**Active Site Threonine Facilitates Proton Transfer during Dioxygen Activation at the Diiron Center of Toluene/o-Xylene Monooxygenase Hydroxylase**

The MIT Faculty has made this article openly available. **Please share** how this access benefits you. Your story matters.

<table>
<thead>
<tr>
<th>Citation</th>
<th>Song, Woon Ju et al. “Active Site Threonine Facilitates Proton Transfer during Dioxygen Activation at the Diiron Center of Toluene/o-xylene Monooxygenase Hydroxylase.” Journal of the American Chemical Society 132.39 (2010): 13582-13585.</th>
</tr>
</thead>
<tbody>
<tr>
<td>As Published</td>
<td><a href="http://dx.doi.org/10.1021/ja1063795">http://dx.doi.org/10.1021/ja1063795</a></td>
</tr>
<tr>
<td>Publisher</td>
<td>American Chemical Society</td>
</tr>
<tr>
<td>Version</td>
<td>Author's final manuscript</td>
</tr>
<tr>
<td>Accessed</td>
<td>Wed Mar 16 11:08:00 EDT 2016</td>
</tr>
<tr>
<td>Citable Link</td>
<td><a href="http://hdl.handle.net/1721.1/67685">http://hdl.handle.net/1721.1/67685</a></td>
</tr>
<tr>
<td>Terms of Use</td>
<td>Article is made available in accordance with the publisher's policy and may be subject to US copyright law. Please refer to the publisher's site for terms of use.</td>
</tr>
<tr>
<td>Detailed Terms</td>
<td></td>
</tr>
</tbody>
</table>
Active Site Threonine Facilitates Proton Transfer During Dioxygen Activation at the Diiron Center of Toluene/o-Xylene Monooxygenase Hydroxylase

<table>
<thead>
<tr>
<th>Journal:</th>
<th>Journal of the American Chemical Society</th>
</tr>
</thead>
<tbody>
<tr>
<td>Manuscript ID:</td>
<td>ja-2010-063795.R1</td>
</tr>
<tr>
<td>Manuscript Type:</td>
<td>Communication</td>
</tr>
<tr>
<td>Date Submitted by the Author:</td>
<td>n/a</td>
</tr>
<tr>
<td>Complete List of Authors:</td>
<td>Song, Woon Ju; Massachusetts Institute of Technology, Chemistry McCormick, Michael; Massachusetts Institute of Technology, Chemistry Behan, Rachel; MIT, Department of Chemistry Sazinsky, Matthew; Pomona College, Chemistry Jiang, Wei; Penn State Univ, Biochemistry and Molecular Biology; Penn State University, Biochemistry and Molecular Biology Lin, Jeffery; Pomona College, Chemistry Krebs, Carsten; Penn State University, Biochemistry and Molecular Biology Lippard, Stephen; Massachusetts Institute of Technology, Department of Chemistry</td>
</tr>
</tbody>
</table>
Toluene/o-xylene monooxygenase hydroxylase (ToMOH), a diiron-containing enzyme, can activate dioxygen to oxidize aromatic substrates. To elucidate the role of a strictly conserved T201 residue during dioxygen activation of the enzyme, T201S, T201G, T201C, and T201V variants of ToMOH were prepared by site-directed mutagenesis. X-ray crystal structures of all the variants were obtained. Steady-state activity, regiospecificity, and single-turnover yields were also determined for the T201 mutants. Dioxygen activation by the reduced T201 variants was explored by stopped-flow UV-vis and Mössbauer spectroscopy. These studies demonstrate that the dioxygen activation mechanism is preserved in all T201 variants; however, both the formation and decay kinetics of a peroxodiiron(III) intermediate, T201\textsubscript{peroxo}, were greatly altered, revealing that T201 is critically involved in dioxygen activation. A comparison of the kinetics of O\textsubscript{2} activation in the T201S, T201C, and T201G variants under various reaction conditions revealed that T201 plays a major role in proton transfer, which is required to generate the peroxodiiron(III) intermediate. A mechanism is postulated for dioxygen activation and possible structures of oxygenated intermediates are discussed.
Active Site Threonine Facilitates Proton Transfer During Dioxygen Activation at the Diiron Center of Toluene/α-Xylene Monooxygenase Hydroxylase

Woon Ju Song,‡ Michael S. McCormick,‡ Rachel K. Behan,‡ Matthew H. Sazinsky,§ Wei Jiang,‡
Jeffery Lin,§ Carsten Krebs,§ Stephen J. Lippard ⊿

1Department of Chemistry, Massachusetts Institute of Technology, Cambridge, Massachusetts 02139, 2Department of Chemistry, Pomona College, Claremont, California 91711, and 3Department of Biochemistry and Molecular Biology and Department of Chemistry, The Pennsylvania State University, University Park, Pennsylvania 16802

Submitted to Journal of the American Chemical Society

A role for a strictly conserved threonine residue at the active sites of bacterial multicomponent monooxygenase (BMM) hydroxylases has been discussed for more than a decade. In particular, it has been conjectured that the threonine might stabilize a hydrogen-bonding network leading from the protein surface to a metal-bound, active dioxygen moiety, facilitating proton transfer required to generate and stabilize oxygenated intermediates. Such speculation for the function of the threonine residue is based in part by analogy to studies of cytochrome P450 monooxygenase (P450), where a similarly positioned threonine in the distal pocket of the heme site facilitates proton transfer to the distal oxygen atom to form an iron(III) hydroperoxo species, which then converts to an oxoiron(IV) porphyrin cation radical. Replacing threonine with alanine uncouples this process, liberating hydrogen peroxide. Unlike P450, however, few studies of diiron monooxygenases have been conducted in sufficient detail to determine whether the threonine residue does play a role during O₂ activation and, if so, whether it operates in a manner similar to that of P450. The present communication addresses these issues.

Toluene/α-xylene monooxygenase hydroxylase (ToMOH) is a carboxylate-bridged nonheme-diiron enzyme in the BMM family. A threonine residue (T201) is located in the region where dioxygen and hydrocarbons bind adjacent to the diiron active site. Previous structural studies of ToMOH and toluene 4-monoxygenase hydroxylase (T4MOH) revealed the hydroxyl group of T201 to be a part of a hydrogen-bonding network within the active site involving a glutamate ligand (E231), a coordinated water molecule, and, in complex with the regulatory protein (T4mOH/T4moD), an ordered water molecule in the cavity. In addition, recent studies of a functionally conservative ToMOH variant, T201S, demonstrated that two oxygenated species, ToMOHα and ToMOHβ (previously designated as T201Sα and T201Sβ in ref. 14), were generated upon the reaction of reduced hydroxylase (ToMOHred) and regulatory protein (ToMOD) with O₂. The latter intermediate exhibits an optical band at ~600-800 nm (ε₂50 nm ~1500 cm⁻¹ M⁻¹). The visible spectrum of T201α allowed us to measure the formation and decay kinetics of the oxygenated species. The ability to probe the formation mechanism of T201α from O₂ provides insight into the oxygenation chemistry of ToMOH. The specificity of this mechanism is particularly useful because kinetic studies of the native enzyme are hampered by the lack of a UV-vis optical transition for ToMOHα.

Figure 1. Overlaid X-ray crystal structures of ToMOH wild-type (PDB 2INC, orange) and T201X variants: T201S (green), T201G (pink), T201C (yellow), and T201V (cyan). Iron atoms and water molecules are represented as orange and red spheres. Ligands and T201X residues are depicted as sticks. Nitrogen and oxygen atoms are colored in blue and red, respectively. T201X residues are rotated counter-clockwise and expanded for clarity.

Here we report the results of our studies of the T201 variants, T201C, T201G, and T201V, prepared by site-directed mutagenesis, which elucidate the effects of the T201 side chain in regulating the kinetics of the T201α and reveal its function during O₂ activation. We also report the X-ray crystal structures, steady-state activities, regiospecificity for toluene and α-xylene oxidation, and single-turnover yields of the T201 variants. X-ray crystal structure determinations of all four T201 variants revealed the absence of any global structural differences or dramatic shift in the geometry of the T201 backbone or side chain atoms (Figure 1 and Supporting Information Table S1 and Figure S1). The overlaid structures of wild-type and T201 variants indicate that the diiron(III) cofactors and coordinating ligands are well aligned. The orientation of the hydroxyl group in T201S is identical to that of the wild-type enzyme, suggesting that the T201S variant retains the capacity to form hydrogen-bonding interactions in the active site similar to those in the native protein. In contrast, the hydrophobic side chains of T201G, T201C, and T201V do not participate in hydrogen bonding. One notable feature is rotation about the Cα-Cβ bond in both the T201C and T201V variants, where thiol and methyl groups, respectively, point in the same direction as the methyl group of the wild-type enzyme toward the hydrophobic cavity.

The steady state activity of the T201 variant was measured using phenol as the substrate (Table 1). The maximum product formation rates, calculated as the specific activity, are comparable for all T201 variants, ranging from 0.3-2.2 times the wild-type activity. Michaelis-Menten kinetics of the T201 variants were also determined (Table 1 and Figure S2). The catalytic rate constants (kcat) for all T201 variants are similar to those of the wild type, indicating that substitution of the T201 side chain does not
significantly perturb the maximum efficiency for aromatic hydroxylation. In contrast, substrate specificity, as revealed by the $k_{cat}/K_m$ values, is diminished by about two orders of magnitude in the T201G, T201C, and T201V variants. This result suggests that the lack of a hydroxyl group at the T201 position in these variants perturbs at least one step during catalysis. A similar effect was observed for toluene 4-monoxygenase hydroxylase (T4mOH)\(^{3}\) and for P450\(^{2,9}\) when the analogous threonine residues were replaced with alanine residues.

The regiospecificity for phenol oxidation in the T201 variants does not change, producing exclusively 2-hydroxyphenol. For toluene oxidation, however, the T201C, T201V, and T201G variants exhibit significant changes in product distribution (Table S2). These results indicate that the T201 substitutions reshape the substrate-binding pocket, altering the orientation of substrate in the active site. The T201 variants also gave different product distributions for $o$-xylene oxidation. The ratio of the two products, 2,3-dimethylphenol and 3,4-dimethylphenol, reflects the steric environment at the T201 site, with the bulkier side chains conveying a higher ratio of 3,4-dimethylphenol vs 2,3-dimethylphenol. This finding suggests that the side chain of the T201 residue may guide and orient hydrocarbon substrates as they enter the diiron active site. It is plausible that the side chain of the T201 residue also interacts with dioxygen during catalysis.

To further characterize the $O_2$ activation reaction in T201 variants of ToMOH and understand which chemical step in this process is perturbed by the T201 substitution, a pre-mixed solution of ToMOH\(_{red}\) T201 variants and ToMOD (ToMOH\(_{oxygen}\) D) was treated with $O_2$-saturated buffer and monitored by Mössbauer and stopped-flow UV-vis spectroscopy. Two of the variants, T201G and T201C, also exhibit broad, transient optical features that absorb maximally at ~675 nm (Figure S3), which are identical to those observed for the T201S variant.\(^{14}\) The Mössbauer spectrum of a sample of the T201G ToMOH\(_{oxygen}\) D, which was allowed to react for 15 s with $O_2$, the time of maximum accumulation of the 675 nm feature observed by stopped-flow UV-vis spectrophotometry, reveals the presence of ~10% of a quadrupole doublet with parameters nearly identical to those reported for T201\(_{chem}\) in T201S ToMOH (Figure S4).\(^{14}\) In addition, the spectrum reveals the presence of other diiron(III) species, which amount to 35 ± 7% of the total intensity of the spectrum.

The kinetics of T201\(_{chem}\) formation and decay in the T201C/G/S variants were investigated by monitoring the absorption at 675 nm over time and fitting the traces based on the model described below (Table 2 and Figure S5). The formation and decay rates of T201\(_{chem}\) in the T201C variant were best fitted to three consecutive, irreversible reactions, consisting of one formation and two consecutive decay processes ascribed to ToMOH\(_{red}\) → T201\(_{chem}\) → T201\(_{chem}\) → ToMOH\(_{ox}\), where T201\(_{chem}\) and H\(_{ox}\) are the sequential decay products of T201\(_{chem}\). Alternatively, the biphase decay in T201C may be the result of two (or more) decay pathways of T201\(_{chem}\). The reaction kinetics for T201\(_{chem}\) in the T201G variant were similarly measured. The $A_{ox}$-ys-time trace is biphase and was fit to a model involving two irreversible reactions corresponding to ToMOH\(_{red}\) → T201\(_{chem}\) → ToMOH\(_{ox}\). The formation rate constant of T201\(_{chem}\) in T201C is nearly identical to that of the T201S variant. The formation rate constant in the T201G variant, however, is ~700 times lower than that in the T201S variant. The similarity in formation kinetics of T201\(_{chem}\) in T201S and T201C implies that the thiol group of T201C may function like a hydroxyl group, at least during formation of T201\(_{chem}\), whereas the lack of a functional group in the T201G variant dramatically reduces the rate of T201\(_{chem}\) formation. The striking difference in T201\(_{chem}\) formation kinetics in the T201 variants indicates that the rate-determining step for T201\(_{chem}\) formation can be greatly perturbed and that the T201 residue is critically involved in controlling the kinetics of dioxygen activation.

Although the formation and decay of T201\(_{chem}\) could be studied for the T201S, T201C, and T201G variants, no optical bands appeared in the UV-vis region upon the addition of $O_2$ to the T201V ToMOH\(_{red}\) protein. This result indicates that the lack of a hydrogen-bonding residue in the T201 position retards a key chemical step involved in formation of T201\(_{chem}\), presumably proton transfer. At present we cannot rule out the possibility that T201\(_{chem}\) is not generated in the T201V enzymes due to the changes in the thermodynamic stability of the various oxygenated species.\(^{15}\)

To elucidate the mechanism of T201\(_{chem}\) formation and the role of the T201 residue in this process, kinetic studies were performed at variable temperature, $O_2$, and $H^+$ concentrations, and with $D_2O$ as solvent. When the dioxygen concentration was varied in its reaction with T201S ToMOH\(_{oxygen}\) D, the formation rate constants ($k_{chem}$) were linearly proportional to $[O_2]$, yielding a second-order rate-constant of $k_2 = 0.22 ± 0.01$ s$^{-1}$M$^{-1}$ (Figure 2A, black squares). The results demonstrate that T201\(_{chem}\) is generated in an irreversible process and that formation of T201\(_{chem}\) is kinetically limited by the bimolecular reaction between ToMOH\(_{ox}\) and $O_2$. From these data, $k_{form}$ of T201\(_{chem}\) in T201S ToMOH enzyme can be represented as $k_O2/k_{form} = k_O2/k_O2$). To our knowledge, this behavior reveals the first example where dioxygen has been determined to be rate-limiting in a pre-steady state reaction of any BMM hydroxylase enzyme. This interpretation is strongly supported by the negative value of $\Delta S^\circ$, -71 ± 2 J/molK (Figure S6). We therefore conclude that chemical steps during $O_2$ activation, such as structural rearrangements in the active site, component interactions, and water/proton translocation, occur much more rapidly than the associative reaction with dioxygen. The rate

| Table 1. Steady-state activity and single-turnover yield of T201X ToMOH variants. |
|---------------------------------|---------|--------|--------|--------|
| WT                             | T201S   | T201G  | T201C  | T201V  |
| Specific activity (mU/mg)      | 1100 ± 100 | 2400 ± 300 | 1900 ± 200 | 1600 ± 200 | 390 ± 20 |
| $k_{cat}/K_m$ (s$^{-1}$M$^{-1}$) | 0.04 ± 0.01 | 0.08 ± 0.04 | 0.08 ± 0.02 | 0.09 ± 0.02 | 0.016 ± 0.009 |
| single-turnover yield (%)      | 56 ± 7 | 59 ± 6 | 2.0 ± 0.9 x 10$^{-4}$ | 2.8 ± 0.8 x 10$^{-4}$ | 1.3 ± 0.9 x 10$^{-4}$ |

\[ \text{Table 2. Formation and decay rate constants of T201}_{chem} \text{ in the reaction of T201X ToMOH}_{oxygen}\text{D with }O_2. \]
| T201S  | 130 ± 7$^a$ | 2.9 ± 0.2 |
| T201C  | 93 ± 3   | 1.9 ± 0.2, 0.23 ± 0.03 |
| T201G  | 0.11 ± 0.01 | 0.035 ± 0.003 |
| T201V  | n.d.     | n.d.$^a$ |

$^a$The reaction was observed by stopped-flow spectrophotometry in 25 mM MOPS pH 7.0 buffer at 4 °C. $^b$This value for the formation rate constant differs slightly from the previously-reported $85 ± 11$ s$^{-1}$ because it varies between batches of purified protein. $^c$Not determined.
observed kinetically.

measured under various pH conditions, more T201 formed compared to total

can estimate the concentration of T201 absor

kinetically involved in this process

consistent

the buffer was prepared with d

reaction of

Figure 2


eution oxide (pL = 7.0 at 5

p peroxo


to MOH

formation rate constant

of O

The ratio of T201, to total protein concentration formed in the

reaction of T201S with O2 under various pH values.

the formation rate constant does not change significantly. When

the buffer was prepared with deuterium oxide (pL = 7.0 at 5°C),

the kinetic solvent isotope effect (KSIE) was 0.92 ± 0.11,

consistent with our hypothesis that proton translocation is not

kinetically involved in this process.

Proton transfer is, however, indispensable to the formation of

T201. By fitting the time-resolved optical spectra to the model

ToMOHred \rightarrow T201_{peroxo} \rightarrow H_2O using the measured molar

absorptivity of T201_{peroxo} from the UV-vis and Mössbauer data, we

can estimate the concentration of T201_{peroxo} generated from the

reaction of ToMOH_red with dioxygen and the fraction of T201_{peroxo}

formed compared to total protein (Figure 2C). When the ratio was

measured under various pH conditions, more T201_{peroxo} was

generated at higher proton concentrations, indicating that proton

transfer is coupled to dioxygen consumption, although not

observed kinetically.

In the T201C variant, the T201_{peroxo} formation rate constants

vary linearly with [O2] over the range ~150–400 μM. At higher O2

concentrations, however, a plateau is reached (Figure 2A, red
circles). The partial O2-dependence for T201_{peroxo} formation in the

T201C enzyme indicates that the rate is not simply limited by the
dioxygen diffusion to the active site (k_{form} \neq k[O2]). Reactions not

involved in the rate-determining step for the T201S variant are

slowed solely due to the T201S \rightarrow T201C mutation. The T201_{peroxo}

formation rate constants for the T201C enzyme display non-linear

Eyring behavior over the 4-16 °C temperature range. This

non-linearity could arise from a temperature-dependent combination of

multiple processes involved in T201_{peroxo} formation in T201C,

rather than a simple associative reaction between the hydroxylase and O2 (Figure S6). The formation rate constant depends on pH,

indicating that higher proton concentration accelerate the

formation of T201_{peroxo} (Figure 2B, red circles). These results

reveal that the rate-determining step of T201_{peroxo} formation in

T201C variant changes from that in the T201S variant to reflect a

combination of O2 associative and proton transfer events.

The T201G mutation evoked an even more substantial effect on

the T201_{peroxo} formation kinetics. The rate constants displayed

do no dependence on O2 concentration over an ~150 – 612 μM range

(Figure 2A, purple triangles), and the ΔS^i value of -35 ± 3 J/mol-

K for T201_{peroxo} formation is significantly diminished as a result

the T201S \rightarrow T201G mutation (Figure S6). These data indicate that

formation of T201_{peroxo} in ToMOH T201G is also controlled by a
different chemical step than occurs in the T201S enzyme. The

KSIE value for the formation process in T201G is 5.7 ± 0.7,

indicating generation of T201_{peroxo} to be limited by a requisite

proton translocation step. The large KSIE value observed for the

T201G enzyme implies that one or more proton translocation

steps are involved in the rate-determining step of T201_{peroxo}

formation, possibly involving the shifting carboxylate (E231, vide

infra). Proton-assisted dissociation of the E231 bridging oxygen

atom may accompany O2 binding and activation at the diiron

center.

These studies demonstrate that several chemical steps

representing the T201_{peroxo} formation are altered by substitution

of T201 in ToMOH. We can assume that an associative reaction of

dioxygen at the diiron center (∆[O2]) occurs for all T201 variants.

In T201S, this step is rate-limiting because proton transfer is

rapid. In T201C and T201G, proton transfer is the slow step, and

is followed by more rapid O2 binding. Removal of the hydroxyl

group at the T201 position disturbs the formation kinetics of

T201_{peroxo} by retarding proton transfer (k_{form} \neq k[H^+]). These data

definitively support the long-standing hypothesis that a hydroxyl

Scheme 1. Proposed mechanism of dioxygen activation in T201 variants

of ToMOH. The oxygen atoms originating from dioxygen are colored in red; the

incoming proton is colored blue. Possible hydrogen-bond between T201, E231, and oxygennated intermediates are represented as dotted lines.
group at residue T201 plays a major role in the facilitation of proton transfer during O₂ activation in ToMO and most likely all BMMs.

Based on these pre-steady state kinetics of T201₆₆₆₆ in T201 variants, we propose a chemical mechanism for dioxygen activation, Scheme 1. Consumption of dioxygen is coupled to the proton transfer into the active site, followed by formation of T201₆₆₆₆. The protonation of the peroxo unit in T201₆₆₆₆, however, is unlikely because it would quench the peroxo-to-iron charge transfer optical bands.¹⁷ Instead, we postulate that the proton is transferred to the adjacent and shifting glutamate residue (E231), a process was recently demonstrated in a synthetic diiron model complex.¹⁸ This proton could then form a hydrogen-bond to the adjacent oxygen atom in the T201₆₆₆₆, possibly a µ-1,2-peroxodiiron(III) unit.¹⁹ In contrast, the proton transfer directly to the peroxo moiety can lead to the formation of ToMO₆₆₆₆, which exhibits no optical bands. A µ-1,1-hydroperoxodiiron(III) unit is proposed as the structure of ToMO₆₆₆₆ based on QM/MM, DFT, and Mössbauer computational studies.¹⁵

The decay rate of T201₆₆₆₆ is also significantly altered in the T201 variants (Table 2). The decay rates decrease in the same order as the formation rates. Since the primary decay pathway of ToMO₆₆₆₆, in the absence of hydrocarbon substrates is protonation of the peroxo unit and subsequent release of hydrogen peroxide,²² the decay rates of the T201 variants may be also related to their abilities to facilitate proton transfer. Further studies to clarify this process are in progress.

In conclusion, steady-state activity, single-turnover yields, and pre-steady state kinetics studies of the dioxygen activation by T201 variants of ToMO support the long-standing but unproven hypothesis that the hydroxyl group in T201 plays a major role in catalysis. The kinetics of T201₆₆₆₆ formation in T201 variants under pre-steady state conditions definitively demonstrate that this threonine residue controls formation and decay rates of T201₆₆₆₆ by facilitating proton translocation during dioxygen activation. The role of T201 in ToMO that we have uncovered here is analogous to that of threonine near the active site of P₄₅₀,²⁴ supporting a general threonine-dependent process as Nature’s strategy to control proton transfer. The proposed mechanism of dioxygen activation explains how a minor change in the proton transfer pathway can lead to the formation of discrete oxygenated intermediates, ToMO₆₆₆₆ and T201₆₆₆₆, in toluene monooxygenases.

ACKNOWLEDGMENT This work was supported by grants GM32134 (to S.J.L.) from the National Institute of General Medical Sciences and by a grant from Research Corporation (to M.H.S.). R.K.B. acknowledges fellowship support from the NIGMS (1 F32 GM084564-02). We thank Ms. C. E. Timberg for comments on the manuscript. Crystallographic coordinates have been deposited in the RCSB databank, with accession codes 3N1X, 3N1Y, 3N1Z, and 3N20, for T201C, T201G, T201S, and T201V, respectively.

SUPPORTING INFORMATION Detailed experimental methods, a table of X-ray data collection, phase determination, and refinement statistics for the structures, a table of product distribution in toluene and o-xylene oxidation, a figure of active sites, a figure of Michaelis-Menten kinetics, time-dependent Mössbauer and UV-visible spectral changes upon the reaction with dioxygen, and the kinetics of T201₆₆₆₆ under various conditions in T201 variants. This material is available free of charge via the Internet at http://pubs.acs.org.

REFERENCES