Noninvasive imaging of immune responses

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At their margins, tumors often contain neutrophils, dendritic cells, and activated macrophages, which express class II MHC and CD11b products. The interplay between stromal cells, tumor cells, and migratory cells such as lymphocytes creates opportunities for noninvasive imaging of immune responses. We developed alpaca-derived antibody fragments specific for mouse class II MHC and CD11b products, expressed on the surface of a variety of myeloid cells. We validated these reagents by flow cytometry and two-photon microscopy to obtain images at cellular resolution. To enable noninvasive imaging of the targeted cell populations, we developed a method to site-specifically label VHVs [the variable domain (Vv) of a camelid heavy-chain only antibody] with 18F or 64Cu. Radiolabeled VHVs rapidly cleared the circulation (t1/2 ∼ 20 min) and clearly visualized lymphoid organs. We used VHVs to explore the possibility of imaging inflammation in both xenogeneic and syngeneic tumor models, which resulted in detection of tumors with remarkable specificity. We also imaged the infiltration of myeloid cells upon injection of complete Freund’s adjuvant. Both anti-class II MHC and anti-CD11b VHVs detected inflammation with excellent specificity. Given the ease of manufacture and labeling of VHVs, we believe that this method could transform the manner in which anti-tumor responses and/or infectious events may be tracked.

PET imaging | non-invasive imaging | inflammation | cancer | camelid single domain antibodies

Myeloid cells, such as neutrophils, dendritic cells, and activated macrophages, express CD11b and/or class II MHC products and are often present at the tumor margin (1, 2). Macrophages, depending on their functional properties (M1 or M2 type) (3), help establish a (micro) environment either detrimental or favorable to tumor growth (3, 4). The ability to image myeloid cells’ presence is thus of diagnostic relevance and may be an approach more generally applicable than detection of tumor cells via unique tumor-specific markers. Current practice either relies on sampling of peripheral blood to enumerate and characterize these cells in the circulation or requires more invasive procedures, such as histological analysis of biopsies or surgical resection specimens. Noninvasive detection of sites of an ongoing inflammatory or immune response might provide clues to the existence and location of tumor foci, which a biopsy could then confirm. PET tracers, such as 18F-2-deoxyfluoroglucose (FDG), visualize differences in metabolic activity that accompany inflammation but lack the discriminatory power that antibodies or ligands, have also been used for PET imaging (6, 7) but do not provide information about immune responses (6, 7). Camelid-derived single-domain VHVs [the variable domain (VH) of a camelid heavy-chain only antibody] (8) present several advantages and characteristics of antibody derivatives (∼60 kDa) (9), Fab fragments (∼50 kDa), or ScFvs (∼25 kDa), and they lack an Fc region. Single-domain VHVs of desired specificity can be readily generated, isolated, and characterized by their amino acid sequence using standard procedures (10). Approaches to modify (11) and humanize (12) these camelid VHVs exist, and early-phase clinical testing is underway (13).

Results

Anti-Mouse Mouse Class II MHC and Anti-Mouse CD11b Stain Secondary Lymphoid Organs. Two VHVs, VH7H7 (anti-mouse class II MHC) and VH1DC13 (anti-mouse CD11b) were generated following standard procedures (SI Materials and Methods) (10). To test whether anti-class II MHC and anti-CD11b–specific VHVs could be used for in vivo labeling of class II MHC- and CD11b-expressing cells, we installed a fluorophore on VH7H7 and antibody is poor. Finally, a labeled, full-sized antibody persists in the circulation because of its comparatively long circulatory half-life. Other scaffolds, such as proteins based on fibronectin type III domains (FN3) and small molecules that serve as receptor-specific ligands, have also been used for PET imaging (6, 7) but do not provide information about immune responses (6, 7). Camelid-derived single-domain VHVs [the variable domain (VH) of a camelid heavy-chain only antibody] (8) present several advantages and characteristics of antibody derivatives (∼60 kDa) (9), Fab fragments (∼50 kDa), or ScFvs (∼25 kDa), and they lack an Fc region. Single-domain VHVs of desired specificity can be readily generated, isolated, and characterized by their amino acid sequence using standard procedures (10). Approaches to modify (11) and humanize (12) these camelid VHVs exist, and early-phase clinical testing is underway (13).

Significance

Tumors are often surrounded and invaded by bone marrow-derived cells. Imaging the infiltration of such immune cells into tumors may therefore be an attractive means of detecting tumors or of tracking the response to anticancer therapy. We show that it is possible to detect these cells noninvasively by positron emission tomography (PET) via the surface markers displayed by them. The ability to monitor the immune response in the course of therapy will enable early determination of the efficacy of treatment and will inform decisions as to whether treatment should be stopped or continued. Noninvasive monitoring could therefore change how therapies are applied and assessed, to the benefit of many patients.

Author contributions: M.R., E.J.K., A.M.B., and H.L.P. designed research; M.R. synthesized the substrates, expressed sortase and VHHs, and modified VHHs with fluorophores and for 64Cu/18F labeling; E.J.K. performed synthesis of 18F/64Cu-labeled VH7H7 and their characterization; M.R., E.J.K., G.R.W., and R.W. performed PET imaging and analysis of the PET data; M.R., M.A.B., J.T.J., and G.D.V. performed and interpreted two-photon microscopy; M.R., A.M.B., and J.N.D. performed FACS experiments; J.N.D., J.C., and L.K.S. identified VH7H7 and DC13; and M.R. and H.L.P. wrote the paper.

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VHHDC13 in a sortase-catalyzed reaction and used this preparation for two-photon microscopy and flow cytometry (14). We have previously established that VHH7 recognizes murine I-A products encoded by the H-2b haplotype and that VHH7 fails to stain class II MHC+ cells (11). VHH7 does not recognize human class II MHC products, as ascertained by immunofluorescence on class II HLA* Mel-Juso cells maintained in culture. We injected 20 μg of Texas Red-labeled VHH7 i.v. into class II MHC-eGFP (15) or class II MHC-deficient mice (16). After 90 min, we explanted lymph nodes and spleen to subject them to two-photon microscopy. The class II MHC-eGFP marker enables visualization of all class II MHC cells without the aid of an antibody, and therefore the efficacy of in vivo immunostaining with VHH7 can be accurately assessed. Two-photon imaging of class II MHC-eGFP mice showed perfect concordance with Texas Red-VHH7-stained cells (Fig. 1 A and B). Class II MHC+/− mice showed no specific staining (Fig. 1C). In the absence of injected Texas Red-VHH7, no staining was apparent in the corresponding channel (Fig. L4 and Fig. S1). These results establish specificity of staining and show that, at 90 min postinjection, VHH7 achieved excellent penetration of spleen and lymph nodes. We similarly labeled VHHDC13, which recognizes CD11b, a marker for neutrophils, macrophages, and dendritic cells. The molecular target of VHHDC13 was identified by mass spectrometry of immunoprecipitates prepared with immobilized VHHDC13. VHHDC13 specifically binds to CD11b+ cells in lymph nodes and spleen (Fig. 1 E1 and E2 and Fig. S1). To further confirm the two-photon experiments, WT, MHC-II-deficient, or CD11b-deficient mice were injected with 2 or 20 μg of VHH7 or VHHDC13-Alexa 647. After 90 min, spleen and brachial lymph nodes were analyzed by flow cytometry after staining with fluorophore-conjugated antibodies to CD3 and CD19, for VHH7, and to CD11b, for DC13. Results clearly confirmed the specificity of the VHHs for their corresponding targets (Fig. 1 G and H). In vivo staining with labeled VHHs obviates the need for fluorescent-protein fusions as the target for multiphoton imaging. A 90-min delay between administration and imaging is sufficient to obtain adequate staining with VHHs in spleen and lymph nodes. Although two-photon microscopy affords cellular resolution and can be used intravitally, it is nonetheless a mostly invasive method. Therefore explored the possibility of noninvasive imaging by positron emission tomography (PET).

Site-Specific 18F or 64Cu Labeling of Single-Domain Antibodies (VHHs) Using Sortase. Antibody-labeling strategies for PET often revolve around modification with metal chelators to enable installation of radioisotopes such as 18F, 68Ga, or 89Zr (17–19). Although the
creation of fluorine–carbon bonds remains a synthetic challenge (20, 21), its use has advantages over 68Ga, 64Cu, 89Zr, and 125I: lower energy positron emission, shorter half-life, reduced cost, and wider availability. 18F-PET imaging is used both for diagnostic purposes and to monitor therapeutic efficacy, but these applications rely on agents that report on metabolic activity, such as FDG or labeled precursors to nucleic acids (22–25). The feasibility of noninvasive imaging of lymphocytes using an antibody fragment was demonstrated only recently, using a ~60-kDa 64Cu-labeled anti-CD8 antibody fragment (26). The half-life (t1/2 ~ 110 min) of 18F requires it to be used shortly after installation. We therefore developed a simple two-step process (Fig. 2) for 18F labeling of VHHs bearing a sortase-recognition motif, using a combination of a click reaction and a sortase-catalyzed modification. This method is robust, reproducible, and site-specific, without compromising the VHH antigen-binding site. This technique is further applicable to any other suitably modified biological entity (11).

The 18F radionuclide was conjugated to the VHHs via a tetrazine (Tz)/transcyclooctene (TCO) reverse-electron demand Diele–Alder cycloaddition. The TCO-tetrazine reaction is the fastest known bioorthogonal reaction to date, with an estimated second-order rate constant of 210 M−1·s−1 (27–29). We synthesized the requisite sortase nucleophile, GGG-tetrazine, which participates in sortase reactions to modify the VHH (Fig. 2A). 18F radiolabeling of tosyl-transcyclooctene (TCO) was achieved with [18F]F/222/K2CO3 in DMSO for ~10 min at 90 °C to produce 18F-TCO (30) (~75% yield, decay-corrected) and purified by HPLC (Fig. 2B). 18F-TCO was then added to the Tz-modified VHH, and the reaction was allowed to proceed for ~20 min at pH 7. Thus, using 18F-TCO, we readily produced >5 mCi of 18F-labeled VHH7 in two steps (Fig. 2C). The 18F-labeled VHH was purified by size-exclusion chromatography in PBS, providing a radiolabeled VHH solution ready for injection. Labeling experiments with 18F-TCO yielded 18F-VHH7 in 61 ± 9% decay-corrected radiochemical yield.

18F-VHH7 Detects Secondary Lymphoid Organs. Administration of 200 μCi/10 μg 18F-VHH7 in vivo in C57BL/6 WT mice (class II MHC-sufficient) allowed us to visualize lymph nodes, spleen, and thymus with a high degree of specificity and with excellent signal-to-noise ratios (Fig. 2G–J). No nonspecific staining was seen in lymphoid tissues when using 18F-VHH7 in class II MHC−/− mice (B6.129S2-H2−/−, B6.129S2-H2−/− (ΔAb1-Eae-1)) (Fig. 2F and K–M and Movies S14, S15, S24, and S28). Biodistribution and circulatory half-life measurements of 18F-VHHs in WT mice showed a t1/2 of ~20 min (Fig. S2).

18F-VHH7 and 18F-VHHDC13 Detect Inflammation. We next used 18F-VHH7 and DC13 to image the behavior of class II MHC+ host cells in a xenograft tumor model. Neither the distribution of class II MHC+ or CD11b+ cells at steady state nor changes in their distribution and location over time have been specifically imaged noninvasively using 18F-labeled tracers. In many tumor models, the margins of the tumor contain macrophages, which, when activated, express class II MHC and CD11b (1, 3). Thus, we asked whether—in addition to the usual lymphoid structures as visualized by 18F-VHH7 and DC13—we could detect the presence of macrophages around and within a xenografted tumor. We created xenografts in NOD-SCID mice by s.c. inoculation of the Mel-Juso human melanoma cell line, representative of a subset of human melanomas. The NOD-SCID hosts contain no B or T cells, but murine class II MHC+ antigen-presenting cells (macrophages and dendritic cells) and CD11b+ cells (macrophages, dendritic cells, and neutrophils) are present and should thus be visible with 18F-labeled VHHs.

One month after injection of 5 × 106 tumor cells, PET imaging using 18F-VHH7 (Fig. 3A–D) or DC13 (Fig. 3F and G) clearly showed the presence of class II+ cells or CD11b+ cells associated with the tumor (Movies S3 and S4). Some tumors, such as a subset of melanomas, express class II MHC products, in which case one might expect an even stronger PET signal (31). To confirm the presence of class II MHC+ cells or CD11b+ cells, we excised the tumors, digested them with collagenase D, and enriched tumor-infiltrating cells on Percoll gradients before flow cytometry. Mouse CD45+ tumor-infiltrating cells stained

![Image](https://www.pnas.org/cgi/doi/10.1073/pnas.1502609112)

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positive for VHH7 and DC13 (Fig. 3 H and I). Postmortem histology of the tumor further verified the presence of inflammatory cells in/around the tumor, with clearly visible infiltrating macrophages and neutrophils (Fig. 3L). We have thus demonstrated noninvasive imaging of class II MHC+ cells and CD11b+ cells, both in normal and in tumor-bearing mice.

**18F-VHH Detects Class II MHC+ Cells Associated with Small Tumors at Early Stages.** Is it possible to image small tumors also at earlier stages of growth? To address this question, we imaged NOD/SCID mice xenografted with 5 × 10⁶ Mel-Juso human melanoma cells at 6 d, 20 d, and 27 d postinjection. We detected inflammation at the site of the malignant growth at the earliest time point after injection, a time when the incipient tumors are neither visible nor detectable by palpation (Fig. 3E). FACS analysis on cell suspensions prepared from the excised tumors again confirmed the presence of tumor-infiltrating class II MHC+ cells (Fig. 3H). It may thus be worthwhile to explore early-stage detection of diseases characterized by an inflammatory signature, such as multiple sclerosis, diabetes, or infections (5).

The longer half-life of ⁶⁴Cu (12.7 h) relative to ¹⁸F (110 min) in principle allows a more extended imaging period to establish tissue penetration and dwell time of VHHs on their targets. We developed a NOTA-(Gly)₃ sortase nucleophile to enable site-specific labeling of VHHs (Fig. 2D and Fig. S3) via this high-affinity copper-chelating agent (Kₘ for Cu²⁺ of ~10⁻⁸ M) (32). The [Cu²⁺-NOTA] complex is kinetically inert, with minimal metal exchange when exposed to other metals present in body (33). We produced site-specifically labeled [(⁶⁴Cu)-NOTA]-VHHs in high radiochemical yield (∼90% decay corrected) and used ⁶⁴Cu-VHH7 to image a C57BL/6 WT mouse at 4 h, 8 h, and 24 h postinjection. VHH7 stayed on its target (secondary lymphoid organs) even after 24 h (Fig. S4) but produced images with an inferior signal-to-noise ratio compared with ¹⁸F-VHH7 (compare Fig. 2 with Fig. S4).

¹⁸F-VHH7 and ¹⁸F-VHHDC13 Detect Infiltration of Immune Cells in B16 Melanoma Tumor-Bearing Mice in a Syngeneic Setting. Next, we explored B16 melanoma-bearing mice and found that here, too, we could readily image the presence of the primary tumor via the presence of CD11b+ and class II MHC+ cells, without a need for melanoma-specific markers (Fig. 4 A and B and Movies S5A, S5B, S5C, S6A, and S6B). We also performed imaging experiments on B16 melanoma-bearing mice using ²⁵⁲F-2-fluorodeoxyglucose (¹⁸F-FDG), a standard means of staging tumor patients by PET. ¹⁸F-FDG revealed the presence of the tumor but produced images of inferior specificity compared with the use of ¹⁸F-VHHs (Fig. 4C and Movie S7).

**Successful Detection of Infiltrated Myeloid Cells Using ¹⁸F-Labeled VHHDC13 upon Injection of Complete Freund’s Adjuvant.** Finally we examined inflammation in response to administration of complete Freund’s adjuvant (CFA). Administration of CFA leads to the influx and accumulation of myeloid cells at the site of the injection and draining lymph nodes within 24 h. When we injected animals with CFA into the left front paw and imaged them 24 h later with ¹⁸F- or ⁶⁴Cu-labeled VHH7, inflammation was readily apparent, with VHHDC13 showing a stronger signal than VHH7 in the inflamed region (Fig. 4 D and E, and Movies S8A, S8B, S9, S10, and S11). ¹⁸F-2-fluorodeoxyglucose (FDG) was used for comparison and failed to detect inflammation evoked by CFA (Fig. 4F and Movies S12A, S12B, and S12C). Accumulation of ¹⁸F-VHHDC13 is consistent with the massive influx of CD11b+ neutrophils generally observed at the site of injection 24 h after administration of CFA.

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*Fig. 3.* ¹⁸F-VHH7 (anti-mouse class II MHC) and ¹⁸F-VHHDC13 (anti-mouse CD11b) detects inflammation. Tumor-associated class II MHC+ cells were visualized using ¹⁸F-VHH7. A NOD-SCID mouse was inoculated subcutaneously on the back of the left shoulder with 5 × 10⁶ human Mel-Juso melanoma cells and imaged 35 days postinjection. (A–C) Coronal PET-CT images, moving from anterior to posterior. In A and B, different sets of lymph nodes are visible. In C, tumor-associated class II MHC+ cells are visible, attributable to influx of host-derived class II MHC+ cells. (D) PET-CT as maximum intensity projections of all slices. See Movie S3 for a 3D visualization of lymph nodes and tumor-associated class II MHC+ cells. (E) ¹⁸F-VHH7 detects class II MHC+ cells associated with small tumors at early stages. NOD/SCID mice were inoculated with Mel-Juso cells as in A, 20 d (Upper) and 6 d (Lower) prior to imaging. Transverse PET and CT images (Left and Right, respectively) are shown after better visualization of the class II MHC+ cells at the site of cancer cells. Images are all window-leveled to the same intensity. Tumors and associated class II MHC+ cells are highlighted with arrows. Axillary (label A), brachial (label B), and mediastinal (label M) lymph nodes and thymus (label T) are shown in E, Lower. (F and G) A NOD-SCID mouse was inoculated subcutaneously on the back of the left shoulder with 5 × 10⁶ human Mel-Juso melanoma cells and imaged 35 days postinjection with ¹⁸F-VHHDC13. Tumor-associated CD11b+ cells are visible, attributable to influx of host-derived CD11b+ cells. See Movie S4 for a 3D visualization. (H and I) FACs analysis of tumor-infiltrating immune cells. The next day, tumors were excised and digested with collagenase D, and tumor-infiltrating cells were obtained after Percoll gradient. Cell suspensions were then stained for FACS analysis. (H) Histograms show the FACs staining of mouse CD45+ tumor-infiltrating cells with VHH7-Alexa 647. Histograms on the Left are gated on CD11c+CD11b+ cells (dendritic cells), and histograms on the Right are gated on CD11c+CD11b+ cells (macrophages and other myeloid cells) for the indicated time points. (I) FACs analysis of tumor-infiltrating cells were harvested 35 d after tumor inoculation as in H and stained with VHHDC13-Alexa 647. Histograms show the levels of CD11b as measured by VHHDC13 on the indicated cell populations. Splenocytes from the same mouse are shown for comparison. Histograms are representative of two to four mice with similar results. (J) PET signals in vivo for the indicated experiment in D and F. (K) PET signals in vivo for the indicated experiment in E. (L) Hematoxylin/eosin (H&E) stains of the tumor from the mice in the part D (human melanoma tumor in NOD-SCID mouse; 35 d postinjection of the cells).
Discussion

Using a combination of single domain antibodies (VHHs), we showed that it is possible to track immune responses with excellent specificity. CD11b+ or class II MHC+ cells can serve as sentinels to detect the presence of the cancerous tissue they surround or invade. Although class II MHC and CD11b are expressed on many activated myeloid cells, such as macrophages and dendritic cells, the observed signal strength for imaging may vary with the extent of activation, and not all tumors may recruit myeloid cells equally efficiently. The approach described here obviates the need for the generation of panels of tumor-specific antibodies, if indeed such information on various forms of therapy. An interesting possibility in the application of antibodies that target immunological checkpoints, there is an urgent need to monitor the progress of such immunotherapies. The advantages of the small size of VHHs in conjunction with PET-compatible isotopes, such as 18F, should enable further refinement of immuno-PET in a clinical setting. Although the present results are geared toward the detection of neutrophils and antigen-presenting cells, an important future goal will be to extend this approach to T cell-specific markers.

Materials and Methods

Synthesis of (Gly)₃-Tetrazine.

The tetrapeptide GGGC was synthesized by standard solid-phase peptide synthesis. Maleimide-tetrazine (ClickChemistryTools) was dissolved in 0.1 M phosphate buffer (pH 7). The tetrapeptide GGGC was added and left to stir at room temperature for 3 h until TLC (1:1 Hex:EtOAc vol/vol) indicated near-complete conversion to the product. The solution was filtered and purified by reverse-phase HPLC with a semipreparative column (C₁₈ column, Phenomenex) at a flow rate of 5.0 mL/min: solvent A, 0.1% TFA in H₂O; solvent B, 0.1% TFA in CH₃CN. (G)₃-Tetrazine eluted at 30–35% (vol/vol) solvent B. Fractions containing pure product were collected and lyophilized. LC-MS calculated for C₂₃H₂₄N₁₀O₁₂[S⁺M+H⁺] was 892.362, found 892.370.

Synthesis of (Gly)₃-NOTA. Maleimide-NOTA (Macromolecules) was dissolved in 0.1 M phosphate buffer (pH 7). The tetrapeptide GGGC was added and left to stir at room temperature for 3 h until TLC (1:1 Hex:EtOAc vol/vol) indicated almost complete conversion to the product. The solution was filtered and purified by reverse-phase HPLC with a semipreparative column (C₁₈ column, Phenomenex) at a flow rate of 5.0 mL/min: solvent A, 0.1% TFA in H₂O; solvent B, 0.1% TFA in CH₃CN. (G)₃-Tetrazine eluted at 30–35% (vol/vol) solvent B. Fractions containing pure product were collected and lyophilized. LC-MS calculated for C₂₃H₂₄N₁₀O₁₂[S⁺M+H⁺] was 892.362, found 892.370.

Accumulation of myeloid cells is a general phenomenon, as suggested by the available evidence (2, 34). With the availability of anti-human VHHs, the ability to monitor the presence or absence of neutrophils and activated macrophages as an indicator of inflammation should be transposable to a clinical setting as a diagnostic tool. The potential immunogenicity of VHHs may limit their therapeutic use, especially when repeated administration would be required, but approaches to humanize VHHs have been described (12). VHH-based PET will likely complement the information provided by 18F-FDG PET. The metabolic status of tumors can be assessed using 18F-FDG PET in ways that VHH-PET or other forms of immuno-PET cannot. In contrast, VHH-PET can in principle reveal the occurrence and extent of an immune response and do so noninvasively. The combination of 18F-FDG-PET and VHH-PET may provide valuable additional information on various forms of therapy. An interesting possibility is the development of single-domain antibodies capable of recognizing markers on T cells with distinct functional properties, including exhausted T cells, for example, by the presence of PD-1, and the presence of PD-L1 on tumors. Given the recent successes
Enzymatic Incorporation of Substrates into Proteins Using Sortase. The pentamutant sortase A, with an improved $K_{\text{cat}}$, was used (35). Reaction mixtures (1 mL) contained Tris-HCl (50 mM pH 7.5), CaCl$_2$ (10 mM), NaCl (150 mM), triglycine-containing probe (500 μM), LPEGT-containing probe (100 μM), and sortase (5 μM) (11, 36). After incubation at 4 °C with agitation for 2 h, reaction products were analyzed by LC-MS, with yields generally >90%. When the yield was below 90%, the reaction was allowed to proceed for an additional 2 h, with addition of sortase to 10 μM and triglycine-containing probe to 750 μM. Ni-NTA beads were added to the reaction mixture with agitation for 5 min at 25 °C, followed by centrifugation to remove sortase and any remaining unreacted His-tagged substrate. The final product—either the tetrazene-labeled protein, NOTA-labeled protein, or fluorophore-labeled protein—was purified by size-exclusion chromatography in PBS or Tris-HCl (50 mM pH 7.5). The labeled protein was stored at −20 °C with 5% (v/v) glycerol and was stable for up to 3 mo.

Synthesis and Characterization of $^{18}$F-VHHs. In a typical reaction, a 1.5-mL centrifuge tube was loaded with VHH7-Tz in 1 mL PBS (40 μL 150 mM), 1× PBS (300 μL), and $^{18}$F-triC in DMSO [4.0 μCi (148.0 MBq), 100 μL]. The tube was sealed and shaken at room temperature for 20 min. The mixture was analyzed by radio-TLC (instant thin layer chromatography (ITLC), 100% MeCN, $R_f^{18}$-F-triC = 0.9, $R_f$-VHH7 = 0.0) showing 90% conversion to $^{18}$F-VHH7. The reaction mixture was loaded onto a PD-10 size-exclusion cartridge (GE Healthcare), and elution with 1× PBS provided 2.3 mCi (85.1 MBq) of $^{18}$F-VHH7 in 75.8% decay-corrected radiochemical yield. Reaction products were analyzed by LC-MS, with yields generally >90%, the reaction was allowed to proceed for an additional 2 h, with addition of sortase to 10 μM and triglycine-containing probe to 750 μM. Ni-NTA beads were added to the reaction mixture with agitation for 5 min at 25 °C, followed by centrifugation to remove sortase and any remaining unreacted His-tagged substrate. The final product—either the tetrazene-labeled protein, NOTA-labeled protein, or fluorophore-labeled protein—was purified by size-exclusion chromatography in PBS or Tris-HCl (50 mM pH 7.5). The labeled protein was stored at −20 °C with 5% (v/v) glycerol and was stable for up to 3 mo.

Synthesis and Characterization of $^{64}$Cu-VHHs. In a typical reaction, a 1.5-mL centrifuge tube was loaded with VHH7-Tz in 1 mL PBS (40 μL 150 mM), 1× PBS (300 μL), and $^{64}$Cu-triC in DMSO [4.0 μCi (148.0 MBq), 100 μL]. The tube was sealed and shaken at room temperature for 20 min. The mixture was analyzed by radio-TLC (instant thin layer chromatography (ITLC), 100% MeCN, $R_f^{64}$Cu-triC = 0.9, $R_f$-VHH7 = 0.0) showing 90% conversion to $^{64}$Cu-VHH7. The reaction mixture was loaded onto a PD-10 size-exclusion cartridge (GE Healthcare), and elution with 1× PBS provided 2.3 mCi (85.1 MBq) of $^{64}$Cu-VHH7 in 75.8% decay-corrected radiochemical yield. Reaction products were analyzed by LC-MS, with yields generally >90%, the reaction was allowed to proceed for an additional 2 h, with addition of sortase to 10 μM and triglycine-containing probe to 750 μM. Ni-NTA beads were added to the reaction mixture with agitation for 5 min at 25 °C, followed by centrifugation to remove sortase and any remaining unreacted His-tagged substrate. The final product—either the tetrazene-labeled protein, NOTA-labeled protein, or fluorophore-labeled protein—was purified by size-exclusion chromatography in PBS or Tris-HCl (50 mM pH 7.5). The labeled protein was stored at −20 °C with 5% (v/v) glycerol and was stable for up to 3 mo.