Reappraisal of hydrocarbon biomarkers in Archean rocks

The MIT Faculty has made this article openly available. Please share how this access benefits you. Your story matters.

<table>
<thead>
<tr>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>As Published</td>
<td><a href="http://dx.doi.org/10.1073/pnas.1419563112">http://dx.doi.org/10.1073/pnas.1419563112</a></td>
</tr>
<tr>
<td>Publisher</td>
<td>National Academy of Sciences (U.S.)</td>
</tr>
<tr>
<td>Version</td>
<td>Final published version</td>
</tr>
<tr>
<td>Accessed</td>
<td>Fri Apr 05 00:11:22 EDT 2019</td>
</tr>
<tr>
<td>Citable Link</td>
<td><a href="http://hdl.handle.net/1721.1/100743">http://hdl.handle.net/1721.1/100743</a></td>
</tr>
<tr>
<td>Terms of Use</td>
<td>Article is made available in accordance with the publisher’s policy and may be subject to US copyright law. Please refer to the publisher’s site for terms of use.</td>
</tr>
<tr>
<td>Detailed Terms</td>
<td>-----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------</td>
</tr>
</tbody>
</table>
Reappraisal of hydrocarbon biomarkers in Archean rocks

Katherine L. French,a,1,2 Christian Hallmannb,c, Janet M. Hoped, Petra L. Schoon,e J. Alex Zumberge,f Yosuke Hoshino,f Carl A. Petersg, Simon C. Georgew, Gordon D. Lovex, Jochen J. Brocksd, Roger Buickf, and Roger E. Summonsb

aJoint Program in Chemical Oceanography, Massachusetts Institute of Technology and Woods Hole Oceanographic Institution, Cambridge, MA 02139; bMax Planck Institute for Biogeochemistry, Jena, Germany; cCenter for Marine Environmental Sciences, University of Bremen, 28359 Bremen, Germany; dResearch School of Earth Sciences, The Australian National University, Canberra, ACT 0200, Australia; eDepartment of Earth Sciences, University of California, Riverside, CA 92521; fDepartment of Earth and Planetary Sciences, Macquarie University, Sydney, NSW 2109, Australia; gDepartment of Earth & Space Sciences and Astrobiology Program, University of Washington, Seattle, WA 98195-1310; and hDepartment of Earth, Atmospheric, and Planetary Sciences, Massachusetts Institute of Technology, Cambridge, MA 02139

Edited by Andrew H. Knoll, Harvard University, Cambridge, MA, and approved March 16, 2015 (received for review October 21, 2014)

Hopanes and steranes found in Archean rocks have been presented as key evidence supporting the early rise of oxygenic photosynthesis and eukaryotes, but the syngeneity of these hydrocarbon biomarkers is controversial. To resolve this debate, we performed a multilaboratory study of new cores from the Pilbara Craton, Australia, that were drilled and sampled using unprecedented hydrocarbon-clean protocols. Hopanes and steranes in rock extracts and hydroprolysates from these new cores were typically at or below our femtogram detection limit, but when they were detectable, they had total hopane (37.9 pg per gram of rock) and total sterane (32.9 pg per gram of rock) concentrations comparable to those measured in blanks and negative control samples. In contrast, hopanes and steranes measured in the exteriors of conventionally drilled and curated rocks of stratigraphic equivalence reach concentrations of 389.5 pg per gram of rock and 1,039 pg per gram of rock, respectively. Polycyclic aromatic hydrocarbons and diamondoids, which exceed blank concentrations, exhibit individual concentrations up to 80 ng per gram of rock in rock extracts and up to 1,000 ng per gram of rock in hydroprolysates from the ultraclean cores. These results demonstrate that previously studied Archean samples host mixtures of biomarker contaminants and indigenous overmature hydrocarbons. Therefore, existing lipid biomarker evidence cannot be invoked to support the emergence of oxygenic photosynthesis and eukaryotes by ~2.7 billion years ago. Although suitable Proterozoic rocks exist, no currently known Archean strata lie within the appropriate thermal maturity window for syngeneic hydrocarbon biomarker preservation, so future exploration for Archean biomarkers should screen for rocks with milder thermal histories.

Significance

The advent of oxygenic photosynthesis set the stage for the evolution of complex life on an oxygenated planet, but it is unknown when this transformative biochemistry emerged. The existing hydrocarbon biomarker record requires that oxygenic photosynthesis and eukaryotes emerged more than 300 million years before the Great Oxidation Event (~2.4 billion years ago (Ga)). We report that hopane and sterane concentrations measured in new ultraclean Archean drill cores from Australia are comparable to blank concentrations, yet their concentrations in the exteriors of conventionally collected cores of stratigraphic equivalence exceed blank concentrations by more than an order of magnitude due to surficial contamination. Consequently, previous hydrocarbon biomarker reports no longer provide valid evidence for the advent of oxygenic photosynthesis and eukaryotes by ~2.7 Ga.


The authors declare no conflict of interest.

This article is a PNAS Direct Submission.

1Present address: Department of Marine Chemistry and Geochemistry, Woods Hole Oceanographic Institution, Woods Hole, MA 02543.

2To whom correspondence should be addressed. Email: kfrench@whoi.edu.

This article contains supporting information online at www.pnas.org/lookup/suppl/doi:10.1073/pnas.1419563112/-/DCSupplemental.

www.pnas.org/cgi/doi/10.1073/pnas.1419563112

PNAS | May 12, 2015 | vol. 112 | no. 19 | 5915–5920
drilling methods (19) and reportedly contains syngenetic Archean biomarkers (3, 4). Spatially separated by less than 1 km, AIDP-2 and RHDH2A both intersept the Carawine Dolomite (<2.63 Ga) of the Hamersley Group and the Jeerinah Formation (20), so they sample the same stratigraphy and have experienced the same regional metamorphic effects (Fig. 1). AIDP-3 (21°46'32''S, 117°34'11''E), which intersected the Marra Mamba Iron Formation and the Jeerinah Formation, was drilled as a time-equivalent core in deeper water facies than AIDP-2. It is stratigraphically equivalent to WRL-1, which was the subject of the first report, to our knowledge, of Archean biomarkers for cyanobacteria and eukaryotes (1).

The AIDP cores were drilled and sampled using protocols chosen to minimize hydrocarbon contamination. At the beginning of the AIDP campaign, all core barrels were cleaned with soap and water, and, subsequently, only new drill bits were used. Water without added lubricants was used as the sole drilling fluid for the entirety of the program. AIDP-1, which mainly intersected metamorphosed igneous rocks, provided negative control samples (i.e., nonsedimentary rocks devoid of syngenetic organic matter) and abrasively removed residual organics that may have accumulated on the drill rods during previous projects. Minutes after recovery from depth, untouched portions of selected core material were rinsed with Type-1 ultrapure water in the aluminum core tray before being placed into organic-clean Teflon bags containing several milliliters of ultrapure water to prevent the core material from drying out. After purging with high purity Ar, the sample bags were sealed and frozen at −20 °C in the dark until the whole cores were sawed. In total, the whole-core samples were packaged, purged, and frozen in less than 20 min after recovery from depth.

**Methods**

Systematic blank analyses using solvent rinses and sawed pieces of a combusted brick demonstrated that the diamond-rimmed blade used with the newly purchased, water-cooled saw constructed entirely of stainless steel and aluminum was a significant vector for hydrocarbon biomarker contamination. After exhausting other methods (see SI Appendix), the saw blade was heated to 300 °C for 1 h and was sonicated in solvent, after which solvent rinses and combusted brick blanks verified that sawing no longer contributed detectable hydrocarbon contaminants. Subsequently, the frozen whole-core samples were sawed into book-matched halves at the Australian National University (ANU), which were distributed to multiple laboratories for independent analyses. The extractable biomarker methods and data described below are specific to the ANU/Massachusetts Institute of Technology (MIT) joint dataset, which analyzed the majority of samples for extractable hydrocarbons (SI Appendix, Table S1). However, all results from all laboratories are consistent with the results described here.

Approximately ~5- to 10-mm-thick slabs of the rock surface were sawed off from the half-core, such that all exposed surfaces were removed (Fig. 1). These rock pieces were collected as the “exterior” sample, while the remaining rock served as the “interior” sample. The interior and exterior samples were prepared in parallel with a combusted sand blank. Between ~10 g and 40 g of the rock powder were extracted with solvents (SI Appendix, Table S2). Each extract was concentrated and redissolved in 100 μL of...
hexane for analyses by gas chromatography mass spectrometry in full scan and metastable reaction monitoring (MRM) modes (see SI Appendix). Each laboratory analyzed either a chert or igneous negative control sample from AIDP-2 or AIDP-1 for extractable hydrocarbons before the analyses of the biomarker target samples. In total, the interiors and exteriors of 10 biomarker target samples across the Carawine Dolomite and Jeerinah Formation of AIDP-2 and 2 biomarker target samples from the Jeerinah Formation of AIDP-3 have been analyzed for extractable hydrocarbons (SI Appendix, Table S1).

Results and Discussion

Hopanes and steranes in the rock extracts from AIDP-2 and AIDP-3 were typically at or below femtogram detection limits, but if they were detectable, the total sterane and total hopane concentrations were comparable to blank and negative control concentrations (Table 1 and Fig. 1). For comparison, the hopane and sterane concentrations that were detectable and measured in the AIDP samples are approximately one to three orders of magnitude less than the hopane and sterane concentrations reported in earlier Archean biomarker studies (7, 21). Moreover, extractable acyclic isoprenoids and a homologous series of extractable n-alkanes (C_{10-23}), both of which were prominent in previous Archean biomarker studies (1–3, 5, 7, 22, 23), were also below detection limits or cumulative laboratory blank concentrations in the AIDP samples.

Hydropyrolysates, which are solvent-soluble hydrocarbons released from the kerogen by cracking under high-pressure hydrogen, were analyzed from preextracted rock powders or kerogens (insoluble organic matter) from eight samples from AIDP-2 and AIDP-3. As in the rock extracts, individual hopanes and steranes in the AIDP-2 and AIDP-3 hydropyrolysates were usually at or below detection limits, but when they were detected, neither the total hopane nor the total sterane concentrations exceeded cumulative blank concentrations (<20 pg per gram of rock). These results are consistent with the absence of hopanes and steranes in previous Archean hydropyrolysis (HyPy) studies (24, 25).

However, a series of C_{10-20} n-alkanes (with a maximum abundance at C_{13}), methylalkanes, and alkylcyclohexanes were detected in hydropyrolysates from two carbonate-rich rocks at 116.09 m and 170.40 m in the Carawine Dolomite of AIDP-2 (SI Appendix, Fig. S6), yet acyclic isoprenoids were not detected in any of the hydropyrolysates. The two samples containing alkanes also hosted the least thermally altered kerogens according to their atomic H/C ratios and pyrolysate polycyclic aromatic hydrocarbon (PAH) ratios. Most samples had H/C ratios of 0.19–0.35 (SI Appendix, Table S5), but these two samples have the highest H/C ratios of 0.54 and 0.46. The same two samples also have the highest abundance of alkylated relative to nonalkylated PAH in the pyrolysates (SI Appendix, Table S10). For example, the methylphenanthrene/phenanthrene (MP/P) ratios for these two samples were 1.0–1.2 compared with the other samples that have MP/P values ranging from 0.2 to 0.5. Similarly, they had methylpyrene/pyrene (MePy/Py) ratios of 0.4–0.6 compared with the other samples that have MePy/Py values close to 0.1. The total n-alkane concentrations in these hydropyrolysates were two orders of magnitude higher than in laboratory blanks (<10 ng per gram of rock), in which methylalkane and alkylcyclohexane series were not detected. The n-alkanes detected in the Carawine Dolomite hydropyrolysate from 116.09 m in AIDP-2 had δ^{13}C values ranging from −38.1‰ to −43.6‰ (SI Appendix, Table S7) compared with the corresponding bulk δ^{13}C of −43.1‰ (SI Appendix, Table S5). This isotopic similarity indicates that these compounds were generated from the kerogen. In contrast, n-alkanes detected in the Jeerinah Formation black shales and more-thermally altered Carawine Dolomite hydropyrolysates were comparable to laboratory blank concentrations (SI Appendix, Table S10).

To further constrain the source of extractable hydrocarbons previously reported in Archean rocks, a Carawine Dolomite quarter core sample from the RHDH2A core (130.2–130.4 m) was analyzed using the same laboratory protocol described for the AIDP samples. Like the AIDP samples, 25–31 g of sample interior and exterior were extracted (SI Appendix, Table S2). The n-alkanes, hopanes, and steranes were enriched on the exterior of the RHDH2A core compared with the interior, where the total hopane and total sterane concentrations in the exterior exceeded the interior and blank concentrations by one to two orders of magnitude (Table 1 and Fig. 1). These patterns substantiate earlier observations of biomarker and bitumen enrichments on the surfaces of conventionally drilled and curated samples relative to the interiors (3, 6, 11). Furthermore, the hopane and sterane concentrations measured in our analysis of the RHDH2A sample are more compatible with the concentrations reported in earlier Archean biomarker studies of the Mount Bruce and Transvaal supergroups (7, 21) than the concentrations measured in the AIDP samples. Comparison of RHDH2A and cumulative blank concentrations indicate that

Table 1. ANU/MIT hopane and sterane concentrations, in picograms per gram of rock

<table>
<thead>
<tr>
<th>Sample</th>
<th>Interior ΣC_{25-34} hopane concentration</th>
<th>Interior ΣC_{26-29} sterane concentration</th>
<th>Exterior ΣC_{30-34} hopane concentration</th>
<th>Exterior ΣC_{26-29} sterane concentration</th>
</tr>
</thead>
<tbody>
<tr>
<td>Range for blanks*</td>
<td>nd to 37.9</td>
<td>nd to 32.9</td>
<td>nd to 37.9</td>
<td>nd to 32.9</td>
</tr>
<tr>
<td>AIDP-2 441.00 m (negative control)</td>
<td>nd</td>
<td>nd</td>
<td>31.4</td>
<td>nd</td>
</tr>
<tr>
<td>AIDP-2 146.20 m</td>
<td>nd</td>
<td>nd</td>
<td>10.7</td>
<td>11.8</td>
</tr>
<tr>
<td>AIDP-2 170.40 m</td>
<td>nd</td>
<td>nd</td>
<td>50.4</td>
<td>49.4</td>
</tr>
<tr>
<td>AIDP-2 197.20 m</td>
<td>nd</td>
<td>nd</td>
<td>7.3</td>
<td>16.5</td>
</tr>
<tr>
<td>AIDP-2 291.28 m</td>
<td>nd</td>
<td>nd</td>
<td>8.0</td>
<td>nd</td>
</tr>
<tr>
<td>AIDP-2 301.40 m</td>
<td>nd</td>
<td>nd</td>
<td>nd</td>
<td>nd</td>
</tr>
<tr>
<td>AIDP-2 325.48 m</td>
<td>nd</td>
<td>nd</td>
<td>nd</td>
<td>nd</td>
</tr>
<tr>
<td>AIDP-2 327.42 m</td>
<td>nd</td>
<td>nd</td>
<td>38.5</td>
<td>24.3</td>
</tr>
<tr>
<td>AIDP-3 98.36 m</td>
<td>nd</td>
<td>nd</td>
<td>nd</td>
<td>nd</td>
</tr>
<tr>
<td>AIDP-3 130.30 m</td>
<td>nd</td>
<td>nd</td>
<td>nd</td>
<td>nd</td>
</tr>
<tr>
<td>RHDH2A interior ‡</td>
<td>3.5</td>
<td>21.1</td>
<td>250</td>
<td>1,092</td>
</tr>
<tr>
<td>RHDH2A sawed surface 1 †</td>
<td>259</td>
<td>509</td>
<td>390</td>
<td>468</td>
</tr>
</tbody>
</table>

*Note that the same blank concentration ranges measured for hopanes and steranes are provided in the interior and exterior columns.

†The RHDH2A 130.2–130.4 m sample surfaces were cut according to Fig. 1.
laboratory procedures cannot fully account for the addition of biomarkers to RHDH2A. Clearly, the hydrocarbons must have been present in, or added to, the RHDH2A quarter core before laboratory preparation.

Two hypotheses have been proposed to explain surficial biomarker enrichments in Archean drill core: contamination and the live oil effect (i.e., the outward migration of hydrocarbons due to drill core depressurization during recovery) (11). If the live oil effect generated the surficial biomarker enrichments, then samples from both AIDP-2 and RHDH2A should display surficial hopane and sterane enrichments in excess of cumulative blanks. Instead, contamination is the only explanation for the reproducibility of the detectable surficial hopanoids and steranes in conventionally drilled and curated Archean samples (3, 6, 7, 11) and the undetectable or blank level-equivalent hopane and sterane concentrations in samples from the ultraclean AIDP drill cores.

The spatial distribution of AIDP-2 and AIDP-3 geographically brackets and nearly spans the entire stratigraphic section from which all of the other Pilbara biomarkers have been previously reported (1, 3, 4, 21). According to currently available H/C ratios and other indicators of thermal history, the metamorphic grades of AIDP-2 and AIDP-3 are similar to (1–4, 6, 7, 15, 21, 26) or lower than (5, 22, 23) the metamorphic grades of the rocks from which biomarkers have been previously reported. Moreover, known vectors and signatures of contamination (i.e., surfactant enrichments) are independent of sampling locality (6, 11). For example, the analyses of the saw blade (before it was heat treated) and the sawed surfaces of the RHDH2A quarter-core highlight that sawing can contribute significant but variable hydrocarbon contaminants to the substrate, yet the cumulative blanks reported in previous studies did not record the hydrocarbon contribution from sawing (1, 4, 6, 7). Furthermore, recent studies have illustrated that petroleum-derived chemical fossils are prevalent in aerosols, providing another potential vector for hydrocarbon contamination (27). Evidently, samples that were the subject of earlier Archean biomarker work accumulated contaminants on surfaces during processes that were not recorded by laboratory blanks (e.g., drilling, sampling, sawing, and/or storage). If true, then trace amounts of these unforeseen hydrocarbon contaminants could have migrated inward as the samples dried out, oxidized, and fractured (11). Alternatively, using solvents to clean surface exteriors could have remobilized the hydrocarbon contaminants into the sample interior (6). Therefore, the results and implications from the new AIDP cores logically extend from the Pilbara Craton to other localities (1, 2, 4, 7, 21–23), so the analysis and implications from the new AIDP cores are independent of sampling locality (6, 11).

Many AIDP samples, however, host extractable PAHs and diamondoids with concentrations up to 80 ng per gram of rock and 4 ng per gram of rock, respectively (SI Appendix, Fig. S4). PAHs and diamondoids were identified by comparison with published mass spectra, published relative retention times, and parallel analysis with standards. PAHs and diamondoids were also detected in the hydropropyrolates, where individual PAH concentrations were up to 1039 ng per gram of rock, and their concentrations were significantly higher than their corresponding HyPy blank concentrations of less than 1 ng per gram of rock for phenanthrene and pyrene (SI Appendix, Table S10). For both rock extracts and hydropropyrolates, the Carawine Dolomite hosted significantly higher concentrations of PAHs and diamondoids than the underlying Jeerinah Formation black shale samples. Like the PAHs and diamondoids that previous studies reported in Archean rock extracts and hydropropyrolates (3, 21, 24, 28), the low molecular mass aromatics (≤C16) dominate over high molecular mass PAHs (>C17), and nonalkylated parent PAHs are in greater abundance compared with alkylated PAH (SI Appendix, Figs. S4 and S5 and Tables S8 and S10).

The PAHs and diamondoids are indigenous (i.e., native to the rock before drilling) based on several lines of evidence. First, their concentrations in the sample material significantly exceed laboratory blank PAH and diamondoid concentrations. For PAHs that are less sensitive to evaporation, the exterior/interior distribution ratios were close to 1. PAHs and diamondoids were detected in both rock extracts and hydropropyrolates. Finally, the PAH and diamondoid distribution and alkylation patterns impart high thermal signatures that match the regional metamorphic grade. More specifically, a predominance of unsubstituted, condensed PAHs is characteristic of hydrothermal petroleum (29, 30), ancient mineral deposits (31, 32), pyrolysis and combustion products (33), and high-maturity organic matter (34). Based on extractable PAHs, the MPI-1 parameter yields calculated vitrinite reflectances (Rv) between 2.63–3.00% (SI Appendix, Table S9) (21, 35) [MPI-1 = 1.5 + (2-MeP + 3-MeP)/(P + 1-MeP + 9-MeP)], where P is phenanthrene and MeP is methyl phenanthrene]. These Rv values are compatible with the measured H/C ratios, which ranged from 0.2 to 0.5 (SI Appendix, Table S5). At a vitrinite reflectance of 2% and H/C ratio of 0.5, catagenesis transitions into metagenesis, which corresponds to the dry gas zone where only methane is generated (12, 36). Although duration of heating and type of organic matter complicate direct conversions between Rv, H/C ratio, and burial temperature, the PAH and diamondoid distributions in tandem with the H/C ratios confirm that these rocks are overmature and are within the prehnite–pumpellyite metamorphic facies (12, 15, 17, 36–38).

Stable carbon isotopic data can be assessed to test whether the indigenous PAHs and diamondoids were generated from the coexisting kerogen and have the same age as the host rock (i.e., syngentic) or, conversely, migrated from underlying or overlying strata. Consistent with earlier work (1, 10, 39, 40), the bulk organic carbon was found to be anomalously 13C-depleted, with 613C values ranging from −32‰ to −50‰ (Fig. 2 and SI Appendix, Table S5). The extractable PAHs from the Carawine Dolomite were typically 13C-enriched by 10–20‰ compared with the host rock organic matter (Fig. 2 and SI Appendix, Table S6), with two exceptions. First, the sample at 133.55 m exhibited an isotopic match (Δ < 0.8‰) between the extractable PAHs (−32.3‰ to −33.2‰) and the associated kerogen (−32.4‰), which was enriched in 13C by >10‰ compared with the bulk δ13C of the other AIDP samples. This anomalously 13C-enriched sample was a 10-cm pyrobituminous styloluculate occurring in stylolitic to brecciated dolarenite of the upper Carawine where dark insoluble matter concentrated as a result of pressure solution and carbonate dissolution (SI Appendix, Fig. S3). Notably, the δ13C values of the PAHs from this sample and the PAHs from the other AIDP Carawine Dolomite samples are equivalent, suggesting that they could share the same allochthonous source. Second, three high molecular mass PAHs (228–252 Da), identified as benzo-naphtho-thiophene, benz[a]anthracene, and benzo[e] pyrene, from the AIDP-2 sample at 116.09 m had isotopic values ranging from −38.9‰ to −41.3‰, yielding an offset from the host rock kerogen of 1.8–4.3‰. These three compounds are the most 13C-depleted PAHs that have been reported from Archean rock extracts (21, 24, 25). Only a single compound-specific isotopic measurement was possible for this sample, so the analytical uncertainty and reproducibility are not well constrained. In contrast to the extractable PAHs, the δ13C of the hydropropyrolate PAHs match the bulk organic matter within a range of 0.1‰ to 2.9‰ (Fig. 2), and they are the most 13C-depleted organic molecules reported from Archean rocks, ranging from −42.6‰ to −45.9‰ (SI Appendix, Table S7). The PyHy hydrocarbon δ13C values reported here are, to our knowledge, the first compound-specific isotopic measurements that are in agreement with the strongly
C-depleted kerogens ($\delta^{13}C_{\text{bulk}} < -39$‰) of the Neoarchean (1, 10, 39, 40). Previous work suggests that the Jeerinah Formation and other organic-rich Archean shales generated and expelled petroleum (17) and that the Pilbara Craton, including the Ripon Hills region, has experienced multiple episodes of hydrothermal fluid flow, particularly around the time of peak regional metamorphism at $\sim 2.1$ Ga (18, 41). The isotopic match between the AIDP bulk organic matter and the hydropyrolysate PAHs supports a syngenetic origin for the hydropyrolysate PAHs generated by thermochemical cleavage of covalent bonds within the host kerogens. In contrast, the $\delta^{13}C$ values of extractable PAHs do not covary with the AIDP bulk organic $\delta^{13}C$ through the carbon isotopic anomaly at 2.6–2.7 Ga, suggesting that the kerogens of the Jeerinah Formation and Carawine Dolomite may not necessarily be the primary source of the cooccurring extractable PAHs, despite strong evidence of indigeneity. The $^{13}C$ enrichment of the bulk organic matter hosted in the stylolitic sample at 133.55 m in tandem with PAHs of equivalent isotopic composition throughout the Carawine Dolomite provides further evidence for emplacement of migrated allochthonous organic matter that was rich in PAHs. Supercritical hydrothermal fluids rich in CO$_2$ or CH$_4$ are efficient solvents for organic compounds, and unsubstituted, condensed PAHs are particularly abundant in hydrothermal fluids (29, 30, 42). Therefore, hydrothermal fluids could account for PAH migration from its original source, where this second generation of migrated overmature hydrothermal PAHs would have diluted or replaced the primary bitumens. Alternatively, lateral oil migration before peak metamorphism could have introduced the extractable organic matter with isotopic compositions distinct from the associated kerogen. The extractable high molecular mass PAHs from 116.09 m that have isotopic compositions approaching the bulk organic carbon isotopic composition hint at mixing between syngenetic high molecular mass extractable PAHs and migrated PAHs containing one to three fused aromatic rings, but future studies are required to test this observation. Lastly, the extractable PAHs could be syngenetic but heavily altered by reactions between hydrothermal fluids and the bitumen (28), but this mechanism may not be able to account for the full $>10$‰ $^{13}C$ enrichment. Regardless of the source of the extractable PAHs, the biomolecular precursors of the PAHs and diamondoids are unknown.

We have demonstrated the feasibility of collecting organic-lean and/or overmature drill core samples without the addition of contaminant biomarker hydrocarbons. Our results verify that hydrocarbon concentration gradients through a sample can be used to distinguish contaminants from indigenous hydrocarbons.


