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Cage assembly of DegP protease is not required for substrate-dependent regulation of proteolytic activity or high-temperature cell survival

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Contributed by Robert T. Sauer, March 22, 2012 (sent for review February 20, 2012)

DegP, a member of the highly conserved HtrA family, performs quality-control degradation of misfolded proteins in the periplasm of Gram-negative bacteria and is required for high-temperature survival of Escherichia coli. Substrate binding transforms DegP from an inactive oligomer containing two trimers into active polyhedral cages, typically containing four or eight trimers. Although these observations suggest a causal connection, we show that cage assembly and proteolytic activation can be uncoupled. Indeed, DegP variants that remain trimeric, hexameric, or dodecameric in the presence or absence of substrate still display robust and positively cooperative substrate degradation in vitro and, most importantly, sustain high-temperature bacterial growth as well as the wild-type enzyme. Our results support a model in which substrate binding converts inactive trimers into proteolytically active trimers, and simultaneously leads to cage assembly by enhancing binding of PDZ1 domains in one trimer to PDZ2’ domains in neighboring trimers. Thus, both processes depend on substrate binding, but they can be uncoupled without loss of biological function. We discuss potential coupling mechanisms and why cage formation may have evolved if it is not required for DegP proteolysis.

E. coli / protein quality control / stress survival

Results

Cage-Defective DegP Variant. Assembly of DegP cages containing 12, 18, or 24, or more subunits is mostly assayed in DegP variants containing the S210A active-site mutation; is stimulated by peptide and protein substrates; and requires binding of the PDZ1 domains in one DegP trimer to the PDZ2’ domains in neighboring DegP trimers (Fig. 1) (8–10). Previously, we found that introducing the Y444A substitution into the PDZ2 portion of the cage interface in DegP*S210A/Y444A resulted in a variant that ran as a hexamer in gel-filtration chromatography at 10 °C (10), both in the absence and presence of a model substrate derived from residues 18–58 of lysozyme (henceforth called 18–58). When we tested the oligomeric form of DegP*S210A/Y444A by sedimentation-velocity experiments performed at 20 °C, the protein was predominantly trimeric irrespective of the absence or presence of 18–58 (Fig. 2). Thus, the Y444A mutation prevents substrate-stabilized cage formation under these conditions. By contrast, 18–58 induces formation of cages of DegP*S210A that are predominantly dodecameric as assayed by gel filtration, sedimentation, and the crystal structure of an 18–58+DegP*S210A complex (10).

DegP*S210A is a variant in which the wild-type cysteines have been mutated to serines and the N296C mutation provides a site for labeling with fluorescent dyes (10). When one batch of DegP*S210A
is labeled with a donor dye and another batch with an acceptor dye, mixing in the presence of substrate results in cage formation that can be monitored by increased FRET. Using this assay, DegP* S210A + 18–58 showed high FRET, indicative of cage formation, at temperatures between 25 and 47 °C (Fig. 2B). As expected, the FRET signal for a mixture of donor- and acceptor-labeled DegP* S210A was much lower in the absence of 18–58 and decreased even more at higher temperatures. The latter effect may reflect dissociation of some metastable cages that form at low temperature in the absence of substrate. Importantly, DegP* S210A/Y444A gave a very low FRET signal at each temperature whether 18–58 was present or absent (Fig. 2B). Thus, multiple assays under a variety of conditions show that the Y444A mutation prevents or greatly destabilizes cage formation.

Cage Assembly Is Not Required for Proteolysis or Cooperative Substrate Binding. To determine if the inability to form cages affects proteolytic activity, we determined initial rates of DegP and DegP* Y444A cleavage of different concentrations of a two-degron substrate with 23 residues (p23) in which cleavage at a single site increases fluorescence by separating a fluorophore and quencher (Fig. 2C). The proteolytic activities of both enzymes were similar. Fitting the DegP data to the Hill form of the Michaelis–Menten equation gave a Vmax of 1.9 min⁻¹ enzyme⁻¹, an apparent Ks of 2.1 μM, and a Hill constant of 1.6. For DegP* Y444A, these values were 2.4 min⁻¹·enz⁻¹, 2.4 μM, and 2.9. Thus, substrate cleavage by both enzymes was positively cooperative, with comparable maximal rates and apparent Ks values. We conclude that cage formation is not required for the intrinsic proteolytic activity of DegP or for cooperative interactions with substrates. Note, however, that apparent Ks are complex functions of the strength of substrate binding to the oligomers present as well as the energetic costs of coupled conformational changes from the inactive to the active conformation and, in the case of wild-type DegP, the cost of the dodecamer assembly reaction. Thus, substrate could bind more efficiently to the active oligomers than would be possible to inactive species.

**Fig. 2.** Cage assembly is not required for proteolysis or cooperative substrate binding. (A) DegP* S210A/Y444A (30 μM monomer equivalent) was predominantly trimeric in the absence or presence of the 18–58 substrate (33 μM) in sedimentation velocity centrifugation experiments. The expected sedimentation positions for trimers, hexamers, and dodecamers are shown as dots (10). (B) Donor and acceptor dye-labeled DegP* S210A or DegP* S210A/Y444A (1 μM total monomer equivalent) plus or minus 18–58 (60 μM) were incubated at different temperatures, and the FRET emission ratio was determined. For DegP* S210A with substrate, FRET was high at all temperatures, indicating formation of stable cages. For DegP* S210A without substrate, FRET was substantially lower and decreased as the temperature was increased. Cages may be marginally stable at low but not high temperatures in the absence of substrate; alternatively, the FRET decrease might correspond to partial dissociation of hexamers. FRET was low under all conditions for DegP* S210A/Y444A, as expected if this variant cannot form cages and is predominantly trimeric. (C) Initial rates of DegP or DegP* Y444A (0.2 μM monomer equivalent) cleavage of different concentrations of the p23 substrate at 25 °C were determined and fitted to the Hill form of a Michaelis–Menten equation (rate = Vmax[S] / (Ks + [S])). The sequence of the p23 substrate sequence is shown above the plot. Abz represents a 2-amino-benzoyl acid fluorophore, and Y444A represents a 3-nitrotyrosine quencher. Error bars are averages ± SD (n = 3). (D) Apparent equilibrium dissociation constants (Kdiss) of Hc18–58 binding to different variants of inactive DegP S210A were determined by titrating protein against a fixed concentration of the substrate (50 or 25 nM), measuring changes in fluorescence anisotropy, and fitting the data to a hyperbolic binding equation. Each variant was assayed in the absence or presence of one-half molar equivalent of nonfluorescent 18–58. Bars represent the error of fitting.
tightly to dodecamers (as required if substrate binding drives assembly), but apparent $K_D$ values could be similar if this difference was offset by higher energetic costs of active dodecamer assembly.

To assay substrate binding in the absence of cleavage, we used fluorescence anisotropy to monitor the binding of DegP$^{S210A}$ or DegP$^{S210A/Y444A}$ to fluorescently labeled 18–58 (Fig. 2A–D). In this assay, positive cooperativity, a hallmark of allosteric enzymes, can be observed by improved binding to the fluorescent substrate in the presence of subsaturating amounts of nonfluorescent 18–58. We found that DegP$^{S210A/Y444A}$ bound 18–58 more tightly in the presence of 0.5 equivalents of nonfluorescent substrate, but binding under both conditions was weaker than observed for the DegP$^{S210A}$ parent (Fig. 2D) (10). We constructed and assayed two additional variants that destabilize the PDZ1 (L276A) or PDZ2 (L446A) portions of the cage interface. Compared with DegP$^{S210A}$, the DegP$^{S210A/L276A}$ and DegP$^{S210A/L446A}$ variants showed a reduced ability to form substrate-stabilized cages (Fig. S1). Moreover, DegP$^{S210A/L276A}$ and DegP$^{S210A/L446A}$ resembled DegP$^{S210A/Y444A}$ in terms of 18–58 binding (Fig. 2D); in each case, binding was weaker than for wild-type DegP but retained positive cooperativity. These results support our conclusion that cage formation is not required for cooperative substrate binding.

The 18–58 substrate bound more tightly to DegP$^{S210A}$ than to DegP$^{S210A/Y444A}$ (Fig. 2D), but the p23 substrate was cleared with similar apparent $K_m$ by DegP and DegP$^{Y444A}$ (Fig. 2C). These variations may arise because of differences in the sequences of the two substrates, the enzymes used (S210A background for binding assays), or the different assay conditions. For example, the substrate titration experiments were performed using 0.2 μM DegP, whereas half-maximal binding of DegP$^{S210A}$ to a fixed substrate concentration occurred at an enzyme concentration of ~0.6 μM. This threefold change in concentration would increase the stability of the dodecamer relative to the hexamer and could account for the tighter apparent binding in the protein titration assay.

**Binding of Isolated PDZ2 Blocks Assembly and Enhances Substrate Binding.** Because the PDZ2 domain plays an essential role in cage formation, we tested if isolated PDZ2 domains could inhibit cage assembly using the DegP* FRET assay. Indeed, when we assembled donor dye- and acceptor dye-labeled DegPs$^{S210A}$ cages in the presence of 18–58 and then added increasing amounts of the purified PDZ2 domain, the FRET signal was reduced in a dose-dependent manner and eventually reached a baseline level expected for complete cage disassembly (Fig. 3A). By contrast, when cage-defective donor and acceptor dye-labeled DegPs$^{S210A/Y444A}$ trimers were mixed with 18–58, the FRET signal remained low in the absence or presence of PDZ2 (Fig. 3A). Thus, high concentrations of the isolated PDZ2 domain can block DegP cage assembly.

We assayed cleavage of a high concentration of the p23 substrate by wild-type DegP in the presence of 2 mM PDZ2 to prevent cage formation. The cleavage rate in this experiment was ~2.4 min$^{-1}$-enzy$^{-1}$ (Fig. 3B). In the absence of added PDZ2, the cleavage rate was ~1.9 min$^{-1}$-enzy$^{-1}$. These results support our conclusion that cage assembly is not required for high-efficiency DegP proteolysis.

Because mutations that prevent or destabilize PDZ1–PDZ2 interactions between trimers weaken 18–58 binding, we reasoned that binding of the isolated PDZ2 domain might increase the substrate affinity of DegPs$^{S210A/Y444A}$ by mimicking PDZ1–PDZ2 interactions that are normally made in cages. This result was observed, and binding was ~fourfold tighter in the presence of 2 mM PDZ2 (Fig. 3C, Upper). By contrast, addition of 2 mM PDZ2 did not change the affinity of DegP$^{S210A}$ for 18–58 (Fig. 3C, Lower). We interpret this observation as indicating that interactions made by binding of the isolated PDZ2 domain have no net effect because they simply replace comparable interactions in wild-type cages. In support of this model, we found that 2 mM PDZ2 also stabilized 18–58 binding by DegP$^{S210A/L276A}$, which contains a different cage-stabilizing mutation in PDZ2, but did not stabilize binding by DegP$^{S210A/L446A}$, which contains a mutation in PDZ1 and thus cannot interact stably with added PDZ2 (Fig. 3D). The Fig. 3C and D experiments were performed in the presence of 0.5 equivalents of unlabeled 18–58, but similar results were observed without added unlabeled substrate (Fig. S2A and B). Thus, PDZ1–PDZ2 binding results in stronger 18–58 binding to DegP even when the PDZ2 domain is not part of a neighboring trimer in a cage.

**Effects of the PDZ1–PDZ2 Linker on Assembly, Substrate Binding, and Proteolysis.** In each DegP subunit, a linker consisting of residues 357–368 connects the PDZ1 and PDZ2 domains. These linkers were ordered in a crystal structure of a symmetric DegP$_2$ cage but were largely disordered in a structure of an asymmetric DegP$_1$ cage, suggesting that changes in linker conformation provide flexibility in cage geometry (9, 10). Iwanczyk et al. (15) constructed a DegP$^{\text{linker}}$ variant (A357–364) and demonstrated that it eluted as a dodecamer in gel filtration and sedimentation equilibrium experiments without added substrate. We confirmed that DegP$^{\text{linker}}$ chromatographed as a dodecamer (Fig. 4A) and also determined that addition of the 18–58 substrate did not change its elution position. However, the DegP$^{\text{linker}}$ variant eluted as a trimer (Fig. 4A), establishing that the DegP$^{\text{linker}}$ dodecamer, like the wild-type dodecamer, is stabilized by PDZ2-dependent interactions. DegP$^{S210A\text{linker}}$ and DegP$^{S210A}$ bound...
Fig. 4. Variants with shorter or longer PDZ1-PDZ2 linkers show altered assembly but maintain robust proteolytic activity. (A) DegP, DegP-linker, and DegP [linker +20 linker] (25 μM loading concentration) eluted from a Superose 6 gel-filtration column at positions expected for a hexamer, dodecamer, and trimer, respectively. (B) DegP S210A+20 linker (25 μM loading concentration) eluted as a hexamer in gel filtration in the absence and presence of 18-58 (38 μM loading concentration). (C) Concentration dependence of the rates of cleavage of the p23 substrate by DegP-linker and DegP [linker +20 linker] (0.2- and 0.1-μM monomer equations, respectively). The curves are fits to the Hill form of the Michaelis-Menten equation. Error bars are averages ±1 SD (n = 3). (D) Temperature dependence of the rate of cleavage of the p23 substrate (60 μM) by DegP, DegP [linker +20 linker], and DegP [linker +20 linker] (0.5-μM monomer equivalent).

Fig. 5. Assembly of DegP cages is not required for high-temperature cell survival. (A) E. coli encoding wild-type DegP, DegP [linker +20 linker], or DegP [linker +20 linker] at the standard chromosomal locus expressed similar levels of DegP protein. Strains were grown to log phase in LB broth at 37 °C, and equivalent numbers of cells were lysed and separated by SDS/PAGE. DegP was identified by Western blotting using an anti-DegP antibody. A ΔdegP strain expressed no DegP. All strains expressed comparable levels of a cross-reacting lower molecular-weight protein. (B) Strains expressing wild-type DegP, DegP [linker +20 linker], or DegP [linker +20 linker] exhibited similar overnight growth on LB agar plates at 37 and 47 °C. The ΔdegP strain grew normally at 37 °C but was inviable at 47 °C.

Discussion

Previous studies have shown that substrate binding drives assembly of DegP cages and activates proteolysis, suggesting a causal relationship (8-10). However, Spiess et al. (13) and Jomaa et al. (14) showed that deletion of the DegP PDZ2 domain results in trimers that retain some proteolytic activity. Our results confirm this observation in that we found that a trimeric variant, DegP Y444A, degrades a model substrate in a positively cooperative fashion with apparent Km and Vmax values similar to wild-type DegP. The Y444A mutation in PDZ2 affects packing interactions that normally occur in cages between the PDZ2 domains of one trimer and the PDZ1 domains of a neighboring trimer. We also found that addition of high concentrations of the isolated PDZ2 domain could disassemble wild-type cages without eliminating proteolysis. Thus, cage formation is not a prerequisite for proteolytic activation, and the molecular mechanisms that drive these reactions must be different, albeit energetically coupled in the wild-type enzyme.

DegS is a DegP paralog that contains a protease domain and one PDZ domain and functions as a stable trimer (16-18). Substrates bind and activate DegS in a positively cooperative reaction that involves a transition from a proteolytically inactive trimer to an active trimer and is fit well by the concerted Monod-Wyman-Changeux model of allostery (19, 20). Because our results establish that trimeric DegP Y444A also shows positive cooperativity in substrate binding and proteolysis, we assume that this trimer undergoes a similar transition between an inactive conformation that predominates in the absence of substrate and an active conformation that is stabilized by substrate binding.

How is substrate binding normally linked to assembly of DegP cages? From an energetic perspective, we found that the 18-58 18 linker with comparable apparent affinities in the presence of 0.5 equivalents of unlabeled substrate (Fig. 2D).

To determine the effects of lengthening the PDZ1-PDZ2 linker, we inserted 20 residues between residues Q356 and S357 in wild-type and S210A backgrounds. Interestingly, DegP S210A+20 linker was still positively cooperative, but binding in the presence of 0.5 equivalents of unlabeled 18-58 was somewhat weaker than observed for the cages of wild-type DegP or DegP-linker (Fig. 2D).

Although their oligomeric states are apparently fixed, DegP [linker +20 linker] cleaved the p23 substrate in positively cooperative reactions with Hill constants of 1.7 and 3.3, Vmax values of 2.9 and 1.9 min -1 × 10 -1 , and apparent Km values of 0.6 and 2.2 μM, respectively (Fig. 4C). Thus, linker length plays an important role in substrate-dependent cage assembly but plays a relatively modest role in determining the steady-state kinetics of substrate cleavage. Moreover, these results show that preformed hexamers and dodecamers still display positively cooperative interactions with the p23 substrate. To test if temperature caused substantial changes in the proteolytic activities of DegP, DegP Y444A, DegP [linker +20 linker], and DegP [linker +20 linker], we assayed cleavage of a single high concentration of the p23 substrate at 25, 37, and 47 °C (Fig. 4D). In each instance, the cleavage rate increased as the temperature was increased. Thus, all of these enzymes would be expected to be active proteases at heat-shock temperatures.

Cage Assembly Is Not Required for High-Temperature Cell Survival.

To determine how DegP mutations affect biological function, we constructed otherwise isogenic bacterial strains in which the Y444A, Δlinker, and +20 linker mutations were recombined into the chromosomal degP gene. Western blots confirmed that each mutant protein was expressed at a level similar to wild-type DegP (Fig. 5A). To test cell viability, cultures were grown to log phase at 37 °C, and serial 10-fold dilutions were spotted onto LB agar plates and incubated overnight at 37 or 47 °C (Fig. 5B). Strikingly, the degP Y444A, degP +20 linker, and degP [linker +20 linker] strains all grew as well as the wild-type strain at both temperatures (Fig. 5B), even though the first two variants do not assemble into cages, and the third forms permanent dodecameric cages. Thus, reversible cage formation does not seem to be an important determinant of high-temperature growth. By contrast, a strain with an in-frame deletion of the DegP coding sequence (ΔdegP) was inviable at 47 °C but grew normally at 37 °C (Fig. 5B).
substrate binds more tightly to wild-type DegP, which can form cages, than to DegP<sub>Y444A</sub> which cannot form cages. Moreover, addition of high concentrations of the isolated PDZ2 domain increased the 18–58 affinity of the DegP<sub>Y444A</sub> trimer ~fourfold. These results indicate that docking of exogenous PDZ2 domains with the PDZ1 domains of DegP<sub>Y444A</sub> stabilizes a trimer conformation with higher substrate affinity. Thermodynamic linkage then ensures that substrate binding to a trimer will also stabilize tighter binding of its PDZ2 domains to exogenous PDZ2 domains, providing the driving force for substrate-dependent assembly of DegP cages (Fig. 6). An important aspect of this model is that geometric constraints imposed by the structure of the trimer prevent its PDZ2 domains from binding to the PDZ1 domains of the same trimer in a way that mimics trimer–trimer interactions within cages. Although a fourfold stabilization for each trimer is a relatively small value, it could be amplified substantially by the presence of multiple trimers in a cage. Indeed, DegP trimers appear to assemble into a range of different geometrically allowed cage- or bowl-like structures in the presence of different substrates and/or membranes (8–10, 21).

Two potential structural mechanisms might link substrate binding to enhanced PDZ1–PDZ2 interactions. The first mechanism is a direct contact of substrate residues between the cleavage-site degron and C-terminal degron in one trimer with a PDZ2 domain in a neighboring trimer. A contact of this type appeared to be present in one subunit of the 18–58 bound dodecamer (Fig. S3A) (10). The second mechanism is an allosteric change in the structure of PDZ1 upon binding to the C terminus of a substrate or to a PDZ2 domain. Indeed, modest changes in PDZ1 structure are observed between substrate-bound cages and substrate-free structures (Fig. S3B) (9), although it is difficult to know if these conformational changes would affect the binding reactions. We note, however, that allosteric coupling between the peptide-binding site and the opposite face of PDZ domains has been proposed based upon patterns of sequence covariation (22).

In DegP variants, the length of the linker between the PDZ1 and PDZ2 domains of each DegP subunit can modulate the stability of higher oligomers. A DegP mutant with a substantially longer PDZ1–PDZ2 linker formed hexamers in the presence or absence of substrate, whereas a mutant with a very short linker formed dodecamers in the presence or absence of substrate. Both of these linker variants had robust proteolytic activity and showed cooperative activation by substrates. These results reinforce our conclusion that the mechanisms that drive substrate-dependent formation of wild-type cages and result in substrate-dependent proteolytic activation are separable. From an evolutionary perspective, altering the length or flexibility of the PDZ1–PDZ2 linker might provide a way to tune the coupling between substrate binding and the assembly of cages or other oligomers.

Multiple studies have shown that DegP is required for survival of E. coli at high temperatures or when misfolded envelope proteins are overexpressed at lower temperatures (4, 5, 23, 24). Importantly, we found that strains expressing DegP<sub>Y444A</sub> or DegP<sub>Y444A +20 linker</sub> or DegP<sub>Y444A linker</sub> from the chromosome were as effective as wild-type DegP in supporting growth at 47 °C. In vitro, these mutants form stable trimers, hexamers, or dodecamers, respectively, and thus the ability of wild-type DegP to form cages in a reversible substrate-dependent manner or to convert from an inactive hexamer to an active cage does not appear to be an important factor in survival at 47 °C. The DegP<sub>Y444A</sub> DegP<sub>+20 linker</sub>, and DegP<sub>Y444A linker</sub> mutants all displayed robust and cooperative proteolysis in vitro, suggesting that this activity is an important aspect of their biological roles in combating protein-folding stress. Indeed, proteolytically inactive DegP<sub>S210A</sub> did not support growth at 47 °C but supported some growth at 42 °C, as expected from the previous work (13, 25).

Why has DegP evolved the ability to form cages in a substrate-dependent fashion if these cages are not required for biological function as assayed by high-temperature growth? There are several possibilities. First, cages may provide additional fitness and thus a strong selective advantage under stress conditions that we have not tested or are difficult to reproduce in laboratory cultures. Second, if cages provided even a marginal growth benefit at high temperatures or under other conditions, then this small advantage exponentially magnified over thousands of generations could provide a strong selective pressure. For example, cages could provide an alternative proteolysis of improper substrates under some stress conditions, providing a fitness benefit. Third, DegP cages may facilitate additional biological activities. For example, degP<sup>ΔdegE</sup> supports growth at 42 °C better than a ΔdegE allele, a property that has been proposed to result from chaperone activity (13) or sequestration of toxic misfolded proteins (26, 27). Finally, DegP cages may allow more efficient degradation of large protein substrates, even though this activity is not required for high-temperature survival.

Materials and Methods

DegP variants, the 18–58 model substrate, and Δ<sup>fl</sup>18–58 were prepared as described (10). The +20 linker variant carries an additional 20 residues (ASGAGGSEGEGSSEGGTSGAT) between Q356 and S357. The p23 substrate was synthesized by the Massachusetts Institute of Technology Biopolymer Laboratory or CHI Scientific, Inc. and was purified by HPLC; its concentration was determined by absorption at 381 nm (ε = 2,200 M<sup>−1</sup> cm<sup>−1</sup>).

Gel filtration chromatography, FRET, fluorescence anisotropy, and analytical ultracentrifugation experiments were performed as described (10) with minor modifications. Sedimentation velocity experiments were carried out at 20 °C and 30,000 rpm using an An-50 Ti rotor in a Beckman XL-A analytical ultracentrifuge (Biophysical Instrumentation Facility, Massachusetts Institute of Technology). Data were fitted with SEDFIT (28) to a model for continuous sedimentation coefficient distribution with a single floating frictional ratio. Kinetic assays for the p23 cleavage were performed with excitation at 320 nm and emission at 435 nm (cutoff filter at 420 nm) using a SpectraMax M5 microplate reader (Molecular Devices). Each reaction was started with the addition of wild-type DegP or variants and terminated by addition of excess wild-type DegP (~25 μM) to determine fluorescence values associated with complete substrate cleavage.

Different degP mutations were introduced into E. coli strain W3110 (wild-type) using a scarless lambda-RED-mediated recombinengineering method, in which the wild-type degP gene is replaced by two successive recombination steps (29) (details are available in SI Materials and Methods and Table S1). The ΔdegE strain constructed bore an in-frame deletion of only protein-coding sequence. The sequences of all chromosomal degE alleles were confirmed by DNA sequencing (GENEWIZ) following PCR amplification using flanking primers.

For Western blotting, cells expressing DegP variants were grown to log phase (OD<sub>600</sub> ~0.6) in LB at 37 °C. A 1-ml sample of the culture was harvested, resuspended with OD<sub>600</sub>~2 μl of SDS sample buffer, heated at 100 °C for 15 min, and centrifuged at 18,000 × g for 10 min. Samples (5 μl) were loaded onto an SDS-PAGE gel transferred to PVDF. The membrane was sequentially incubated in protein-free 2× Blocking buffer (Thermo Scientific), anti-DegP antibody (30,000:1 dilution), and anti-rabbit IgG-HRP (30,000:1 dilution; GE Healthcare), and then blotted with...
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SuperSignal West Pico reagent (Thermo Scientific). To test viability, cells were at 37 °C grown to OD_{600} = 0.2, and dilutions of 10, 10^2, 10^3, and 10^5 were prepared using LB broth. A 5-μL sample of each dilution was spotted onto LB agar plates and incubated at 37 °C and 42 °C for ~11 h.


ACKNOWLEDGMENTS. We thank D. Barthelme, A. Olivesares, D. Pheasant, A. de Regt, and J. Sohn for helpful discussions and assistance. Anti-DegP antibody was a generous gift from T. Silhavy. Support for this work was provided by National Institutes of Health Grant AI-16892.
