Structural colour from helicoidal cell-wall architecture in fruits of Margaritaria nobilis

The MIT Faculty has made this article openly available. Please share how this access benefits you. Your story matters.

Citation

As Published
http://dx.doi.org/10.1098/rsif.2016.0645

Publisher
Royal Society

Version
Final published version

Accessed
Wed Feb 13 08:51:56 EST 2019

Citable Link
http://hdl.handle.net/1721.1/107402

Terms of Use
Creative Commons Attribution 4.0 International License

Detailed Terms
http://creativecommons.org/licenses/by/4.0/
Structural colour from helicoidal cell-wall architecture in fruits of *Margaritaria nobilis*

Silvia Vignolini¹, Thomas Gregory², Mathias Kolle⁴, Alfie Lethbridge³, Edwige Moyroud⁵, Ullrich Steiner⁶, Beverley J. Glover⁵, Peter Vukusic³ and Paula J. Rudall²

¹Chemistry Department, University of Cambridge, Lensfield Road, Cambridge CB2 1EW, UK
²Royal Botanic Gardens Kew, Richmond, Surrey TW9 3AB, UK
³Thin Film Photonics, School of Physics, Exeter University, Exeter EX4 4QL, UK
⁴Massachusetts Institute of Technology, 77 Massachusetts Avenue, Cambridge, MA 02139-4307, USA
⁵Department of Plant Sciences, University of Cambridge, Downing Street, Cambridge CB2 3EA, UK
⁶Adolphe Merkle Institute, Chemin des Verdiers 4, 1700 Fribourg, Switzerland

The bright and intense blue-green coloration of the fruits of *Margaritaria nobilis* (Phyllanthaceae) was investigated using polarization-resolved spectroscopy and transmission electron microscopy. Optical measurements of freshly collected fruits revealed a strong circularly polarized reflection of the fruit that originates from a cellulose helicoidal cell wall structure in the pericarp cells. Hyperspectral microscopy was used to capture the iridescent effect at the single-cell level.

1. Introduction

In some plants, the cell walls of selected tissues exhibit helicoidal architecture, in which multiple adjacent wall layers are composed of aligned cellulose fibrils that rotate along a helical screw [1]. Despite this regular construction, considerable flexibility exists in the dimensions and geometry of the multi-layered structure [2]. In the special case when the dimension of the helicoid, defined by the distance between two planes with closely similar fibril orientation (half of a full 360° rotation, pitch *p*), is comparable to the wavelength of visible light and is constant within the cell wall, these structures are capable of selectively reflecting coloured light that may be polarized. In particular, they reflect circularly polarized light at a wavelength defined by *λ* = *np* (where *n* is the mean refractive index of the medium) and with optical handedness that depends on the handedness of the helicoid [3].

Helicoidal cell-wall architecture has been reported in a broad range of land plants, including mosses, ferns, gymnosperms and angiosperms [2,4], but they are also common in beetle exoskeletons [5]. This apparently complex cell-wall structure occurs in tissues that include thick-walled cells [6], including epidermis, sclerenchyma and xylem and in many different plant organs, including leaves, stems and fruits [1,7–11]. For example, structural colour obtained from helicoidal architecture has been reported in leaves of plants from a range of different habitats [12–15]. However, with a few exceptions (e.g. hazelnut [16], *Pollia* [11,17]), this structure has rarely been studied in fruits and seeds, which often possess thick-walled tissues that are resistant to desiccation. Most fruit colour is produced by pigmentation [18], but a few plant species produce highly metallic and intensely coloured fruits by means of a nanostructured multi-layered cell wall, including the commelinid monocot *Pollia condensata* [11,17] and the rosid eudicot *Margaritaria nobilis* [19,20].

© 2016 The Authors. Published by the Royal Society under the terms of the Creative Commons Attribution License http://creativecommons.org/licenses/by/4.0/, which permits unrestricted use, provided the original author and source are credited.
In this paper, we use both polarization-resolved spectroscopy and electron microscopy to present a detailed optical analysis of fresh fruits of *M. nobilis* (Phyllanthaceae), a forest tree from tropical Central and South America. In this species, the fruits possess a green exocarp, which splits after they become detached and fall to the forest floor [19,20]. The remaining exposed inner part of the fruit wall exhibits a metallic greenish-blue colour, particularly in humid environments, that is attractive to birds such as jays and doves [19]. These birds consume the fruits and hence act as dispersal agents. The results obtained here demonstrate that the strong intense coloration of *M. nobilis* fruits is due to a helicoidal cellulose structure in the endocarp cell walls. The optical measurements are confirmed by high-resolution electron microscopy of the tissue showing a Bouligand pattern typical of helicoidal architectures [21].

The fruits of *M. nobilis* are only the second example of a plant species that has been conclusively demonstrated to use helicoidal cell-wall architecture to produce structural color. The first example was of the fruits of the commelinid monocot *P. condensata* [11,17]. This is a surprising discovery because of the evolutionary distance separating *Margaritaria* and *Pollia*. The use of a cellulose helicoidal architecture to produce colour has clearly evolved independently and convergently in these two species, which are estimated to have diverged over 100 Ma. This finding suggests that helicoidal structures represent a possible strategy for convergent evolution of structural colour in plants.

2. Material and methods

2.1. Plant material

For optical and microscopic analysis, fresh fruits were collected in Panama under permit SEX/P-59-13 to Dr Edmund Tanner (issued 23 October 2013 by the Direccion de Areas Protegidas y Vida Silvestre). Fruits were refrigerated and then sent directly to Cambridge, UK. For examination, using light microscopy, scanning electron microscopy (SEM) and transmission electron microscopy (TEM), fruits were also obtained from the Royal Botanic Gardens, Kew, either from alcohol-preserved specimens (collected from Brazil by Milliken in 2011) or dried herbarium specimens from two separate collections, the first collected by Spruce in 1855, and the second collected by Belem and Mendes in 1964.

2.2. Microscopy

Optical imaging was performed using a customized Zeiss optical microscope equipped with epi-illumination and a 10× objective. Unpolarized light from a halogen lamp served as illumination for imaging. A polarizer and a quarter-waveplate mounted onto independent motorized rotation stages were selectively inserted into the optical path to perform polarization-resolved imaging.

For SEM imaging, dried fruit material was fractured, mounted on an aluminium stub, coated with platinum using a sputter coater (Quorum Q150T ES) and examined using a Hitachi S-4700 SEM at 2 kV.

For TEM imaging, fruits were cut into small fragments and fixed in 3% phosphate-buffered glutaraldehyde followed by immersion in 1% osmium tetroxide. Fixed samples were taken through a graded ethanol and London resin (LR) medium white resin series prior to embedding in an epoxy resin. Ultrathin sections (30–100 nm) were cut using an ultramicrotome (Reichert-Jung Ultrcut E) and collected on Formvar-coated copper slot grids. Initial results using post-staining with uranyl acetate and lead citrate (as used for fruits of *P. condensata*, [11]) failed to reveal a helicoidal ultrastructure. This could only be resolved when these staining stages were omitted (see electronic supplementary material, figure S1). Samples were imaged using a Hitachi H-7650 TEM equipped with an AMT XR41 digital camera.

2.3. Spectroscopic characterization

Reflectance spectra of the fruit surface were measured on a microspectroscopic scale (spot size: >10 μm) which allowed the collection of optical signals from individual cells. The halogen lamp of the microscope served as light source in bright-field configuration. Light reflected from the sample passed back into the objective and was coupled in confocal configuration with a 100 m core optical fibre connected to a spectrometer (QE65000, Ocean Optics, 200–880 nm). The reflection spectra were normalized with respect to a silver mirror (Thorlabs). Spectra and images were collected using unpolarized illumination and a circularly polarizing filter consisting of a superachromatic quarter waveplate (B. Halle) combined with a linear polarizer (Thorlabs) for right-handed (RH) and left-handed (LH) light detection. The hyperspectral images were collected in the same configuration using an additional liquid crystal filter (CRI, Varispec) that was inserted in front of the CCD imaging chip. Images were collected with a camera and carefully normalized if recorded with different exposure times, considering also the nonlinearity of the camera response.

3. Results

3.1. Fruit anatomy

Each fruit of *M. nobilis* consists of several (four to six) segments, each containing a single seed (figures 1 and 2; electronic supplementary material, figure S2). The entire structure is enclosed in a pericarp that consists of two layers: an outer papery exocarp that dehisces at fruit maturity (clearly visible in figure 1 of [19]) and an endocarp consisting of three or
four layers of thick-walled cells (figure 2a–c). The endocarp is about 1 mm thick, and the average thickness of the cell wall is about 10–15 $\mu$m. When the fruit is fresh or well hydrated, the colour of the remaining fruit is metallic blue or green. Fruits have a more pearly white appearance when they are completely dry (figure 1b).

Transverse sections of fresh fruits (figure 2a,b) show that the blue-green coloration of the fruits comes from the endocarp, which consists of thick-walled cells (figure 2d–g). When the fruit is fresh, the seeds are hydrated and adhere perfectly to the endocarp. In the dry state, the seeds shrink, and the endocarp is separated from the seeds by an air layer that prevents light absorption and therefore decreases the contrast and the saturation of the structural coloration [22].

A schematic drawing in the electronic supplementary material, figure S3, illustrates this effect and describes the mechanism of the scattering induced by the presence of the air layer. The change in macroscopic appearance of the fruit is completely reversible. By leaving the fruit in a closed environment with saturated humidity (such as in a sealed vial containing a wet tissue, not in contact with the fruit) or simply by immersing it in water, the blue coloration reappears as the fruit rehydrates. To further demonstrate that the structural colour is not lost in the dehydrated state, the micrograph in electronic supplementary material, figure S5, reveals that the colour is visible in the pericarp layer alone.

SEM and TEM cross-sectional images show the multi-layered helicoidal architecture of the cell wall structure of the endocarp cells (figure 2). At low magnification, the structure appears as a simple multilayer (figure 2d,f). At higher magnification and resolution (figure 2e,g), a Bouligand arch pattern is visible. The twist of the individual cellulose microfibrils allows to infer their organization in a helicoidal morphology.

While this helicoidal structure is readily visualized by SEM imaging, it could not be resolved by high-resolution TEM imaging of fresh material, but only in the dry state.

3.2. Optical characterization

Figure 3 shows the optical response of a fresh fruit illuminated at different polarization configurations. In particular, figure 3a shows an optical micrograph of the fruit with polarization filters in collection or illumination. In figure 3a, the colour reflected from the cell wall and an additional reflection that originates from the air–fruit interface can be observed. Between cross-polarizers (illuminating with polarizing light and collecting with linear polarization perpendicular to the illumination), only the reflection from the multi-layered structure is collected. The colours remain unchanged, but the image contrast sharpens (figure 3b).

The nature of the multilayer morphology is revealed when placing the sample between circularly polarizing filters.
was not resolved by TEM imaging, possibly as a result of
found inside individual cells [20], but the helical structure
beyond the scope of this work. However, using the extrapolated
measured from the TEM images is tricky and therefore
reflectivity of each cell with the anatomical parameters
a stack of several cells. Performing a detailed correlation of the
than the cell size, because the collected signal typically traverses
indices, in this case, the interface between air and the outer
layer of the endocarp. Using the Fresnel equations for unpolar-
ized illumination (equal reflectance in both polarization
channels), a reflectivity of 2% is predicted for each channel,
assuming a refractive index of the reflecting medium of 1.5.

In order to capture iridescence at the single-cell level,
we investigated the fruit using hyperspectral microscopy.
The shape of the epidermal cells of M. nobilis can be approxi-
mated as cylinders. As observed by Kolle et al. [20], when
illuminating the cells with an objective with numerical aper-
ture NA = 0.45, light is reflected from the different cells in a
range of colours. This arises from the cell’s curved geometry.
This effect, typical of every multilayer structure with ellipsoidal
or cylindrical geometry, reveals the iridescent nature of
the colour, as shown in figure 4. This is however averaged
out when the fruit is illuminated with diffuse light, and the iri-
descence disappears, leaving only an intense ‘metallic’ colour
appearance. See also electronic supplementary material, figure
S5 and the electronic supplementary material, movie S6.

In this configuration, colour is observed only in the left-
handed (LH) circular polarization channel (figure 3c), and
only very little light is collected in the right-handed (RH)
channel, probably scattered from inner cells tilted with
respect to the surface of the fruit (figure 3d). It is interesting
to note that all cells reflect only left-handed circularly polar-
ized light, in contrast with cells of P. condensata, in which both
handednesses were observed [11].

Earlier work on the structural characterization of
M. nobilis described a concentrically layered architecture
found inside individual cells [20], but the helical structure
was not resolved by TEM imaging, possibly as a result of
the staining issue described in §2.2.

Similar to many other examples of structural colour in
nature, different cells reflect slightly different colours, as it
is evident from figure 3c. The measured spectra therefore
differ between imaged areas, even if the collection spot is smaller
than the cell size, because the collected signal typically traverses
a stack of several cells. Performing a detailed correlation of the
reflectivity of each cell with the anatomical parameters
measured from the TEM images is tricky and therefore
beyond the scope of this work. However, using the extrapolated
averaged pitch from TEM and the refractive index of the
cellulose (n = 1.53), a reflection peak in the blue-green region
of the spectrum is predicted, in agreement with (figure 3).

Bright-field spectra taken at the single-cell level using a 20×
magnification objective are shown in figure 3e. In the left polar-
ization channel (red line), several peaks are visible in the
spectral region between 500 and 550 nm. In the opposite chan-
nel, only a wavelength-independent response of about 2% was
recorded. This signal arises from the ubiquitous specular reflec-
tion from the interface of two media with differing refractive
indices, in this case, the interface between air and the outer
layer of the endocarp. Using the Fresnel equations for unpolar-
zized illumination (equal reflectance in both polarization
channels), a reflectivity of 2% is predicted for each channel,
assuming a refractive index of the reflecting medium of 1.5.

In order to capture iridescence at the single-cell level,
we investigated the fruit using hyperspectral microscopy.
The shape of the epidermal cells of M. nobilis can be approxi-
mated as cylinders. As observed by Kolle et al. [20], when
illuminating the cells with an objective with numerical aper-
ture NA = 0.45, light is reflected from the different cells in a
range of colours. This arises from the cell’s curved geometry.
This effect, typical of every multilayer structure with ellipsoidal
or cylindrical geometry, reveals the iridescent nature of
the colour, as shown in figure 4. This is however averaged
out when the fruit is illuminated with diffuse light, and the iri-
descence disappears, leaving only an intense ‘metallic’ colour
appearance. See also electronic supplementary material, figure
S5 and the electronic supplementary material, movie S6.

4. Discussion

Our results demonstrate that the intense blue-green coloration
of the fruits of M. nobilis is a structural effect, resulting from a
helicoidal cellulose structure in the multi-layered cell walls
of the pericarp. The results of our optical measurements are con-
firmed by high-resolution electron microscopy of the tissue
showing Bouligand patterns typical of helicoidal architectures
[21]. The chiral nature of the optical response of the fruit of
M. nobilis resembles that of the fruit of P. condensata, except
that in M. nobilis only left-handed polarization is reflected,
whereas both LH and RH circular polarization are detected in

These two species are relatively distantly related among
flowering plants: P. condensata is a commelinid monocot
and M. nobilis is a rosid eudicot. Therefore, the detailed heli-
coidal cellulose structure in the fruits of these two species is
clearly an example of convergent evolution of metallic fruit
colour. Both species produce fruits lacking soft tissues, and
therefore offer little nutritional reward to potential seed dis-
persers [11,19]. Although the diversity and evolution of
fruit colour remains imperfectly understood [18], some
studies suggest that brightly coloured non-nutritious fruits
are likely to be mimetic, where the plant deceives potential
dispersers such as birds by mimicking the colour of other
species with fleshy nutritious fruits that grow in the same
habitat [19]. This form of mimicry may allow efficient seed
dispersal without the energetic cost of providing a food
reward to the disperser.

Interestingly, a related example of helicoidal architecture
facilitating seed dispersal occurs in some plant species with
muclaginous seed coats that adhere to passing animals. For
example, in the seed coat of quince, the outer cell layers
possess helicoidal thickenings that produce a slime consisting of scattered microfibrils that result from unravelling helicoidal arrays [23].

5. Conclusion

Our study provides a detailed correlation between the anatomy of the fruit of *M. nobilis* and its optical response. Our results demonstrate that, as in the case of *P. condensata* [11], the intense blue-green coloration of this fruit is a structural effect resulting from a helicoidal cellulose structure in the multi-layered cell walls of the pericarp. This helicoidal architecture is common, and interestingly, a related example of helicoidal architecture facilitating seed dispersal occurs in some plant species with mucilaginous seed coats that adhere to passing animals. Future studies on the internal geometry of cell walls in a diverse range of plant tissues could provide further clues concerning the construction and properties of this highly ordered and multifunctional cell-wall architecture. Even though the development of such structures in nature is not yet fully understood, material scientists have been inspired by such bright colour appearance and bioinspired photonic fibres [20] and films [24] have been produced using different strategies.

Data accessibility. Relevant experimental data are available online in the supplementary material.

Competing interests. We declare we have no competing interests.

Funding. This work was supported by the Leverhulme Trust (F/09-741/G) and a BBSRC David Phillips fellowship (BB/K014617/1). P.V. acknowledges support from the US Air Force Office of Scientific Research under award number FA9550-10-1-0020. U.S. acknowledges support from the Adolph Merkle foundation and the Swiss National Science Foundation through the National Centre of Competence in Research Bio-Inspired Materials.

Acknowledgements. We thank Prof. Richard Bateman, Prof. Jeremy Baumberg and Dr Bodo Wilts for useful discussions. We are grateful to Dr Edmund Tanner and Dr Chadtip Rodtassana (Department of Plant Sciences, University of Cambridge) for help with the collection of fresh fruits of *M. nobilis*. All the research data supporting the publication are included in the publication and in the supplementary material.

References


