The protein phosphatase 2A functions in the spindle position checkpoint by regulating the checkpoint kinase Kin4

The MIT Faculty has made this article openly available. Please share how this access benefits you. Your story matters.

<table>
<thead>
<tr>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>As Published</td>
<td><a href="http://dx.doi.org/10.1101/gad.1804609">http://dx.doi.org/10.1101/gad.1804609</a></td>
</tr>
<tr>
<td>Publisher</td>
<td>Cold Spring Harbor Laboratory Press</td>
</tr>
<tr>
<td>Version</td>
<td>Author's final manuscript</td>
</tr>
<tr>
<td>Accessed</td>
<td>Wed Jan 02 21:17:50 EST 2019</td>
</tr>
<tr>
<td>Citable Link</td>
<td><a href="http://hdl.handle.net/1721.1/77594">http://hdl.handle.net/1721.1/77594</a></td>
</tr>
<tr>
<td>Terms of Use</td>
<td>Creative Commons Attribution-Noncommercial-Share Alike 3.0</td>
</tr>
<tr>
<td>Detailed Terms</td>
<td><a href="http://creativecommons.org/licenses/by-nc-sa/3.0/">http://creativecommons.org/licenses/by-nc-sa/3.0/</a></td>
</tr>
</tbody>
</table>
The Protein Phosphatase 2A functions in the spindle position checkpoint by regulating the checkpoint kinase Kin4.

Leon Y. Chan and Angelika Amon

David H. Koch Institute for Integrative Cancer Research and Howard Hughes Medical Institute
Massachusetts Institute of Technology, E17-233
40 Ames Street
Cambridge MA 02139
USA

1 To whom correspondence should be addressed.
e-mail: angelika@mit.edu
Abstract

In budding yeast, a surveillance mechanism known as the spindle position checkpoint (SPOC) ensures accurate genome partitioning. In the event of spindle mis-position the checkpoint delays exit from mitosis by restraining the activity of the mitotic exit network (MEN). To date, the only component of the checkpoint to be identified is the protein kinase Kin4. Furthermore, how the kinase is regulated by spindle position is not known. Here, we identify the protein phosphatase PP2A in complex with the regulatory subunit, Rts1, as a component of the SPOC. Loss of PP2A-Rts1 function abrogates the spindle position checkpoint but not other mitotic checkpoints. We further show that the protein phosphatase functions upstream of Kin4, regulating the kinase’s phosphorylation and localization during an unperturbed cell cycle and during spindle position checkpoint activation thus defining the phosphatase as a key regulator of SPOC function.
Introduction

Many cell divisions are polarized, with intrinsic and/or extrinsic cues dictating the directionality of cell division. To ensure that the genetic material is segregated accurately during polarized division, the mitotic spindle must be positioned according to these polarization cues. There is mounting evidence to suggest that this process of spindle positioning is coupled to the cell cycle. In cultured rat kidney cells, a delay in anaphase onset occurs when the metaphase spindle is mis-positioned (O'Connell and Wang 2000). In Drosophila male germline stem cells, cell cycle progression also appears delayed when the spindle is mis-positioned (Cheng, Turkel et al. 2008). Such coupling is suggestive of a feedback mechanism that delays the cell cycle in response to defects in spindle position.

A surveillance mechanism that delays the cell cycle in response to defects in spindle position has been described in the budding yeast, *Saccharomyces cerevisiae*, and is termed the spindle position checkpoint (SPOC) (Yeh, Skibbens et al. 1995). Each cell division of budding yeast is asymmetric and thus inherently polarized. Budding yeast cells divide by forming a bud and hence determine the site of cytokinesis during entry into the cell cycle long before the mitotic spindle is formed. This manner of dividing requires the active positioning of the mitotic spindle along this predetermined division axis, which is known in yeast as the mother-bud axis. The spindle position checkpoint ensures proper spindle position and thus chromosome segregation by delaying exit from mitosis, specifically anaphase spindle disassembly and cytokinesis, until the spindle is correctly positioned along the mother-bud axis (Yeh, Skibbens et al. 1995). The function of the SPOC is most apparent in cells deficient in cytoplasmic microtubule dynamics, guidance and capture. Such cells fail to position the mitotic spindle along the division axis resulting in anaphase spindle elongation in the mother cell (Yeh, Skibbens et al. 1995). Failure of this checkpoint results in the formation of anucleated and multinucleated cells.

The spindle position checkpoint delays exit from mitosis by inhibiting the mitotic exit network (MEN). The MEN is a signal transduction pathway whose activity is controlled by the small GTPase, Tem1 (Shirayama, Matsui et al. 1994). Tem1 regulates a kinase

Several proteins have been identified that regulate Tem1 during the cell cycle and in response to spindle position defects. Tem1 is controlled positively by the bud cortex restricted protein, Lte1 (which resembles a guanine nucleotide exchange factor; GEF) and negatively by the two-component GTPase activating protein (GAP), Bub2-Bfa1 (Shirayama, Matsui et al. 1994; Lee, Hwang et al. 1999; Li 1999; Bardin, Visintin et al. 2000; Bloecher, Venturi et al. 2000; Pereira, Hofken et al. 2000; Geymonat, Spanos et al. 2002). In the absence of \textit{LTE1}, cells fail to activate the MEN and do not exit from mitosis at low temperatures (Shirayama, Matsui et al. 1994). The absence of the GAP leads to inappropriate MEN activation and mitotic exit in cells arrested in mitosis due to activation of the spindle assembly checkpoint (SAC) or the SPOC (Hoyt, Totis et al. 1991; Fesquet, Fitzpatrick et al. 1999; Bardin, Visintin et al. 2000; Bloecher, Venturi et al. 2000; Pereira, Hofken et al. 2000). A single spindle position checkpoint specific regulator of the MEN has also been identified. The protein kinase Kin4 is only required for MEN inhibition in response to spindle position defects (D'Aquino, Monje-Casas et al. 2005; Pereira and Schiebel 2005). Kin4 localizes to the mother cell cortex throughout most of the cell cycle. During anaphase the protein kinase also localizes to the spindle pole body (SPB) that remains in the mother cell (D'Aquino, Monje-Casas et al. 2005; Pereira and Schiebel 2005). In cells with mis-positioned spindles, Kin4 associates with both SPBs where it phosphorylates Bfa1 (Pereira and Schiebel 2005; Maekawa, Priest et al. 2007). This phosphorylation protects the GAP from inhibitory phosphorylation by the Polo kinase, Cdc5, effectively locking Bub2-Bfa1 in an active state thus inhibiting the MEN (Hu, Wang et al. 2001; Geymonat, Spanos et al. 2003; Maekawa, Priest et al.)
How Kin4 itself is controlled during the cell cycle or in response to spindle position defects is not understood.

Here, we identify the protein phosphatase PP2A-Rts1 as a regulator of Kin4 function. PP2A-Rts1 is required for the dephosphorylation of Kin4 during cell cycle entry and to maintain Kin4 in the dephosphorylated state during S-phase and mitosis. Furthermore the phosphatase controls the association of Kin4 with SPBs both during the cell cycle and in response to spindle position defects. The importance of this control is underlined by the finding that PP2A-Rts1 is essential for spindle position checkpoint function but not for other mitotic checkpoints. We propose that PP2A-Rts1 is a SPOC component that facilitates Kin4 localization thereby restraining MEN activity.

Results

**PP2A-Rts1 regulates Kin4 phosphorylation.**

Kin4 is essential for spindle position checkpoint function. To determine how the protein kinase is controlled, we focused on its previously observed cell cycle regulated phosphorylation. Phosphorylated Kin4 can be detected as slower migrating species by SDS-PAGE (D'Aquino, Monje-Casas et al. 2005). Kin4 is phosphorylated in G1, but upon release from a pheromone induced G1-arrest, Kin4 phosphorylation is rapidly lost. The protein remains in a hypophosphorylated state throughout S phase and mitosis, but is rapidly rephosphorylated during exit from mitosis (Figure 1A and (D'Aquino, Monje-Casas et al. 2005)).

Kin4 is in the dephosphorylated state during the stages of the cell cycle when Kin4 activity is needed to inhibit the MEN in response to potential spindle position defects. This correlation prompted us to investigate the significance of Kin4 phosphorylation by identifying phosphatases responsible for dephosphorylating Kin4. We screened mutants defective in known cell cycle associated phosphatases for effects on Kin4 phosphorylation. Kin4 phosphorylation was examined in cells harboring temperature sensitive alleles of CDC14 (cdc14-3), SIT4 (sit4-102), (Wang, Wang et al. 2003), Protein Phosphatase 1 (PP1; glc7-12), and a combination of alleles that renders the Protein
Phosphatase 2A (PP2A) temperature sensitive (\(pph3\Delta, pph21\Delta, pph22-12\); henceforth temperature sensitive PP2A, (Evans and Stark 1997)). We grew the cells to mid exponential phase at permissive temperature (23°C) and followed Kin4 phosphorylation upon shift to the restrictive temperature (37°C). Temperature shift led to a transient accumulation of hyperphosphorylated Kin4 even in wild-type cells, for unknown reasons, which was particularly evident when the ratio of phosphorylated to unphosphorylated Kin4 was determined (Figure 1B, C). After the transient accumulation of hyperphosphorylated Kin4, however, both hyper and hypophosphorylated Kin4 were detected in wild-type and most phosphatase mutants. The \(sit4-102\) mutant displayed hyperphosphorylation after temperature shift, which was progressively lost during incubation at 37°C (Figure 1B, C). Temperature sensitive PP2A mutants showed a persistence of hyper-phosphorylated Kin4 (Figure 1B-C). Though other phosphatases, possibly Sit4, contribute to Kin4 dephosphorylation (see also Figure 3B), we conclude that PP2A is primarily required for Kin4 de-phosphorylation.

Budding yeast PP2A is a heterotrimeric enzyme composed of a single scaffolding subunit, Tpd3, a catalytic subunit, Pph21 or Pph22 and a regulatory subunit, Cdc55 (B type-PP2A) or Rts1 (B’ type-PP2A) (reviewed in (Jiang 2006)). To determine which regulatory subunit is required for Kin4 dephosphorylation, we examined the phosphorylation status of Kin4 in the \(cdc55\Delta\) and \(rts1\Delta\) mutants. In \(cdc55\Delta\) cells, Kin4 phosphorylation status resembled that of wild-type. In contrast, in the \(rts1\Delta\) strain, Kin4 phosphorylation resembled that of the PP2A mutant (Figure 1B, C). We conclude that PP2A-Rts1 is primarily responsible for Kin4 dephosphorylation.

**PP2A-Rts1 is a component of the spindle position checkpoint.**

If dephosphorylation of Kin4 is important for its checkpoint function, mutants in which Kin4 dephosphorylation is impaired should exhibit spindle position checkpoint defects. To test this possibility, we examined how \(PP2A-RTS1\) mutants respond to spindle mis-position. Cells lacking cytoplasmic dynein (\(dyn1\Delta\)) exhibit spindle position defects particularly at low temperature (14°C)(Li, Yeh et al. 1993). As a result, chromosome segregation frequently occurs within the mother cell. This in turn leads to activation of
the SPOC, which causes inhibition of the MEN and cell cycle arrest in late anaphase (“arrested” morphology; Figure 2A). *dyn1Δ* cells with an impaired SPOC fail to delay mitotic exit resulting in anucleated and multinucleated cells (“bypassed” morphology; Figure 2A).

After growth at 14°C for 24 hours, 27% of *dyn1Δ* mutants exhibit the “arrested” morphology and only 10% the “bypassed” morphology (Figure 2B), indicating that the SPOC is functional and that the MEN is inhibited. When the SPOC is inactivated by deletion of *KIN4*, 35% of *dyn1Δ kin4Δ* double mutant cells exhibit the “bypassed” morphology. Deletion of *RTS1* in the *dyn1Δ* mutant also lead to inactivation of the SPOC with 42% of cells showing the “bypassed” morphology (Figure 2B). Importantly, the *rts1Δ* single mutant did not exhibit spindle position defects indicating that PP2A-Rts1 likely does not play a role in the actual positioning of the spindle. The spindle position checkpoint defect was specific to PP2A-B’ as the *cdc55Δ dyn1Δ* mutant displayed no SPOC defect (Figure 2B). We observed a defect in SPOC function in *dyn1Δ pph21Δ pph22-12 pph3Δ* quadruple mutants although this defect was not as pronounced as in the *dyn1Δ rts1Δ* mutant possibly due to the proliferation defect of the quadruple mutant (Figure 2B). Lastly, we found that deletion of *KIN4* did not enhance the SPOC defect of *dyn1Δ rts1Δ* mutants. Taken together, these data indicate that PP2A-Rts1 is a component of the SPOC that likely functions in the same pathway as *KIN4*.

**PP2A-Rts1 functions in the SPOC by controlling Kin4 localization to spindle pole bodies.**

Having established that PP2A-Rts1 is required for SPOC function, we wished to determine whether PP2A-Rts1 acts through Kin4. To this end we examined the effects of deleting *RTS1* on the phenotype associated with *KIN4* overexpression. Overexpression of *KIN4* from the *GAL1-10* promoter terminally arrests cells in anaphase (D’Aquino, Monje-Casas et al. 2005). Deletion of *RTS1* suppressed the lethality caused by *GAL-KIN4* (Figure 3A) indicating that overexpressed Kin4 requires PP2A-Rts1 function to exert its inhibitory effects on mitotic exit. Importantly, deletion of *RTS1* did not suppress the lethal anaphase arrest induced by *GAL-BFA1* (Lee, Hwang et al. 1999; Li 1999)
suggesting that \( rts1\Delta \) is not a general suppressor of anaphase arrest but instead shows specificity towards \( KIN4 \). The rescue of the \( GAL-KIN4 \) lethality by the deletion of \( RTS1 \) was not as complete as that brought about by the deletion of \( BUB2 \), a gene known to function downstream of \( KIN4 \), suggesting that the mechanisms of suppression may not be the same ((D’Aquino, Monje-Casas et al. 2005); Figure 3A).

To determine the relationship between PP2A-Rts1 and Kin4 we examined the effects of deleting \( RTS1 \) on Kin4 phosphorylation, activity and localization. We found that Kin4 is hyper-phosphorylated throughout the cell cycle in the \( rts1\Delta \) mutant (Figure 3B). While we observed a consistent 10 to 15 minute delay in cell cycle progression prior to metaphase spindle assembly, inactivation of \( RTS1 \) did not affect the phosphorylation state of the Bub2-Bfa1 complex, nor the MEN GTPase Tem1, nor the Tem1 activator Lte1, as judged by changes in electrophoretic mobility (Figure 3C – F). These findings suggest that not all SPOC components and its targets are substrates of PP2A-Rts1.

Next we examined whether PP2A-Rts1 affects Kin4 activity, which is essential for checkpoint function (Supplemental Figure 1A). Kin4 kinase activity was not decreased when isolated from \( rts1\Delta \) mutant cells (Supplemental Figure 1B-C) indicating that PP2A-Rts1 does not affect Kin4’s \textit{in vitro} enzymatic activity.

An intact SPOC not only requires Kin4 kinase activity but also binding of the protein kinase to SPBs (Maekawa, Priest et al. 2007). During an unperturbed cell cycle Kin4 localizes to the mother cell cortex. During anaphase, Kin4 also associates with the SPB that remains in the mother cell (mSPB). In cells with mis-positioned anaphase spindles, Kin4 associates with both SPBs. Kin4 mutants that fail to localize to SPBs are checkpoint defective (Maekawa, Priest et al. 2007). Furthermore, ectopically targeting Kin4 to SPBs suffices to delay mitotic exit (Maekawa, Priest et al. 2007). We monitored the ability of Kin4-GFP to associate with the mSPB during anaphase and observed a strong reduction in mSPB localization in \( rts1\Delta \) cells (Figure 4). Whereas Kin4-GFP localized to the mSPB during anaphase in 54% of wild-type cells, Kin4-GFP was found on mSPBs of only 11% of \( rts1\Delta \) anaphase cells (Figure 4C). Furthermore, in the \( rts1\Delta \) mutant cells that showed
Kin4 localization at the mSPB, the signal was weaker (Figure 4A). The loss of Kin4 localization to SPBs was not due to a reduction in KIN4-GFP expression in the rts1Δ mutant, as levels of Kin4-GFP in these cells are similar to that observed in wild-type cells (Figure 4B). The cortical localization and asymmetry of Kin4 was also reduced in rts1Δ cells which was most pronounced in small budded cells (Figure 4A, 5A, D). The basis of this loss in asymmetric localization of Kin4 is currently unclear. Localization of Bub2, Bfa1 and Tem1 was not affected by the deletion of RTS1 (Supplemental Figures 2A-C) indicating that loss of RTS1 function does not alter the overall structure of SPBs nor affects the localization of other SPB-associated MEN proteins. Bud restricted cortical localization of Lte1 was also not affected by deletion of RTS1 suggesting that overall cell polarity and integrity of the bud neck is maintained in the mutant (Supplemental Figure 2D). We conclude that PP2A-Rts1 is required for Kin4 function and association of the protein with the mSPB during anaphase.

**RTS1 is Required for Efficient SPB Loading of Kin4 in response to SPOC activation.**

Having determined that RTS1 was required for Kin4 localization during an unperturbed cell cycle we next investigated whether RTS1 also affected Kin4 loading onto SPBs when the spindle is depolymerized or mis-positioned, situations where the SPOC is active and KIN4 function would be most important. We first examined cells treated with the spindle depolymerizing drug nocodazole. Under these conditions Kin4 associates with SPBs ((Pereira and Schiebel 2005); Figure 5A, B). In contrast, Kin4 association with SPBs was significantly reduced in rts1Δ mutants (Figure 5A-B). In the few cells where Kin4 was detected on SPBs the signal intensity was reduced. This loss of SPB association could not be explained by changes in Kin4 expression as Kin4-GFP protein levels were similar in the two strains (Figure 5C). Similar results were obtained in dyn1Δ cells with mis-positioned spindles. In such cells Kin4 localizes to both SPBs ((Pereira and Schiebel 2005); Figure 5D, E). In the absence of RTS1, Kin4 localization to SPBs was severely impaired (Figure 5D, E).
Overexpression of *KIN4* only partially suppressed the Kin4 SPB loading defect of *rts1Δ* mutants, providing a possible explanation for why deleting *RTS1* suppressed the lethality associated with high levels of Kin4 (Figure 3A). Over-expression of *KIN4* from the *MET25* promoter, which inhibits proliferation (Figure 5F), allowed some Kin4 to associate with SPBs in *rts1Δ* mutants but the number of cells that exhibit Kin4 localization to SPBs was reduced compared to wild-type cells (Figure 5G). Thus in all situations in which Kin4 is known to load onto SPBs (correctly positioned anaphase spindles, spindle depolymerization, mis-positioned anaphase spindles and overexpressed Kin4), we observe a defect in SPB loading in the *rts1Δ* mutant.

In cells with a mis-positioned spindle and an intact SPOC, both Bub2 and Bfa1 localize to both SPBs but Tem1 fails to load onto SPBs. When *KIN4* is deleted, Tem1 loads onto SPBs and thus presumably allows for premature exit from mitosis ((D’Aquino, Monje-Casas et al. 2005); Figure 5J). Consistent with previously observed effects of loss of *KIN4* function on the localization of Tem1 complex components, both Bub2 and Bfa1 localization were unaffected by deletion of *RTS1* but Tem1 was observed to improperly load on to SPBs of cells with mis-positioned spindles (Figure 5H-J). This improper loading of Tem1 onto SPBs of cells with mis-positioned spindles in the *rts1Δ* mutant indicates that *RTS1* is required for normal *KIN4* function.

**RTS1 inhibition of the MEN is spindle position checkpoint specific.**

Our data show that PP2A-Rts1 inhibits the MEN when the SPOC is active. We next wished to determine whether this inhibition was SPOC specific or whether PP2A-Rts1 was a general MEN inhibitor. To address this question we first examined the importance of MEN activity on SPOC and spindle assembly checkpoint (SAC) activity. We constructed a series of hypermorphic alleles of *TEM1* and tested their behavior in response to challenges to both spindle assembly and spindle positioning. We isolated a hypermorphic allele of *TEM1, TEM1-eGFP*. *TEM1-eGFP* suppresses the growth defect of *lte1Δ* cells at 16°C. Other C-terminal tagged alleles of *TEM1* do not display this suppression (Figure 6A). This allele produces about two fold more protein than wild-type *TEM1* (Figure 6B). In addition we examined two overexpression constructs of
TEM1. TEM1 expressed from a multi copy 2micron plasmid expressed approximately six fold more than wild-type TEM1 and TEM1 expressed from the GAL1-10 promoter produced about 50 fold more protein (Figure 6B). We then tested the ability of these TEM1 alleles to hyperactivate the MEN and thus bypass the cell cycle arrests caused by the SAC and the SPOC. All three hypermorphic alleles of TEM1 bypassed the cell cycle arrest caused by spindle position defects to a similar degree (Figure 6C). Cells treated with the spindle poison, nocodazole, activate the SAC which in turn prevents both entry into anaphase and MEN activation in response to microtubule – kinetochore attachment defects (reviewed in (Lew and Burke 2003)). Indeed, deletion of BUB2 leads to inappropriate activation of the MEN and exit from mitosis in cells treated with nocodazole (Figure 6D; (Fraschini, Formenti et al. 1999)). This is evident from examining the ability of cells to form small buds (termed rebudding) in the presence of nocodazole (Figure 6D; (Hoyt, Totis et al. 1991; Li and Murray 1991; Fraschini, Formenti et al. 1999)). Tem1-eGFP failed to bypass the nocodazole-induced cell cycle arrest (Figure 6D) whereas the intermediate strength allele, 2micron-TEM1, displayed an intermediate level of bypass (Figure 6E). The strongest allele, GAL-TEM1, displayed the greatest degree of bypass of the nocodazole induced arrest (Figure 7F).

Having established this series of hyperactive TEM1 alleles, we then compared these alleles to mutants of known SPOC components. Cells lacking RTS1 or KIN4 resembled TEM1-eGFP cells. The SAC was intact in these cells but the SPOC was not ((D'Aquino, Monje-Casas et al. 2005; Pereira and Schieber 2005); Figure 6D). In contrast, bub2Δ cells most resembled GAL-TEM1 cells, exhibiting both SAC and SPOC defects (Figure 6F). This finding indicates that like KIN4, RTS1 is not a general inhibitor of the MEN but only exerts its inhibitory function in the SPOC. Additionally, our results suggest that higher levels of MEN activity are required to bypass the SAC than the SPOC.

Discussion

PP2A-Rts1 is a component of the spindle position checkpoint.
Previous studies identified the protein kinase Kin4 as an inhibitor of MEN activity in response to spindle position defects. How the protein kinase was controlled however, was not understood. Based on the correlation between phosphorylation status and presumptive time of KIN4 function during the cell cycle, we hypothesized that dephosphorylated Kin4 was active and that phosphatases that promoted accumulation of this form would be required for Kin4 function. To test this hypothesis we screened known phosphatases implicated in cell cycle control and identified PP2A and its regulatory subunit Rts1 as being required for Kin4 dephosphorylation. Consistent with the idea that dephosphorylation of Kin4 is indeed required for the protein’s function we found that cells lacking PP2A-Rts1 failed to delay exit from mitosis in response to spindle position defects. Our studies also shed light on the mechanism whereby PP2A-Rts1 restrains the MEN when spindles are mis-positioned. The phosphatase does not appear to affect Kin4 kinase activity but instead promotes its association with SPBs, which was previously shown to be essential for SPOC activity (Maekawa, Priest et al. 2007).

Several lines of evidence indicate that PP2A-Rts1 is not a general inhibitor of the MEN but specifically functions in the SPOC by regulating Kin4 localization. Deletion of RTS1, like deletion of KIN4, leads to a bypass of the spindle position checkpoint arrest but not the nocodazole-induced cell cycle arrest. In contrast, deletion of the MEN inhibitor and target of Kin4, Bub2-Bfa1, leads to a bypass of both checkpoint arrests. The specificity of PP2A-Rts1 towards Kin4 regulation is revealed by the observations that (i) deletion of KIN4 does not enhance the SPOC defect of the dyn1Δ rts1Δ mutant, that (ii) loss of RTS1 function only alters the phosphorylation state and localization of Kin4 and not other MEN components and that (iii) loss of RTS1 function suppresses the lethality of overexpression of KIN4 but not BFA1. Attempts to test whether targeting Kin4 to SPBs was the sole function of PP2A-Rts1 in the spindle position checkpoint failed because a KIN4 allele previously described to constitutively localize to SPBs (KIN4-SPC72(177-622); (Maekawa, Priest et al. 2007)) is not functional in the checkpoint (Supplemental Figure 3).
**PP2A-Rts1 regulation of Kin4 – an additional layer of control in the spindle position checkpoint.**

Based on the localization patterns of the MEN activator Lte1 and MEN inhibitor Kin4, we and others previously proposed that bud restricted Lte1 creates a zone of MEN activation in the bud and mother cell restricted Kin4 generates a zone of MEN inhibition in the mother cell ((Bardin, Visintin et al. 2000; Pereira, Hofken et al. 2000; D’Aquino, Monje-Casas et al. 2005); Figure 6G). Because components of the MEN localize to SPBs, movement of the MEN-bearing SPB into the bud would lead to MEN activation and exit from mitosis. In principle, this division of the cell into mitotic exit restrictive and permissive zones would be sufficient to ensure that exit from mitosis only occurs when one SPB and hence half of the nucleus moves into the bud. Our studies described here show that an additional layer of control exists where PP2A-Rts1 controls the association of Kin4 with SPBs that are located in the mother cell. This additional layer of control could provide temporal control over Kin4, restricting access of the kinase to its target to the time when a spindle is present in cells, specifically mitosis. This hypothesis would be consistent with the observed cell cycle regulation of Kin4 phosphorylation.

We do not yet understand how *RTS1* promotes Kin4 loading onto SPBs. PP2A-Rts1 could mobilize Kin4 at the cortex and facilitate its association with SPBs (Figure 6G). Whether the phosphatase does so by dephosphorylating Kin4 or a Kin4 receptor at SPBs is also not known. PP2A-Rts1 not only affects the ability of Kin4 to associate with SPBs in the mother cell but also its association with the mother cell cortex. In wild-type cells Kin4 is restricted to the mother cell cortex during S-phase and early mitosis. In cells lacking *RTS1* less Kin4 is observed at the cortex and localization is not restricted to the mother cell. It is therefore also possible that PP2A-Rts1 plays an important role in the establishment or maintenance of specific Kin4 localization to the mother cell cortex and that perhaps this localization is a prerequisite for Kin4 to load onto SPBs (Figure 6G). Indeed we observe that Kin4 mutants that fail to associate with the mother cell cortex also fail to associate with SPBs during anaphase (L. Y. C. unpublished observations).
A key question raised by the observation that Kin4 phosphorylation changes during the cell cycle is whether PP2A-Rts1 itself is cell cycle and SPOC regulated or whether it is a yet to be identified kinase(s) whose ability to phosphorylate Kin4 is regulated. Despite intense efforts, we have been unsuccessful in identifying kinases that affect phosphorylation and function of Kin4. Regardless of whether the Kin4 kinase(s) is cell cycle regulated, is there evidence that PP2A-Rts1 activity changes during the cell cycle? We do not detect any changes in Rts1 binding to the scaffolding subunit of PP2A, Tpd3, during the cell cycle (Supplemental Figure 4), arguing against cell cycle regulated changes in PP2A subunit composition. There is however evidence that PP2A-Rts1 localization is cell cycle regulated. The protein localizes to centromeric DNA upon cell cycle entry and remains there until anaphase entry. The phosphatase also localizes to the bud neck during late mitosis where it regulates septin dynamics (L. Y. C. unpublished observations, (Gentry and Hallberg 2002; Dobbelaeere, Gentry et al. 2003)). Rts1 is not detected on SPBs (L. Y. C. unpublished observations, (Gentry and Hallberg 2002)) arguing against a model where Rts1 association with its targets at the SPB is part of SPOC control. Rather we favor the model where Rts1 is required for localization of Kin4 to the mother cell cortex, which is itself a requirement for SPB localization.

**PP2A – a key regulator of chromosome segregation fidelity.**

PP2A has previously been implicated in mitotic checkpoint control. The phosphatase together with its targeting subunit Cdc55 is required for cell cycle arrest upon triggering the SAC (Minshull, Straight et al. 1996). Our studies now show that the same phosphatase but with a different targeting subunit is essential for SPOC-induced cell cycle arrest. PP2A however has many other roles in mitotic progression. In controlling chromosome segregation, PP2A-Cdc55 controls the activity of the protein phosphatase Cdc14 during early anaphase as part of the FEAR network and also potentially controls the activity of the APC/C at the metaphase to anaphase transition (Wang and Burke 1997; Queralt, Lehane et al. 2006; Tang and Wang 2006; Yellman and Burke 2006; Chiroli, Rossio et al. 2007). During meiosis, PP2A-Rts1 controls the chromosome segregation factor Sgo1 (Riedel, Katis et al. 2006). In all instances where mechanisms are at least partly understood, PP2A control involves the targeted action of the phosphatase at specific
locations in the cell. In the case of the SPOC it is possibly the cell cortex, during meiosis the kinetochore and in the case of the FEAR network the nucleolus. It thus appears that the recurring theme in PP2A control of mitosis is regulation by targeted localization.

Acknowledgements:
We are grateful to James Broach, Steve Elledge, Marco Geymonat, Yu Jiang, Elmar Schiebel and Michael Stark for strains and reagents. We thank Frank Solomon, Fernando Monje-Casas and members of the Amon lab for comments on the manuscript. This work was supported by the National Institutes of Health GM056800 to A.A. and a NSF Pre-doctoral Fellowship to L. Y. C.. A. A is also an investigator of the Howard Hughes Medical Institute.

Materials and Methods:
Yeast Strains and Growth Conditions
All strains are derivatives of W303 (A2587) with the following exceptions: A20126 and A21574 are derivatives of DEY100 (A19130) and A20176 is a derivative of Y398 (A19926). All strains are listed in Supplemental Table 1. Lte1-GFP, Tem1-GFP, Rts1-3HA, rts1Δ, Spc42-mCherry and MET-GFP-KIN4 were constructed by standard PCR based methods (Longtine, McKenzie et al. 1998; Hentges, Van Driessche et al. 2005; Snaith, Samejima et al. 2005). kin4T209A was constructed by a two-step gene replacement using the URA3 gene from Kluyveromyces lactis. Growth conditions are described in the figure legends.

Plasmid Construction
The plasmid used to construct the HIS3MX6:MET-GFP-KIN4 allele, pFA6a-HIS3MX6:pMET25-GFP (A1758), was constructed by digesting pFA6a-HIS3MX6:pGAL-GFP (Longtine, McKenzie et al. 1998) and a PCR product containing the MET25 promoter with BglII and PacI and ligating the two fragments together.
**Immunoblot Analysis**

Immunoblot analysis to determine total amount of Kin4-3HA, 3HA-Bfa1, Tem1-3MYC and Lte1-13MYC were performed as described (Bardin, Visintin et al. 2000; Seshan, Bardin et al. 2002; D'Aquino, Monje-Casas et al. 2005). For immunoblot analysis of Kin4-GFP, 3HA-Bub2 and Tem1, cells were incubated for a minimum of 10 minutes in 5% trichloroacetic acid. The acid was washed away with acetone and cells were pulverized with glass beads in 100 μL of lysis buffer (50 mM Tris-Cl pH7.5, 1 mM EDTA, 2.75 mM DTT and complete protease inhibitor cocktail [Roche]) using a bead mill. Sample buffer was added and the cell homogenates were boiled. Kin4-GFP was detected using an anti-GFP antibody (Clontech, JL-8) at 1:1000, 3HA-Bub2 was detected using an anti-HA antibody (Covance, HA.11) at 1:1000 and Tem1 was detected with an anti-Tem1 antibody at 1:1500. Semi-quantitative estimates of relative protein levels of Tem1 were made using ECL Plus (GE Healthcare) and fluorescence imaging. Quantification was performed using NIH Image Quant software. Rts1-3HA was detected using the same anti-HA antibody at 1:1000 and Tpd3 was detected with a rabbit anti-Tpd3 antibody at 1:2000.

**Antibody generation**

An anti-Tem1 antibody was raised in rabbits against the peptide, CKKLTPIEINEIGDPLLKYKHL. The antibody was then affinity purified using immobilized antigen peptides (Covance).

**Fluorescence Microscopy**

Indirect in situ immunofluorescence methods and antibody concentrations for Tub1 and Tem1-13MYC were as previously described (Kilmartin and Adams 1984; D'Aquino, Monje-Casas et al. 2005). For simultaneous visualization of nuclei and the mCherry-Tub1 fusion protein, cells were prepared as described (Monje-Casas, Prabhu et al. 2007) with the following modifications: cells were permeabilized with 1% Triton X-100 for 5 minutes and the cells were resuspended in 1 μg/mL Hoechst 3342. For live cell microscopy, cells were grown in YePAD, harvested, resuspended in SC and immediately imaged using a Zeiss Axioplan 2 microscope and a Hamamatsu OCRA-ER digital.
camera. Deconvolution was performed with Openlab 4.0.2 software. For quantifications, Z-stacks were taken and the localization was determined.

**Spindle position checkpoint assay**

Cells were grown to mid-exponential phase and then incubated at 14°C for 24 hours. Cells fixed, stained for the nuclei and spindle and examined for endpoint morphology. Cells that were anucleated, multinucleated or budded with two nuclei in the mother cell body but with a disassembled spindle were counted as bypassed. Budded cells with two nuclei in the mother cell body with an intact anaphase spindle were counted as arrested.

**Kin4 Kinase Assays**

Kin4 kinase assays were performed as previously (D'Aquino, Monje-Casas et al. 2005) with the following modifications: 30 µL of anti-HA affinity matrix bead slurry (Roche) were used, 3.5 mg of total protein were used per immunoprecipitation, kinase reactions were allowed to run for one hour and the substrate was a mix of 1 µg myelin basic protein and about 5 µg of recombinant MBP-BFA1. MBP-BFA1 was purified as previously described (Geymonat, Spanos et al. 2002). Kinase signal was determined by phosphorimaging and immunoblot signal was determined using ECL Plus (GE Healthcare) and fluorescence imaging. Quantification was performed using NIH Image Quant software.

**Co-Immunoprecipitation assays**

Approximately 12 OD units of cells were collected, washed once with 10mM Tris-Cl pH7.5 and then lysed with glass beads in a bead mill with lysis buffer (50mM Tris-Cl pH7.5, 150mM NaCl, 1% NP-40, 60mM β-glycerolphosphate, 0.1mM sodium orthovanadate, 15mM para-nitrophenylphosphate, 1mM DTT and complete protease inhibitor cocktail [Roche]). 1 µg anti-HA antibody (Covance, HA.11) was added to 900 µg of total protein and incubated for 1 hour. 10 µL of pre-swelled protein-G bead slurry (Pierce) were then added to each sample and agitated for 2 hours at 4°C. Beads were then washed six times with wash buffer (50 mM Tris-Cl pH7.5, 150 mM NaCl, 1% NP-40). Sample buffer was added to the beads that were then boiled.
Figure Legends:

Figure 1. PP2A-Rts1 is necessary for Kin4 dephosphorylation

(A) Wild-type cells expressing a Kin4-3HA fusion (A11779) were arrested in G1 with 5 μg/mL α factor and released into pheromone free media. After 70 minutes, 10 μg/mL α factor was added to prevent entry into a subsequent cell cycle. Cell cycle stage was determined by spindle and bud morphology. Kin4-3HA was monitored by western blot. An asterisk indicates a cross-reacting band with the HA antibody.

(B) Wild-type (A11779), cdc14-3 (A19111), glc7-12 (A19808), sit4-102 (A20176), pph3Δpph21Δpph22-12 (A20126), cdc55Δ (A19804) and rts1Δ (A20187) cells expressing a Kin4-3HA fusion were grow to exponential phase at room temperature and then shifted to 37°C (t=0). Samples were taken at the indicated times to examine Kin4-3HA mobility. Vph1 was used as a loading control.

(C) Quantification of (B). The phosphorylation ratio is the intensity of the slower migrating band divided by the intensity of the faster migrating band.

Figure 2. PP2A-Rts1 functions in the spindle position checkpoint

(A) A dyn1Δ rts1Δ mutant (A21725) carrying a mCherry-Tub1 marker (red) was grown at 14°C for 24 hours. The DNA was visualized by Hoechst staining (blue). Cells with the arrested morphology are budded and have two DNA masses spanned by an anaphase spindle in the mother cell. Cells with the bypassed morphology have no DNA masses or multiple DNA masses but with disassembled spindles or spindles indicative of cell cycle progression after improper mitotic exit.

(B) Wild-type (A2587), dyn1Δ (A17349), dyn1Δkin4Δ (A17351), dyn1Δ rts1Δ (20310), rts1Δ (A20312), dyn1Δ cdc55Δ (A21520), cdc55Δ (A15396), dyn1Δ
pph3Δpph21Δpph22-12 (A21574), pph3Δpph21Δpph22-12 (A19130) and 
dyn1Δrts1Δkin4Δ (A21538) were grown as described in (A). Cells were 
stained for tubulin by indirect immuno-fluorescence and the DNA was stained 
with DAPI. n > 100 cells per sample. Gray bars represent the percentage of 
cells with the arrested morphology and white bars represent the percentage of 
cells with the bypassed morphology. Error bars represent SEM.

Figure 3. PP2A-Rts1 affects Kin4 phosphorylation but not other MEN components

(A) GAL-GFP-KIN4 (A11997), GAL-GFP-KIN4 rts1Δ (A20893), GAL-GFP-
BFA1 (A3487), GAL-GFP-BFA1 rts1Δ (A20892), rts1Δ (A20312), GAL-GFP-
KIN4 bub2Δ (A18792) and bub2Δ (A1863) cells were spotted on plates 
containing either dextrose or galactose and raffinose. The first spot represents 
growth of approximately 3x10^4 cells and each subsequent spot is a 10 fold 
serial dilution.

(B-F) Wild-type and rts1Δ cells expressing (B) Kin4-3HA (A11779 and A20187), 
(C) 3HA-Bfa1 (A4378 and A21540), (D) 3HA-Bub2 (A21921 and A22064), 
(E) Tem1-3MYC (A1828 and A21539) or (F) Lte1-13MYC (A22669 and 
A22669) were analyzed as described in Figure 1A. Vph1, Pgk1 and a cross 
reacting band with the HA antibody (labeled with an asterisk) were used as 
loading controls.

Figure 4. Localization of Kin4 to the mother SPB during anaphase requires RTS1

(A-C) Wild-type (A19900) and rts1Δ (A20918) cells expressing Kin4-GFP and 
mCherry-Tub1 fusion proteins were treated as described in Figure 1A. 
Samples were taken every 15 minutes for 2 hours after release and the cells 
were imaged live. Serial sections spanning the entire cell were collected to 
ensure imaging of all spindle poles. Panels in (A) show deconvolved images 
from 20 serial sections. Kin4-GFP is in green and mCherry-Tub1 is in red. 
Levels of Kin4-GFP for equal ODs of culture are shown in (B). Pgk1 was used
as a loading control. Quantification of Kin4-GFP colocalization with spindle poles is shown in (C). \( n \geq 100 \) for interphase, metaphase and anaphase cells. \( n \geq 40 \) for the rarer telophase cells. Error bars represent SEM.

Figure 5. SPOC induced Kin4 localization to spindle poles requires \( RTS1 \)

(A-C) Wild-type (A19902) and \( rts1\Delta \) (A21732) cells expressing Kin4-GFP and SPC42-mCherry fusion proteins were treated with 1.5 \( \mu \)g/mL nocodazole for 90 minutes. Cells were analyzed as described in Figure 5A. Kin4-GFP is in green and Spc42-mCherry is in red. Quantification of Kin4-GFP colocalization with the SPC42-mCherry marker is shown in (B). \( n \geq 50 \) for each strain. Error bars represent SEM. Levels of Kin4-GFP for equal ODs of culture are shown in (C). Pgk1 was used as a loading control.

(D-E) \( dyn1\Delta \) (A21720) and \( dyn1\Delta rts1\Delta \) (A22878) cells expressing Kin4-GFP and mCherry-Tub1 proteins were grown to exponential phase and then shifted to 14\(^\circ\)C for 24 hours. Cells were collected and analyzed as described in Figure 5A. Quantification of Kin4-GFP colocalization with spindle poles is shown in (E). \( n \geq 50 \). Error bars represent SEM.

(F) Wild-type (A2587) and \( MET\text{-}GFP\text{-}KIN4 \) (A23232) were spotted on SC medium supplemented with 8 mM methionine or SC medium lacking methionine and incubated at 30\(^\circ\)C for 24 hours. The first spot represents growth of approximately 3x10\(^4\) cells and each subsequent spot is a 10 fold serial dilution. This overexpression allele was used instead of \( GAL\text{-}GFP\text{-}KIN4 \) for single cell analysis of the effects of \( KIN4 \) overexpression due to the low viability of \( rts1\Delta \) mutants when grown in raffinose.

(G) \( MET\text{-}GFP\text{-}KIN4 \) (A23358) and \( MET\text{-}GFP\text{-}KIN4 rts1\Delta \) (A23357) cells expressing a mCherry-Tub1 fusion protein were grown and arrested with \( \alpha \) factor in YePAD + 8 mM methionine. After 2.5 hours of arrest, cells were washed and resuspended in SC–\( \alpha \) factor media and the arrest was held for an additional 30 minutes. Cells were then released into pheromone free SC–
MET media. Anaphase cells were counted for co-localization of GFP-Kin4 with the ends of the spindle. n ≥ 100. Error bars represent SEM.

(H-I) dynam1Δ and dynam1Δ rts1Δ cells expressing mCherry-Tub1 and (F) Bfa1-eGFP (A21723 and A21722) or (G) Bub2-eGFP (A21724 and A21725) were analyzed as described in (D) and (E). n ≥ 50. Error bars represent SEM.

(J) dynam1Δ (A12123), dynam1Δ kin4Δ (A12122) and dynam1Δ rts1Δ (A22636) cells expressing a Tem1-13MYC fusion protein were grown as described in (D) and stained for tubulin and Tem1-13MYC by indirect immuno-fluorescence. n ≥ 50. Error bars represent SEM.

Figure 7. The SPOC and SAC are bypassed by differing levels of MEN activation

(A) Wild-type (A2587), lte1Δ (A18591), lte1Δ TEM1-eGFP (A21483), lte1Δ TEM1-GFP (A22567) and lte1Δ TEM1-3MYC (A4365) cells were spotted on YePAD plates and incubated at 30°C and 16°C. Pictures shown represent growth from 2 days for the 30°C condition and 4 days for 16°C. The first spot represents growth of approximately 3x10^4 cells and each subsequent spot is a 10 fold serial dilution.

(B) Whole cell lysates from 0.13 ODs of cells were analyzed by western blot for Tem1 levels in the wild-type (A2587), TEM1-eGFP (A21089), Yep13-TEM1 (A23122) and GAL-TEM1 (A2441) strains. A23122 was grown in SC-LEU for plasmid maintenance and A2441 was pregrown in YePA + 2% raffinose and induced with addition of 2% galactose for 2 hours.

(C) dynam1Δ (A17349), dynam1Δ TEM1-eGFP (A22811), dynam1Δ Yep13 (A23125), dynam1Δ Yep13-TEM1 (A19104) and dynam1Δ GAL-TEM1 (A23117) were grown and analyzed as described in Figures 2A and 2B with the following modifications: strains carrying Yep13 plasmids were pregrown in SC-LEU and then transferred to YePAD medium prior to temperature shift and strains A17349 and A23117 for the experiment in the third panel were pregrown in YePA + 2% raffinose and the GAL promoter was induced two hours prior to temperature shift with the addition of 2% galactose. n ≥ 100 and error bars represent SEM.
(D) Wild-type (A2587), mad1Δ (A928), bub2Δ (A1863), kin4Δ (A17865), rts1Δ (A20312) and TEM1-eGFP (A21089) cells were grown and arrested in G1 as described in Figure 1A and subsequently released into media containing 1.5 µg/mL nocodazole. Additional 0.75 µg/mL nocodazole was added at 220 minutes after release to maintain the metaphase block. Samples were taken every 20 minutes for microscopic analysis. 

(E) YEp13 (A23121), Yep13-TEM1 (A23122) and YEp13 bub2Δ (A23120) cells were grown and analyzed as in 1A and 7D with the following modification: strains were pregrown in SC-LEU to maintain the plasmid and then transferred to YePAD medium prior to pheromone arrest.

(F) Wild-type (A2587), GAL-TEM1 (A2441) and bub2Δ (A1863) cells were grown and analyzed as in 1A and 7D with the following modifications: strains were grown and arrested in YePA + 2% raffinose and the GAL promoter was induced half an hour prior to release with the addition of 2% galactose.

(G) Model for the role of RTS1 in the control of KIN4. See discussion for further details.

Supplemental Figure 1. Loss of RTS1 function does not affect the kinase activity of Kin4

(A) dyn1Δ (A17349), dyn1Δ kin4Δ (A17351) and dyn1Δ kin4T209AΔ (A22736) cells were grown for 24 hours at 14°C. The DNA was visualized by DAPI staining and microtubules were stained by indirect immuno-fluorescence. Gray bars represent the percentage of cells with the arrested morphology and white bars represent the percentage of cells with the bypassed morphology.

(B) Wild-type cells expressing either a Kin4-3HA or a kin4T209A-3HA (kinase dead) fusion (A11779 or A22119) and rts1Δ cells expressing either a Kin4-3HA or a kin4T209A-3HA fusion (A20187 or A22120) were grown to exponential phase and arrested with 1.5 µg/mL nocodazole for 2 hours. Kin4 associated kinase activity (top, Kin4 kinase), immunoprecipitated Kin4-3HA (second row, IPed Kin4), total amount of Kin4-3HA in extracts (third row, input Kin4) and levels of Bfa1 substrate (as monitored by Coomassie stain) added to the kinase reaction
(bottom, Bfa1) is shown. The band that is shown for Kin4 associated kinase activity and total Bfa1 substrate is the first major degradation product of MBP-Bfa1 as described in Maekawa et al., 2007 and was the dominant signal. An asterisk indicates a cross reacting band with the HA antibody.

(C) Normalized activity of (B) was calculated as kinase activity divided by immunoprecipitated Kin4. Relative activity was calculated as normalized activity of the sample divided by the normalized activity of the wild-type sample.

Supplemental Figure 2. Tem1 and its regulators display normal localization in the rts1Δ mutant

(A-C) Wild-type and rts1Δ cells expressing mCherry-Tub1 and (A) Bfa1-eGFP (A21608 and A21729), (B) Bub2-eGFP (A21610 and A21730) or (C) Tem1-GFP (A22556 and A22667) were analyzed as described in Figures 4A and 4C.

(D) Wild-type (A22632) and rts1Δ (A22631) cells expressing Lte1-GFP and mCherry-Tub1 were analyzed as described in Figure 4A. Panels display typical cells and all localization trends were completely penetrant in all cells examined.

Supplemental Figure 3. KIN4-SPC72(177-622) is a loss of function allele of KIN4

(A) dyn1Δ (A17349), dyn1Δ kin4Δ (A17351) dyn1Δ rts1Δ (A20310) dyn1Δ rts1Δ KIN4-SPC72(177-622) (A22062) and dyn1Δ KIN4-SPC72(177-622) (A22063) cells were grown for 24 hours at 14°C. The DNA was visualized by DAPI staining and microtubules were stained by indirect immuno-fluorescence. Gray bars represent the percentage of cells with the arrested morphology and white bars represent the percentage of cells with the bypassed morphology.

Supplemental Figure 4. Rts1 and Tpd3 are associated throughout the cell cycle
(A-B) Wild-type cells expressing an Rts1-3HA (A20757) fusion protein were grown as described in Figure 1A. Rts1-3HA was immunoprecipitated using an HA antibody and the ability to co-immunoprecipitate Tpd3 was examined by western blot with an antibody against Tpd3. Cell cycle progression was monitored by spindle morphology (B).
Supplemental Table 1: Strain List

<table>
<thead>
<tr>
<th>Strain</th>
<th>Relevant genotype</th>
<th>Source</th>
</tr>
</thead>
<tbody>
<tr>
<td>928</td>
<td>MATa mad1Δ::URA3</td>
<td></td>
</tr>
<tr>
<td>1828</td>
<td>MATa TEM1-3MYC</td>
<td></td>
</tr>
<tr>
<td>1863</td>
<td>MATa bub2Δ::HIS3MX6</td>
<td></td>
</tr>
<tr>
<td>2441</td>
<td>MATa ura3::GAL-TEM1::URA3</td>
<td></td>
</tr>
<tr>
<td>2587</td>
<td>MATa ade2-1 leu2-3 ura3 trp1-1 his3-11,15 can1-100 GAL+ psi+</td>
<td></td>
</tr>
<tr>
<td>3487</td>
<td>MATa HIS3MX6::GAL-GFP-BFA1</td>
<td></td>
</tr>
<tr>
<td>4365</td>
<td>MATa lte1Δ::KanMX6 TEM1-3MYC</td>
<td></td>
</tr>
<tr>
<td>4378</td>
<td>MATa 3HA-BFA1</td>
<td></td>
</tr>
<tr>
<td>11779</td>
<td>MATa KIN4-3HA::KanMX6</td>
<td></td>
</tr>
<tr>
<td>11997</td>
<td>MATa HIS3MX6::GAL-GFP-KIN4</td>
<td></td>
</tr>
<tr>
<td>12122</td>
<td>MATa dyn1Δ::URA3 TEM1-13MYC::HIS3MX6 3HA-CDC14 kin4Δ::KanMX6</td>
<td></td>
</tr>
<tr>
<td>12123</td>
<td>MATa dyn1Δ::URA3 TEM1-13MYC::HIS3MX6 3HA-CDC14</td>
<td></td>
</tr>
<tr>
<td>15396</td>
<td>MATa cdc55Δ::KanMx6</td>
<td></td>
</tr>
<tr>
<td>17349</td>
<td>MATa dyn1Δ::URA3</td>
<td></td>
</tr>
<tr>
<td>17351</td>
<td>MATa kin4Δ::KanMX6 dyn1Δ::URA3</td>
<td></td>
</tr>
<tr>
<td>17865</td>
<td>MATa kin4Δ::KanMX6</td>
<td></td>
</tr>
<tr>
<td>18591</td>
<td>MATa lte1Δ::KanMX6</td>
<td></td>
</tr>
<tr>
<td>18792</td>
<td>MATa HIS3MX6::GAL-GFP-KIN4 bub2Δ::HIS3MX6</td>
<td></td>
</tr>
<tr>
<td>19104</td>
<td>MATa dyn1Δ::URA3 Yep13-TEM1</td>
<td></td>
</tr>
<tr>
<td>19111</td>
<td>MATa KIN4-3HA::KanMX6 cdc14-3</td>
<td></td>
</tr>
<tr>
<td>19130</td>
<td>MATa pph21Δ1::HIS3 pph22-12 pph3Δ1::LYS2 ssd1-d2</td>
<td>DEY100¹</td>
</tr>
<tr>
<td>19804</td>
<td>MATa KIN4-3HA:TRP1 cdc55Δ::KanMX6</td>
<td></td>
</tr>
<tr>
<td>19808</td>
<td>MATa trp1-1:glc7-12:TRP1 glc7Δ::LEU2 KIN4-3HA::KanMX6</td>
<td></td>
</tr>
</tbody>
</table>
19900 MATa ura3:mCherry-TUB1:URA3 KIN4-GFP:HIS3MX6
19902 MATa SPC42-mCherry:NatMX6 KIN4-GFP:HIS3MX6
19926 MATa sit4Δ::HIS3 ssd1-d1 Ycp50-sit4-102 (URA/CEN) 
20126 MATa pph21Δ1::HIS3 pph22-12 pph3Δ1::LYS2 ssd1-d2 
KIN4-3HA:KanMX6
20176 MATa sit4Δ::HIS3 ssd1-d1 KIN4-3HA:KanMX6 
Ycp50-sit4-102 (URA/CEN)
20187 MATa KIN4-3HA::KanMX6 rts1Δ::NatMX6
20310 MATa rts1Δ::NatMX6 dyn1Δ::URA3
20312 MATa rts1Δ::NatMX6
20757 MATa RTS1-3HA::KanMX6
20892 MATa HIS3MX6::GAL-GFP-BFA1 rts1Δ::NatMX6
20893 MATa HIS3MX6::GAL-GFP-KIN4 rts1Δ::NatMX6
20918 MATa ura3:mCherry-TUB1:URA3 KIN4-GFP:HIS3MX6 
rts1Δ::NatMX6
21089 MATa TEM1-yEGFP:HIS3MX6
21483 MATa lte1Δ::KanMX6 TEM1-yEGFP:HIS3MX6
21520 MATa cdc55Δ::KanMX6 dyn1Δ::URA3
21538 MATa rts1Δ::NatMX6 kin4Δ::TRP1 dyn1Δ::URA3
21539 MATa TEM1-3 rts1Δ::NatMX6
21540 MATa rts1Δ::NatMX6 3HA-BFA1
21574 MATa pph21Δ1::HIS3 pph22-12 pph3Δ1::LYS 
ssd1-d2 dyn1Δ::NatMX6
21608 MATa ura3:mCherry-TUB1:URA3 BFA1-yEGFP:KanMX6
21610 MATa ura3:mCherry-TUB1:URA3 BUB2-yEGFP:KanMX6
21720 MATa ura3:mCherry-TUB1:URA3 KIN4-GFP:HIS3MX6 
dyn1Δ::URA3
21722 MATa ura3:mCherry-TUB1:URA3 BFA1-yEGFP:KanMX6 
dyn1Δ::URA3 rts1Δ::NatMX6
21723 MATa ura3:mCherry-TUB1:URA3 BFA1-yEGFP:KanMX6 
dyn1Δ::URA3

Y398²
21724  MATa ura3:mCherry-TUB1:URA3 BUB2-yEGFP:KanMX6
dyn1Δ::URA3
21725  MATa ura3:mCherry-TUB1:URA3 BUB2-yEGFP:KanMX6
dyn1Δ::URA3 rts1Δ::NatMX6
21729  MATa ura3:mCherry-TUB1:URA3 BFA1-yEGFP:KanMX6
rts1Δ::NatMX6
21730  MATa ura3:mCherry-TUB1:URA3 BUB2-yEGFP:KanMX6
rts1Δ::NatMX6
21732  MATa SPC42-mCherry:NatMX6 KIN4-GFP:HIS3MX6
rts1Δ::NatMX6
21921  MATa 3HA-BUB2  yFH649-2³
22062  MATa dyn1Δ::URA3 rts1Δ::NatMx6 LEU2:KIN4
-SPC72(177-622):kin4Δ::KL-URA3:Kin4'3'UTR:KanMX6
22063  MATa dyn1Δ::URA3 LEU2:KIN4-SPC72(177-622):
kin4Δ::KL-URA3:Kin4'3'UTR:KanMX6
22064  MATa 3HA-BUB2 rts1Δ::NatMX6
22119  MATa Kin4T209A-3HA:KanMX6
22120  MATa Kin4T209A-3HA:KanMX6 rts1Δ::NatMX6
22556  MATa ura3:mCherry-TUB1:URA3 TEM1-GFP:HIS3MX6
22567  MATa lte1Δ::NatMX6 TEM1-GFP:HIS3MX6
22631  MATa ura3:mCherry-TUB1:URA3 LTE1-GFP:KanMX6
rts1Δ::NatMX6
22632  MATa ura3:mCherry-TUB1:URA3 LTE1-GFP:KanMX6
22636  MATa dyn1Δ::URA3 TEM1-13MYC:HIS3MX6
rts1Δ::NatMX6
22667  MATa ura3:mCherry-TUB1:URA3 TEM1-GFP:HIS3MX6
rts1Δ::NatMX6
22668  MATa LTE1-13MYC:KanMX6
22669  MATa LTE1-13MYC:KanMX6 rts1Δ::NatMX6
22736  MATa KIN4T209A:Kin4-3'UTR:KanMX6 dyn1Δ::URA3
22811  MATa dyn1Δ::URA3 TEM1-yEGFP:HIS3MX6
22878  MATa ura3:mCherry-TUB1:URA3 KIN4-GFP:HIS3MX6
dyn1Δ::HIS3MX6 rts1Δ::NatMX6
23117  MATa dyn1Δ::URA3 ura3::GAL-TEM1::URA3
23120  MATa Yep13 bub2Δ::HIS3MX6
23121  MATa Yep13
23122  MATa Yep13-TEM1
23125  MATa dyn1Δ::URA3 Yep13
23232  MATa HIS3MX6::MET-GFP-KIN4
23357  MATa HIS3MX6::MET-GFP-KIN4 ura3::mCherry-TUB1:URA3
         rts1Δ::NatMX6
23358  MATa HIS3MX6::MET-GFP-KIN4 ura3::mCherry-TUB1:URA3

References:

type 2A protein phosphatase catalytic subunit reveal roles in cell wall
integrity, actin cytoskeleton organization and mitosis." Genetics 145(2):
227-41.

2 Wang, H., X. Wang, et al. (2003). "Interaction with Tap42 is required for the
essential function of Sit4 and type 2A phosphatases." Mol Biol Cell 14(11):
4342-51.


**Figure 2**

(A) Confocal microscopy images showing mCherry-Tub1, Hoechst, merge, and merge + phase for wild-type and various mutant strains.

(B) Bar graph showing the percent cells for different strains: wild-type, dyn1Δ, dyn1Δ kin4Δ, dyn1Δ rts1Δ, rts1Δ, dyn1Δ cdc55Δ, cdc55Δ, dyn1Δ pph3Δ, pph21Δ pph22-12, and dyn1Δ rts1Δ kin4Δ.
A) Dextrose vs. Galactose

GAL-KIN4
GAL-KIN4 rts1Δ
GAL-BFA1
GAL-BFA1 rts1Δ
rts1Δ
GAL-KIN4 bub2Δ
bub2Δ

B) Time (min) vs. % Metaphase vs. % Anaphase

Kin4-3HA
wild-type
rts1Δ

C) Time (min) vs. % Metaphase vs. % Anaphase

3HA-Bfa1
wild-type
rts1Δ

D) Time (min) vs. % Metaphase vs. % Anaphase

3HA-BUB2
wild-type
rts1Δ

E) Time (min) vs. % Metaphase vs. % Anaphase

Tem1-3MYC
wild-type
rts1Δ

F) Time (min) vs. % Metaphase vs. % Anaphase

Lte1-13MYC
wild-type
rts1Δ
A  

Wild-type  
mCherry-Tub1  

G1  
small  
budded  
metaphase  
anaphase  
telophase  

Kernel4-GFP  

B  

wild-type  

0 SPB labeled  
1 SPB labeled  

C  

Percent cells  
0  
20  
40  
60  
80  
100  
wild-type  
rts1Δ  

Interphase cells  
Metaphase cells  
Anaphase cells  
Telophase cells  

Wild-type  

0 SPB labeled  
1 SPB labeled  

rt1Δ  

Kin4-GFP  
Pgk1
Wild-type

\[ \text{Spc42-mCherry} \]

\[ \text{merge} \]

\[ \text{rts1} \Delta \]

\[ 0 \quad 80 \quad 60 \quad 40 \quad 20 \quad 100 \]

Percent cells with Kin4 at SPBs

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ +\text{nocodazole} \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]

\[ \text{wild-type} \]

\[ \text{rts1} \Delta \]

\[ \text{Kin4-GFP} \]