Dynamic association of the replication initiator and transcription factor DnaA with the Bacillus subtilis chromosome during replication stress

The MIT Faculty has made this article openly available. Please share how this access benefits you. Your story matters.

<table>
<thead>
<tr>
<th>Citation</th>
<th>Breier, Adam M., Grossman, Alan D. Dynamic Association of the Replication Initiator and Transcription Factor DnaA with the Bacillus subtilis Chromosome during Replication Stress J. Bacteriol. 2009 191: 486-493</th>
</tr>
</thead>
<tbody>
<tr>
<td>As Published</td>
<td><a href="http://dx.doi.org/10.1128/jb.01294-08">http://dx.doi.org/10.1128/jb.01294-08</a></td>
</tr>
<tr>
<td>Publisher</td>
<td>American Society for Microbiology</td>
</tr>
<tr>
<td>Version</td>
<td>Author's final manuscript</td>
</tr>
<tr>
<td>Citable Link</td>
<td><a href="http://hdl.handle.net/1721.1/57484">http://hdl.handle.net/1721.1/57484</a></td>
</tr>
<tr>
<td>Terms of Use</td>
<td>Article is made available in accordance with the publisher's policy and may be subject to US copyright law. Please refer to the publisher's site for terms of use.</td>
</tr>
<tr>
<td>Detailed Terms</td>
<td></td>
</tr>
</tbody>
</table>

Dynamic association of the replication initiator and transcription factor DnaA with the \textit{Bacillus subtilis} chromosome during replication stress

Adam M. Breier and Alan D. Grossman*

Department of Biology

Massachusetts Institute of Technology

Cambridge, MA 02139

running title: Dynamics of \textit{B. subtilis} DnaA binding to DNA in vivo

Key words: \textit{Bacillus subtilis}; DnaA; replication stress; transcription

*Corresponding author:
Department of Biology
Building 68-530
Massachusetts Institute of Technology
Cambridge, MA 02139
phone: 617-253-1515
fax: 617-253-2643
E-mail: \texttt{adg@mit.edu}
Abstract

DnaA functions as both a transcription factor and the replication initiator in bacteria. We characterized the DNA binding dynamics of DnaA on a genomic level. Based on crosslinking and immunoprecipitation (ChIP) data, DnaA binds at least 17 loci, 15 of which are regulated transcriptionally in response to inhibition of replication (replication stress). Six loci, each of which has a cluster of at least 9 potential DnaA binding sites, had significant increases in binding by DnaA when replication was inhibited, indicating that the association of DnaA with at least some of its target sites is altered after replication stress. When replication resumed from oriC after inhibition of replication initiation, these high levels of binding decreased rapidly at origin-proximal and origin-distal regions, well before a replication fork could pass through each of the regulated regions. These findings indicate that there is rapid signaling to decrease activation of DnaA during replication and that interaction between DnaA bound at each site and the replication machinery is not required for regulation of DnaA activity in response to replication stress.
Introduction

Cells use multiple mechanisms to detect and respond to perturbations in replication. In bacteria, the well-characterized RecA-dependent SOS response affects expression of many genes after disruptions in replication elongation and DNA damage (13). Additionally, there are recA-independent mechanisms that affect gene expression in response to alterations in replication. A significant part of the recA-independent response to perturbations in replication (replication stress) appears to be mediated directly by DnaA (15).

DnaA is widely conserved in bacteria and is best known as the replication initiator [reviewed in (23, 33, 40)]. It binds to a 9 bp site that appears multiple times in the origin of replication in bacterial chromosomes. Binding sites are also found upstream of many genes and throughout the chromosome. At replication origins, DnaA mediates melting of DNA (open complex formation) and helps to recruit proteins required for replication. DnaA is a member of the AAA+ family of ATPases and the ATP-bound form (DnaA-ATP) is required for replication initiation (36).

Newly synthesized DnaA is predominantly in the ATP-bound form and nucleotide hydrolysis converts DnaA-ATP to the inactive DnaA-ADP after replication initiation. In E. coli, the levels of DnaA-ATP and DnaA-ADP are linked to replication status and there can be high levels of DnaA-ATP in cells that are unable to replicate (27).

DnaA is also a transcription factor. DnaA activates or represses transcription of various genes, including its own (1, 3, 26, 34, 39). It is thought that, as is the case for replication initiation, DnaA-ATP is the active form for transcriptional regulation [reviewed in (23, 37, 40)], and its ability to function as a transcription factor appears to increase when replication initiation or elongation is inhibited (14, 15). Thus, accumulation of DnaA-ATP during replication stress
may result in activation of the transcriptional regulatory activity of DnaA as well as its initiator activity.

In *Bacillus subtilis*, DnaA appears to control expression of >50 genes in ~20 operons in response to perturbations in replication (15). Transcription of these operons changes in response to inhibition of either replication elongation or replication initiation, independently of the well-characterized RecA-mediated SOS response. Furthermore, these operons contain putative DnaA binding sites in their regulatory regions and DnaA appears to associate with at least six of these in vivo based on results from chromatin immunoprecipitation (ChIP) or affinity purification (ChAP-chip) experiments (15, 20). Alterations in DnaA levels during exponential growth also affect expression of several genes and a genome-wide analysis of DnaA binding to chromosomal DNA demonstrated strong binding to eight intergenic regions, five of which appear to regulate gene expression (20).

In this study, we used ChIP-chip and ChIP-PCR to monitor binding of *B. subtilis* DnaA to DNA in vivo during exponential growth and after inhibition of replication. We detected association of DnaA with at least 17 chromosomal regions, 15 of which correspond to operons whose expression responds to perturbations in DNA replication. Following inhibition of replication elongation or initiation, DnaA binding increased significantly at 6 chromosomal regions each of which contains at least 9 potential DnaA binding sites. We found that passage of a replication fork through a regulatory region was not required for release of bound DnaA from that region following the resumption of replication. Our results support the notion that DnaA regulates a global response to perturbations in replication and indicate that alterations in DnaA binding to target promoters contributes to the changes in gene expression following inhibition of replication.
Materials and Methods

Media and growth conditions

For all experiments, cells were grown in S750 defined minimal medium (18, 21) with 0.1% glutamate and supplemented with required amino acids (typically trp and phe). Glucose (1%) was the carbon source except for strains bearing PxyldnaN, for which arabinose (1%, together with xylose at 0.1% as an inducer) was used. Cells were typically grown at 32°C and shifted to 47°C for temperature sensitive mutants.

Strains and alleles

Strains used in this study were the lab wild type, AG174 (trp, phe; a.k.a., JH642); KPL69 \{dnaB134ts- zb8-3::Tn917, trp, phe (29, 30, 38)\}; KPL73 \{dnaD23ts chr::Tn917ΩHU151(mls), trp, phe (29)\}; and AIG200 \{Δ(dnaA-oriC-dnaN)::spc, amyE::(PxyldnaN, cat), spoIIIJ::(oriN, repN, kan), phe, trp\+ (15)\}.

dnaD23ts and dnaB134ts are temperature sensitive mutations that prevent initiation of replication at nonpermissive temperature (7, 8, 10, 24, 30).

dnaA is normally essential, but it can be deleted in strains capable of initiating chromosomal replication from a heterologous origin (2, 19, 22, 32). Therefore, in strain AIG200, the plasmid origin of replication, oriN, along with the plasmid initiator gene repN, was integrated into the chromosome at spoIIIJ, near oriC. This strain also contains a deletion-insertion that removes oriC and the dnaA-dnaN operon, and inserts spc. dnaN, the essential gene encoding the β-processivity clamp of DNA polymerase that is normally in an operon with dnaA, is expressed at a heterologous locus from the xylose-inducible promoter PxyI (PxyldnaN) (2, 15).
Chromatin immunoprecipitation

Chromatin immunoprecipitation of DNA bound to DnaA was done essentially as described (15), with minor modifications. Briefly, protein and DNA were crosslinked with formaldehyde, lysed and sheared with 1 mg/ml lysozyme and sonication, and immunoprecipitated with chicken anti-DnaA primary antibodies, donkey anti-chicken secondary antibodies, and protein A-sepharose beads. Samples were washed and crosslinks reversed and DNA eluted followed by further washing of the beads with 50 mM Tris pH 8, 10 mM EDTA, 0.8% SDS for 5 min at 37°C. The eluate and wash were combined, treated with proteinase K (0.1 mg/ml, 30 minutes, 37°C), extracted with phenol-chloroform, precipitated with ethanol, and resuspended in 15 µl ddH₂O. Samples of total DNA were taken prior to incubation with primary antibodies and treated with proteinase K (0.1 mg/ml, 30 minutes, 37°C) in the presence of 0.8% SDS, then subjected to crosslink reversal, phenol-chloroform extraction and ethanol precipitation in parallel with the immunoprecipitates. There were at least 3 biological replicates for each experiment.

ChIP-PCR

For ChIP-PCR, samples were diluted (typically between 1:10 and 1:150) to give an appropriate level of signal. PCRs were done for 28 cycles and products were run on 1.5% agarose gels containing ethidium-bromide and analyzed and quantified with AlphaEaseFC software v4.0 (Alpha Innotech). The locus yabM was used as a negative control site for background reference. Enrichment at this locus was at background levels in ChIP-chip data (Fig. 1) and it behaved similarly in ChIP-PCR experiments to other loci where DnaA was not expected to bind (not shown). Despite not having specific binding by DnaA, there was a background signal and this signal was used for normalization. Enrichment was calculated as a ratio of ratios in order to normalize to the background level, as represented by \( \frac{IP}{Total} \).
(IP<sub>yabM</sub>/Total<sub>yabM</sub>) where IP<sub>i</sub> and Total<sub>i</sub> represent the dilution-adjusted band intensities at the locus of interest.

**ChIP-chip**

To generate sufficient signal in ChIP-chip experiments, we typically took samples of 100 ml of mid-exponential phase culture (grown in S<sub>750</sub> defined minimal medium). DNA samples were prepared for microarray analysis by labeling samples of the resuspended immunoprecipitated DNA or total DNA with aminoallyl-dUTP using 13 U Sequenase (USB Corp.) and 5 µg random nonamer as primers. There was not an amplification step. The labeled samples were purified and conjugated to Cy3 or Cy5. We used DNA microarrays containing PCR products representing >95% of the open reading frames in the *B. subtilis* genome and nearly all of the 295 intergenic regions that are >364 bp in length, as described (4, 5, 16). Microarray hybridization and data acquisition were performed as described (4).

Enrichment of DnaA at each genomic locus represented on the microarrays was calculated as the relative amount of DNA in the immunoprecipitate divided by the amount of total DNA for that locus without immunoprecipitation. This value was normalized to the background signal calculated as the median value for all chromosomal loci of the amount of DNA in the immunoprecipitate divided by the amount of total DNA. Where presented, enrichment data are plotted according to chromosome position.

**Results**

**Binding of DnaA to several chromosomal regions increases following replication stress**

Previously, using ChIP-PCR, we found that DnaA is associated with several putative target operons whose mRNA levels change in response to perturbations in replication and that contain
potential DnaA binding sites (15). Since blocking replication elongation or preventing
replication initiation (while allowing ongoing replication to finish) elicits transcriptional
responses controlled by DnaA (15), we postulated that binding of DnaA to some targets might
increase when replication is inhibited. To test this hypothesis, we monitored DnaA binding at
three regulatory targets, dnaA, sda, and ywlC, using ChIP-PCR during replication and after
inhibition of replication. Binding was quantified by measuring the enrichment of a given DNA
region in the crosslinked and precipitated material relative to that of other chromosomal regions
(see Experimental Procedures). We inhibited replication elongation by treating cells with
HPUra, a DNA Polymerase III inhibitor (6). We inhibited replication initiation by shifting the
temperature sensitive replication initiation mutants dnaBts and dnaDts to non-permissive
temperature and allowing ongoing rounds of replication to finish.

During replication in wild type and the temperature sensitive mutants at permissive
temperature, there was low but detectable enrichment for each of the three regions (dnaA, sda,
and ywlC) in the DnaA immunoprecipitates, relative to other chromosomal regions (Table 1).
Sixty minutes after treatment with HPUra to block replication elongation, there was a significant
increase in enrichment of each of these three regions, indicating that association of DnaA with
these regions was increased (Table 1). There was also a significant increase in enrichment of
these regions 90 min after shifting the dnaBts and dnaDts mutants to non-permissive (47°C)
temperature (Table 1) and the dnaDts typically gave higher enrichment levels than the dnaBts or
treatment with HPUra. There was no increase in enrichment of these regions in wild-type cells
incubated at 47°C (Table 1), indicating that the increased binding by DnaA to DNA in the
dnaBts and dnaDts mutants at non-permissive temperature (47°C) was not due to the temperature
shift per se.
Analysis of genome-wide DnaA binding

We used ChIP-chip to identify additional chromosomal regions bound by DnaA in vivo. Based on the increase in association of DnaA with dnaA, sda, and ywlC after replication stress, we analyzed DnaA binding by ChIP-chip under these conditions as well as during exponential growth. We detected high levels of DnaA binding (~10-70-fold enrichment) at 6 chromosomal regions following arrest of either replication elongation or initiation (Figs. 1, 2). The regions include dnaA, sda, ywlC, yydA, and the intergenic regions between the 3’ ends of gcp and ydiF, and the 5’ ends of ywcI and vpr. All six of these regions contained at least nine instances of the DnaA box sequence (allowing one mismatch) clustered within a 500 bp region (Fig. 3), and we refer to these as "clustered" sites. Four of the regions with high binding (dnaA, sda, ywlC, yydA) are associated with operons whose transcription changes in response to perturbations in replication {Fig. 3 and (15)}. We postulate that the increased binding of DnaA following inhibition of replication could directly affect transcription of these target operons.

DnaA was also bound at the six clustered sites in exponentially growing wild-type cells (Figs. 1C, 2), and in dnaBts and dnaDts cells at permissive temperature (not shown). However, the enrichment values of ~2-8-fold over background indicated that DnaA binding was significantly lower than that following replication inhibition (~10-70-fold). As above, binding did not increase in wild-type cells incubated at 47°C (not shown).

The increase in DnaA binding at clustered sites after replication was blocked was not due to an increase in the intracellular concentration of DnaA. The increase in binding occurred despite transcriptional auto-repression of dnaA (15, 34), which should decrease DnaA levels in response to replication inhibition. As expected, we found that levels of DnaA, as a fraction of total protein, had decreased by approximately 35-50% 60 min after inhibition of replication.
Breier and Grossman

eelongation with HPUra (data not shown). Thus, the increase in DNA binding by DnaA is not
due to increased levels of DnaA, but rather is likely due to increased activity of DnaA in
response to replication inhibition.

To verify that the signal in the ChIP-chip experiments was specific to DnaA, we did similar
experiments in the presence and absence of HPUra in a dnaA null mutant, in which DnaA has
been rendered nonessential by the substitution of oriC with the plasmid replication origin oriN
(2, 15, 19, 22, 32). There was no detectable enrichment of the six clustered loci in the absence of
DnaA (Figs. 1E, 2). There was enrichment detected from yutF in both dnaA+ and dnaA null cells
(Fig. 1E), indicating that this signal is not due to specific immunoprecipitation of DnaA and is
perhaps due to a cross-reacting protein. The enrichment value appeared higher in dnaA null
cells, probably due to the reduced background signal throughout the chromosome caused by the
absence of DnaA and the subsequent lack of non-specific binding throughout the genome. We
have excluded yutF from further analyses and discussion.

In addition to the six strong DnaA binding regions identified above, the ChIP-chip data
indicated that there were other regions that were weakly and/or inconsistently enriched.
Comparison of ChIP-chip data from three conditions, exponentially growing cells, after
inhibition of replication initiation, and after inhibition of replication elongation, indicated that
only two additional regions had enrichment levels >2-fold (relative to both the genomic
background and the ChIP-chip data from the dnaA null mutant) under at least two of the three
conditions. One region, near yrhC, is in the middle of an operon and its transcription does not
appear to change in response to replication stress and is not analyzed further. The other, near
spo0J, does respond transcriptionally to replication stress (15) and is discussed further below. An
independent genome-wide analysis of DnaA binding in untreated cells also identified the six
clustered sites plus dnaN and thdF (20). We have included dnaN in the analysis of dnaA as the
two genes constitute an operon and the sites are also part of oriC. thdF did not qualify as
significantly enriched in our analysis due to variability, although it was 2.05-fold enriched in
arrested dnaBts cells. It is possible that our microarray probe for thdF was poorly placed to
detect bound DnaA.

Analysis of DnaA binding to regions affected by inhibition of replication by ChIP-PCR

Several chromosomal regions previously found to be enriched in DnaA immunoprecipitates
in ChIP-PCR experiments (15) were not identified in our ChIP-chip experiments nor in
chromatin affinity precipitation (ChAP-chip) experiments (20). This discrepancy indicates that
the ChIP-PCR experiments are probably more sensitive than the ChIP-chip or ChAP-chip
experiments. Previously we had observed a difference in sensitivity between ChIP-PCR and
ChIP-chip for the DNA binding protein Spo0J (4). If the difference in sensitivity between ChIP-
chip and ChIP-PCR exists for DnaA, as we expect, then there are likely to be chromosomal
regions bound by DnaA that were not detected by the ChIP-chip and ChAP-chip approaches.

Using ChIP-PCR, we tested for DnaA binding at almost all of the operons previously
postulated to be regulated by DnaA in response to inhibition of replication (15) for which ChIP-
chip results were negative. mRNA levels from these operons change in response to inhibition of
replication in a RecA-independent manner, and the regulatory regions have at least two matches
to the DnaA box consensus (allowing for one mismatch per box) within 500 bp of the start codon
of the first gene (15). We found that nine of the 14 loci tested had statistically significant
enrichment in the DnaA immunoprecipitates during replication and/or after inhibition of
replication elongation (Table 2). These regions include: citZ, lysC, yurY, yclN, dnaB, ypA,
kdgR, and ykuN (Table 2). Additionally, spo0J, which responds to perturbation of replication
stress but has only one DnaA box within 500 bp of the beginning of its operon (15), was found to bind DnaA in the ChIP-chip experiments (Table 2 and see above). The coding sequence of the \textit{spo0J} operon contains three DnaA boxes.

DnaA binding changed notably at only three of these nine regions following inhibition of replication elongation. The \textit{lysC} region was reproducibly increased in enrichment after inhibition of replication elongation (Table 2). In contrast, the enrichment in the \textit{dnaB} and \textit{yurY} regions significantly decreased after inhibition of replication elongation and was quite high during ongoing replication. This decrease might indicate modulation of DnaA by factors that do not affect binding at other regions. Alternatively, it might indicate different binding specificities for the two different nucleotide bound forms of DnaA, DnaA-ATP and DnaA-ADP.

In contrast to the chromosomal regions with clustered DnaA boxes that had significant increases in DnaA binding after inhibition of replication, most of these regions (Table 2) did not consistently show significant changes in levels of DnaA binding following treatment with HPUra. However, given the uncertainty in the data, we cannot distinguish whether DnaA binding is relatively constant regardless of replication status, or whether increases occur that are not detectable with this approach.

The ChIP-PCR results for the \textit{dnaB} and \textit{yurY} regions differed from the ChIP-chip results. Enrichment levels for these regions were approximately 20-fold in exponentially growing cells in the ChIP-PCR experiments, but neither was found to be significantly enriched in the ChIP-chip experiments; DnaA enrichment at \textit{dnaB} was generally ~2-fold, but it was not statistically significant due to variability, and no enrichment was detected at \textit{yurY}. It is possible that these loci had unusually large differences in sensitivity between the two techniques for reasons such as the location of the probes relative to DnaA binding sites, as the ChIP-PCR primers targeted the
upstream region containing binding sites, and the array probes corresponded to ORFs. The
hybridization intensity for the dnaB spot on the microarrays was also below average and not of
high quality, perhaps contributing to the unreliability of the data.

Five of the 14 loci tested gave inconclusive results. They had enrichment levels at least 1.6-
fold above background, but the levels were not statistically significant (Table 2).

**Rapid changes in DnaA binding upon resumption of replication**

In *E. coli*, Hda is required for the regulatory inactivation of DnaA (RIDA) during replication
elongation \((23, 27)\) and references therein. It is not known if RIDA activity is completely
distributive or requires passage of a replication fork through chromosomal regions bound by
DnaA-ATP. We suspected that the increases in DnaA binding and activity that occur when
replication is blocked in *B. subtilis* were likely to be reversible. We directly tested binding of
DnaA after resumption of replication and found that DnaA was released from DNA and that
release occurred in a distributive manner at all the regions tested before passage of a replication
fork through each region.

We monitored binding of DnaA to five regions with clusters of binding sites—*dnaA, yydA, ywlC, ywcI*,
and *sda*—using ChIP and quantitative PCR while replication was blocked and when
replication resumed. Samples were taken: 1) during exponential growth, 2) after inhibition of
replication initiation, and 3) at various times following release of the replication block during a
synchronized round of replication. If replication forks must interact with DnaA bound at the
various regions to facilitate release of DnaA, then DnaA binding should decrease at regions
closer to the replication origin before decreasing at distal regions. Alternatively, if DnaA bound
at these regions is able to exchange rapidly in response to the presence of active replication
forks, then DnaA binding should decline similarly at all regions, before the replication fork reaches each region.

To block replication, we shifted exponentially growing dnaDts cells to the restrictive temperature and waited for 60 minutes to allow most ongoing rounds of replication to finish. As expected, enrichment of the five tested regions in the DnaA immunoprecipitates increased 60 min after inhibition of replication initiation (Fig. 4). Replication was allowed to re-initiate from oriC by shifting the dnaDts cells back to permissive temperature (32°C). By five minutes after the temperature shift-down, binding of DnaA to the chromosomal regions had already decreased by about 50% or more. After 15 min at permissive temperature, binding of DnaA at each region was near its original level (Fig. 4). The positions of replication forks were estimated based on previous work with synchronized B. subtilis cultures (38). The rate of replication elongation is ~0.5 kb/s or ~2.6°/min. A fully synchronous round of bidirectional replication initiating at oriC (0°/360°) would result in fork positions at approximately 13° and 347° at 5 minutes, 39° and 321° at 15 minutes, and 104° and 256° at 40 minutes after replication initiation (Fig. 4, inset).

However, cells do not actually initiate in a fully synchronous manner. Rather, initiation occurs over a period of 5-10 min after release. Thus, in the cell population, replication forks are spread over the approximately 15°-20° trailing the positions given above.

The similar behavior of these five sites irrespective of chromosomal location indicates that direct interaction between the replication machinery and the DnaA-DNA nucleoprotein complex at each site along the chromosome is not required to control the level of DnaA binding to DNA at each region. Rather, it seems likely that DnaA can exchange rapidly in vivo and respond globally to the presence of an active replication fork.
Discussion

We found that DnaA associates with multiple regions of chromosomal DNA in vivo and that association at the regions that have clustered DnaA boxes increases in response to inhibition of replication. We readily detected binding at the clustered regions, and most were detected previously using ChIP-PCR (15) and ChAP-chip (20). DnaA association with other regions was more difficult to detect with the genomic approaches (ChIP-chip and ChAP-chip) but was detected using ChIP-PCR. We did not detect significant changes in the level of DnaA association with most of these regions in response to perturbations in replication. Clearly, the current techniques are limited in their ability to detect many of the DnaA binding sites in vivo. Nonetheless, there is useful information from analysis of the available data from in vivo experiments.

DnaA binding sites

A combination of in vivo and in vitro analyses demonstrated that the DnaA binding site is a somewhat degenerate 9 bp sequence, TT^AT/TNCACA (23, 31), found in multiple copies in bacterial origins of chromosomal replication. Allowing for one mismatch in any of the 9 positions, there are ~12,000 potential binding sites throughout the B. subtilis genome. Comparative DNA sequence analyses have been used to estimate the conservation, and hence significance, of some of these sequences (9, 15, 20). In addition, studies of physical association of DnaA with chromosomal DNA revealed binding to several regions that contain multiple potential binding sites (15, 20). The set of DnaA-bound sites that we observed by ChIP-chip and ChIP-PCR very clearly fell into two groups, distinguished on both a functional and sequence level (Fig. 3).
Clustered DnaA binding sites

Clustered DnaA binding sites, which had greater than 9 potential DnaA binding sites with at most 1 mismatch, behaved differently with respect to DnaA binding than regions with only a few potential DnaA binding sites. Enrichment of the regions with clustered DnaA binding sites increased dramatically in response to inhibition of replication. Increased enrichment of these regions with clusters of sites indicates that either the number of bound DnaA molecules increased, or that the crosslinking efficiency of individual molecules increased, perhaps due to a conformational change. These possibilities are not mutually exclusive. DnaA appears to form a polymeric nucleoprotein filament \(^{(12)}\), and it is quite possible that this type of structure allows for more efficient binding and/or crosslinking to DNA. Such filamentation was originally proposed for DnaA at oriC, but it is reasonable to extend this notion to other chromosomal regions with clusters of DnaA binding sites, analogous to the number of sites in oriC. (Other than oriC, the regions with clustered sites do not function as replication origins). Both increased binding and a conformational change could be affected by the nucleotide-bound state of DnaA (see below), and changes in binding and conformation are not mutually exclusive; we suspect that both occur.

Non-clustered DnaA binding sites

The other targets of DnaA that we analyzed have fewer potential binding sites, less than nine in a 500-bp region (Fig. 3). We focused on those regions upstream of genes whose expression changes in response to replication stress \(^{(15)}\). In most of these regions, there was little or no detectable change in DnaA binding following inhibition of replication. Because the levels of DnaA binding at these regions were generally low relative to the detection limit of the technique, we cannot conclude that the level of DnaA binding did not change. A change of around 50% or
less would be very difficult to detect by ChIP because of the variability between replicates, but potentially could cause a significant change in gene expression. If the amount of binding does not actually change at these regions, then another mechanism must be involved in controlling the genes downstream from these regulatory regions. For example, the nucleotide-bound state of DnaA could change and affect gene expression perhaps via interactions with the transcriptional machinery or alterations in the DNA. Alternatively, there could be other regulatory proteins involved in expression of some of the target operons.

The 9-bp DnaA binding site consensus sequence is quite common when one mismatch is allowed (11,983 occurrences in the genome), and thus there are many potential binding sites throughout the B. subtilis genome. A 500 bp region has a 5.5% chance (Poisson statistics) of containing at least four matches. In contrast, the chance of a cluster of sites with 9 or more boxes in a 500 bp region is 0.0018%. Thus, throughout the genome, there are many collections of potential binding sites that are similar in sequence and number to those in the lower part of Figure 3. Given the limited sensitivity of the ChAP-chip (20) and ChIP-chip data, it is likely that some (perhaps many) of these sites are bound by DnaA in vivo. For many transcription factors, degenerate binding sequences are more common than demonstrable regulatory activity (see, e.g., (11, 17, 28)), and DnaA is no different. If these sites are in fact bound by DnaA and serve any function, then it is possible that regulation of expression of nearby genes occurs under specific conditions that have not been tested. Alternatively, these sites may play a more general role in some aspect of DnaA function such as titrating the levels of DnaA available to bind sites in regulatory regions that directly influence gene expression or replication.
Nucleotide binding and regulation of DnaA

DnaA is a member of the AAA+ family of ATPases (25) and, based on work with *E. coli* DnaA, changes in the nucleotide binding state of DnaA serve as a regulatory switch; the ATP bound form is fully active, and the ADP bound form is not, although it binds DNA (23, 37, 40). Applying this model to transcriptional regulatory sites leads to the notion that inhibition of replication causes the accumulation of DnaA in its active, ATP bound form, thereby causing an increase in binding and/or a conformational change resulting in changes in transcription of target operons.

When replication resumes, association of DnaA with chromosomal regions with clusters of sites rapidly returns to normal levels, well before a replication fork has had time to traverse each of the clustered binding regions. These findings indicate that either there is a diffusible signal that controls the activity of DnaA in response to ongoing replication, and/or that DnaA rapidly associates with and dissociates from DNA in vivo. The off rate of DnaA dissociation from its binding site in vitro is a few minutes (35), consistent with the possibility of this type of regulation. For example, if the nucleotide bound state of DnaA is affected by passage of a replication fork through sites to which DnaA is bound, then there would be local conversion of DnaA-ATP to DnaA-ADP. Rapid association and dissociation in vivo could allow for such local changes in DnaA to quickly equilibrate with the pool of DnaA throughout the cell and affect sites far from the replication fork. We are not aware of any measurements of the in vivo exchange of DnaA on and off DNA.

Whereas much is known in *E. coli* about regulation of the activity of DnaA at oriC and the regulatory factors involved {(23, 40) and references therein}, many of these factors are not present in *B. subtilis* and other Gram-positives, nor in many Gram-negatives (40). Our findings
Breier and Grossman indicate that the association of DnaA with some of its target sites significantly changes in response to replication stress in *B. subtilis*. One of the present challenges is to determine how the activity of DnaA is modulated in response to replication stress.

Acknowledgements

This work was supported by NIH grant GM41934 to A.D.G. and NIH Kirschstein NRSA F32 G-0712950 fellowship to A.M.B. We thank Stephen P. Bell, Catherine A. Lee, Wiep Klaas Smits, Richard B. Weart, and Lyle A. Simmons for critical reading of the manuscript, Alexi I. Goranov and Melanie M. Berkmen for helpful conversations, and Hannah Blitzblau for technical advice.

References


Table 1. DnaA binding during exponential growth and replication stress.

<table>
<thead>
<tr>
<th>locus</th>
<th>32°C dnaBts</th>
<th>47°C dnaBts</th>
<th>32°C dnaDts</th>
<th>47°C dnaDts</th>
<th>HP-Ura</th>
<th>wt</th>
</tr>
</thead>
<tbody>
<tr>
<td>dnaA</td>
<td>2.2 ± 2.3, 42.2 ± 9.5 (18.8)</td>
<td>8.8 ± 2.4, 180 ± 130 (20.4)</td>
<td>4.8 ± 0.9, 61.4 ± 1.9 (12.7)</td>
<td>6.4 ± 4.0, 6.3 ± 3.4 (1.0)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>sda</td>
<td>1.7 ± 0.4, 6.8 ± 1.0 (4.0)</td>
<td>2.2 ± 0.5, 15.7 ± 4.2 (7.1)</td>
<td>2.0 ± 0.5, 4.4 ± 0.1 (2.2)</td>
<td>1.3 ± 0.04, 1.5 ± 0.4 (1.2)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>ywlC</td>
<td>2.3 ± 0.3, 9.7 ± 0.8 (4.3)</td>
<td>3.4 ± 1.2, 70.0 ± 3.1 (20.7)</td>
<td>2.8 ± 0.4, 14.4 ± 5.9 (5.1)</td>
<td>1.7 ± 0.38, 1.6 ± 0.1 (0.9)</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Values presented are the mean (from three independent biological replicates) enrichments of the indicated DNA regions (dnaA, sda, ywlC) measured by ChIP-PCR analysis ± the standard error. Data are normalized to the yabM locus, which does not bind DnaA specifically (Fig.1 and data not shown). Values in parentheses are the fold-changes for the indicated pair. Strains used were KPL69 (dnaBts), KPL73 (dnaDts), and AG174 (wt; ±HP-Ura). Cells were grown at 32°C and treated with HP-Ura for 60 min. Temperature shifts were to 47°C and samples were taken after 90 min at that temperature.

1 Enrichment value ± standard error (fold-change)
Table 2. ChIP-PCR analysis of DnaA binding at putative regulatory targets

<table>
<thead>
<tr>
<th>locus</th>
<th>- HPUrA</th>
<th>+ HPUrA</th>
<th>+/-HPUrA</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>enrichment ± SE</td>
<td>p-value</td>
<td>enrichment ± SE</td>
</tr>
<tr>
<td>citZ</td>
<td>1.3 ± 0.8</td>
<td>(0.72)</td>
<td>2.5 ± 0.4</td>
</tr>
<tr>
<td>dnaB</td>
<td>26.4 ± 9.2</td>
<td>(0.01)</td>
<td>0.9 ± 0.4</td>
</tr>
<tr>
<td>flgB</td>
<td>2.0 ± 1.1</td>
<td>(0.32)</td>
<td>1.0 ± 0.3</td>
</tr>
<tr>
<td>kdgR</td>
<td>1.8 ± 0.1</td>
<td>(0.01)</td>
<td>3.3 ± 1.6</td>
</tr>
<tr>
<td>lysC</td>
<td>1.7 ± 0.4</td>
<td>(0.16)</td>
<td>4.7 ± 1.1</td>
</tr>
<tr>
<td>ndrI (ndrEF ymaB)</td>
<td>1.8 ± 0.3</td>
<td>(0.07)</td>
<td>2.2 ± 1.1</td>
</tr>
<tr>
<td>pyrP</td>
<td>2.6 ± 0.6</td>
<td>(0.04)</td>
<td>3.5 ± 2.4</td>
</tr>
<tr>
<td>spo0J (soj)</td>
<td>3.9 ± 0.4</td>
<td>(0.001)</td>
<td>6.3 ± 1.7</td>
</tr>
<tr>
<td>sunA (sunT)</td>
<td>4.4 ± 1.7</td>
<td>(0.06)</td>
<td>8.6 ± 4.8</td>
</tr>
<tr>
<td>yciN (yciOPQ)</td>
<td>5.4 ± 1.6</td>
<td>(0.03)</td>
<td>6.8 ± 2.1</td>
</tr>
<tr>
<td>ykuN (ykuOP)</td>
<td>5.4 ± 1.2</td>
<td>(0.02)</td>
<td>4.8 ± 3.3</td>
</tr>
<tr>
<td>yhlB (ylxA fisL pbpB)</td>
<td>1.8 ±0.5</td>
<td>(0.18)</td>
<td>1.7 ± 0.9</td>
</tr>
<tr>
<td>ypvA</td>
<td>3.1 ± 0.7</td>
<td>(0.05)</td>
<td>2.7 ± 0.3</td>
</tr>
<tr>
<td>yurY (yurX csd yurVU)</td>
<td>21.7 ± 5.2</td>
<td>(0.01)</td>
<td>5.2 ± 1.1</td>
</tr>
<tr>
<td>ywzC (ywfO ywgA)</td>
<td>4.2 ± 1.7</td>
<td>(0.05)</td>
<td>3.2 ± 1.6</td>
</tr>
</tbody>
</table>

1 Wild-type cells (AG174) untreated (-HPUrA) and treated with HPUrA (+HPUrA) for 60 min were harvested and analyzed by ChIP-PCR for DnaA binding at the indicated loci. Values presented are the mean enrichments ± standard error from at least three independent biological replicates, normalized to the yabM locus, which is not specifically bound by DnaA. p-values are in parentheses (t-test). p-values comparing the results with (+) and without (-) HPUrA are indicated in the last column.

2 Operons that respond transcriptionally to replication stress and have at least 2 potential DnaA binding sites were chosen for analysis. Loci are sorted alphabetically by the first gene. Additional genes in a given operon that are also affected by replication stress are indicated in parentheses.

3 Data for spo0J are taken from the ChIP-chip results (Fig. 1).
Figure Legends

Figure 1. Genome-wide DnaA ChIP-chip profiles in arrested, replicating, and \textit{dnaA null} cells. The enrichment of each chromosomal region in the DnaA ChIP relative to total DNA is plotted versus chromosomal position relative to \textit{oriC} at 0°, in the center of the graphs. For each data set, at least three biological replicates were collected, and the median for each locus on the arrays is shown.

A. Wild-type cells (AG174) in exponential growth were treated with HPUra for 60 min at 32°C, crosslinked, and harvested for ChIP-chip.

B. \textit{dnaDts} (KPL73) cells in exponential growth at 32°C were shifted to the restrictive temperature (47°C) for 90 minutes, crosslinked, and harvested for ChIP-chip.

C. \textit{dnaBts} (KPL69) cells in exponential growth at 32°C were shifted to the restrictive temperature (47°C) for 90 minutes, crosslinked, and harvested for ChIP-chip.

D. Wild-type cells (AG174) in exponential growth at 32°C were crosslinked and harvested for ChIP-chip.

E. \textit{dnaA} null mutant cells (AIG200) in exponential growth at 32°C were crosslinked and harvested for ChIP-chip. The signal at 283° (-77° in the figure) is \textit{yutF}; there appears to be a protein bound here that interacts nonspecifically with the anti-DnaA antibody (see text).
Figure 2. DnaA binding at loci with clustered DnaA boxes. Data are shown for specific regions from Fig. 1, plotted for ~8 kb regions centered around clustered DnaA boxes where DnaA binding increased substantially upon inhibition of replication. Chromosomal positions are listed as distance (in degrees) from oriC and enrichment is as described in Fig. 1. DnaA box locations are indicated by diamonds below the data traces; filled and open diamonds represent perfect and single-mismatch boxes, respectively. Gene locations and orientations are indicated with arrows. Expression of genes with asterisks is altered after inhibition of replication (15). Expression of all of the genes indicated here with asterisks decreases after replication inhibition, except for that of sda, which increases. Data are shown in the vicinities of dnaA (A), ywlC (B), ywcI (C), yydA (D), gcp (E), and sda (F), respectively. Filled squares with solid lines represent wild type cells (AG174) 60 min after treatment with HPylra; filled circles with dotted lines represent untreated wild type cells (AG174); gray open circles and lines represent dnaA null mutant cells (AIG200); and filled triangles with solid lines represent dnaDts cells (KPL73) after 90 min at 47°C.
Figure 3. Distribution of DnaA boxes at sites bound by DnaA. Shown are locations of perfect (solid circles) and single mismatch (triangles) DnaA boxes occurring from 750 bp upstream to 375 bp downstream of the start codon of genes where DnaA binding was detected, either in ChIP-chip or ChIP-PCR experiments. The first six regions indicated (above the thick line) all have at least 9 potential DnaA binding sites with ≤1 mismatch from consensus and have increased enrichment after inhibition of replication (Table 1; Figs. 1, 2). Regions indicated below the thick line are those with fewer than 9 potential DnaA binding sites. Enrichment at these regions did not consistently increase when replication was inhibited, and DnaA enrichment at flgB, ndrI, sunA, and yllB was greater than at the background locus yabM but not statistically significant (Table 2). At right are the transcriptional changes in each of these genes due to inhibition of elongation with HPUra and inhibition of initiation by high-temperature incubation of dnaDts cells. Listed in parentheses are genes in the same operon that also responded transcriptionally. Data are from (15). Bracketed values are not statistically significant.

Functions of the genes potentially regulated by DnaA have been summarized previously (15, 20) and include genes involved in: DNA replication (dnaA, dnaN, dnaB, and perhaps ypvA), cell division (ftsL, pbpB), nucleotide metabolism (ndrI, ndrE, F), pyrimidine biosynthesis (pyrP), lantibiotic biosynthesis (sunA, T), sporulation (sda, spo0J), and several unknowns (“y” genes).

†sda and the yqeGH-aroD-yqelJK operon are divergently transcribed and both respond to inhibition of replication; the yqeG operon decreases ~2-3 fold.

*The region shown for gcp is downstream of its 3' end.

**Gene expression values are listed for ywfO; ywzC did not respond to replication inhibition.
Figure 4. Release of DnaA binding after resumption of replication. *dnaDts* cells (KPL73) in exponential growth were shifted to restrictive temperature for 60 minutes and then shifted back to permissive temperature in order to generate a synchronous round of replication initiation. Samples were taken for ChIP-PCR analysis immediately before and at the end of the incubation at restrictive temperature, and 2, 5, 10, 15, and 40 minutes after the shift back to permissive temperature. DnaA binding was analyzed at *oriC/dnaA* (solid diamonds, black lines); *yydA* (gray asterisks, gray dashed lines); *ywcI* (solid triangles, black lines); *ywlC* (black X characters, black dashed lines); and *sda* (filled gray squares, solid gray lines). Data were normalized according to the basal and maximal level of signal detected to facilitate comparison of binding kinetics at sites with different levels of DnaA binding (see, for example, levels at *dnaA/oriC* and *sda* in Fig. 2). The inset shows the positions of each analyzed site on the chromosome and the approximate extent of replication at 5, 15, and 40 minutes after release.
Fig. 1

A. Elongation-arrested cells

B. Initiation-arrested dnaDts cells

C. Initiation-arrested dnaBts cells

D. Exponentially growing cells

E. Exponentially growing dnaA- cells
Fig. 2

A. dnaA
B. ywIc
C. ywcl
D. yydA
E. gcp
F. sda

Position on Chromosome (kb)

Relative enrichment
Fig. 3

DnaA boxes:

<table>
<thead>
<tr>
<th>Gene</th>
<th>Distance relative to ATG (bp)</th>
<th>Fold change in gene expn.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Perfect</td>
<td>-750 -375 0 375</td>
<td></td>
</tr>
<tr>
<td>Single mismatch</td>
<td></td>
<td></td>
</tr>
<tr>
<td>dnaA (dnaN)</td>
<td>-7.7</td>
<td>-7.4</td>
</tr>
<tr>
<td>ywlC</td>
<td>-3.2</td>
<td>-3.2</td>
</tr>
<tr>
<td>sda†</td>
<td>2.0</td>
<td>2.0</td>
</tr>
<tr>
<td>yydA (yydBCD)</td>
<td>[-1.5]</td>
<td>-1.6</td>
</tr>
<tr>
<td>ywcI</td>
<td>[-1.2]</td>
<td>[1.1]</td>
</tr>
<tr>
<td>gcp*</td>
<td>[1.2]</td>
<td>[1.1]</td>
</tr>
<tr>
<td>citZ</td>
<td>2.1</td>
<td>1.8</td>
</tr>
<tr>
<td>dnaB</td>
<td>-2.4</td>
<td>N/A</td>
</tr>
<tr>
<td>flgB</td>
<td>-2.3</td>
<td>[-1.1]</td>
</tr>
<tr>
<td>kdgR</td>
<td>-2.3</td>
<td>-1.8</td>
</tr>
<tr>
<td>lysC</td>
<td>-5.6</td>
<td>-4.3</td>
</tr>
<tr>
<td>nrdl (nrdEF ymaB)</td>
<td>[1.5]</td>
<td>2.8</td>
</tr>
<tr>
<td>pyrP</td>
<td>-3.0</td>
<td>[-1.5]</td>
</tr>
<tr>
<td>spo0J (soj)</td>
<td>[-1.4]</td>
<td>-1.9</td>
</tr>
<tr>
<td>sunA (sunT)</td>
<td>-3.2</td>
<td>-2.0</td>
</tr>
<tr>
<td>yclN (yclOPQ)</td>
<td>-3.8</td>
<td>-1.8</td>
</tr>
<tr>
<td>ykuN (ykuOP)</td>
<td>-16.4</td>
<td>[1.1]</td>
</tr>
<tr>
<td>yllB (ylxA pbpB ftL)</td>
<td>-2.5</td>
<td>-3.3</td>
</tr>
<tr>
<td>ypvA</td>
<td>1.8</td>
<td>1.7</td>
</tr>
<tr>
<td>yurY (yurX csd yurVU)</td>
<td>2.3</td>
<td>[1.7]</td>
</tr>
<tr>
<td>ywzC (ywfo ywgA)**</td>
<td>2.8</td>
<td>2.3</td>
</tr>
</tbody>
</table>

Distance relative to ATG (bp)
Fig. 4

Percent of maximum increase in DnaA binding

Time from shift-down (min)

Approx. extent
1C of replication

oriC yydA
ywCI ywcI
ywIC
sda

dnaA 0°/360°
yydA 353°
ywcI 334°
ywlC 324°
sda 226°