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Longitudinal spread of mechanical excitation through tectorial membrane traveling waves

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The mammalian inner ear separates sounds by their frequency content, and this separation underlies important properties of human hearing, including our ability to understand speech in noisy environments. Studies of genetic disorders of hearing have demonstrated a link between frequency selectivity and wave properties of the tectorial membrane (TM). To understand these wave properties better, we developed chemical manipulations that systematically and reversibly alter TM stiffness and viscosity. Using microfabricated shear probes, we show that (i) reducing pH reduces TM stiffness with little change in TM viscosity and (ii) adding PEG increases TM viscosity with little change in TM stiffness. By applying these manipulations in measurements of TM waves, we show that TM wave speed is determined primarily by stiffness at low frequencies and by viscosity at high frequencies. Both TM viscosity and stiffness affect the longitudinal spread of mechanical excitation through the TM over a broad range of frequencies. Increasing TM viscosity or decreasing stiffness reduces longitudinal spread of mechanical excitation, thereby coupling a smaller range of best frequencies and sharpening tuning. In contrast, increasing viscous loss or decreasing stiffness would tend to broaden tuning in resonance-based TM models. Thus, TM wave and resonance mechanisms are fundamentally different in the way they control frequency selectivity.

Significance

The sharp frequency selectivity of auditory nerve fiber responses to sound is a hallmark of mammalian cochlear function. This remarkable signal processing originates in the cochlear signal processing chain (1–7), as evidenced by measured motions and mechanical properties of the basilar membrane (BM) (2–9) and tectorial membrane (TM) (10–24). Although the hydromechanical mechanisms underlying BM motions have been characterized based on experimental and theoretical studies, the mechanisms underlying TM motions remain unclear.

The TM is an acellular matrix that overlies the hair bundles of sensory receptor cells. Based on its strategic position above the organ of Corti, conventional cochlear models (25–29) have implicated local mechanical properties (i.e., mass, stiffness) of the TM in stimulating the sensory hair bundles of hair cells and in cochlear tuning. Recent dynamic measurements of the TM, in vitro (17, 30–33) and in vivo (34), suggest that the TM supports longitudinal coupling, with large spatial extents across a broad range of frequencies. This longitudinal coupling manifests in the form of propagating traveling waves that are thought to contribute to hearing mechanisms (17, 21, 30, 35–40). Genetic modification studies provide further support that the spatial extent of TM waves may play a significant role in cochlear tuning (30, 32). Although these measurements, models, and genetic modification studies have confirmed the importance of TM mechanical properties in hearing, they have not isolated the distinct roles of TM stiffness and viscosity in generating longitudinally propagating traveling waves of the TM.

To understand the contributions of TM material properties on traveling waves better, we developed chemical manipulations to alter the stiffness and viscosity of the TM selectively and reversibly. Because the TM is poroelastic (32, 41), we expect that changes in bath composition can have a direct effect on the mechanical properties of the TM mechanical matrix and its interstitial fluid, which makes up 97% of TM wet weight (42). The addition of PEG has previously been shown to generate an osmotic response that could be accounted for by the permeability of these molecules through the matrix rather than by direct changes to the matrix itself (41). In contrast, changing bath pH has little effect on the osmotic pressure or viscosity of the bath but has been shown to have a direct effect on the macromolecular matrix (43). In this paper, we apply these physicochemical manipulations to alter TM material properties reversibly, and thereby probe their role in controlling longitudinal spread of excitation through the TM.

Results

Chemically Altering TM Stiffness and Viscosity. The sensory receptor cells in the inner ear are mechanically stimulated by shear motions of the overlying TM. To understand the relative contributions of TM shear stiffness and viscosity in determining motion of the TM, we developed chemical manipulations to alter these material properties selectively. Using microfabricated shearing probes, we measured TM shear impedance in three different solutions of artificial endolymph (AE) (Fig. 1A). Materials and Methods, and SI Materials and Methods). Fig. 1B (Left) shows results when the TM is bathed in AE at physiological and reduced pH.

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Effects of Chemical Manipulations on TM Waves. Point impedance measurements (e.g., Fig. 1) characterize local relations between force and displacement. However, motions of adjacent parts of the TM are also coupled, so that motions at one point can generate motions and forces at other points. To characterize the roles of stiffness and viscosity on coupling, we measured wave properties of isolated TMs in AE with reduced pH and added PEG (Materials and Methods). TM segments were excised from the basal turn, suspended between two supports, and immersed in a physiological AE bath (Fig. 2A). Audio frequency vibrations of a piezoelectric crystal attached to one of the supports generated radial vibrations of the TM that propagated longitudinally as traveling waves (e.g., Movie S1). Traveling waves were visualized using motion magnification algorithms (44) and quantified using a previously published computer microvision technique (45) (Materials and Methods and SI Materials and Methods). The magnitude and phase of displacement at each longitudinal position were determined from stroboscopic images at eight phases of the sinusoidal stimulus (Fig. S2). Results across longitudinal distance z (Fig. 2B) were fit to complex exponentials of the form $A e^{i(\omega t - \lambda z)/k}$ to determine wavelength $\lambda$, wave speed $V = f \lambda$ (Fig. 2C), and wave decay constant $\sigma$ (Fig. 2D), where $f$ represents frequency. Fig. 2B shows snapshots of representative TM waves under physiological conditions, in a bath with reduced pH, and in AE with PEG [15 mM and 8 kDa, which increases bath viscosity by 8.9-fold (46)].

Fig. 1B (Right) shows results when the TM is bathed in AE with and without the addition of PEG. The frequency dependence of TM impedance in AE with reduced pH (Fig. 1C, Left; $n = 4$ preparations) and with added PEG (Fig. 1C, Right; $n = 4$ preparations) provides insight into the changes in mechanical properties induced by chemical manipulations. Decreasing bath pH had little effect on the real (lossy) component of shear impedance but significantly reduced the imaginary (stiffness) component by $\sim 2.2$-fold of shear impedance across frequencies. In contrast to pH, adding PEG significantly increased the real component of TM shear impedance by $\sim 4.4$-fold and had little effect on the imaginary component of TM shear impedance. The effects of reducing pH and adding PEG to the bath surrounding the TM were largely reversed upon reequilibration to physiological AE (Fig. S1). These results show that effects of pH and PEG on TM viscoelastic properties are complementary, and provide useful tools for probing TM wave properties.

Viscoelastic Model of Traveling Waves and Material Properties of the TM. We observed a simple relationship between the chemical manipulations applied to the TM and impedance measurements: (i) Adding PEG to the bath increased the real part of the point impedance with little change in the imaginary part, suggesting that the effect was predominantly viscous, and (ii) lowering bath pH decreased the imaginary part with little change in the real part, suggesting that the effect was predominantly stiffness. However, both chemical manipulations altered both the speed and decay of TM waves. To understand the relation between these results, we analyzed an analytical model of viscoelastic gels. In this model, TM waves are represented by shearing motions of a semi-infinite viscoelastic gel. This analytical model has the advantage that wave motions can be described by two simple equations (47):

\[
\sigma = \frac{2(G'' + \omega^2 \eta^2)}{\rho \omega^3 \left( \sqrt{G'^2 + \omega^2 \eta^2} - G' \right)} \tag{1}
\]

\[
\nu = \frac{2(G'' + \omega^2 \eta^2)}{\rho \left( \sqrt{G'^2 + \omega^2 \eta^2} + G' \right)} \tag{2}
\]

where $\rho$ is the density of water, $G'$ is the shear storage modulus of the TM, $\eta$ is the shear viscosity of the TM, and $\omega$ is the angular frequency.

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We used this model to compute TM material properties from TM wave measurements (Fig. 3). Results were similar to results for the shearing probe (Fig. 1): Reducing bath pH tends to reduce shear storage modulus (with little change in shear viscosity), and adding PEG to the bath tends to increase shear viscosity of the TM (with little change in storage modulus). Although these results show that TM wave properties can be understood by thinking about the TM as a viscoelastic solid, they give little insight into the relatively complex dependence of TM wave properties on TM material properties. To understand this dependence better, we computed contour plots to relate wave and material properties at low (1 kHz) and high (18 kHz) frequencies (Fig. 4).

At low frequencies, wave speed depends almost entirely on shear storage modulus $G'$ and very little on shear viscosity $\eta$ (Fig. 4B). At high frequencies, the reverse is true: wave speed is affected very little by $G'$ and significantly by $\eta$ (Fig. 4C). In both low- and high-frequency cases, the wave decay constant depends on both $G'$ and $\eta$. To illustrate the use of these contour plots to understand our measurements, we have plotted the mean and SDs of the wave and material properties as ellipses (Fig. 4C). Reducing pH decreases shear storage modulus with little change to shear viscosity, which reduces the wave decay constant but has little effect on wave speed. By contrast, adding PEG increases shear viscosity with little change to shear storage modulus, which decreases wave decay constant and increases wave speed. These results highlight the complex dependence of TM wave parameters on TM material properties.

To investigate effects of cochlear loads on TM waves, we calculated the relation between wave parameters and TM material properties before and after the addition of springs to represent the mechanical loading of hair cells and dashpots to represent fluid loading in the subtectorial space (Fig. S3). Adding stiffnesses on the order of the measured stiffness of hair bundles (48) and adding damping that would result from a 2-μm subtectorial gap change wave speed and decay constants by less than ~10%.
Although it is widely accepted that the TM is a viscoelastic structure (13, 16, 32, 49), the relative contributions of viscosity and stiffness to TM dynamics have not previously been examined systematically. In this study, we have developed a method to modulate TM dynamic properties by altering the pH and viscosity of the bath surrounding the TM. Results (Fig. 4) show that decreasing pH caused the shear storage modulus of the TM to decrease by nearly a factor of 2 (from 49.8 ± 14.5 to 22.1 ± 11.2 kPa, mean ± SD; range: 17–19 kHz) with little change in shear viscosity (from 0.21 ± 0.040 to 0.27 ± 0.044 Pa·s). In contrast, increasing bath viscosity increased the loss modulus of the TM by nearly a factor of 3 (from 0.19 ± 0.07 to 0.51 ± 0.13 Pa·s) with little change in shear storage modulus (from 34.9 ± 14.0 to 28.4 ± 14.0 kPa). Furthermore, both of these manipulations are almost completely reversible (Fig. S1). Thus, we can alter the chemical composition of the bath to alter TM dynamic material properties selectively and reversibly.

**Frequency-Dependent Effects of Stiffness and Viscosity on TM Waves.** By chemically modulating TM material properties, we demonstrated that changes in material properties have a profound effect on wave speeds and decay constants. In particular, we showed that at high frequencies, reducing TM stiffness primarily results in a decrease in TM wave decay constant, whereas increasing TM viscosity results in an increase in TM wave speed and a decrease in TM wave decay constant (Fig. 2). These measurements fit our model predictions of shear viscosity and shear storage modulus on wave behavior (Fig. 4). The viscoelastic contour plots at low frequencies (Fig. 4B) reveal that wave speed is controlled almost exclusively by shear storage modulus at physiological conditions. In contrast, at high frequencies, speed is primarily determined by shear viscosity (Fig. 4C). This result is counterintuitive. Typically, the storage element in a transmission line controls the speed; here, we see that the loss element plays a more important role in determining wave speed at high frequencies. Thus, at high frequencies, viscous coupling of the TM must be precisely maintained to allow the TM wave speeds to match the wave speeds of the BM wave for effective cochlear amplification (50, 51).

**TM Stiffness and Viscosity Are Both Essential for Maintaining Longitudinal Spread of Mechanical Excitation.** The viscoelastic contour plots (Fig. 4 B and C) reveal that both TM shear viscosity and shear storage modulus play significant roles in controlling TM wave decay constants at all frequencies. Small changes from the physiological values of either \( G' \) or \( \eta \) cause nearly proportional changes in wave decay constants. Changes to both TM shear storage modulus and shear viscosity alter longitudinal coupling at all frequencies, and thereby change longitudinal spread of mechanical excitation through TM waves. These wave decay constant contour plots thus provide a basis for evaluating alterations to TM dynamics, and their effects on spread of mechanical excitation in the cochlea.

**Effects of Cochlear Attachments.** In this study, we investigated response properties of TMs that were isolated from their normal cochlear attachments. In vivo, we expect cochlear attachments to stimulate motions of the TM and present loads that could alter wave propagation. We investigated these possibilities with a model in which the TM was represented as a distributed series of masses coupled by viscous and elastic elements, the BM was represented by an underlying parallel plate, hair bundles were represented as discrete springs, and subtectorial fluid was represented as Couette flow (Fig. S4A). In the absence of BM motion, the hair bundles and subtectorial fluid had little effect on properties of TM waves (Fig. S3 B–D).

In addition to generating loads, the subtectorial fluid and hair bundles couple the BM and TM, thereby providing a means by which the BM can launch TM waves. Oscillating a portion of the BM with constant radial velocity launched waves on the TM (Fig. S4) with similar decay constants to those decay constants observed in Fig. 2 for the isolated preparation at physiological conditions (Fig. S4B), reduced stiffness (Fig. S4C), and increased viscosity (Fig. S4D). These results show that the impedance of a portion of the TM is comparable to impedance of the hair bundles and subtectorial space in that portion. In a sense, the impedance of the TM is “matched” to the impedance of the structures that couple the TM to the organ of Corti (i.e., the impedance of the TM is small enough to be stimulated by subjacent structures but large enough to resist the decay of traveling waves).

**Implications for Cochlear Tuning Mechanisms.** Previous cochlear models have represented the TM as a resonant structure that plays an important role in cochlear tuning (25–29). However, the presence of TM waves demonstrates substantial longitudinal coupling, which has an impact on response properties at each longitudinal location. Consider a simple model of the cochlea that consists of a bank of resonators tuned for higher frequencies at the base and progressively lower frequencies near the apex. In such a model, longitudinal coupling would tend to increase the
sensitivity of each resonator to neighboring best frequencies, and thereby broaden the apparent tuning of each resonator. The TM wave decay constant provides a measure of the distance over which TM coupling is significant. This distance spans some range of best frequencies as given by the cochlear map, and this range of best frequencies determines an effective quality of tuning $Q$ (Fig. 5A). For physiological bath conditions, the $Q_{\lambda_{\text{out}}}$ predicted from TM wave decay constants closely matches measurements from neural recordings (52).

Because wave decay constants depend on stiffness (Fig. 4) and the $Q_{\lambda_{\text{out}}}$ depends on wave decay constants (Fig. 5A), it follows that $Q_{\lambda_{\text{out}}}$ will depend on stiffness as well. Estimates of $Q_{\lambda_{\text{out}}}$ as a function of shear storage modulus show that increasing TM stiffness broadens cochlear tuning (Fig. 5B, Left). Strikingly, the opposite trend is predicted in resonant models of the TM, where increasing TM stiffness would give rise to sharper cochlear tuning.

TM wave decay constants also depend on shear viscosity, which suggests that in addition to stiffness, $Q_{\lambda_{\text{out}}}$ would depend on TM shear viscosity. Estimates of $Q_{\lambda_{\text{out}}}$ as a function of TM shear viscosity show that increasing shear viscosity sharpens cochlear tuning (Fig. 5B, Right). This finding further contradicts resonant models, in which increasing viscous damping would tend to broaden cochlear tuning. Thus, the effects of stiffness and viscosity in controlling tuning via TM waves are opposite in direction and fundamentally different from resonant models of the TM.

**Materials and Methods**

**Isolated TM Preparations.** The cochleae of adult mice (strains 129SvEv/C57Bl/6J, B6129F1, and CD-1; 4–8 wk old) were excised using a previously published surgical technique (53). No strain-dependent differences were found. The organ of Corti and TM were exposed by chipping away the bony enclosure of the cochlea. We used transmitted light illumination to visualize the TM spiraling around the cochlear turns. Using a sterilized eyelash, segments of the TM were teased apart from the basal turn of the cochlea. These TM segments were kept in an AE solution containing 174 mM KCl, 5 mM Hepes, 3 mM dextrose, 2 mM NaCl, and 0.02 mM CaCl2 (equilibrated at pH 7.3 at room temperature). The care and use of animals in this study were approved by the Massachusetts Institute of Technology Committee on Animal Care.

**Measuring TM Point Impedance.** Radial TM shear impedance was measured using microfabricated probes (18). The probe design consisted of a base structure that was driven by a piezoactuator, a 30 × 30-μm shearing plate, and flexible arms that connected the base structure to the probe. To measure TM point impedances, basal TM samples were first immersed in AE and adhered to a glass slide using commercially available bioadhesive (Cell-Tak; Collaborative Research). The microfabricated shear plate’s shearing plate was then engaged on the surface of the TM using a micromanipulator (Rucker and Kollos). The relative motion of the shearing plate and the base depends on the relative impedance of the TM and the cantilever arms. The shearing plate was designed to approximate the shear forces exerted by a cluster of hair bundles. To reduce variability caused by radial gradients in stiffness, the shearing plate was centered on Hensen’s stripe. The base of the probe was stimulated in the radial direction using the piezoelectric actuator across a broad range of frequencies (5–35 kHz) with displacements of ~0.5–1 μm. The impedance of the TM was determined by analyzing the relative motions of the TM and probe (SI Materials and Methods, Calculating TM Impedance).

**Measuring TM Wave Properties.** Isolated TM segments were suspended between vibrating and stationary supports in a wave chamber (17, 30). The vibrating support consisted of a piezoelectric actuator (Thorlabs) that delivered oscillatory motions at audio frequencies (10–20 kHz). The stationary support was firmly attached to the underlying glass slide. Both supports were coated with 2 μL of tissue adhesive (Cell-Tak) and perfused with AE. The TM was then injected into the AE bath and carefully attached to the surfaces of the supports in the regions coated with Cell-Tak. TM wave motions were generated by stimulating the piezoelectric actuator coupled to the vibrating supports. These motions launched longitudinally propagating waves in the radial direction. TM motions were then fit with a decaying sinusoid to extract wave motion parameters. These fits had two free parameters: wavelength ($\lambda$; distance the wave travels while going through a full cycle of motion) and wave decay constant ($\sigma$; distance the wave travels before dissipating by a factor of e in amplitude).

**TM Point Impedance and Wave Properties in PEG-Buffered AE.** We added PEG (15 mM of 8-kDa PEG; Sigma–Aldrich) to the AE bath surrounding the TM to alter the shear viscosity of the TM. To ensure equilibration of PEG, the bath (5 mL) was exchanged four times over the course of ~5 min. The final solution was equilibrated for 5 min before TM shear impedance and wave measurements. Once measurements were completed, the bath was reequilibrated to normal AE and wave measurements were repeated.

**TM Point Impedance and Wave Properties in pH 4-Buffered AE.** In a separate set of experiments, we altered the pH of the bath surrounding the TM by exchanging AE equilibrated at pH 7.3 with AE at pH 4. To ensure equilibration, the bath surrounding the TM was perfused continuously for 5 min and monitored for changes in pH levels. TM shear impedance and wave measurements were completed under physiological conditions (pH 7.3) and at pH 4. Each test bath was perfused twice to test for repeatability.

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**Fig. 5.** Relationship between TM material properties and tuning. (A) TM wave decay constants, in conjunction with the slope of a place-frequency map, are used to calculate quality of tuning. The solid black line represents the relation between best place and best frequency (S4). Horizontal lines and single-sided arrows denote the spatial extent of TM waves (i.e., wave decay constants) for TMs at physiological conditions (mean of 17–19 kHz, blue, 249 μm), with increased viscosity (orange, 146 μm), and reduced pH (red, 118 μm). Vertical dashed lines and single-sided arrows denote the frequency bandwidth around the best frequency. The ratio of frequency bandwidth and best frequency yields $Q_{\lambda_{\text{out}}}$ in physiological conditions (~7), with increased bath viscosity (~12), and with reduced bath pH (~15). (B) Tuning quality factor ($Q_{\lambda_{\text{out}}}$) estimates determined from the viscoelastic model as a function of shear storage modulus and shear viscosity at 18 kHz. Open circles indicate $Q_{\lambda_{\text{out}}}$ predicted from experimental conditions (mean of 17–19 kHz). Right $Q_{\lambda_{\text{out}}}$ estimates increase with increasing shear viscosity, while the shear storage modulus is fixed (40 KPa). Left In contrast, $Q_{\lambda_{\text{out}}}$ estimates decrease with increasing shear storage modulus, G′, while shear viscosity is fixed (0.20 Pa s).


