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Molecular vibration-sensing component in Drosophila melanogaster olfaction

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A common explanation of molecular recognition by the olfactory system posits that receptors recognize the structure or shape of the odorant molecule. We performed a rigorous test of shape recognition by replacing hydrogen with deuterium in odorants and asking whether Drosophila melanogaster can distinguish these identically shaped isotopes. We report that flies not only differentiate between isotopic odorants, but can be conditioned to selectively avoid the common or the deuterated isotope. Furthermore, flies trained to discriminate against the normal or deuterated isotopes of a compound, selectively avoid the corresponding isotope of a different odorant. Finally, flies trained to avoid a deuterated compound exhibit selective aversion to an unrelated molecule with a vibrational mode in the energy range of the carbon–deuterium stretch. These findings are inconsistent with a shape-only model for smell, and instead support the existence of a molecular vibration-sensing component to olfactory reception.

Olfactory systems perform remarkable feats of molecular recognition, but although much is known about the neurophysiology of olfaction (1–5), how olfactory receptors “read” molecular structure remains unknown. Parts of odorant molecules (odotopes) have been proposed to engage particular receptors in a “lock-and-key” manner and this molecular shape recognition mechanism is thought sufficient for odor discrimination (2). An alternative hypothesis (6) posits that molecular vibrations of all atoms, or of particular functional groups of odorant molecules, contribute to odor recognition, and odorants with similar vibrational spectra should elicit similar olfactory responses (7). Molecules in which deuterium replaces nonexchangeable hydrogens constitute appropriate probes to test these alternatives because deuteration does not alter atom size or bond length or stiffness (8). Thus, the conformation of a deuterated molecule should be identical to that of a hydrogen-only (i.e., normal) odorant and, according to the molecular recognition theory, the two isotopes should smell identical. However, atoms in a molecule vibrate in normal modes at particular energies that depend on the molecular structure. The doubling of nuclear mass upon deuteration will change all the vibrational modes of an odorant molecule to differing extents. The second hypothesis predicts that deuteration will alter its smell in comparison with the hydrogen-only isotope. A recent attempt to test the latter hypothesis in humans was rather inconclusive, potentially confounded by interference from previous experience, olfactory training or habituation of the subjects, or shortcomings of human olfaction as suggested by the authors (9). A major experimental difficulty is that, in general, isotopes are prepared and purified differently, and therefore perceived differences in smell could be attributed to different impurities. Purity can, in principle, be ensured by smelling single-molecule peaks exiting a gas chromatographic column, but this makes simultaneous two-compound comparisons difficult and animal experiments harder still.

Here we describe a different approach to test whether molecular vibrations contribute to recognition of odor character. We assumed that, if molecular vibrations are indeed detected, the deuterated isotopes of different odorants may share a common odor character, as deuteration shifts particular peaks—e.g., the carbon–deuterium (C–H) stretch—of the compound-specific vibrational spectra to lower frequencies. If so, the deuterium odor character could be distinct and identifiable, irrespective of the structure and chemical properties of the odorant molecules that carry it. Significantly, we used Drosophila as unbiased and objective subjects to address this issue. They possess a relatively well understood olfactory system (10–13), exhibit keen olfactory discrimination (14–16), and can be conditioned to selectively avoid or seek odors with the use of established methodology (17, 18). We ask whether Drosophila can detect deuterium as a distinguishing molecular feature in odorant isotopes and a salient cue for conditioning. The results of these experiments provide support for the notion that flies can smell molecular vibrations.

Results

Spontaneous Differential Responses to Deuterated Odorants. Although deuteration does not appreciably change molecular shape, atom size, or bond length or stiffness, it doubles hydrogen mass, thus affecting the overall vibrational modes of an odorant. Therefore, if recognition of molecular shape alone was the sole determinant for odor character (2, 3), then flies should not respond differentially to deuterated [d, where d30 denotes replacement of x nonexchangeable hydrogens with deuterium atoms] and nondeuterated/normal (i.e., H-) odorants. To address this hypothesis, we took advantage of the commercial availability of acetophenone (ACP) carrying three, five, or eight deuterium atoms (d3, d5, and d8) in place of the respective hydrogens in the normal molecule (h-ACP). Equal amounts (75 μL) of each odorant were diluted to 1 mL in isopropyl myristate and we quantified (Fig. 1A) the response of groups of flies to each odorant versus unscented air traversing the arms of a standard T-maze (Materials and Methods) (19, 20). When given a choice between normal ACP (i.e., h-ACP) and air, the flies present in the odor-delivering maze arm indicated that, at the dilution used, the odorant was attractive. However, replacing only three hydrogen atoms with deuterium eliminated this spontaneous preference as the flies distributed nearly equally in the arms of the maze. The d3-ACP was mildly aversive and the fully substituted d8-ACP was the most aversive of the four isotopes. Moreover, flies consistently avoided d3, d5, and d8 ACP against equal concentration of h-ACP (Fig. S1). Therefore, a spontaneous switch in osmotaxis (14) to ACP was precipitated by substituting hydrogen atoms with deuterium.

To verify that flies are generally capable of spontaneous discrimination between airborne deuterated odorants and their normal counterparts, we additionally presented them with equal concentrations of normal and deuterated 1-octanol, normal and d8-benzaldehyde, or a normal–perdeuterated ACP pair. In confirmation of the previous results, h-ACP was preferred versus an equal concentration of the d8 odorant (Fig. 1B). Reducing the concentration of the deuterated compound by 50% resulted in

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**Conditioned Discrimination of Isotopes.** If the deuterated and normal isotopes of a pair exhibit salient odor character differences, then *Drosophila* should be able to associate either odorant with a punishing stimulus (20). To eliminate bias, the amounts of each odorant in a pair were adjusted as shown in Fig. 1 B–D to yield spontaneous preference as near zero as possible (i.e., balanced maze), so any postconditioning distribution changes would be a consequence of training. Flies were conditioned to associate electric foot-shock punishment with the presence of the deuterated or normal odorant of a pair. *Drosophila* successfully associated either odorant with punishment, demonstrated by the conditioned selective aversion of the shock-associated odor upon testing (Fig. 2). Flies shocked in the presence of the normal odorant distributed preferentially in the arm carrying its deuterated counterpart and vice versa. The shock-associated learning extended to h-benzaldehyde and d5-benzaldehyde, for which the flies exhibited no spontaneous preference, suggesting, as expected (24), that conditioned discrimination is independent from spontaneous preference.

This was not an exclusive property of the *w*1118 control strain (14), as experiments were repeated with Canton-S, a different WT strain, with identical results (Figs. S2 A and B). We also reversed the order of stimulus presentation, such that the shock-associated odor was delivered immediately before testing. Flies continued to selectively avoid the shock-associated odor (Fig. 2C), eliminating the possibility that they were simply attracted to the odor presented last in the absence of the shock reinforcer. Thus, deuterated and normal odorants present salient differences to the fly olfactory system, which can be used to predict punishment or its absence by association with either isotope.

**Generalization of Conditioned Isotope Discrimination Across Odorant Pairs.** The results so far are consistent with an unambiguous difference between deuterated and normal odorants for all three pairs tested. Is the difference associated with the majority (≥99%) compound or with impurities? Although the odorants used were of the highest purity available (Figs. S3 and S4), they
neithertheless may contain small amounts of impurities that, in principle, could account for the spontaneous preferences and the shock-associated learning. To examine this possibility, we asked whether Drosophila could recognize deuterium as a salient feature from one odorant pair to another (i.e., generalize). If different impurities were present, salient features(s) of the training odors allowing their discrimination would be absent if testing with a different odorant pair. Hence, the flies would be unable to generalize (25) and thus distribute equally in the maze arms, as if faced with novel odors. In contrast, if the deuterium odorants had a common salient feature or odor character, then flies could, in principle, generalize between pairs.

To address this hypothesis, Drosophila were trained to selectively discriminate against deuterated or normal 1-octanol or deuterated versus normal ACP, but tested against the novel pair h-benzaldehyde versus d₈-benzaldehyde. We found that flies trained to avoid h-1-octanol avoided h-benzaldehyde selectively when tested with the h-benzaldehyde/d₈-benzaldehyde pair. Conversely, conditioning to discriminate against d₈-1-octanol resulted in selective avoidance of d₈-benzaldehyde (Fig. 3A).

Similarly, flies trained to selectively avoid h-ACP discriminated preferentially against benzaldehyde and those trained against d₈-ACP avoided the deuterated isotope selectively (Fig. 3B). Because h-ACP is attractive, the task of discriminating against the aversive h-benzaldehyde was expectedly more difficult than the converse choice, as indicated by the modest selective discrimination (Fig. 3B). To further substantiate these findings, we also trained with the h-benzaldehyde/d₈-benzaldehyde pair, which do not elicit differential naive discrimination, and tested against normal and deuterated ACP. Again, flies exhibited differential avoidance of d₈-ACP if conditioned with d₈-benzaldehyde and of h-ACP if trained with h-benzaldehyde (Fig. 3C). Therefore, generalization occurred irrespective of the odorant pairs used for training and testing, suggesting that salient differences independent of their chemical identity and shape allow learned selective aversion to be transferred from the training to the testing pair.

Fig. 2. Conditioned avoidance of normal and deuterated chemically identical odorants. The mean relative distribution of flies in the arms of the maze (% excess flies) carrying the indicated odorants ± SEM is shown. The total number of flies tested in each group is shown and mean was derived from at least six repetitions per group. Drosophila was conditioned to selectively avoid the indicated member of an odorant pair by coupling it to electric foot shock (indicated by the lightning symbol). The complementary odor of each pair was used as the unpunished control. Each experiment included a naive group (open bar) to establish balancing of the odors used for testing and was used as control for all statistical comparisons on how conditioning resulted in subsequent selective avoidance of the punished odorants. Asterisks indicate significant differences that are significant. (A) Flies selectively avoided h-ACP or d₈-ACP as expected based on the conditioning scheme and illustrated by their differential distribution in the arms of the maze. The performance of both groups was significantly different from that of naive animals (P < 0.001, Dunnett test). (B) Selective discrimination (P < 0.001 vs. naive distribution; open bars) against h-octanol and d₈-1-octanol in accord with the punished odorant during training. (C) Conditioned discrimination against h-BZA (P = 0.001 vs. naive) and d₅-BZA (P < 0.001 vs. naive, Dunnett test).

Significantly, even if impurities were present, it is not expected that they would be similar in all three odorant pairs and account for differential conditioning, because the syntheses of the six compounds are different. Furthermore, the graded switch from attraction to aversion for a single molecule, ACP, clearly depended on the number of deuterium atoms it carried (Fig. 1A). Hence, it would be remarkable for an aversive impurity to fortuitously follow the number of deuterium atoms in that odorant. Nevertheless, could a single aversive impurity present only in every deuterated compound conceivably account for these results? If this were the case, it is difficult to explain how this impurity is ineffective in d₈-benzaldehyde, consistent with the observed lack of spontaneous avoidance (Fig. 1D), but renders d₅-ACP and d₅-1-octanol aversive. Furthermore, it is possible that benzaldehyde lacks the putative impurity, a potential explanation for the equivocal aversion of its two isotopes. However, if the putative impurity was masked, then, in contrast to our results (Fig. 3A and B), the benzaldehyde isotopic would not elicit differential discrimination in flies conditioned with 1-octanol/d₈-1-octanol or ACP/d₅-ACP. Collectively, then, impurities cannot account for the observed across odorant differential conditioned discrimination. In contrast, the generalization results are explained if we hypothesize that the salient cue(s) is a common property that differentiates the isotopes of each pair and distinguishes all deuterated odorants from their normal counterparts. As this cannot be molecular shape, differential evaporation, or impurities, it is likely that the common salient cue is the presence or absence of deuterium.

Odorant Molecular Vibrations Are Salient Cues for Conditioning. The doubling of hydrogen mass by deuterium will affect every molecular vibration in which movement of the hydrogen atoms occurs. When the vibrational mode involves mostly heavy atoms (i.e., C, N, O), the change in frequency is modest because of the relatively small change in mass of the deuterated versus normal compounds. In contrast, when most of the mode motion is in the hydrogen atoms themselves, as is the case for C=H stretch, wag, and scissor modes, the effect of deuteration on mode frequency will be large. A plausible biophysical mechanism for detection of molecular vibrations has been proposed previously (6), and a detailed study of the underlying physics was recently presented.
The fact that the C-D stretch occurs in the 2,200 cm⁻¹ region is useful because only a few triple-bonded functional groups possess vibrations in this region, such as acetylenes, nitriles, isonitriles, and azides. If our hypothesis is correct, flies should be able to generalize the C-D stretch vibration in deuterated compounds to a similar vibration not carried by the C-D stretch, but instead by a triple-bonded functional group similar only in stretch frequency.

Therefore, the test pair should be molecules of similar odor, only one of which includes a triple bond. We took advantage of the fact that, at least to humans, many nitriles possess odors similar in character to the corresponding aldehydes. A well known such aldehyde–nitrile pair is citronellal (3,7-dimethyloct-6-enal) and citronellyl nitrile (3,7-dimethyloct-6-enonitrile), both citrus compounds with a lemongrass smell, with the nitrile slightly more metallic/oily (27). Consistent with what we perceive as their similar odor character, flies did not show spontaneous preference for or against citronellal or citronellyl nitrile (Fig. 4C). Moreover, in agreement with these empirical observations, the calculated IR spectra of the two compounds in the fingerprint region are very similar (Fig. 4B). The main difference lies in the vibrations of the aldehyde with a C=O stretch around 1,740 cm⁻¹ and the aldehydic C-H stretch around 2,765 cm⁻¹. The nitrile lacks these two vibrations, replaced by a –CN stretch near 2,265 cm⁻¹. The 2,200-cm⁻¹ region is the only one shared by d₁₇-1-octanol and citronellyl nitrile but not by citronellal or octanol. Therefore, in conjunction with the d₁₇-1-octanol/h-1-octanol pair, we now have two pairs of chemically unrelated molecules in which vibrations in the 2,200-cm⁻¹ region are present or absent.

If the salient character recognized in the deuterated odor is indeed the C-D stretch vibrations, flies should be able to generalize the odor character from C-D to C≡N and vice versa. Because the C-D vibration is at the same energy as the nitrile’s C≡N stretch, the nitrile should be selectively avoided by animals conditioned to avoid d₁₇-1-octanol. The results of experiments designed to test this idea are shown in Fig. 4C and D. Flies trained to discriminate against d₁₇-1-octanol selectively avoided the citronellyl nitrile, whereas flies trained to avoid h-1-octanol did not show preferential avoidance (Fig. 4C). The latter outcome was not unexpected, as both citronellyl compounds have similar spectra aside from the functional group region, which do not overlap that of h-octanol, thus lacking recognizable salient features to guide behavioral responses. Furthermore, flies trained to avoid citronellyl nitrile discriminated selectively against d₁₇-1-octanol, whereas discrimination against citronellal did not yield selective avoidance of h-1-octanol for the reasons outlined earlier. Therefore, in accord with our hypothesis, the presence of nitrile or deuterium resulted in cross-learning in either direction. Moreover, these results further constraint possible explanations invoking a hypothetical impurity, because they can be explained only if d₁₇-1-octanol contains traces of citronellyl nitrile. This seems unlikely, not least because no nitrile solvent was used during its synthesis. As the nitrile is not deuterated, these results also make it very unlikely that the isotope effects we observe are caused by a property of isotopes unrelated to their vibrational modes.

Discussion

Flies, like humans, perceive odor quality and intensity and can be conditioned to discriminate differences in its concentration (28, 29), and at sufficiently different concentrations, the same odorant may appear as distinct qualities (30). Therefore, it can be argued that flies discriminate d₈-ACP and d₁₇-1-octanol from their normal isotopes based on putative (small) concentration

![Figure 3](https://www.pnas.org/doi/10.1073/pnas.1012293108)  
**Fig. 3.** *Drosophila* can be conditioned to selectively avoid deuterium. The mean relative distribution of flies in the arms of the maze (% excess flies) carrying the indicated odorants ± SEM is shown and the total number of flies in each group is denoted. The mean was derived from at least six repetitions per group. *Drosophila* was conditioned with the indicated pairs of isotopes, but tested against a different pair as indicated. The test odorants were adjusted such that avoidance of naive flies was as near zero as possible (open bar) and used to compare the distribution of conditioned animals. (A) Animals conditioned with h-1-octanol, but tested after training with h-BZA versus d₅-BZA exhibited significantly different distribution than that of naive flies (P < 0.001 vs. naive for both, Dunnett test). Animals trained with d₁₇-1-octanol selectively avoided the deuterated test odorant and vice versa. (B) Animals punished to h-ACP discriminated selectively against h-BZA, albeit not as robustly as previously described (P = 0.004 vs. naive, Dunnett test), but ones trained to avoid d₈-ACP avoided the deuterated BZA efficiently (P < 0.001 vs. naive, Dunnett test). (C) Flies conditioned to with h-BZA/d₅-BZA exhibited efficient selective avoidance of d₈-ACP and h-ACP, respectively (P < 0.001 vs. naive for both, Dunnett test).
Individual olfactory receptors in *Drosophila* and other species can mediate excitatory and inhibitory responses to different odors (10). Our data suggest that the vibrational mode and frequency of particular atoms and active groups of odorant molecules may also provide discriminatory cues that could broaden the recognition repertoire of odorant receptors, but still retain specificity. Currently, we do not know whether the same or different olfactory receptors are recruited to sense the normal and deuterated versions of the odorant molecules, and this question is currently under investigation broadly guided by the following considerations. A single receptor may recognize, by broad odotope features, a given odorant whose particular vibrational resonance may contribute to the odor-specific activity patterns of odor receptor neurons (39), potentially modifying its particular odor character. This hypothesis predicts that receptors with distinct shape selectivities must also recognize similar vibrational modes and frequencies to explain our data of selective avoidance of citronellyl nitrile after conditioning with deuterated versions of the odorant molecules (36). The third objection has been that no known biological mechanism could behave as the equivalent of a vibrational spectroscopy. However, the proposal that olfaction uses inelastic electron tunneling spectroscopy (IETS) has made the idea physically plausible. IETS is a quantum mechanism whereby electrons move from a donor to an acceptor site at constant total energy, although the acceptor is energetically lower than the donor (37). To satisfy conservation of energy, tunneling occurs only if a molecule is present between donor and acceptor, possessing vibrational mode(s) at or near this excess energy, absorbing it, and becoming excited. A modified IETS mechanism appropriate to proteins was recently described (26), suggesting a testable model applicable to olfactory receptors. Finally, it was objected that enantiomers with identical vibrations should always smell the same, whereas some smell different (38). Given that proteins are chiral, a shape-only theory cannot account for the identical odors of most enantiomer pairs. In contrast, the objection is easily answered by IETS because it exhibits the pronounced polarization effects expected when sensing molecules in fixed orientations such as enantiomers (6).
For example, we would expect to find several receptors in the 2,150-cm\(^{-1}\) band, able to sense deuterated molecules but not their undeuterated counterparts.

If the proposed vibrational sensing component is arranged in broad frequency bands of 100 to 200 cm\(^{-1}\) wide, 10 to 20 receptors would suffice to cover the entire vibrational range. However, why then do flies (and mammals) have many more receptors (2, 3)? A possible answer may be that the olfactory system has evolved to reconcile conflicting requirements, namely, to sense a wide range of odorants, thus being nonspecific, but also to possess high affinity to detect them in small concentrations. Therefore, multiple receptors may serve to ensure that one or more will always bind with adequate affinity to any molecule to provide broad olfactory recognition through the combined activities of multiple receptors. Testing these and other predictions is well within reach of available behavioral, imaging, and genetic techniques.

Materials and Methods

*Drosophila* were cultured as described previously (40) under a 12-h dark, 12-h light cycle. Mixed-sex groups of 2- to 3-d-old flies were lightly anesthetized and segregated in groups of 50 to 70 animals in vials containing food. Twenty-four hours later they were changed to fresh vials and placed in the dark before behavioral experiments commenced. Details of the behavioral procedures can be found in SI Materials and Methods.

All behavioral experiments were performed at 25 °C and 80% to 85% relative humidity. Training and testing were performed in complete dark to adapt 2 to 3 h before behavioral experiments commenced. Details of the behavioral procedures are presented in SI Materials and Methods.

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