Particulates from hydrophilic-coated guiding sheaths embolise to the brain

The MIT Faculty has made this article openly available. Please share how this access benefits you. Your story matters.
Particulates from Hydrophilic-Coated Guiding Sheaths Embolize to the Brain

James R.L. Stanley, DVM, MS*, Abraham R. Tzafriri, PhD†, Kathryn Regan, HT(ASCP)CM*, Alan LaRochelle, BS*, Gee Wong, BS, HT*, Brett G. Zani, PhD†, Peter M. Markham, MS*, Lynn Bailey, AS, LATg*, Anna Spognardi, BA†, Gregory A. Kopia, PhD†, and Elazer R. Edelman, MD, PhD‡

*CBSET, Inc. 500 Shire Way, Lexington, MA – 02421
†Kopia Consulting, 61-4D Taurus Drive, Hillsborough, NJ 08844
‡Institute for Medical and Engineering Science, Massachusetts Institute of Technology, E25, 45 Carleton St., Cambridge, MA, 02139, and Department of Medicine, Brigham and Women’s Hospital and Harvard Medical School Boston, MA 02115

Abstract

Aims—We sought to evaluate the incidence of embolic material in porcine brain following vascular interventions using hydrophilic-coated sheaths.

Methods and results—A new self-expanding stent and delivery system (SDS) were deployed through a hydrophilic-coated (Cook® Flexor Ansel) Guiding Sheath into iliac and/or carotid arteries of 23 anesthetized Yucatan miniswine. Animals were euthanized at 3, 30, 90 and 180 days and brains were removed for histological analysis. In an additional single control animal, the guiding sheath was advanced but no SDS was deployed.

Advancement of the coated guiding sheath with or without the SDS was associated with frequent foreign material in the arterioles of the brain. The embolic material was amorphous, non-refractile, non-crystalline, and non-birefringent and typically lightly basophilic with a slight stippled appearance on hematoxylin and eosin (H&E) stain. Material was observed at all time points involving 54% of all study animals (ie, test and control) and in vitro after incubation in 0.9% saline.

Conclusions—The hydrophilic coating on a clinically used guiding sheath readily avulses and embolizes to the brain during deployment in a porcine model. Further documentation of this effect and monitoring in clinical scenarios is warranted.

Keywords

hydrophilic coating; guiding sheath; porcine brain

Send all correspondence to: Rami Tzafriri, 500 Shire Way, Lexington, MA – 02421, Phone: 781-541-5589, Fax: 781-541-5656, rtzafriri@cbset.org.

DISCLOSURES

No conflicts of interest to disclose.
2. Introduction

Medical catheters are used ubiquitously in diagnostic and therapeutic interventional procedures in large numbers of patients each year. These catheters permit external access to deep body spaces for monitoring, imaging, sampling and delivery of therapeutic drugs such as local thrombolysis, and/or deployment of medical devices, such as endovascular stents, aneurysm coils, ablation catheters, and transcatheter valves. Such catheter-based procedures are generally far less traumatic alternatives to surgical means.

Yet, percutaneous catheter insertion is not without risk. Patients undergoing percutaneous interventional procedures are extensively anticoagulated raising the specter of imbalance between hemorrhage (bleeding) and thrombosis (clotting). As larger catheters are employed for increasingly complex interventions in the periphery and across or within valves and ventricles, the risk of vascular injury rises. Guiding sheaths which have been modified by the addition of a hydrophilic coating on their exterior enhance catheter delivery and advancement. Though in clinical use for more than a decade, there have been reports of complications with these hydrophilic-coated devices. The majority of reported cases of adverse events with hydrophilic-coated catheters involve local inflammation following radial artery catheter insertion (1, 2). Such adverse events have been histologically correlated with intralesional foreign material that appeared filamentous, non-birefringent, and blue-grey, with hematoxylin-eosin (H&E) staining and similar in appearance to hydrophilic coating scraped from the exterior surface of involved devices (1).

However, foreign material adverse clinical events have not been limited to the radial artery. Indeed, non-refractile, non-polarizable, granular basophilic foreign material was found in either biopsy or autopsy samples of tissues including lung, foot and brain (3, 4). Another more recent report has described the finding of foreign material following histopathological analyses of brain sections from three patients in whom fatal ipsilateral intracerebral hemorrhages developed several days after uneventful Pipeline Embolization Device treatment of supraclinoid aneurysms (5). In each case, a basophilic intra-arterial filamentous foreign material was observed in brain autopsy histology sections that not only resembled shavings of the coatings shed during benchtop manipulation of the hydrophilic-coated guiding sheath, but were spectroscopically identical and identified as poly(vinylpyrrolidone) (PVP) (5).

As the above studies were conducted in patients with some underlying vascular pathology with numerous and different interventional devices, additional instructive data could be gained via use of hydrophilic material-coated catheters in experimental animals where both the access devices and the test devices are consistent among the study population. Here we report on a hydrophilic-coated guiding sheath that was associated with foreign material in brain sections of treated animals and material shedding in vitro. To our knowledge this is the first such preclinical report of foreign material findings in animal brains as is the correlation of animal findings with the propensity for in vitro shedding.
3. Methods

Studies were conducted at CBSET, Inc. (Lexington, MA) and adherent to the Guide for the Care and Use of Laboratory Animals (6) under an approved Institutional Animal Care and Use Committee (IACUC) protocol, in compliance with the Animal Welfare Act and the Food and Drug Administration Regulations and their amendments.

In Vivo

Twenty-four adult, healthy Yucatan mini-swine weighing 39.4–45.5 Kg were pretreated with 650 mg aspirin and 300 mg clopidogrel orally to reduce the potential for thrombosis, and thereafter 81 mg aspirin and 75 mg clopidogrel orally each day until euthanized. On the day of surgery, animals were sedated with Telazol (4–6 mg/kg, i.m.) and anesthetized with isoflurane in 100% oxygen. Following intubation and pre-treatment with nifedipine (10 mg sublingually) a 90 cm 6F guiding sheath (Cook Flexor Ansel – Check-Flo Valve. Product Code G29989) with a hydrophilic coating (AQ® hydrophilic coating) was placed in either the right common carotid artery (for iliac stent deployment) or right femoral artery (for carotid artery deployment). Animals were pretreated with heparin (50–200 U/kg) to achieve a target activated clotting time of approximately 200–275 seconds. Under fluoroscopic guidance, a guiding catheter was advanced through the guiding sheath over a guide wire to either the carotid or iliac arteries. Stent and delivery systems (SDS) compatible with a 6 Fr introducer sheath were then advanced through the guiding sheaths to a predetermined arterial site and deployed. Two stents were overlapped in the carotid arteries while deployed singly in the iliac arteries. Stents dimensions (6x60 mm or 7x60 mm) were selected for use based on vessel size. Nitroglycerin (50–200 μg) was administered intra-arterially prior to obtaining angiograms. Following the interventional procedure, the accessed vessel was either ligated or repaired and the surgical incision closed in layers with suture. Animals were observed until recovered and monitored until full recovery from anesthesia. Total dwell time for the guiding sheath ranged from 20 to 25 minutes. Animals were then euthanized post-stenting at 3 days (n=4), 30 days (n=7), 90 days (n=8) and 180 days (n=4). In one animal the preliminary angiographic assessment indicated that the target vessels were inadequate to meet the size criteria for the intended stents. Since this animal received an identical guiding sheath but no stent deployment, it was maintained on study for 30 days to serve as an unstented control.

At the respective study end-points, all animals underwent a complete physical examination by the institutional veterinarian (including evaluation of neuro-muscular systems) followed by angiography, euthanasia, and necropsy. Stented vessels were pressure perfused (100 mmHg) with 10% neutral buffered formalin (NBF) and removed. The entire brain was collected and placed in 10% NBF for histopathologic processing and assessment.

In Vitro

Tip segments of a 9F guiding sheath with a hydrophilic coating (Cook Flexor Ansel AQ® hydrophilic coating; Product Code G49102) were placed in 0.9% saline and serially photographed over 60 minutes. Material was then scraped from the surfaces of the catheter segment and placed into the lumen of a swine carcass aorta (to retain the material during

EuroIntervention. Author manuscript; available in PMC 2016 March 23.
paraffin histologic processing), formalin fixed, embedded, sectioned, stained with H&E and examined microscopically.

**Histopathology**
Brains were fixed in NBF for at least 7 days after which they were embedded in paraffin. Six sections were obtained from each brain of the test animals and 28 sections obtained from the single control animal. Sections were mounted on glass slides, stained with H&E and examined microscopically by a board-certified veterinary pathologist using an Olympus B41 microscope.

**Statistical analysis**
Experimental values are reported descriptively as mean ± standard deviation. Categorical data based on intravascular access point (Carotid vs. Femoral) and the evidence of brain emboli (“Yes” when present, “No” when not) were pooled from all examined brain sections and statistically compared using a Fisher Exact Test (SigmaPlot 11.0).

4. Results

**A. Brain Histopathology**
Of the 24 animals treated with the hydrophilic-coated guiding sheath, the brains of 13 (54%) exhibited evidence of an acellular intra-arterial foreign material (Figure 1), which, collectively, involved all major regions of the brain (i.e., forebrain, midbrain, hindbrain, brain stem and cerebellum). With H&E stain, the material was amorphous (i.e., smeared and conforming), non-refractile, non-crystalline, non-birefringent and typically lightly basophilic, and occasionally eosinophilic, with a slight stippled appearance. Frequently associated with the foreign material was the presence of variable chronic inflammatory infiltrates (i.e., lymphohistiocytic and/or multinucleated giant cells [MNGCs]), regardless of time point, which were minimal in magnitude and typically occurred around the arterioles containing the foreign material, and less frequently observed in the pia mater, choroid plexus, and/or neuropil. There was no evidence of acute inflammation (e.g., neutrophils, suppuration) and no trends related time. Evidence of any significant adverse effects (e.g., infarction) was limited to a focus of resolving malacia in the brain of one treated animal (Figure 2). Similar to affected test animals, intra-arterial foreign material, and/or associated minimal lymphohistiocytic and multinucleated giant cells, was also observed in the control animal which underwent preliminary angiography with access using the hydrophilic-coated guiding sheath but did not receive any stents (Figure 3).

**B. Incidence and Brain Distribution**
Of the 24 animals exposed to the guiding sheath, arteriolar foreign material was observed in brain sections from 1 of 5 (20%) animals accessed via the right femoral artery while 12 of 19 (63%) animals accessed via the right common carotid artery demonstrated brain foreign material. However, the trend for higher incidence of brain emboli with carotid versus femoral access was not statistically significant (P=0.122, Fisher Exact Test).
Foreign material was observed in the brains of animals from all time points (Table 1). Fifty-four percent of all study animals (i.e., all test and one non-stented control) had at least one fragment of foreign material observed in each brain section. Of the 138 histologic sections of brain tissue examined from all SDS animals, representing major regions of the brain (n=6 per animal), 15 (11%) contained fragments of foreign material. By contrast, of the 28 histologic sections of brain tissue examined from the control animal (exposed only to hydrophilic-coated sheath), 9 (32%) contained foreign material. When all animals are combined across treatments and time points (n=24), the greatest incidence of foreign material was observed in the cerebellum with a lesser incidence in the forebrain, right midbrain, hindbrain and brain stem with the least incidence observed in the left midbrain (Figure 4).

C. In vitro Histopathology

To better assess the nature of the foreign material, a distal portion of a hydrophilic-coated guiding sheath was placed in 0.9% saline for 60 minutes. Avulsion of the hydrophilic coating from the sheath was noted when incubated in saline for as little as 15 minutes. Over time, the material (presumed hydrophilic coating) was observed to have increasingly separated/shed from the surface of the catheter. When samples of this material were scrapped off the 60 minute-incubation sheath sample (Figure 5) and placed in a cadaver porcine aorta, the histomorphologic appearance of the material was interpreted to be consistent with that observed in the porcine brain sections (Figure 6).

D. Physical Examination

Despite the embolization of the hydrophilic coating, none of the affected animals exhibited abnormal clinical signs indicative of central neurologic impairment (e.g., mentation, craniofacial deficits, proprioceptive deficits, and ataxia) during the conduct of the study and had no remarkable findings upon physical examination by the institutional lab animal veterinarian at their respective terminal time points.

5. Discussion

To our knowledge, this is the first animal study to demonstrate the presence of embolic foreign material in the brain following the use of a hydrophilic material-coated guiding sheath. While the overall purpose of the present studies was to examine the safety of test therapeutic devices, the use of the accessory guiding sheaths in a controlled and consistent environment, with preset durations and follow up times, allowed us to make relevant in vivo observations of the potential clinically negative impact of these devices and to correlate them with in vitro experiments.

The observations from the porcine histologic brain sections and the in vitro assessments strongly suggest that the non-biologic material observed in porcine brains originated from the guiding sheath, in particular, shed and embolized pieces of its hydrophilic PVP coating (AQ®) as previously described (5). Foreign body emboli observed in the control animal in which only the guiding sheath was interventionally deployed (i.e., animal was never exposed to the test or control delivery systems/stents) appeared morphologically and tinctorially
consistent with emboli that were observed in stented animals. Separated/shed material obtained in vitro directly from the surface of a representative segment of the guiding sheath appeared morphologically and tinctorially consistent with that observed in study animals (stented or unstented) and reported in clinical and preclinical publications (4,5,7,8). In addition, the potential of emboli originating from the SDS is generally precluded as these lacked any coating and were largely shielded from blood flow in route to the stenting site (e.g., iliac arteries). Together these observations indicate that the source of the foreign material emboli was the guiding sheath. The trend towards increased incidence of foreign material found among animals accessed via the carotid versus the femoral artery was not statistically significant (P=0.122) and could simply reflect the larger number of animals accessed via carotid only (n=18) versus femoral only (n=3, two other animals were accessed by both femoral and carotid). However, based on the guiding sheath’s propensity to shed at the vascular access site as indicated in the radial artery inflammation literature (see below), it could reflect the potential for more antegrade shedding of material to the brain following carotid access which would not be possible with femoral access.

Despite a relatively low per plane incidence of foreign material in the histologic brain sections of test animals, the overall incidence of foreign material in test animals over the 180 day study duration was high (52%). The low per plane incidence in the histologic brain sections of test animals was interpreted to likely reflect the low number of sections examined per test brain (n=6/brain) as increased regional and histologic sampling of the brain, as performed in the one unstented control animal (n=28/brain), was associated with an increase in foreign material incidence. The generally benign inflammatory response observed with the foreign material in these animals, the rare incidence of significant adverse effects (e.g., infarction) and the clinically silent nature is consistent with what has been previously reported (5).

A review of the literature has provided many examples of hydrophilic material-coated catheters associated with percutaneous intervention-associated adverse events. Most numerous are clinical reports of post-intervention radial artery inflammation unassociated with bacterial infection (1,2,9,10,11,12). Adverse event reports obtained from a search of the FDA Manufacturer and User Facility Device Experience (MAUDE) database demonstrated 174 cases of radial artery inflammation over a period of 10 years (2). All were associated with the same or similar type of PVP coated Cook sheaths examined herein. Hydrophilic coating material has also been identified histologically in heart (7,8,13,14), lung (4,15), brain (1,3,4,5), kidney (8), AV graft associated (8), and foot (4) following percutaneous access with a catheter having a hydrophilic coating. The histomorphologic and tinctorial appearance of the embolic material reported for the hydrophilic PVP coating in the literature both ex vivo (1,3,4,7) and in patient autopsy or biopsy samples (1,3,8,13,14) was consistent with that observed in the brain sections of stented or unstented swine in the current study (Figures 1–4). The reporting of fewer adverse events with hydrophilic coated catheters that do not employ PVP (4,11,16) suggests that the risk for coating embolization can be minimized through the use of less abrasive coatings and highlights the urgent need for further evaluating this class of devices in preclinical animal and benchtop models.
Study limitations

Caution is warranted in translating findings to humans as the arteries in young, healthy swine may be different from the arteries of patients in respect to mechano-anatomic properties which could potentiate catheter abrasion and risk for coating embolization (e.g., calcification, compliance, length and tortuosity). In addition, the cerebral blood supply of humans is generally interpreted as comparable to that of swine (17) though some differences exist, most notably a vascular meshwork/plexus (i.e., rete mirabile) situated between the carotid arteries and cerebral vessels of swine (18) which could serve to filter hydrophilic coating emboli resulting in a lesser amount of cerebral emboli in swine compared to humans (19). Regardless, the incidence of embolic material observed in our swine model, despite the absence of vascular disease and the presence of an intervening vascular meshwork, suggests prudent observation in comparable clinical scenarios.

Clinical implications

Hydrophilic coatings add to the ease of sheath use and are increasingly used in sheath technology. Yet, there appears to be a price to pay – these materials are not as adherent as more hydrophobic material and can avulse in vitro and embolize in animal models to critical organs like the brain. In the present study, greatest incidence of foreign material was observed with hydrophilic guiding sheaths involved in carotid arterial access. Though generally avoided in the past carotid access is resurging with carotid stenting (20–21) and percutaneous aortic valve placement (22–24). The lack of physical examination sequelae in the pig even in the face of high incidence of emboli in the cerebellum may speak to a legitimate lack of effect from the reserve capacity of the porcine brain to withstand embolic insult, and the marked anatomic differences between pigs and human cerebral vasculature. The cerebral circulation in the pig is far more developed and anastomotic than in the human which may not only reduce the incidence of particles reaching the brain but also their distribution patterns within different brain regions. The concern for marked hypersensitivity in a small subset of hypersensitive individuals further amplifies the potential risks associated with our animal findings. Validation and scrutiny in clinical settings is therefore warranted and especially for neuromuscular and cognitive dysfunction.

6. Conclusions

The histomorphologic similarity of the foreign material found in porcine brain to material taken from a sample guiding sheath and to foreign material reported in the literature indicates that this material originated from the hydrophilic coating on the guiding sheaths used in the present study. Distribution to the brain in experimental animals has not previously been reported but raises another potential clinical concern around the use of this device thus warranting further documentation of effect and monitoring in clinical scenarios.

Acknowledgments

Funding: ERE is supported in part by a grant from the NIH (R01 GM 49039).
References


Figure 1.
Intra-arterial Foreign Material (arrows) in Porcine Brain Sections. Example of intra-arterial amorphous, stippled, basophilic (A) to eosinophilic (B, C, D) foreign material with associated peri-arterial inflammatory cells in sections of 30 day porcine right midbrain (A, B), and 180 day forebrain (C) and cerebellum (D) following deployment of the hydrophilic-coated guiding sheath followed by distal peripheral stenting.
Figure 2.
Foreign Material in a Brain Section from the Control Animal. Intra-arterial accumulation of amorphous, stippled basophilic to eosinophilic material (arrows) from a non-stented control animal that received only a hydrophilic-coated guiding sheath 30 days previously. Note the associated lymphohistiocytic infiltrates (A) and additional multinucleated giant cell infiltrates (B).
Figure 3.
Example of Local Infarction. Note focally extensive area of infarction illustrated by chronic, resolving malacia (within dashed lines, 1.25× image) characterized by fibrous astrocytosis and microglial hemosiderosis (4× image).
Figure 4.
Distribution of Hydrophilic Coating Material. Regional distribution of foreign material in porcine brains following use of guiding sheath. Incidence represents number of animals with material observed in the specified region divided by all animals (n=24).
Figure 5.
Hydrophilic Coating Separation. *In vitro* shedding of the hydrophilic coating from the tip of a 9F guiding sheath placed in 0.9% saline for 60 minutes.
Figure 6.
In Vitro Coating-Derived Foreign Material. Representative image of material removed from surface of a 9F hydrophilic-coated guiding sheath following in vitro submersion in saline as presented in Figure 4 and then placed in a porcine cadaver aorta and processed for histological analysis. Note the amorphous, stippled basophilic material and it’s resemblance to material shown in Figures 1 and 2.
Table 1

Incidence of foreign material observed in porcine brains from animals treated with the Cook guiding sheath.

<table>
<thead>
<tr>
<th>Group</th>
<th>Time Point</th>
<th>Animal Incidence</th>
<th>Planar Incidence</th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Total Animals</td>
<td>Animals with Foreign Material</td>
<td>%</td>
</tr>
<tr>
<td>Cook Medical Guiding Sheath</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Control</td>
<td>Day 30</td>
<td>1</td>
<td>1</td>
<td>100%</td>
</tr>
<tr>
<td>Test</td>
<td>Day 3</td>
<td>4</td>
<td>1</td>
<td>25%</td>
</tr>
<tr>
<td>Test</td>
<td>Day 30</td>
<td>7</td>
<td>4</td>
<td>57%</td>
</tr>
<tr>
<td>Test</td>
<td>Day 90</td>
<td>8</td>
<td>4</td>
<td>50%</td>
</tr>
<tr>
<td>Test</td>
<td>Day 180</td>
<td>4</td>
<td>3</td>
<td>75%</td>
</tr>
<tr>
<td>Total Test Animals</td>
<td></td>
<td>23</td>
<td>12</td>
<td>52%</td>
</tr>
<tr>
<td>All Animals</td>
<td></td>
<td>24</td>
<td>13</td>
<td>54%</td>
</tr>
</tbody>
</table>