of the RBC membrane at different osmolalities (Fig. 4). We also extracted the viscosity of the cytosol by fitting the fluctuation data at two frequencies; the viscosity generates the only frequency-dependent stress. The shear modulus of spectrin network $\mu$ in hypotonic media shows a significant increase compared to the normal and hypertonic cases. Interestingly, $\mu$ does not change above 300 mOsm/kg.

As expected, the spectrin network becomes significantly less compliant ($\mu \approx 13 \text{ mN}^{-1}$) under tension. We model the spectrin network as a perfect triangular lattice of semiflexible wormlike chain (WLC) filaments, using a standard value for the lattice constant, lower osmolality leads to a uniform extension of the spectrin network as a perfect triangular lattice of semiflexible wormlike chain (WLC) filaments, using a standard value for the lattice constant of the spectrin network at physiological osmolality $300 \text{ mOsm/kg}$.

In the former case, we suggest the higher value may be attributed to elastic nonlinearities. In the latter case, the higher $\mu$ is the measured $V/A$, cytosol viscosity $\eta$, and bending modulus $\kappa$ vs different osmotic pressure. Error bar represents standard deviation for 20 RBCs. (b) The observed increase in $\mu$ of the membrane (symbols) compared to WLC predictions for the nonlinear increase in $\mu$ associated with the tending of the membrane (line). Contour lengths used for the WLC model are 180, 173, 167, and 167 nm, for $\mu = 6.0, 7.0, 8.0, 9.0$, and 9.5 $\text{ mN/m}$, respectively.

FIG. 4. (Color online) (a) Shear modulus $\mu$, cytosol viscosity $\eta$, and bending modulus $\kappa$ vs different osmotic pressure. Error bar represents standard deviation for 20 RBCs. (b) The observed increase in $\mu$ of the membrane (symbols) compared to WLC predictions for the nonlinear increase in $\mu$ associated with the tending of the membrane (line). Contour lengths used for the WLC model are 180, 173, 167, and 167 nm, for $\mu = 6.0, 7.0, 8.0, 9.0$, and 9.5 $\text{ mN/m}$, respectively.
curvature of the RBC membrane at the edge may effectively suppress fluctuations rendered a higher measured value.

Finally, the viscosity of the cytosol $\eta_c$ increases monotonically with increasing osmolality since water leaves the cell (through aquaporin-1 channels), increasing the cytoplasmic Hb concentration and resulting in an increase of cytosolic viscosity [38]. We speculate that large changes in the cytosolic viscosity may have a physiologically protective effect. By increasing viscous dissipation when RBCs pass through the small capillaries of the kidney, where the osmolality can be as high as 1200 mOsm/kg, the viscous stresses in cytosol may reduce in-plane membrane stresses and prevent cell lysis as they undergo large-scale deformation.

**IV. SUMMARY AND CONCLUSIONS**

In summary, we have used the dynamic fluctuations of RBCs under varying osmotic stress to measure the nonlinear response of the composite membrane’s shear modulus, bending modulus, and area compression modulus. By examining data at different frequencies, we also obtain the cytosolic viscosity. We find that the composite membrane is strongly strain hardening under tension in both shear and area moduli. The shear modulus enhancement is easily understood in terms of the WLC model. Further work is needed to understand the nonlinear stiffening of $K_A$ under tension. Previous measurements of RBC mechanics report two sets of elastic moduli separated by about four orders of magnitude. Fluctuation data are consistent with our smaller measurements, while micropipette aspiration techniques consistently report the larger values. We propose that consideration of the nonlinear mechanics of the membrane may resolve much of this discrepancy. Detailed measurements of precisely those elastic nonlinearities, such as reported here, are the key route to testing this proposal.

As expected, the bending modulus is highly insensitive to imposed osmotic stress and the cytosolic viscosity increases with Hb concentration. These elastic constants are all relatively insensitive to membrane relaxation in hypertonic solutions. Understanding this point may require more sophisticated models that include steric relaxation in hyper- tonic solutions. Finally, we note that the RBC moduli reach their minimal values at physiological osmolality, perhaps reflecting some evolutionary tuning of these elastic nonlinearities.

**ACKNOWLEDGMENTS**

This research was supported by the National Institutes of Health (Grant No. P41-RR02594-18-24). G.P. was partially supported by the National Science Foundation (Grant No. 08-46660 CAREER) and National Cancer Institute (Grant No. R21 CA147967-01). A.J.L. acknowledges partial support from National Science Foundation Grant No. NSF-DMR-0907212. The authors acknowledge Hawoong Jung and Kyomin Jung of the Korean Advanced Institute of Science and Technology for statistical analysis, and Ramachandra R. Dasari of the Massachusetts Institute of Technology for fruitful discussions.

---