Kinetics of radical intermediate formation and deoxynucleotide production in 3-aminotyrosine-substituted Escherichia coli ribonucleotide reductases

The MIT Faculty has made this article openly available. Please share how this access benefits you. Your story matters.

<table>
<thead>
<tr>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>As Published</td>
<td><a href="http://dx.doi.org/10.1021/ja201640n">http://dx.doi.org/10.1021/ja201640n</a></td>
</tr>
<tr>
<td>Publisher</td>
<td>American Chemical Society (ACS)</td>
</tr>
<tr>
<td>Version</td>
<td>Author's final manuscript</td>
</tr>
<tr>
<td>Accessed</td>
<td>Sun Jan 13 08:02:28 EST 2019</td>
</tr>
<tr>
<td>Citable Link</td>
<td><a href="http://hdl.handle.net/1721.1/72365">http://hdl.handle.net/1721.1/72365</a></td>
</tr>
<tr>
<td>Terms of Use</td>
<td>Article is made available in accordance with the publisher's policy and may be subject to US copyright law. Please refer to the publisher's site for terms of use.</td>
</tr>
<tr>
<td>Detailed Terms</td>
<td></td>
</tr>
</tbody>
</table>
Kinetics of radical intermediate formation and deoxynucleotide production in 3-aminotyrosine-substituted *Escherichia coli* ribonucleotide reductases

Ellen C. Minnihan†, Mohammad R. Seyedsayamdost†,§, Ulla Uhlin‡, and JoAnne Stubbe†,‡,*

†Department of Chemistry, Massachusetts Institute of Technology, 77 Massachusetts Avenue, Cambridge, MA 02139

‡Department of Biology, Massachusetts Institute of Technology, 77 Massachusetts Avenue, Cambridge, MA 02139

§Department of Molecular Biology, Swedish University of Agricultural Science, Uppsala Biomedical Center, Box 590, SE-75124 Uppsala, Sweden

Abstract

*Escherichia coli* ribonucleotide reductase is an αβ2 complex and catalyzes the conversion of nucleoside 5′-diphosphates (NDPs) to 2′-deoxynucleotides (dNDPs). The reaction is initiated by the transient oxidation of an active-site cysteine (C439) in αβ by a stable diferric tyrosyl radical (Y122•) cofactor in β2. This oxidation occurs by a mechanism of long-range proton-coupled electron transfer (PCET) over 35 Å through a specific pathway of residues: Y122• → W48 → Y356 in β to Y731 → Y730 → C439 in α. To study the details of this process, 3-aminotyrosine (NH2Y) has been site-specifically incorporated in place of Y356 of β. The resulting protein, Y356NH2Y-β2, and the previously-generated proteins Y731NH2Y-α2 and Y730NH2Y-α2 (NH2Y-RNRs) are shown to catalyze dNDP production in the presence of the second subunit, substrate (S), and allosteric effector (E) with turnover numbers of 0.2–0.7 s−1. Evidence acquired by three different methods indicates that the catalytic activity is inherent to NH2Y-RNRs and not the result of co-purifying wt enzyme. The kinetics of formation of 3-aminotyrosyl radical (NH2Y•) at position 356, 731, and 730 have been measured with all S/E pairs. In all cases, NH2Y• formation is biphasic (kfast of 9–46 s−1 and kslow of 1.5–5.0 s−1) and kinetically-competent to be an intermediate in nucleotide reduction. The slow phase is proposed to report on the conformational-gating of NH2Y• formation, while the kcat of ~0.5 s−1 is proposed to be associated with rate-limiting oxidation by NH2Y• of the subsequent amino acid on the pathway during forward PCET. The Xray crystal structures of Y730NH2Y-α2 and Y731NH2Y-α2 have been solved and indicate minimal structural
changes relative to wt-α2. From the data, a kinetic model for PCET along the radical propagation pathway is proposed.

INTRODUCTION

The class Ia ribonucleotide reductase (RNR) from Escherichia coli catalyzes the reduction of four nucleoside 5'-diphosphate substrates (S, where S is UDP, CDP, ADP and GDP) to 2 '-deoxynucleoside 5'-diphosphates (dNDPs) in a highly orchestrated fashion.1–3 Allosteric effectors (E, where E is ATP, dGTP, TTP, and dATP) bind to the specificity site (S-site) and ATP/dATP bind to the activity site (A-site), dictating the substrate reduced and the overall enzyme activity, respectively. The active site and both allosteric sites are located in α2, whereas the essential diferric tyrosyl radical (Y122• in E. coli) cofactor resides in β2. This stable Y• serves as the radical initiator for the transient oxidation of the active-site cysteine (C439) in α2 which initiates nucleotide reduction.

The active form of the E. coli RNR is proposed to be an α2β2 complex.4 While the individual structures of α2 and β2 have been solved, no structure of the α2β2 complex has been determined to date. Thus, a docking model of the two subunits has been generated based on shape complementarity of the individual subunits, the superposition of their two-fold symmetry axes, and the relative locations of strictly conserved residues.5 The most provocative feature of the docking model is the 35 Å distance it places between the redox-coupled residues C439 in α2 and Y122• in β2.5 Uhlin and Eklund proposed a mechanism to rationalize this long-range oxidation in which conserved, redox-active aromatic amino acids constitute a specific pathway for radical propagation: Y122• → W48 → Y356 in β2 to Y731 → Y730 → C439 in α2. Our hypothesis for radical propagation (Figure 1) has developed from this original model, and invokes orthogonal proton-coupled electron transfer (PCET) within β2 and co-linear PCET within α2.6 In the advent of emerging technology for the site-specific incorporation of unnatural amino acids into proteins, we have begun to provide convincing evidence in support of this hypothesis.

Studies from our laboratory have established that the rate-limiting step in NDP reduction in wt E. coli RNR is associated with conformational changes triggered by binding of S and E to α2.7 These changes occur prior to radical propagation, are intricately linked to its initiation, and mask observation of any radical intermediates generated during long-range propagation.7 To study the details of this process and to characterize radical intermediates, our laboratory has focused on subtly perturbing the wt enzyme through the site-specific incorporation of tyrosine analogs with different reduction potentials and/or phenolic pKₐs in place of the Ys on the pathway. Studies conducted on these mutants have provided evidence for our working model shown in Figure 1.

The role of Y356 in β2 has been most studied extensively, with more than a half dozen tyrosine analogs substituted at this position using expressed protein ligation (EPL). These include the thermodynamic radical trap 2,3-dihydroxyphenylalanine (DOPA),8,9 the radical block and pKₐ probe 3-nitrotyrosine (NO₂Y),10,11 and the dual pKₐ/reduction potential reporter fluorotyrosines (FₙYs, n = 1–4).12,13 The data from these studies have provided convincing evidence that Y356 is a redox-active participant on the pathway. This result is particularly important as the C-terminal 35 amino acids of β, including Y356, constitute a thermally disordered tail that provides key recognition features for the α2/β2 interaction.14 The docking model predicts a distance of >25 Å between W48 in β2 and Y731 in α2 (Figure 1), and our collective results support the participation of Y356 as a relay between these two residues across the subunit interface.
More recently, Y730 and Y731 of a2 have been targeted for site-specific replacement using the in vivo nonsense codon suppression technology. Employing a specific, orthogonal tRNA/tRNA-synthetase (RS) pair, the radical trap 3-aminotyrosine (NH7, E° -0.64 V, pH 7) was incorporated into each position. When combined with β2, CDP, and ATP, Y730NH2Y-a2 (or Y731NH2Y-a2) generated an aminotyrosyl radical (NH2Y•) in a kinetically-competent fashion. These studies, in conjunction with complementary studies of the “off-pathway” mutant Y413NH2Y-a2, provided evidence for a specific radical propagation pathway in α, involving Y730 and Y731. Our initial results also reported that both Y730NH2Y-a2 and Y731NH2Y-a2 were active in dNDP formation and thus indicated that the decrease in reduction potential of NH2Y relative to Y is insufficient to shut down radical propagation. Using an E° for Y of -0.83 V (pH 7), the difference between Y and NH2Y is estimated to be ~190 mV; however, potentials up to 0.93 V have been reported for Y, suggesting the difference may be greater. The activities of NH2Y-a2s were surprising, as this observation contrasts with previous studies on Y356DOPA-β2 (E° -0.57 V, pH 7), for which no catalytic activity was detectable (<1/10 wt). Additionally, our previous studies on Y356NO2Y-β2 and Y356F6Y-β2s indicated that a 200 mV increase in potential relative to Y is sufficient to shut down nucleotide reduction.

In this work, we report the expression, isolation, and characterization of Y356NH2Y-β2. This is the first mutant at position 356 of β2 generated by the in vivo nonsense suppression method. The data obtained with this mutant is free of many of the complexities described previously for the EPL-generated mutant β2s, in which two additional mutations were required to make sufficient protein for biophysical studies. As was previously observed with NH2Y-a2s, Y356NH2Y-β2 catalyzes dNDP production. Steady-state assays of NH2Y-RNRs demonstrate they possess 3–12% the activity of the respective wt subunit (kobs -0.2–0.7 s⁻¹). Three types of experiments were conducted to establish that the catalytic activity measured is associated with the NH2Y-RNRs and not with contaminating wt enzyme that co-purifies with each mutant. The X-ray crystal structures of Y730NH2Y-α2 and Y731NH2Y-α2 have been solved and demonstrate that NH2Y introduces minimal perturbation to the structural integrity of a2 and the conformation of residues involved in radical propagation.

The kinetics of NH2Y• formation have been examined with each of the three NH2Y-RNRs in the presence of the physiologically relevant S/E pairs and are all biphasic. The slower phase is invariant with position of NH2Y in the pathway and with the S/E pair studied, and is proposed to report on the rate-limiting conformational change in the wt enzyme. A comparison of the rate constants for NH2Y• formation with S or E alone indicates that S is responsible for triggering radical propagation. From these studies, a general model is proposed for long distance radical propagation that may be used to understand mechanistic details of this process relevant to the wt enzyme.

**MATERIALS AND METHODS**

**Materials**

Wt-α2 (2500 nmol/min/mg) and wt-β2 (1.2 Y122/β2, 7600 nmol/min/mg) were expressed from pMJ1-nrdA and pTB2-nrdB, respectively, and purified as previously described. Y731NH2Y-α2 and Y730NH2Y-α2 were co-expressed from pTrc-nrdA-TAG731 or pTrc-nrdA-TAG730 and pAC-NH2Y, and purified as described. All α2 proteins were pre-reduced prior to use. E. coli thioredoxin (TR, 40 U/mg) and thioredoxin reductase (TRR, 1400 U/mg) were isolated as described. 2´-Azido-2´-deoxycytidine 5´-diphosphate (N3CDP) was synthesized from uridine by known procedures. [5,3H]-CDP was purchased from ViTrax (Placentia, CA). Nucleotide primers were purchased from Invitrogen, Pfu Ultra II polymerase from Stratagene, and restriction enzymes from New England Biolabs. Assay buffer consists of 50 mM Hepes, 1 mM EDTA, and 15 mM J Am Chem Soc. Author manuscript; available in PMC 2012 June 22.
MgSO$_4$, pH 7.6. Generation of pTrc-nrdB, pTrc-nrdB(TAG$_{356}$), pET-nrdA(wt), pET-nrdA(TAG$_{730}$), and pEVOL-NH$_2$Y, and expression and purification of N-Strep-Y$_{356}$NH$_2$Y-$\alpha_2$, N-Strep-Y$_{356}$NH$_2$Y-$\beta_2$, and (His)$_6$-Y$_{356}$NH$_2$Y-$\beta_2$ are described in detail in the Supporting Information (SI).

**Expression and purification of Y$_{356}$NH$_2$Y-$\beta_2$**

Y$_{356}$NH$_2$Y-$\beta_2$ was expressed in *E. coli* DH10B cells (Invitrogen) from pTrc-nrdB(TAG$_{356}$) and pAC-NH$_2$Y in a fashion identical to that of NH$_2$Y-substituted $\alpha_2$s and gave ~1.5 g wet cell paste per L of culture. Purification by the previously reported protocol using two anion exchange chromatography steps gave a mixture of three species: Y$_{356}$NH$_2$Y-$\beta_2$ (where $\beta^*$ is $\beta$ truncated after residue 355), Y$_{356}$NH$_2$Y-$\beta_\beta^*$, and Y$_{356}$NH$_2$Y-$\beta_2$.

Fractions containing primarily full-length Y$_{356}$NH$_2$Y-$\beta_2$ were pooled, concentrated using a YM-30 membrane (Amicon), and subjected to FPLC on a Poros HQ/20 column (Applied Biosystems, 1.6 × 10 cm, 20 mL). The column was equilibrated in buffer B (50 mM Tris, 5% glycerol, pH 7.6) and was loaded with Y$_{356}$NH$_2$Y-$\beta_2$ (~20 mg). The column was washed with one column volume (CV) of 150 mM NaCl in buffer B at 1.5 mL/min, then eluted with a gradient of 150 mM to 450 mM NaCl (60 mL × 60 mL) at the same flow rate. This protocol gave Y$_{356}$NH$_2$Y-$\beta_2$ of >95% purity, as judged by SDS-PAGE. Protein contained ~0.3 Y$_{122}•/\beta_2$ as isolated.

**ESI-MS characterization of Y$_{356}$NH$_2$Y-$\beta_2$**

ESI-MS analysis was conducted at the Proteomics Core Facility in the Koch Center for Integrative Cancer Research (MIT). The protein was absorbed on a protein microtrap (Michrom BioResources) and desalted by HPLC (50% water/50% acetonitrile/0.1% formic acid) immediately prior to MS analysis. Molecular weight measurements were made by LC-MS on a QSTAR Elite quadrupole-TOF mass spectrometer, which had been externally calibrated to a resolution of ~50 ppm.

**Iron chelation and reconstitution of di-iron Y• cofactor in Y$_{356}$NH$_2$Y-$\beta_2$**

FPLC-purified Y$_{356}$NH$_2$Y-$\beta_2$ (225 µM, 1.0 mL, 0.3 Y•/β2) was placed in a pear-shaped flask fitted with a vacuum adaptor, degassed on a Schlenk line, and brought into an anaerobic chamber at 4 °C. Sodium dithionite and methyl viologen were added to the stirring protein solution to final concentrations of 8 mM and 20 µM, respectively. The resulting pale blue solution was stirred for 1 h at 4 °C. A 50-fold molar excess of ferrozine was added, and the solution was stirred for an additional 15 min. The protein solution was removed from the anaerobic chamber, applied to a G-25 Sephadex column (45 mL), and eluted with reconstitution buffer (50 mM Hepes, 5% glycerol, pH 7.6). The protein-containing fractions were concentrated to give apo-Y$_{356}$NH$_2$Y-$\beta_2$ in >90% yield. The diiron-Y• cofactor was then reconstituted as described. Radical quantitation by UV-vis absorption and EPR spectroscopies indicated 0.5 Y$_{122}•/\beta_2$.

**Determining the specific activity of Y$_{356}$NH$_2$Y-$\beta_2$**

The activity of Y$_{356}$NH$_2$Y-$\beta_2$ was determined using the spectrophotometric and radioactive assays. Y$_{356}$NH$_2$Y-$\beta_2$ (0.1 or 0.5 µM) was assayed in the presence of a 5-fold excess of wt-α2 (0.5 µM or 2.5 µM) in assay buffer at 25 °C with [5-$^3$H]-CDP (5500 cpm/nmol).

**Expression, purification, and activity assays of (His)$_6$-Y$_{730}$NH$_2$Y-$\alpha_2$**

*E. coli* BL21(Star)-DE3 cells (Invitrogen) were co-transformed with pET-nrdA(TAG$_{730}$) and pEVOL-NH$_2$Y and plated on LB-agar plates with 50 µg/mL kanamycin (Km) and 50 µg/mL chloramphenicol (Cm) at 37 °C. Starter (5 mL) and intermediate (100 mL) cultures
were grown in 2XYT at 37 °C with Km and Cm. Enriched GMML\textsuperscript{17} (500 mL) with the appropriate antibiotics was inoculated with the saturated intermediate culture (1:50 dilution) and grown at 37 °C, 225 rpm until reaching an OD\textsubscript{600} ~0.75, at which point NH\textsubscript{2}Y and DTT were added to the culture to concentrations of 1 mM and 0.1 mM, respectively. After 15 min, the NH\textsubscript{2}Y-RS was induced with L-arabinose to a final concentration of 3.3 mM. After an additional 20 min, \textit{nrdA} expression was induced with 1 mM IPTG. Cells were harvested by centrifugation 3.5 h after induction to give 2 g cell paste/L culture.

The cell pellet was resuspended in Buffer H (50 mM sodium phosphate, 5% glycerol, pH 7.2) with 0.5 mM PMSF and 10 mM β-mercaptoethanol (β-ME) in 5 mL buffer/g cell paste, homogenized, and lysed via two passes though a French pressure cell at 14,000 psi. Cell debris was cleared by centrifugation (40,000g, 25 min, 4 °C), and DNAse (NEB) was added to the supernatant at 10 U/mL. The resulting solution was incubated with rocking for 30 min at 4 °C. The supernatant was added to Ni-NTA agarose (Qiagen, 1 mL/g) and buffer H was adjusted to contain 300 mM NaCl. The resulting slurry was incubated with rocking for 1 h at 4 °C, then loaded into a column, which was washed with 15 CVs of wash buffer (buffer H with 300 mM NaCl and 10 mM imidazole, pH 7.2) with 0.5 mM PMSF and 10 mM β-ME. The protein was eluted with a linear gradient of 10 to 250 mM imidazole in wash buffer. Fractions were pooled, diluted with buffer H, and concentrated using an Amicon YM 30 membrane to give ≥5 mg per g cell paste.

The protein was subjected to anion-exchange FPLC to remove a small amount of co-purifying 729-truncated α. A Poros HQ/20 column was equilibrated in assay buffer and was loaded with 10–20 mg of protein. The column was washed with one CV of assay buffer at a flow rate of 4 mL/min, then eluted with a linear gradient of 50 to 450 mM NaCl (60 mL × 60 mL) in the same buffer at the same flow rate. Fractions containing the desired protein, determined by SDS-PAGE, were pooled and concentrated with an Amicon YM-30 membrane. The activity of (His)\textsubscript{6}-Y\textsubscript{730}NH\textsubscript{2}Y-α2 was determined using the radioactive assay.\textsuperscript{13}

**Characterization of NH\textsubscript{2}Y• formation in (His)\textsubscript{6}-Y\textsubscript{730}NH\textsubscript{2}Y-α2**

The formation of NH\textsubscript{2}Y• in the reaction of (His)\textsubscript{6}-Y\textsubscript{730}NH\textsubscript{2}Y-α2 with wt-β2, CDP, and ATP was studied by SF UV-vis absorption and EPR spectroscopy as described previously, as was the reaction between (His)\textsubscript{6}-Y\textsubscript{730}NH\textsubscript{2}Y-α2, wt-β2, N\textsubscript{3}CDP, and ATP.\textsuperscript{17}

**Crystal structure determination of Y\textsubscript{730}NH\textsubscript{2}Y-α2 and Y\textsubscript{731}NH\textsubscript{2}Y-α2**

Proteins were crystallized at 4 °C using the hanging drop vapor diffusion method in EasyXtal Tool plates (Qiagen) in the presence of a 20-amino acid peptide corresponding to the C-terminus of the \textit{E. coli} β.\textsuperscript{5} Hanging drops consisted of 2 μL of a 1:1 mixture of protein (8–9 mg/mL, final concentration) and peptide (30 mg/mL) solutions in assay buffer and 2 μL of a solution of 25 mM sodium citrate, 1.5 M LiSO\textsubscript{4}, 2 mM DTT, pH 6 to a final mixture pH of 6.0–6.5. Crystals were grown for one week in a cold room, washed with 1.5 M LiSO\textsubscript{4} in 20% ethylene glycol, mounted in fiber loops, and flash-frozen in liquid N\textsubscript{2}.

Data sets were collected at 100 K at the European Synchrotron Radiation Facility. A 2.3 Å structure of the \textit{E. coli} wt α2 (PDB-ID 2x0x)\textsuperscript{11} was used as the initial model for the refinement of data for the mutant proteins. Processing and scaling were done with the program MOSFLM/SCALA,\textsuperscript{29} refinement with the program Refmac,\textsuperscript{30} and model building with the program O.\textsuperscript{31} Additional details on data collection and refinement are given in Table S3. Structures have been deposited to the PDB with ID codes 2xo4 (Y\textsubscript{730}NH\textsubscript{2}Y-α2) and 2xo5 (Y\textsubscript{731}NH\textsubscript{2}Y-α2).
Reaction of $Y_{730}NH_2Y–\alpha_2$ (or $Y_{731}NH_2Y–\alpha_2$) and wt-$\beta_2$ with various S/E pairs monitored by EPR spectroscopy

Pre-reduced $Y_{730}NH_2Y–\alpha_2$ (or $Y_{731}NH_2Y–\alpha_2$) and E were mixed rapidly with $\beta_2$ and S in assay buffer at 25 °C. The reaction was hand-quenched at 20 s in liquid N$_2$ and its EPR spectrum recorded (see below). Reactions were carried out with 15 µM of each subunit and the following S/E pairs: CDP/ATP (1 mM, 3 mM), GDP/TTP (1 mM, 0.2 mM), UDP/ATP (1 mM, 3 mM), and ADP/dGTP/ATP (1 mM, 0.2 mM, 3 mM). The reaction between (His)$_6$-$Y_{730}NH_2Y–\alpha_2$ and wt-$\beta_2$ with CDP/ATP was studied in an identical fashion.

EPR spectra were recorded at 77 K on a Bruker ESP-300 X-band spectrometer equipped with a quartz finger dewar containing liquid N$_2$ in the Department of Chemistry Instrumentation Facility. EPR parameters were as follows: microwave frequency = 9.34 GHz, power = 30 µW, modulation amplitude = 1.5 G, modulation frequency = 100 kHz, time constant = 5.12 ms, scan time = 41.9 s. Spin quantitation and analysis of composite reaction spectra were carried out as described previously.17

Reaction of $Y_{730}NH_2Y–\alpha_2$ (or $Y_{731}NH_2Y–\alpha_2$) and wt-$\beta_2$ with S, E, or S/E pairs monitored by stopped-flow (SF) UV-vis spectroscopy

SF kinetics were performed on an Applied Photophysics DX 17MV instrument equipped with the Pro-Data upgrade. All reactions were carried out in assay buffer at 25 °C. In all cases, pre-reduced $Y_{730}NH_2Y–\alpha_2$ (or $Y_{731}NH_2Y–\alpha_2$) and E (if present) in one syringe were mixed rapidly with $\beta_2$ and S (if present) in a second syringe to yield a final concentration of 5 µM $NH_2Y–\beta_2$ and S (if present) in a second syringe to yield a final concentration of 5 µM $NH_2Y–\beta_2$. The following S and/or E combinations (final concentrations) were studied: CDP/ATP (1 mM, 3 mM), GDP/TTP (1 mM, 0.2 mM), UDP/ATP (1 or 2 mM, 3 mM), ADP/dGTP (1 mM, 0.2 mM), ADP/dGTP/ATP (1 mM, 0.2 mM, 3 mM), CDP (1 mM), GDP (1 mM), UDP (1 mM), ATP (3 mM), TTP (0.2 mM), and dGTP (0.2 mM). The reactions were monitored at 325 nm for $NH_2Y_{730}•$ ($\varepsilon \sim 10,500 M^{-1} cm^{-1}$) and 320 nm for $NH_2Y_{731}•$ ($\varepsilon \sim 11,000 M^{-1} cm^{-1}$) using PMT detection. Averaged kinetic traces generated from >5 individual traces were fit iteratively using OriginPro or KaleidaGraph software until residuals were minimized. The reaction between (His)$_6$-$Y_{730}NH_2Y–\alpha_2$ and wt-$\beta_2$ with CDP/ATP was examined in an analogous fashion.

Reaction of $Y_{356}NH_2Y–\beta_2$ and wt–$\alpha_2$ with various S/E pairs monitored by SF UV-vis and EPR spectroscopy

SF UV-vis experiments were conducted by mixing $Y_{356}NH_2Y–\beta_2$ and pre-reduced wt-$\alpha_2$ in a 1:1 ratio (final concentration of 7.5–10 µM per subunit, 3.75–5 µM total Y$_{122}•$) in the presence of one of the four S/E pairs or CDP alone, as described above. Reactions were monitored at 324 nm for $NH_2Y_{356}•$ and 410 nm for Y$_{122}•$. Data analysis was conducted as described above. The reaction of $Y_{356}NH_2Y–\beta_2$ (25 µM) with wt-$\alpha_2$ (25 µM), CDP (1 mM) and ATP (3 mM) at 25 °C was hand-quench at 20 s and analyzed by EPR spectroscopy as described above.

Reaction of $Y_{356}NH_2Y–\beta_2$, wt–$\alpha_2$, N$_3$CDP, and ATP monitored EPR spectroscopy

Wt-$\alpha_2$ and ATP were mixed with $Y_{356}NH_2Y–\beta_2$ and N$_3$CDP in assay buffer to give final concentrations of 30 µM, 3 mM, 30 µM (15 µM Y•), and 250 µM, respectively, in a reaction volume of 240 µL. The reaction mixture was incubated at 25 °C for 1 min, then hand-quenched in liquid N$_2$. Acquisition of EPR spectra and spin quantitation were conducted as described above, and deconvolution of the three radical species was performed as described previously.17
Single-turnover reaction of \( \text{NH}_2\text{Y-}\alpha_2\text{ys} \) with wt-\( \beta_2 \), CDP, and ATP

In a total volume of 500 \( \mu \text{L} \) was combined wt-\( \beta_2 \) (10 \( \mu \text{M} \)), \( [\text{\textsuperscript{3}H}]\text{-CDP} \) (0.3–0.5 mM, specific activity of 6700 or 21,300 cpm/nmol), and ATP (3 mM) in assay buffer at 25 \( ^\circ \text{C} \). The assay was initiated by addition of pre-reduced \( \alpha_2 \) (wt, \( \text{Y}_{730}\text{NH}_2\text{-Y-}\alpha_2 \), (His)\( _6 \)-\( \text{Y}_{330}\text{NH}_2\text{-Y-}\alpha_2 \), or \( \text{Y}_{731}\text{NH}_2\text{-Y-}\alpha_2 \), 2.0 \( \mu \text{M} \)). The reaction was quenched in a boiling water bath after the time required to consume all the substrate under steady-state conditions. Sample workup and scintillation counting was conducted as described previously.\(^{13}\)

RESULTS

Expression, purification, and characterization of \( \text{Y}_{356}\text{NH}_2\text{-Y-}\beta_2 \)

A number of expression systems, growth conditions, and purification techniques were investigated to maximize the production and isolation of full-length \( \text{Y}_{356}\text{NH}_2\text{-Y-}\beta_2 \), generated by the \textit{in vivo} nonsense codon suppression method. This method results in the expression of a mixture of full-length \( \beta \) (residues 1–375) and truncated \( \beta \) (1–355, designated \( \beta' \)) and, since \( \beta \) is an obligate dimer, a statistical distribution of \( \beta_2 \), \( \beta'\beta' \), and \( \beta'\beta' \) is generated. Ultimately, the pTrc-\( \text{nrdB}(\text{TAG}_{356}) \) and pAC-\( \text{NH}_2\text{Y} \)\(^{17}\) expression system was chosen, as it resulted in the highest yield of purified \( \text{Y}_{356}\text{NH}_2\text{-Y-}\beta_2 \). A purification protocol employing three anion-exchange chromatography steps was required to separate \( \text{Y}_{356}\text{NH}_2\text{-Y-}\beta_2 \) from \( \beta'\beta' \) and \( \beta'\beta' \) and to isolate the desired species in >95% purity (Figure S1 of SI). ESI-MS analysis of the protein indicated a single major species of 43,402 Da (predicted 43,401 Da, Figure S2 of SI). Addition of variable amounts of wt-\( \beta_2 \) (predicted 43,386 Da) to \( \text{Y}_{356}\text{NH}_2\text{-Y-}\beta_2 \) and analysis by ESI-MS allowed the lower limit of detection of contamination by wt-\( \beta_2 \) to be set to <5% the total \( \text{Y}_{356}\text{NH}_2\text{-Y-}\beta_2 \).

The radical content of \( \text{Y}_{356}\text{NH}_2\text{-Y-}\beta_2 \) as isolated was 0.3 \( \text{Y}^\bullet/\beta_2 \). Efforts to increase the amount to levels observed in the wt enzyme (1.2 \( \text{Y}^\bullet/\beta_2 \)) via standard reconstitution methods resulted in a maximum of 0.5 \( \text{Y}^\bullet/\beta_2 \). The reduced level of radical is likely associated with perturbation of the cofactor assembly pathway by the introduction of an easily-oxidized \( \text{Y} \) analog at position 356. Similar results were observed in previous attempts to increase radical content in \( \text{Y}_{356}\text{DOPA-}\beta_2 \).\(^{8,32}\)

Characterization of \( \text{NH}_2\text{Y}^\bullet \) at position 356

\( \text{Y}_{356}\text{NH}_2\text{-Y-}\beta_2 \) was reacted with wt-\( \alpha_2 \), CDP, and ATP and the reaction quenched at 20 s and examined by EPR spectroscopy (Figure 2A). The reaction spectrum (black) is a composite of two radical species. Subtraction the \( \text{Y}_{122}^\bullet \) spectrum (red) yields a spectrum suggestive of an \( \text{NH}_2\text{Y}^\bullet \) (blue),\(^{33}\) with more pronounced hyperfine features than observed for the \( \text{NH}_2\text{Y}^\bullet \)s at positions 730 and 731. Spin quantitation indicates 20% total spin is lost during the reaction time with the remaining spin distributed between \( \text{NH}_2\text{Y}^\bullet \) (41%) and \( \text{Y}^\bullet \) (59%). Thus, 33% of the total spin at t=0 was trapped as \( \text{NH}_2\text{Y}^\bullet \) at t=20 s. For comparison, the reaction with \( \text{Y}_{731}\text{NH}_2\text{-Y-}\alpha_2 \) gave 30% spin loss over 20 s, with the remaining radical distributed between \( \text{NH}_2\text{Y}^\bullet \) (45%) and \( \text{Y}^\bullet \) (55%).

The kinetics of \( \text{NH}_2\text{Y}^\bullet \) formation and \( \text{Y}_{122}^\bullet \) loss were examined under similar conditions by SF UV-vis spectroscopy (Figure 2B). The data were best fit to two exponentials, giving \( k_{\text{fast}} \) of 36 s\(^{-1}\) and \( k_{\text{slow}} \) of 2.1 s\(^{-1}\). Twenty-five percent of the starting \( \text{Y}^\bullet \) was converted to \( \text{NH}_2\text{Y}^\bullet \) in the fast phase, and an additional 5% in the slow phase. A complete kinetic comparison of \( \text{NH}_2\text{Y}^\bullet \) formation in the three \( \text{NH}_2\text{Y-}\)RNRSs with all S/E pairs is described below.
Catalytic activity of \( \text{Y}_{356}\text{NH}_2\text{Y-B2} \)

Initial experiments indicated that \( \text{Y}_{730}\text{NH}_2\text{Y-}\alpha2 \) and \( \text{Y}_{731}\text{NH}_2\text{Y-}\alpha2 \) catalyze dNDP production, despite the introduction of a ~190 mV thermodynamic hole in the PCET pathway.\(^{17} \) Activity assays revealed that \( \text{Y}_{356}\text{NH}_2\text{Y-B2} \) also catalyzes dCDP formation with 4–5% the activity of wt-\( \beta2 \) (Table 1). When scaled for radical content (\( \text{Y}_{356}\text{NH}_2\text{Y-B2} \) contains 0.5 \( \text{Y}^\bullet/\beta2 \) vs. 1.2 \( \text{Y}^\bullet/\beta2 \) in wt), the activity is 10–12% that of wt.

Since RNR is essential for \( E. \text{coli} \) viability, expression of \( \text{NH}_2\text{Y-RNRs} \) is always accompanied by endogenous levels of wt-\( \alpha2 \) or \( \beta2 \).\(^{34} \) Additional contamination by wt RNR can arise from the misincorporation of \( \text{Y} \) in response to the nonsense codon due to imperfect substrate specificity by the \( \text{NH}_2\text{Y-RS} \). Thus, three types of experiments were carried out to ensure that the reductase activity measured is associated with \( \text{NH}_2\text{Y-RNRs} \).

(A) \( \text{N}_3\text{CDP} \) inactivation of \( \text{NH}_2\text{Y-RNRs} \)—Studies with wt RNRs have shown that \( \text{N}_3\text{CDP} \) is a stoichiometric inhibitor which requires \( \text{C}_{430}^{\bullet} \)-mediated 3'–hydrogen atom abstraction from the nucleotide prior to RNR inactivation. A nitrogen-centered nucleotide radical (\( \text{N}^\bullet \)) is formed and covalently bound to the active site.\(^{23,26} \) Complete enzyme inactivation results after a 50% conversion of \( \text{Y}_{122}^{\bullet} \) to \( \text{N}^\bullet \), in agreement with the proposed half-site reactivity of RNR.\(^{7,8} \) Thus, if the 4–5% activity measured with \( \text{Y}_{356}\text{NH}_2\text{Y-B2} \) is associated with contaminating wt-\( \beta2 \), 2–2.5% of the \( \text{Y}^\bullet \) would be converted to \( \text{N}^\bullet \). Observation of a larger percentage of \( \text{N}^\bullet \) would be indicative of activity inherent to \( \text{Y}_{356}\text{NH}_2\text{Y-}\beta2 \). Thus, \( \text{Y}_{356}\text{NH}_2\text{Y-}\beta2 \) was reacted with wt-\( \alpha2 \), \( \text{N}_3\text{CDP} \), and ATP, quenched at 1 min, and analyzed by EPR spectroscopy (Figure S3 of SI). No spin was lost during the reaction, with 56% of the spin associated with \( \text{Y}^\bullet \), 28% with \( \text{NH}_2\text{Y}^\bullet \), and 16% with \( \text{N}^\bullet \) (Table 1). The amount of \( \text{N}^\bullet \) is six times higher than predicted on the basis of wt contamination, providing support for \( \text{Y}_{356}\text{NH}_2\text{Y-}\beta2 \)’s activity. This result is consistent with previous inactivation assays of \( \text{Y}_{730}\text{NH}_2\text{Y-}\alpha2 \) and \( \text{Y}_{731}\text{NH}_2\text{Y-}\alpha2 \) (Table 1).\(^{17} \)

(B) Affinity purification and catalytic activity of N-terminally tagged \( \text{NH}_2\text{Y-RNRs} \)—In an attempt to remove endogenous levels of wt subunit contamination, N-terminally affinity-tagged \( \text{NH}_2\text{Y-RNRs} \) were constructed. Since \( \alpha \) is an equilibrium mixture of monomer and dimer in the absence of nucleotides, recombinant, tagged \( \text{NH}_2\text{Y-}\alpha2 \) should be separable from wt-\( \alpha2 \) via affinity chromatography (Figure 3). A number of StrepII- or (His)\(_6\)-tagged \( \text{nrdA} \) expression constructs with variable linker regions (0–10 amino acids) were constructed and the encoded proteins expressed and purified (Table S1 of SI). An N-terminal (His)\(_6\) tag and 10 amino acid linker gave the maximum yield, purity, and activity of all the tagged wt-\( \alpha2 \)s investigated. This construct and the improved pEVOl\(_{35} \) vector encoding the tRNA/\( \text{NH}_2\text{Y-RS} \) were used to express (His)\(_6\)-Y\(_{730}\text{NH}_2\text{Y-}\alpha2 \), and the resulting protein was purified to homogeneity (Figure S4 of SI). MALDI-TOF MS of the purified protein gave a major peak corresponding to the full-length protein with a single \( \text{NH}_2\text{Y} \) incorporated (observed, 87,975 Da, predicted + Na\(^+ \), 87,976 Da). The only other observable peak corresponded to 729-truncated protein. Further characterization of (His)\(_6\)-Y\(_{730}\text{NH}_2\text{Y-}\alpha2 \) by EPR (Figure S5 of SI) and SF UV-vis experiments monitoring \( \text{NH}_2\text{Y}^\bullet \) formation gave results almost identical to those with the untagged mutant.\(^{17} \) Thus, this construct was adopted for all future experiments. The affinity tagging strategy was extended to the \( \beta2 \) subunit (Table S2 of SI), but the yield and/or radical content of the N-terminally StrepII- and (His)\(_6\)-tagged \( \text{Y}_{356}\text{NH}_2\text{Y-}\beta2 \) mutants was low relative to that of untagged \( \text{Y}_{356}\text{NH}_2\text{Y-}\beta2 \) and thus study of tagged \( \text{NH}_2\text{Y-}\beta2 \)s was not pursued further.

Tagged \( \text{NH}_2\text{Y-RNRs} \) were assayed for nucleotide reductase activity and were found to be catalytically active, with the results summarized in Table 1 and Tables S1 and S2 of the SI. (His)\(_6\)-Y\(_{730}\text{NH}_2\text{Y-}\alpha2 \) has ~50% the activity of the untagged mutant and demonstrates a
narrower range of activities. Thus, the tagging procedure appears to have been successful in removing varying levels of contaminating wt. Additionally, (His)₆-Y⁷₃₀NH₂Y-α₂ generates seven times more N• in the N₃CDP assay than can be rationalized on the basis of wt contamination (Table 1 and Figure S6 of the SI).

(C) Single turnover assays for dCDP formation—As a final test of catalysis, a hand-quenched single-turnover experiment was conducted using Y⁷₃₀NH₂Y-α₂ (or (His)₆-Y⁷₃₀NH₂Y-α₂), wt-β₂, [⁵³H]CDP, and ATP. Under the reaction conditions employed, wt-α₂ generates ~3 dCDP/α₂.⁷ Thus, a sample of Y⁷₃₀NH₂Y-α₂ with 4% wt activity, in which all of the activity is associated with contaminating wt RNR, should generate 0.12 dCDP/α₂. Both Y⁷₃₀NH₂Y-α₂ and (His)₆-Y⁷₃₀NH₂Y-α₂ generated 0.7 dCDP/α₂, or six-fold more product than would be predicted for activity originating solely from contaminating wt (Table 1). A similar experiment with Y⁷₃₁NH₂Y-α₂ yielded 0.65 dCDP/α₂, whereas a control experiment using a redox-inactive Y⁷₃₁F mutant generated 0.09 dCDP/α₂, consistent with 3% contaminating wt in the latter sample. At present, the basis for sub-stoichiometric formation of dCDP by NH₂Y-RNRs under single-turnover conditions is not understood. However, this observation is consistent with the results of single-turnover N₃CDP assays, in which only ~1/3 of the NH₂Y• formed is converted to N• during a one minute reaction, as described above. Collectively, the steady-state and single-turnover activity assays of both untagged and tagged NH₂Y-RNRs (Table 1 and Tables S1 and S2 of the SI) and the high accumulation of N• in all mutants (Table 1) provide convincing evidence that NH₂Y-RNRs catalyze dNDP formation.

Structural characterization of Y⁷₃₀NH₂Y-α₂ and Y⁷₃₁NH₂Y-α₂

To determine whether incorporation of NH₂Y structurally perturbs the PCET pathway, Y⁷₃₀NH₂Y-α₂ and Y⁷₃₁NH₂Y-α₂ were crystallized, their structures determined by molecular replacement, and refined to 2.5 and 2.7 Å resolution, respectively (Table S3 of SI). As the starting model, a 2.3 Å structure of wt-α₂ crystallized under similar conditions was used.¹¹ In all cases, the asymmetric unit contains α₂ (molecules A and B), to which is appended a third α (molecule C). This third α also forms a true dimer in the crystal lattice. The major conformation assumed by Y⁷₃₀NH₂Y-α₂ (Figure 4A) is one in which the NH₂ substitution is oriented to the left when looking toward C₄₃₉ from Y₇₃₁ and the protein surface. Thus, the NH₂ group is situated toward the sterically less-dense side, and its opportunities for intermolecular hydrogen bonding to other residues are minimized. A similar conformation has been reported for the NO₂ group in the crystal structure of Y⁷₃₀NO₂Y-α₂.¹¹ Interestingly, additional electron density in the structure of Y⁷₃₀NH₂Y-α₂ suggests that Y₇₃₁ can undergo a flipping motion away from Y⁷₃₀NH₂Y toward the protein surface, placing a distance of 9.5 Å between the phenolic oxygens of NH₂Y₇₃₀ and Y₇₃₁ (Figure 4B, green). Concomitant with this flipping are reorientations of N₇₃₃ and R₄₁₁. This reorientation is provocative in that it highlights the dynamic flexibility of residues at the proposed α₂/β₂ interface and suggests motions that may be possible upon subunit interactions. In the deposited structure, one molecule (C) in the asymmetric unit is built in the flipped conformation. A water molecule, present in the wt structure and hydrogen bonded to Y₇₃₀, Y₄₁₃ and D₃₃₄, is also present in this structure, as well as an additional water hydrogen-bonded to Y₇₃₁.

The major conformation of the residues in the PCET pathway observed in the Y₇₃₁NH₂Y-α₂ crystal is shown in Figure 4C. Similar to the 730 mutant, the NH₂ group is oriented to the left and does not interact with any surrounding residues. An analogous conformation has been reported for the NO₂ group of Y₇₃₁NO₂Y-α₂.¹¹ However, in one of the three α monomers (B), the NH₂ substituent is oriented to the right, within hydrogen-bonding distance (2.5 Å) of the adjacent Y₄₁₃. In molecule C, a water with high occupancy is

*J Am Chem Soc.* Author manuscript; available in PMC 2012 June 22.
hydrogen-bonded to the OH groups of residues 730 and 731 as well as to the NH$_2$ group of NH$_2$Y$_{731}$. Higher resolution structures are necessary to determine the role(s) for ordered waters in the PCET pathway.

An overlay of the wt structure with the most common conformations of the Y$_{730}$NH$_2$Y-α2 and Y$_{731}$NH$_2$Y-α2 structures is shown in Figure 4D and reveals no significant perturbations among the three structures in the distance between the phenolic oxygens of the residues at 730 and 731 (3.2–3.4 Å) and the distance between the phenolic oxygen of residue 730 and the sulfur of C$_{439}$ (3.5–3.7 Å). In general, the structures reveal no major surprises and suggest that the redox pathway remains intact upon NH$_2$Y substitution. However, catalysis involving PCET is dependent on 0.1 Å changes and thus functionally-informative structures require the presence of β, S and E, all of which are absent from the structures reported herein.

**NH$_2$Y$^\cdot$ formation with S/E pairs monitored by EPR spectroscopy**

We have previously reported NH$_2$Y$^\cdot$ formation and its detection by EPR methods in the reaction of Y$_{730}$NH$_2$Y-α2 or Y$_{731}$NH$_2$Y-α with wt-β2, CDP and ATP quenched at 10–20 s.$^{17,33}$ We now report similar studies on the reaction of Y$_{730}$NH$_2$Y-α2 and Y$_{731}$NH$_2$Y-α2 with wt-β2 and the physiologically relevant S/E pairs (GDP/TTP, ADP/ATP, dGTP, UDP/ATP), with the samples hand-quenched at 20 s. The concentrations of S and E were chosen to saturate the nucleotide binding sites on α2. When examining ADP, both dGTP and ATP effectors were used,$^{36}$ as the presence of ATP makes the behavior of ADP/dGTP more closely mimic all other S/E pairs.

The spectra for the reactions with Y$_{730}$NH$_2$Y-α2 are shown in Figure S7a of the SI and are composites of Y$_{122}^\cdot$ and NH$_2$Y$^\cdot$. Subtraction of the Y$_{122}^\cdot$ spectrum yields an NH$_2$Y$^\cdot$ spectrum, with the latter species accounting for 47–53% of the total spin at 20 s. The NH$_2$Y$^\cdot$ spectrum shows very little variation with different S/E pairs. The spectra for the reactions with Y$_{731}$NH$_2$Y-α2 are similar, with 27–45% of the spin at 20 s associated with NH$_2$Y$^\cdot$ (Figure S7b of SI). In all cases, 20–30% of the total initial spin was lost in the first 20 s, suggesting that rapid freeze-quench (RFQ) techniques are preferable for future EPR analysis of these and similar reactions.

**Kinetics of NH$_2$Y$^\cdot$ formation with S/E pairs monitored by SF UV-vis spectroscopy**

SF UV-vis kinetic experiments monitoring NH$_2$Y$^\cdot$ formation and Y$^\cdot$ loss were carried out with all three NH$_2$Y-RNRs and all S/E pairs. In all cases, NH$_2$Y$^\cdot$ formation was biphasic and occurred concomitantly with Y$_{122}^\cdot$ loss (Table 2). The fast rate constant, $k_{\text{fast}}$, varied from 9 – 45 s$^{-1}$ and demonstrated a small dependence on position within the pathway with both $k_{\text{fast}}$ and the conversion amplitude decreasing slightly with increasing distance between NH$_2$Y and the Y$_{122}^\cdot$. The average $k_{\text{fast}}$ for all S/E pairs with NH$_2$Y at position 356, 731, and 730 is 38 s$^{-1}$ (22% conversion), 19 s$^{-1}$ (20%), and 14 s$^{-1}$ (15%), respectively. $k_{\text{fast}}$ also demonstrates a small dependence on the S/E pair, with purines giving rise to faster rate constants than pyrimidines (GDP/TTP $\approx$ ADP/dGTP/ATP $>$ CDP/ATP $>$ UDP/ATP). In contrast to the fast phase, the slow phase is nearly invariant with position and S/E pair, with an average $k_{\text{slow}}$ of ~ 2.5 s$^{-1}$. We believe that both rate constants measure protein conformation changes rather than chemical events; our hypothesis is that $k_{\text{slow}}$ reports on conformational changes preceding catalysis that are rate-determining in the wt enzyme,$^{7}$ as described in the Discussion.

In the SF UV-vis experiments, 30–40% of Y$_{122}^\cdot$ is converted to NH$_2$Y$^\cdot$ over the two phases. This conversion is in good agreement with the conversion of total initial Y$_{122}^\cdot$ to NH$_2$Y$^\cdot$ (31–41%) in the hand-quench EPR experiments described above, and in a single RFQ EPR
experiment with $Y_{731}$NH$_2$-a2, wt-b2, CDP, and ATP. In the case of previous studies with $Y_{356}$DOPA-β2, a 50% conversion of initial $Y_{122}•$ to DOPA• was reported, which is the maximum conversion one would predict on the basis of the enzyme’s proposed half-site reactivity. The difference in accumulation between DOPA• and NH$_2$Y•s is not understood, but may reflect the difference in reduction potentials between the two amino acids.

Similar SF kinetic experiments were carried out on NH$_2$Y-RNRs with S or E alone to gain insight about how nucleotide binding conformationally gates PCET. All three NH$_2$Y-RNRs (Table S4 of SI) form NH$_2$Y• with S alone, but both $k_{\text{fast}}$ and $k_{\text{slow}}$ are reduced relative to those with the corresponding S/E pair. In the absence of E, purine substrates experience an average reduction in rate constants that is two-fold greater than that of pyrimidines. Total amplitudes of NH$_2$Y• formation are reduced only moderately, with an average of 22% conversion over two phases, compared to a 30% average in the presence of S/E. In presence of E alone (Table S4), rate constants and amplitudes of NH$_2$Y• formation are dramatically decreased, supporting a role for S as the key factor in triggering radical propagation. Recent studies from our laboratory have suggested that the role of E is to maximize the amount of active enzyme complex and enhance $k_{\text{cat}}$ by lowering the $K_d$ for subunit interactions in the presence of a properly matched S/E pair (Hassan, Yokoyama, and Stubbe, unpublished results). The nature of the conformational changes induced upon E binding are not known, but it is likely that the NH$_2$Y probe is sensitive to these changes, resulting in modest enhancements of $k_{\text{cat}}$ and conversion amplitude for NH$_2$Y• formation in the presence of S/E relative to S alone.

Finally, a comparison of the biphasic kinetics of NH$_2$Y$_{356}•$ formation to the predominantly triphasic kinetics previously reported for DOPA$_{356}•$ formation (Table S5 of SI) supports our hypothesis that the additional mutations required for the ELP method have increased the kinetic complexity of the $Y_{356}$DOPA-β2 reaction. The striking similarities in the rate constants and amplitudes between the fast phase for NH$_2$Y• formation and fastest phase for DOPA• formation suggest that this phase reports on a purely conformational event, as an ET event would show correlation between the rate constant for radical formation and the redox potential of the unnatural amino acid at position 356.

Thus, NH$_2$Y fulfills two roles in studying PCET in E. coli RNR. First, its incorporation into three different positions on the pathway (Figure 1) has allowed for the kinetic characterization of intermediates formed during long-range radical propagation. Second, NH$_2$Y acts as a unique conformational probe, providing evidence for the role of S/E in conformational gating that is undetectable in the wt system.

**DISCUSSION**

**NH$_2$Y-RNRs are catalytically active**

When studies on NH$_2$Y-RNRs were initiated, it was hypothesized that NH$_2$Y, ~190 mV easier to oxidize than Y at pH 7, would act as a thermodynamic sink, effectively trapping the radical during propagation and shutting down nucleotide reduction. This hypothesis was supported by previous studies on $Y_{356}$DOPA-β2, in which conversion of DOPA (~260 mV easier to oxidize than Y) to DOPA• rendered the enzyme completely inactive. It was further supported by experiments on $Y_{356}$F$_{Y}$-β2S$_{13}$ and NO$_2$-Y-RNRs ($Y_{356}$NO$_2$-β2, $Y_{731}$NO$_2$Y-α2, and $Y_{730}$NO$_2$Y-α2), which indicated that a 200 mV increase in potential relative to Y prevents catalysis.

Thus, it was unexpected when all three NH$_2$Y-RNRs showed considerable catalytic activity (Table 1), as this implies that NH$_2$Y• is capable of the thermodynamically uphill oxidation of the next residue (Y or C) on the pathway (Figure 1). Given the important implications this
observation has on the PCET mechanism, we made a considerable effort to establish that the observed activity is inherent to NH₂Y-RNRs and not associated with contaminating wt RNR. Since the genes coding for the subunits of RNR are essential in *E. coli*, the host organism for protein expression, endogenous levels of the wt subunits are always present at some level and thus can co-purify with the recombinantly expressed protein. NH₂Y-RNR expression is sensitive to growth conditions and results in heterogeneity in the quality of NH₂Y-RNR isolated, as manifested by the large standard deviations in NH₂Y-RNR activities (Table 1). We hypothesize that the activities at the higher end of these ranges (Table 1) are associated with protein isolated from growths with less successful expression, and contains a higher fraction of endogenous RNR. A similar argument has been made with NO₂-Y-α2s.¹¹

An affinity purification protocol for an optimized tagged-α2, His₆-Y7₃₀NH₂Y-α2, was developed (Figure 3) and the isolated protein assayed. This protein gave 50% lower activity and a narrower range of activities relative to untagged Y7₃₀NH₂Y-α2 (Table 1). These results support our proposal that the endogenous wt-α2 that contaminates the latter sample has been largely removed from the former.

In application of the in vivo nonsense codon suppression method, the fidelity of the evolved tRNA/RS pair can introduce an additional mechanism by which wt RNR is produced. Both the *Kₘ* of the RS for the unnatural amino acid (UAA) and the relative abundance and availability of the UAA in the cell can influence the RS’s selectivity for the UAA relative to Y. The fidelity of the tRNA/RS is also influenced by the expression system for the protein of interest, the position of UAA incorporation, and the growth medium. Since this source of contaminating wt protein is not removed by affinity purification, single-turnover experiments utilizing a native substrate (CDP) or a mechanism-based inhibitor (N₃CDP) were conducted on both the tagged and untagged NH₂Y-RNRs. In all cases, at least fourfold more product (dCDP or N•, respectively) was formed than can be rationalized on the basis of contaminating wt (Table 1, Figures S3 and S6). These single-turnover assays provide the strongest support for the high fidelity of the NH₂Y-RS/tRNA pair and for the catalytic activity of NH₂Y-RNRs.

**Kinetic model for NH₂Y-RNRs**

Our current model for catalysis by NH₂Y-RNRs is shown in Scheme 1. While NH₂Y is drawn at position 730 of α, a similar mechanism incorporating an additional oxidation step(s) may be drawn if NH₂Y is located at position 731 of α or 356 of β. The model indicates that, in the presence of both α2 and β2, the binding of S and E initiates radical propagation, as demonstrated by SF studies described above and previously.¹⁷ NH₂Y• formation occurs by two distinct pathways (steps 1 and 8) as revealed by its biphasic kinetics (Figure 2). We assign the observed *k*₇₃₀ to step 8 (green pathway, Scheme 1) and propose that this rate constant reports on PCET between a pathway residue and Y₁₂₂•, resulting in the formation of an NH₂Y• that is not in an appropriate conformation to oxidize C₄₃₉. It is likely that this electron excursion or “rattling” occurs in the wt RNR, but that the equilibrium lies strongly in favor of reoxidation of Y₁₂₂, the proposed thermodynamic minimum on the pathway. While this process is kinetically invisible in the wt enzyme, in the NH₂Y-RNRs, introduction of a new thermodynamic minimum on the pathway results in the rapid formation of a stable NH₂Y• with its rate constant for formation and accumulation dependent on position and S/E pair (Table 2). A similar event has been proposed to occur in the adenosylcobalamin-dependent class II RNR. Using a prochirally labeled 5´-deoxyadenosyl moiety of the cofactor, fast stereochemical scrambling of the label occurs, indicating rapid cleavage of the carbon-cobalt bond despite the inability to detect cob(II)alam in by rapid kinetic techniques.³⁷
NH₂Y• is also formed by step 1, with a k_{slow} of ~2.5 s⁻¹ that we assign to the slow conformational gating step subsequent to S/E binding in which the protein assumes a conformation optimized for C₄₃₉ oxidation. This assignment is supported by k_{slow} being independent of position and S/E pair (Table 2). This number falls within the lower range of values reported for the rate-determining conformational change in wt RNR (2–10 s⁻¹),⁷ and may reflect changes induced by the NH₂ substitution that could slow conformational priming, such as disruption of ordered water molecules or H-bonding networks. It is not yet known whether the two populations of NH₂Y•s formed in steps 1 and 8 interconvert (steps 9 and -9), but if they do, the interconversion must be slow (< 2.5 s⁻¹) based on the observation of two distinct kinetic phases. It is possible that these states may interconvert on the time scale of steady-state turnover, and if so, both populations would be relevant to dCDP formation. RFQ methods in conjunction with EPR and ENDOR spectroscopies may be used to probe the existence, and possible interconversion, of the two NH₂Y• populations. The nature of the conformational changes associated with k_{slow} (1) and k_{fast} (8) are unknown, but they likely involve subtle reorientations to optimize radical propagation. Temperature dependence and viscogen-dependence studies may provide further evidence supporting the assignment of these rate constants to conformational changes.³⁸

We will argue subsequently that the next step, oxidation by NH₂Y• of the subsequent residue in the pathway (step 2), is the rate-limiting step in the steady state (0.2 to 0.7 s⁻¹). But first let us consider the remaining steps in Scheme 1. The chemistry of nucleotide reduction (steps 3 and 4) occurs rapidly (>100 to 300 s⁻¹) based on kinetic modeling of the wt system⁷ and recent experimental evidence using the mutant Y₁₂₂NO₂Y•-β2 as a radical initiator.³⁹ In this mutant, ET is decoupled from both PT and conformational gating, and dCDP is formed at 100–300 s⁻¹. This rate constant is similar to the pre-steady state rate constant for dCTP formation of 55 s⁻¹ that has been measured in the L. leichmanii RNR.⁴⁰ Once dCDP is formed (step 4), reformation of the NH₂Y• by the reverse PCET should be rapid (>100 s⁻¹), magenta pathway, step 5. Evidence from single-turnover experiments using Y₁₂₂NO₂Y•-β2 also indicate that Y₃₅₆• is re-formed with a rate constant of at least 100–300 s⁻¹ following nucleotide reduction,³⁹ and kinetic simulations of the wt RNR mechanism require a comparably fast (if not faster) rate constant for reverse PCET.⁷

Our model provides two possible fates for the NH₂Y• generated upon reverse PCET (step 5). It can either re-initiate the nucleotide reduction process directly (step 7), or it can regenerate the Y₁₂₂• through step 6, or steps 7 and -1. The latter two mechanisms would involve one or two transient Y•s and a W• as intermediates, depending on the position of NH₂Y• on the pathway.⁵,⁶ While steps -1 and 6 are likely to be endergonic, we have previously demonstrated reverse PCET and slow re-oxidation of Y₁₂₂ by DOPA₃₅₆• and NH₂Y₃₅₆• only with the homodimers Y₃₅₆DOPA-ββ ⁹ and Y₃₅₆NH₂Y-ββ’ (Minnihan and Stubb, unpublished results) with wt-α2, CDP and ATP. Thus, steps -1 and/or 6 have thermodynamic precedent. However, re-oxidation of Y₁₂₂• has never been observed in the relevant homodimers and thus a pathway in which NH₂Y• becomes the radical initiator for all subsequent turnovers (step 7 to step 2) must also be considered.

We now return to the slow steady-state rate constant for dNDP production by NH₂Y-RNRs (0.2 to 0.7 s⁻¹) and consider two steps to which it may be assigned. It may be associated with reduction of the active-site disulfide (not shown in Scheme 1) formed concomitant with dCDP production (step 4), or with oxidation by NH₂Y• of the subsequent amino acid on the radical propagation pathway (step 2). Let us consider the first possibility. Under our assay conditions, wt RNR has a turnover number of >2 s⁻¹, more than four times faster than the steady-state rate constant of NH₂Y-RNRs. We think it is unlikely that substitution of NH₂Y for Y would alter the kinetics of disulfide reduction or conformational changes accompanying re-reduction. Disulfide reduction has been proposed to be rate-limiting in E.
coli wt RNR when assayed at high protein concentration ($k_{\text{cat}} = 1 \text{ s}^{-1}$ at [$\alpha 2$] > 1$\mu$M). Thus, steady-state assays were performed on Y$_{730}$NH$_2$$\alpha 2$ over a 20-fold concentration range, encompassing the regimes in which disulfide reduction is and is not rate limiting for wt-$\alpha 2$. All concentrations yielded identical specific activity measurements for the mutant. Thus, the chemistry of disulfide reduction and the conformational reorganization necessary to prepare RNR for a second turnover seem unlikely to limit $k_{\text{cat}}$ in NH$_2$Y-RNRs.

Thus, our favored candidate for the rate-determining step in the overall scheme is step 2, oxidation of subsequent residues in the pathway by NH$_2$$\alpha \cdot$. One problem with this proposition arises from the three different locations of NH$_2$$\alpha$, and the resulting differences in the amino acid oxidized in step 2. As drawn in Scheme 1, NH$_2$Y$_{730}$$\alpha \cdot$ oxidizes C$_{439}$. However, NH$_2$Y$_{356}$$\alpha \cdot$ and NH$_2$Y$_{731}$$\alpha \cdot$ will oxidize Y$_{731}$ and Y$_{730}$, respectively. We predicted that the oxidation of C$_{439}$ by NH$_2$Y$_{730}$$\alpha \cdot$ would be the most endergonic of the three, and that this would be reflected in position-dependent differences in $k_{\text{cat}}$. This is not the case (Table 1), with the rate constants for Y$_{730}$NH$_2$Y-$\alpha 2$ and Y$_{731}$NH$_2$Y-$\alpha 2$ being identical within error. If step 2 is rate-determining, it is likely that the protein environment modulates the relative oxidation potentials of the pathway Ys and/or that the uphill oxidation of C$_{439}$ is driven by coupling to rapid irreversible chemical step(s) during nucleotide reduction.

Finally, for the sake of completeness, Scheme 1 also includes the slow reduction of the NH$_2$$\alpha \cdot$ by a number of non-specific pathways (step 10). This decay process ($k_{\text{red}}$ of 0.004 to 0.007 s$^{-1}$) is 100-fold slower than the steady-state turnover number and does not contribute to significant radical loss on the timescale of our experiments.

Our model (Scheme 1) makes testable predictions about radical formation and dNDP production in NH$_2$Y-RNRs. For instance, if step 2 is rate-limiting, a solvent kinetic isotope effect (KIE) should be apparent when any of the NH$_2$Y-RNRs is assayed in D$_2$O. While interpretation of solvent isotope effects is always complicated, the ability to measure them at each step in the pathway will be mechanistically informative. Similarly, the rate constant for dCDP formation in the first turnover, determined by a rapid chemical-quench experiment, should be identical to that in the steady-state. Alternatively, if dCDP is formed with a rate constant similar to NH$_2$$\alpha \cdot$ formation (~2.5 s$^{-1}$), a step subsequent to dCDP formation (step 6 or 7, or disulfide reduction) must be rate-limiting. Finally, the ability to detect a catalytically-active NH$_2$$\alpha \cdot$ at each position in the pathway is allowing high-field ENDOR spectroscopic experiments to be conducted to elucidate hydrogen-bonding networks relevant to coupling PT and ET events. These and other experiments are underway to challenge and refine the kinetic model for catalysis in NH$_2$Y-RNRs.

**Supplementary Material**

Refer to Web version on PubMed Central for supplementary material.

**Acknowledgments**

We thank Dr. Kenichi Yokoyama for cloning pEVOl-NH$_2$$\alpha$ and for thoughtful discussions. This work was supported by the NIH grant GM29595 (to J.S.).

**REFERENCES**

20. Tommos C, Skalicky JJ, Pilloud DL, Wand AJ, Dutton PL. Biochemistry. 1999; 38:9495–9507. [PubMed: 10413257] (b) Throughout this manuscript, we cite 0.83 V as the \( E_p \) of \( Y \) at pH 7 as measured in reference 20a, as this is the value that has been independently confirmed in our lab (ref 10). It may be compared to a value of 0.93 V, as reported in reference 16 and references therein.


FIGURE 1.
The Stubbe/Nocera model for long-range (~35 Å), reversible PCET by a radical hopping mechanism in *E. coli* class Ia RNR. Evidence suggests orthogonal PCET is operative in the \( \beta \) subunit, and co-linear PCET is operative in the \( \alpha \) subunit. Residue Y\(_{356}^\beta\) has not been observed in any crystal structure of \( \beta 2 \), and thus its location relative to the other residues is unknown.
FIGURE 2.
Reaction of $Y_{356}NH_2Y-\beta2$ with wt-α2, CDP, and ATP. (A) EPR spectrum of the reaction mixture hand-quenched after incubation for 20 s at 25 °C. The reaction spectrum (black) is a composite of two species. Subtraction of a spectrum of the $Y_{122}^•$ (blue) gives the spectrum of the $NH_2Y_{356}^•$ (red). (B) SF UV-vis spectroscopy was used to determine the kinetics of $NH_2Y_{356}^•$ formation (324 nm, red) and $Y_{122}^•$ loss (410 nm, blue) under single-turnover conditions at 25 °C. Biexponential fits to the data are indicated by black lines.
FIGURE 3.
Strategy employed for removal of endogenous levels of wt-α2 and heterodimers of α and mutant α from NH2-Y-α2 samples. An N-terminal affinity tag on the mutant protein allows separation of the recombinant α from endogenous α via affinity chromatography under conditions in which the protein exists in an equilibrium between monomer (α) and dimer (α2).
FIGURE 4.
Crystal structures of NH₂Y-α₂s. Oxygens are colored red, nitrogens, blue, and sulfurs, yellow. (A) The primary conformation assumed by residues of the PCET pathway in Y₇₃₀NH₂Y-α₂ (2.5 Å). (B) Electron density suggests a secondary conformation (green) present in the Y₇₃₀NH₂Y-α₂ crystal in which the phenol of Y₇₃₁ is oriented away from NH₂Y₇₃₀ and toward the protein surface, placing a distance of 9.5 Å between the phenolic oxygens. This movement is accompanied by changes at R₄₁₁ and N₇₃₃ (shown in sticks), and highlights the dynamic motion possible at the surface believed to participate in α₂/β₂ subunit interactions. (C) The primary conformation assumed by residues of the PCET pathway in Y₇₃₁NH₂Y-α₂ (2.7 Å). A second conformation, in which the NH₂ group is oriented on the right side of the phenol, is observed in one of the three molecules in the asymmetric unit. (D) An overlay of the structures of Y₇₃₀NH₂Y-α₂ (cyan) and Y₇₃₁NH₂Y-α₂ (magenta) with wt-α₂ (yellow, 2.3 Å) solved under the same conditions demonstrates minimal perturbation of the PCET pathway in the predominant conformations assumed by the two mutants. Distances of 3.2–3.4 Å separate the phenolic oxygens of residues 730 and 731, and distances of 3.5–3.7 Å separate the sulfhydryl of 439 and the phenol of 730. Figure generated in PyMol from PDB-IDs 2x0x, 2xo4, and 2xo5.
SCHEME 1.
Kinetic model for NH$_2$Y• formation and nucleotide reduction in NH$_2$Y-RNRs. The model for Y$_{730}$NH$_2$Y-α2 is shown, but similar models can be drawn for the 731 and 356 mutants by the addition of one or two Y oxidation steps within step 2 below. Enzyme species are colored according to three different pathways – forward PCET (blue), reverse PCET (magenta), and off-pathway PCET and/or radical quenching (green). The rate constants for individual steps, if available, are given in the text. Substrate (NDP) and product (dNDP) are shown in red. Effector is omitted from the scheme for clarity.
Nucleotide reductase activities of NH\textsubscript{2}Y-RNRs.

<table>
<thead>
<tr>
<th>mutant</th>
<th>Y\textsuperscript{•} / \beta\textsuperscript{2}</th>
<th>Spec Act nmol/min/mg\textsuperscript{a}</th>
<th>turnover (s\textsuperscript{−1})</th>
<th>activity (% wt)\textsuperscript{b}</th>
<th>% wt (scaled)\textsuperscript{c}</th>
<th>N\textsuperscript{•} formed % initial Y\textsuperscript{•}</th>
<th>dCDP /α\textsuperscript{2}</th>
</tr>
</thead>
<tbody>
<tr>
<td>Y\textsubscript{730}NH\textsubscript{2}Y- α\textsubscript{2} \textsuperscript{f}</td>
<td>-</td>
<td>156 ± 36</td>
<td>0.3–0.6</td>
<td>4 – 8</td>
<td>-</td>
<td>16</td>
<td>0.70</td>
</tr>
<tr>
<td>(His)\textsubscript{6}Y\textsubscript{730}NH\textsubscript{2}Y-α\textsubscript{2}</td>
<td>-</td>
<td>78 ± 12</td>
<td>0.2–0.3</td>
<td>3 – 4</td>
<td>-</td>
<td>14</td>
<td>0.70</td>
</tr>
<tr>
<td>Y\textsubscript{731}NH\textsubscript{2}Y-α\textsubscript{2} \textsuperscript{f}</td>
<td>-</td>
<td>175 ± 50</td>
<td>0.4–0.7</td>
<td>5 – 9</td>
<td>-</td>
<td>15</td>
<td>0.65</td>
</tr>
<tr>
<td>Y\textsubscript{356}NH\textsubscript{2}Y-β\textsubscript{2}</td>
<td>0.5</td>
<td>305 ± 38</td>
<td>0.4–0.5</td>
<td>4 – 5</td>
<td>10 – 12</td>
<td>15</td>
<td>ND\textsuperscript{g}</td>
</tr>
</tbody>
</table>

\textsuperscript{a}The average and standard deviation of 3 – 5 assays conducted on 2 or more independent enzyme isolations;

\textsuperscript{b}2500 nmol/min/mg for wt-α\textsubscript{2}, 2200 nmol/min/mg for (His)\textsubscript{6}-α\textsubscript{2}, 7000 nmol/min/mg for wt-β\textsubscript{2};

\textsuperscript{c}Scaled for radical content (0.5 Y\textsuperscript{•}/β\textsuperscript{2} in mutant vs 1.2 Y\textsuperscript{•}/β\textsuperscript{2} in wt).

\textsuperscript{d}N\textsubscript{3}ADP was the substrate for Y\textsubscript{730}NH\textsubscript{2}Y-α\textsubscript{2} and Y\textsubscript{731}NH\textsubscript{2}Y-α\textsubscript{2}, while N\textsubscript{3}CDP was the substrate for (His)\textsubscript{6}Y\textsubscript{730}NH\textsubscript{2}Y-α\textsubscript{2} and Y\textsubscript{356}NH\textsubscript{2}Y-β\textsubscript{2}. Error in EPR spin quantitation is ~3% of the initial Y\textsubscript{122}\textsuperscript{•};

\textsuperscript{e}Determined by a hand-quench single turnover assay, as described in the text;

\textsuperscript{f}First described in ref 17;

\textsuperscript{g}Not determined.
TABLE 2

Kinetics of NH$_2$Y• formation in β2 and α2 at 25 °C monitored by SF UV-vis kinetics with different S/E pairs.

<table>
<thead>
<tr>
<th>Substrate/ Effector</th>
<th>NH$<em>2$Y$</em>{358}$•</th>
<th>NH$<em>2$Y$</em>{359}$•</th>
<th>NH$<em>2$Y$</em>{730}$•</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>$k$ (s$^{-1}$)$^a$</td>
<td>A (%)</td>
<td>$k$ (s$^{-1}$)</td>
</tr>
<tr>
<td>CDP/ATP</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Fast phase</td>
<td>36 ± 4</td>
<td>25 ± 2</td>
<td>18 ± 2</td>
</tr>
<tr>
<td>Slow phase</td>
<td>2.1 ± 0.6</td>
<td>5 ± 1</td>
<td>2.5 ± 0.3</td>
</tr>
<tr>
<td>UDP/ATP</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Fast phase</td>
<td>30 ± 7</td>
<td>11 ± 1</td>
<td>9.3 ± 1.6</td>
</tr>
<tr>
<td>Slow phase</td>
<td>1.5 ± 0.8</td>
<td>2 ± 1</td>
<td>2.0 ± 0.2</td>
</tr>
<tr>
<td>GDP/TTP</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Fast phase</td>
<td>38 ± 6</td>
<td>25 ± 2</td>
<td>28 ± 3</td>
</tr>
<tr>
<td>Slow phase</td>
<td>1.8 ± 0.3</td>
<td>7 ± 1</td>
<td>2.3 ± 0.3</td>
</tr>
<tr>
<td>ADP/GTP/ATP</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Fast phase</td>
<td>46 ± 9</td>
<td>25 ± 2</td>
<td>20 ± 6</td>
</tr>
<tr>
<td>Slow phase</td>
<td>3.6 ± 2.0</td>
<td>5 ± 1</td>
<td>5.0 ± 1.8</td>
</tr>
</tbody>
</table>

$^a$Errors reported as the standard deviation of >5 SF traces.