Aneuploid yeast strains exhibit defects in cell growth and passage through START
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ABSTRACT
Aneuploidy, a chromosome content that is not a multiple of the haploid karyotype, is associated with reduced fitness in all organisms analyzed to date. In budding yeast aneuploidy causes cell proliferation defects, with many different aneuploid strains exhibiting a delay in G1, a cell cycle stage governed by extracellular cues, growth rate and cell cycle events. Here, we characterize this G1 delay. We show that 10 of 14 aneuploid yeast strains exhibit a growth defect during G1. Furthermore, 10 of 14 aneuploid strains display a cell cycle entry delay that correlates with the size of the additional chromosome. This cell cycle entry delay is due to a delayed accumulation of G1 cyclins that can be suppressed by supplying cells with high levels of a G1 cyclin. Our results indicate that aneuploidy frequently interferes with the ability of cells to grow and, as many other cellular stresses, entry into the cell cycle.

INTRODUCTION
In most eukaryotes, the decision of whether or not to enter the cell cycle is made in G1 and governed by extracellular and intracellular cues (reviewed in (Turner et al., 2012)). Identifying and characterizing the pathways that regulate this decision is critical not only for understanding normal cell division, but also the abnormal cell divisions that are seen in cancer. The molecular events governing entry into the cell cycle – known as START – are well characterized in budding yeast (Jorgensen and Tyers, 2004). During early G1, the cyclin-dependent kinase (CDK) Cdc28 is activated by the Cln3 cyclin subunit in a manner that is intimately coordinated with growth rate and extracellular signals such as nutrient availability and the presence of mating pheromone (reviewed in
In turn, Cln3-CDKs regulate the transcriptional inhibitor Whi5. During G1, Whi5 localizes to the nucleus where it inhibits the SBF transcription factor complex (Costanzo et al., 2004; de Bruin et al., 2004). This complex (composed of Swi4 and Swi6) and a similar complex MBF (composed of Mbp1 and Swi6), activate the transcriptional program essential for DNA replication and bud formation (Futcher, 2002). Key among SBF’s targets are the two G1 cyclin-encoding genes CLN1 and CLN2 (Cross et al., 1994; Stuart and Wittenberg, 1994). Cln1/2-CDKs trigger S phase and budding.

Cln3-CDKs phosphorylate Whi5 (Costanzo et al., 2004; de Bruin et al., 2004), triggering nuclear export of this transcriptional inhibitor and allowing SBF-dependent transcription to commence and Cln1/2-CDKs to accumulate. This sets in motion a positive feedback loop where Cln1/2 CDKs further phosphorylate Whi5, promoting its nuclear export and hence SBF-mediated transcription of CLN1 and CLN2 (Skotheim et al., 2008). Once cells have accumulated enough Cln-CDK activity, proteins critical for bud formation and initiation of DNA replication are phosphorylated, driving entry into the cell cycle (Bloom and Cross, 2007). Pathways parallel to CLN3 also promote entry into the cell cycle. BCK2 promotes passage through START in parallel to CLN3 by inducing CLN1/2 transcription by an unknown mechanism (Epstein and Cross, 1994; Di Como et al., 1995).

Activation of the Cln3-CDK cascade is controlled by macromolecule biosynthesis rates (reviewed in (Turner et al., 2012)). Cell growth controls cell cycle progression by influencing the critical size, which is the volume at which cells enter the cell cycle (Rupes, 2002). In general, slow-growing cells enter the cell cycle at smaller volumes than cells growing at faster rates (Jorgensen and Tyers, 2004; Ferrezuelo et al., 2012). In other words, a higher growth rate leads to an increase in critical cell size. How growth rate controls the G1 cell cycle machinery is poorly understood. So far, only one mechanism, translational regulation of Cln3, has been described whereby growth rate affects the G1 cell cycle machinery (Polymenis and Schmidt, 1997). However, additional mechanisms must exist that link growth rate to cell cycle entry, as the consequences of interfering with this CLN3 regulatory mechanism are subtle (Polymenis and Schmidt, 1997). Whatever these additional mechanisms are, it is clear that Cln-CDKs must be the target, as modulating Cln-CDK activity affects the critical cell size. For example, overexpression of any of the G1 cyclins (CLN1, CLN2, or CLN3) leads to a “small cell” phenotype, while their deletion causes a “large cell” phenotype (Tyers et al., 1992; Tyers et al., 1993).

We previously constructed a series of haploid budding yeast strains that are disomic for one or two chromosomes (henceforth disomes; (Torres et al., 2007)). All strains were found to exhibit proliferation defects compared to a wild-type strain. Analysis of cell cycle progression of aneuploid yeast strains released from a pheromone-induced G1 arrest showed that most disomes exhibit a delay in CLN2 expression and cell cycle entry (Torres et al., 2007). Here, we further characterize the defects underlying the G1 delay in these strains. We find that most of the disomes (10 of 14) exhibit a growth defect during G1. Entry into the cell cycle was also delayed in 10 of 14 strains, resulting in an increase in critical cell size that correlated with the degree of aneuploidy. Our analyses further show that a defect in G1 cyclin accumulation underlies the G1 delay of disomic strains that can be suppressed by supplying cells with high levels of CLN2. Our results indicate that aneuploidy causes a G1 delay in most strains analyzed by interfering with cell growth and/or the activation of the Cln3-CDK cascade.
RESULTS

G1 is extended in most disomic yeast strains

The 20 disomic budding yeast strains we previously generated (Torres et al., 2007) share a number of phenotypes, prominent among them a delay in entry into S phase after release from an α-factor pheromone-induced G1 arrest. 16 of 20 aneuploid yeast strains exhibited a delay in bud formation and initiation of DNA replication, ranging from 10-20 minutes (Torres et al., 2007). Treatment of cells with pheromone inhibits cell cycle progression but cell growth continues, disrupting the co-ordination of cell growth and division. Thus, pheromone block-release experiments do not allow one to determine whether cell growth or cell cycle entry (or both) is impaired in aneuploid yeast cells.

To characterize the effects of aneuploidy on cell growth and cell cycle entry, we isolated small, unbudded cells using centrifugal elutriation and examined their growth and cell cycle entry properties. We chose two aneuploid yeast strains for this analysis that did not exhibit a G1 delay upon pheromone release (strains disomic for chromosome II and V) and 12 strains that showed cell cycle entry delays after release from an α-factor arrest (strains disomic for chromosome IV, VIII, X, XI, XII, XIII, XIV, XV, XVI, VIII+XIV, XI+XV, and XI+XVI; Supplemental Figure 1). Consistent with our previous studies (Torres et al., 2007), strains disomic for chromosome II and V did not exhibit a cell cycle entry delay as judged by the time to bud formation following isolation of isovolumetric populations of wild-type and disomic cells (Supplemental Figure 1A, C). Cell cycle entry was delayed in all other disomic yeast strains tested, with delays ranging from ~5 min for disome XIII to ~110 min for disome IV. We conclude that most of the disomic yeast strains tested are delayed in cell cycle entry.

Critical cell size is increased in many disomic strains

A delay in cell cycle entry can be caused by a cell cycle defect that culminates in a delay in the accumulation of Cln-CDK activity, slowed growth, or a combination of both. Cell cycle entry delays caused by defects in the cell cycle machinery promoting the G1 – S transition result in an increase in critical cell size (Rupes, 2002). We found that 10 of 14 aneuploid strains analyzed showed an increase in critical cell size, defined by the modal cell volume corresponding to half-maximal budding (Figure 1A-N; Table 1). This increase in critical size ranged from 3 fL for disome VIII+XIV to 22 fL for disome XI+XV. Interestingly, the increase in critical size correlates with the size of the additional chromosome (Figure 1O), suggesting that some aspect of aneuploidy proportional to the amount of additional genetic material contributes to the increase in critical size.

To determine whether the delay in cell cycle entry was proportional to the amount of additional DNA or proportional to other factors dependent on chromosome size such as the amount of yeast gene products produced from the additional chromosomes, we examined the critical sizes of yeast strains containing human DNA carried by yeast artificial chromosomes (YACs). We do not know whether transcription occurs from the human DNA; however, due to the fundamentally different splicing mechanisms in yeast and mammals, any mRNA produced from the YAC is likely to produce few, if any, functional gene products. This allowed us to examine the impact of excess chromosomal DNA on G1 – S phase progression, without excess gene products.

Introduction of an 850 kb, 670 kb or 620 kb YAC did not delay cell cycle entry as judged by budding (Figure 2). We conclude that additional functional yeast
chromosomes, but not chromosome-sized amounts of DNA that do not produce yeast gene products, cause an increase in critical size. This increase in critical size is proportional to the size of the additional chromosome and thus the amount of additional genes, as chromosomes in yeast are densely packed with genes.

**Cell volume accumulation in G1 is slowed in aneuploid yeast strains**

Following elutriated cells as they progress through G1 also allowed us to assess the effects of disomy on cell growth during G1, monitored by cell volume accumulation. This was of interest because slow growth can delay entry into the cell cycle. For example, cells lacking Sfp1, a transcription factor required for ribosome biogenesis (Fingerman *et al.* 2003), exhibit a severe G1 delay (Jorgensen *et al.*, 2002; Hoose *et al.*, 2012). *sfp1Δ* cells obtained by centrifugal elutriation take a longer time to enter the cell cycle than wild-type cells as judged by bud formation (Supplemental Figure 2A; (Jorgensen and Tyers, 2004; Hoose *et al.*, 2012)). Cell volume measurements during G1 show that *sfp1Δ* cells have a cell volume accumulation rate ~50% of wild-type when G1 time points (<20% budded) were fit to an exponential function, and a critical size smaller than wild-type, consistent with other reports (Supplemental Figure 2B-D; (Jorgensen *et al.*, 2002; Hoose *et al.*, 2012)).

To determine whether a growth defect contributes to the G1 delay observed in aneuploid cells, we examined rates of volume increase of aneuploid and euploid cells during G1. We isolated small, unbudded cells using centrifugal elutriation and monitored cell volume as a function of time. Of the 14 aneuploid strains analyzed, we found 10 exhibited a defect in volume accumulation (Figure 3; Supplemental Figure 3). This is easily seen when size distributions of wild-type and aneuploid cells of the same volume are compared with size distributions after growth in rich medium for 60 minutes (Figure 3B, D, F, H).

To determine the growth rate during G1, we fit the plots of volume increase vs. time to exponential functions, restricting this analysis to time points in which less than 20% of the cells in the population were budded. Growth rates were reduced in 10 of 14 aneuploid strains; this reduction is best observed when the growth rate is shown as a percentage of the wild-type control (Figure 3I, Table 1). Three strains (disomes IV, XVI and XI+XVI) showed growth rates of less than 75% of wild-type (Figure 3I, red); 7 strains (disomes V, VIII, XI, XII, XIII, XIV, and XI+XV) exhibited growth rates between 75-90% of wild-type (Figure 3I, yellow); four strains (disomes II, X, XV, and VIII+XIV) grew as well as wild-type cells (Figure 3I, green). The decreases in growth rates of aneuploid strains were significant (p=0.0007, paired Student’s t-test); however, the extent of the growth defect did not correlate with the size of the additional chromosome (Supplemental Figure 3K).

The growth properties of disome XVI cells are particularly notable, as the delay in S phase entry observed in this strain is entirely due to a defect in cell volume accumulation. Critical cell size was not affected in disome XVI, yet budding was delayed for almost 40 minutes (Table 1; Figure 1K; Supplemental Figure 1K). Cell volume measurements showed that growth was impaired in disome XVI cells (Figure 3G, H), providing an explanation for the delay in bud formation. It is possible that the additional copy of *CLN2* located on chromosome XVI masks any cell cycle defect, as G1 cyclin levels are rate-limiting for cell cycle entry (Futcher, 1996). In summary, our results
indicate that most aneuploid strains analyzed show a reduced growth rate in G1. In
contrast to the increased critical size observed in aneuploid cells, the severity of the cell
volume accumulation defect is not correlated with the amount of additional DNA
(Supplemental Figure 3K). These findings suggest that gene-specific effects, and not
general features of aneuploidy, are responsible for the cell volume accumulation defect
seen in the disomic strains.

Decreased growth rates in aneuploid cells are not due to gross amino acid
biosynthesis defects

Our data show that aneuploid yeast strains exhibit both growth defects and cell
cycle entry delays. We decided to first characterize the growth defect in more detail. To
determine whether the G1 growth defect was due to a lack of amino acids, we measured
pools of free intracellular amino acids in aneuploid cells. sfp1Δ cells were analyzed for
comparison as this mutant is able to synthesize amino acids but grows slowly due to
defects in ribosome biogenesis (Fingerman et al., 2003).

To measure intracellular amino acid pools, we grew cells in medium
supplemented with the essential amino acids for these strains (lysine, leucine, tryptophan,
and methionine). Levels of the amino acids alanine, asparagine, aspartate, glycine,
histidine, isoleucine, phenylalanine, proline, serine, threonine, tyrosine and valine as well
the intermediates of the TCA cycle were quantified using time-of-flight mass
spectrometry. Intracellular amino acid pools were not lower in disomic strains IV, VIII,
XI, XV, or XVI (Figure 4), indicating that amino acid abundance (at least of the amino
acids measured) was not affected in aneuploid strains. In fact, disome IV showed
consistently higher levels of most amino acids (except aspartate and isoleucine) and TCA
cycle intermediates. Increased amino acid levels in disome IV cells may be related to the
significant proliferation defect of this disome as this phenotype is also seen in the
sfp1Δ strain, which proliferates slowly (Figure 4R).

Consistent with the conclusion that free amino acids are not limiting in aneuploid
cells is the observation that aneuploid cells do not exhibit a starvation response
(Supplemental Figure 4). GCN4 encodes a transcription factor that controls the
expression of 30 amino acid biosynthetic genes (Hinnebusch, 2005). Its abundance is
translationally regulated; upon amino acid starvation, GCN4 translation is increased
(Hinnebusch, 2005). We monitored a GCN4-LacZ reporter construct (Hinnebusch, 1985)
by LacZ activity in the absence or presence of amino acid starvation induced by the
addition of 3-amino-1,2,4-triazole (3-AT), a competitive inhibitor of an intermediate step
in histidine synthesis. In the absence of 3-AT, all disomes tested (IV, VIII, XI, XV, and
XVI) showed similar levels of LacZ activity to the euploid control (Supplemental Figure
4, gray bars). In the presence of 3-AT, disomic cells exhibited an increase in LacZ
activity due to GCN4-LacZ translational up-regulation consistent with the euploid control
(Supplemental Figure 4, white bars). Therefore, we conclude that the disomes analyzed
do not exhibit a starvation response under normal growth conditions and are not defective
in eliciting a starvation response. Thus, the slower growth rate seen in aneuploid cells is
not the result of limiting amounts of amino acids, but is likely due to decreased rates of
biomass production.
Effects of disomy XVI on translation

Next we examined whether defects in translation are responsible for the growth defects observed in the disomic yeast strains. For this analysis we chose disome XVI as this strain exhibits one of the most dramatic growth defects of the disomes, yet does not show a cell cycle entry delay as judged by critical cell size measurements. $^{35}$S-methionine incorporation studies did not reveal any significant differences between wild-type and disome XVI cells (data not shown), yet we were able to detect defects in $^{35}$S-methionine incorporation in sfp1Δ cells (data not shown), which exhibit a severe growth defect ((Jorgensen et al., 2002; Hoose et al., 2012; Supplemental Figure 2). Although a decrease in growth rate of disome XVI was detected by cell volume measurements (Figure 3; Table 1), changes were not evident when translation was examined using polysome profiling (Figure 5A). These results indicate that this disomic strain does not suffer from dramatic translation defects.

Since disome XVI cells did not show major translation defects by polysome profiling or $^{35}$S-methionine incorporation, we measured translation efficiency in this strain to determine whether translation efficiencies of individual genes were altered. Consistent with previous studies (Torres et al., 2007; Torres et al., 2010), we found that disome XVI cells have twice the amount of mRNA of genes located on chromosome XVI as wild-type cells (Figure 5B), and these additional mRNAs are translated as judged by their association with ribosomes (Figure 5C). Importantly, we observed no major shifts in relative translation efficiency between wild-type and disome XVI cells (Figure 5D). In summary, our data indicate that many disomic yeast strains exhibit a subtle growth defect that is detected only by cell volume measurements. As all chromosomes are efficiently translated in disome XVI cells, we conclude that the decreased growth rate of this disome is not due to major translational defects or changes in translation efficiencies of individual genes.

Aneuploid cells exhibit delays in the transcriptional program governing the G1 – S transition

Most disomic yeast strains are defective in cell cycle entry, having a cell volume larger than that of wild-type at the time of budding (Figure 1; Table 1). We next wanted to determine the basis for this defect. The genetic program governing entry into the cell cycle culminates in the expression of the G1 cyclins CLN1 and CLN2 (Dirick et al., 1995). To determine whether the increased cell volume of disomic cells at the time of budding was due to a defect in cyclin expression, we examined CLN2 mRNA levels in disomes IV, XI and XV, which have larger critical sizes than wild-type cells (Figure 1B, F, J). Small, unbudded cells were isolated by centrifugal elutriation and CLN2 mRNA levels were measured as a function of cell volume. These analyses showed that transcription of CLN2 occurred at a larger cell volume in the three disomic strains tested than in wild-type cells (Figure 6A, B, D). Maximal CLN2 transcription occurred at 42 fL, 48-50 fL and 43 fL in disome IV, XI and XV cells, respectively, whereas CLN2 levels were maximal in wild-type cells at a size of 26-35 fL. We conclude that CLN2 transcription occurs at a larger cell volume in disome IV, XI and XV strains. This finding is consistent with our previous studies of CLN2 expression in disomic strains released from a pheromone-induced G1 arrest, which also revealed delays in CLN2 transcription (Torres et al., 2007).
SBF and MBF, two transcription factor complexes with overlapping specificities, control transcription of approximately 200 genes important for events in late G1 and S phase (Ferrezuelo et al., 2010). SBF and MBF targets are cell cycle regulated, peaking at the G1 – S transition (Futcher, 2002). To determine whether the entire G1 – S transcriptional program was delayed in aneuploid cells with a G1 delay, we analyzed the expression of three SBF targets (CLN1, CLN2, and PCL1) and three MBF targets (POL1, CLB5, and RNR1) in disomes XI and XV as a function of cell volume. Peak expression of all SBF and MBF targets tested occurred at a larger cell volume in both disomic strains (Figure 6C-H), suggesting that SBF and MBF activation is delayed in these aneuploid strains.

Xbp1 is a stress-induced transcriptional repressor that can inhibit transcription of G1 cyclins, delaying cell cycle entry under stressful conditions (Mai and Breeden, 1997, 2000). Deletion of XBP1 had no significant effect on the critical size of wild-type or disome VIII and XI cells (Supplemental Figure 5), suggesting that inhibition of cyclin expression by Xbp1 is not causing the G1 delay seen in the disomes. We conclude that the SBF and MBF transcription program is delayed in disomic yeast cells, and this delay is not caused by Xbp1-mediated inhibition of G1 cyclin expression.

The SBF and MBF transcriptional program is delayed, suggesting aneuploidy interferes with the G1 – S transition upstream of CLN2 expression; however, this does not exclude the possibility of aneuploidy also interfering with entry into S phase at a step following CLN1/2 expression. To address this possibility, we examined whether overexpression of CLN2, which is known to accelerate entry into the cell cycle (reviewed in (Turner et al., 2012)), suppresses the increase in critical size of cells disomic for chromosome IV, VIII or XI. CLN2 was overexpressed from the galactose-inducible GAL1-10 promoter for 1 hour and then small, unbudded cells were isolated by centrifugal elutriation. Because cells were grown in medium supplemented with raffinose and galactose as the carbon source, euploid cells expressing wild-type levels of CLN2 budded at a smaller critical size of 29 fl (Figure 7A). As expected, overexpression of CLN2 reduced the critical size of cells: wild-type cells overexpressing CLN2 exhibited a critical size of 22 fl (Figure 7B; Supplemental Table 1). Overexpression of CLN2 suppressed the cell cycle entry defect of disomes IV, VIII, and XI (Figure 7B), with critical sizes of 19, 20, and 25 fl compared to critical sizes of 34, 44, and 50 fl, respectively, for those not overexpressing CLN2. We conclude that the increase in critical size in aneuploid cells is due to a delay in CLN2 expression.

**Whi5 nuclear exit is delayed in aneuploid cells**

SBF, the transcription factor complex that promotes CLN1/2 expression, is composed of Swi4 and Swi6 and is negatively regulated by the transcriptional inhibitor Whi5. Whi5 binds to Swi4/6 at the promoters of genes required for the G1 – S transition, inhibiting transcription (Costanzo et al., 2004; de Bruin et al., 2004). In G1, Cln3-CDKs phosphorylate Whi5, promoting its export from the nucleus (Costanzo et al., 2004; de Bruin et al., 2004). In wild-type cells, Whi5-GFP exits the nucleus just before budding. In three disomes tested (IV, XI, and XV), Whi5-GFP protein levels were not dramatically altered and Whi5-GFP nuclear exit occurred, as in wild-type, just before budding (Figure 8). However, both budding and Whi5-GFP nuclear exit were delayed compared to the euploid control (Figure 8; Supplemental Table 1). A fourth aneuploid strain, disome XVI,
exhibits a growth defect but not a critical size increase (Figure 1K; Supplemental Figure 1K; Table 1); as expected, Whi5 nuclear exit also occurred just before budding in this strain, and Whi5-GFP levels were not dramatically altered. We conclude that the relationship between Whi5 nuclear exit and budding is not altered in aneuploid cells; however, the entire program of Whi5 nuclear exit, SBF/MBF activation, and budding is delayed in the disomic strains analyzed.

To further investigate the misregulation of START control in disomic cells, we examined the consequences of WHI5 deletion in disomes IV, VIII and XI. The critical size of wild-type cells is 39 fL, but when WHI5 is deleted the critical size decreases to 30 fL (Figure 8F). Deletion of WHI5 did not suppress the critical size increase in aneuploid cells; it affected wild-type and disomic cells similarly, decreasing the critical size by 6-13 fL (Figure 8F-I; Supplemental Table 1). Together, our data indicate that the timing of Whi5 nuclear exit is delayed in the disomic strains analyzed, but time of budding is also perturbed through parallel pathways. Perhaps disomy IV, VIII and XI interfere with the positive feedback loop regulating Whi5 nuclear export, alter SBF/MBF activity, or affect parallel pathways.

Accumulation of Cln3 is delayed in disome VIII and XI cells.

Whi5 export from the nucleus is promoted by Cln3-CDKs. To determine whether lower Cln3 protein levels could be responsible for the delayed exit of Whi5 from the nucleus in the disomic yeast strains, we measured Cln3-13Myc levels in elutriated cells. Cln3 protein levels fluctuated significantly during G1 in wild-type cells. Cln3 protein levels were low in small G1 cells, accumulated as cells reached a size of 21 fL, and declined shortly before cells entered the cell cycle (Figure 9A). This fluctuation in protein levels was not due to changes in CLN3 RNA levels (Figure 9A), indicating that posttranscriptional mechanisms are responsible for Cln3 protein fluctuations. The CLN3-13MYC allele is hypermorphic, causing a decrease in critical size from 38 fL to 31 fL in wild-type cells (Figure 9A). This is most likely because the C-terminal tag stabilizes the protein by interfering with degradation of Cln3, which is mediated by the PEST sequences located in the C-terminus of the protein (Tyers et al., 1992). Thus, the fluctuations in untagged Cln3 are likely to be, if anything, more dramatic.

Overall Cln3 levels were comparable between wild-type and disomic strains, but Cln3 protein levels peaked at a larger volume in both disome VIII and XI cells as compared to wild-type (Figure 9, Supplemental Figure 6). This finding indicates that Cln3 translation and/or stability is affected in these disomic strains. The effect of the CLN3-13MYC allele on the critical sizes of the two disomic yeast strains was variable. The basis of this variability is presently unclear (Figure 9B; Supplemental Figure 6A). We conclude that the delay of START in disome VIII and XI cells is at least in part due to a delay in the accumulation of Cln3 protein.

Aneuploidy impairs cell cycle entry in parallel to BCK2

Cln3-CDK activity is not essential for cell cycle entry, indicating that parallel pathways exist that activate Cln1/2-CDKs (Cross, 1988; Nash et al., 1988). Bck2 is one such factor; cells lacking BCK2 are large and exhibit a delay in CLN1 and CLN2 transcription (Di Como et al., 1995). In addition, deletion of BCK2 causes lethality or severe growth retardation in cells lacking CLN3 (Epstein and Cross, 1994; Di Como et
al., 1995). To determine whether aneuploidy interferes with cell cycle entry through BCK2, we examined the critical size of disomic cells lacking BCK2. If aneuploidy inhibits cell cycle entry through mechanisms that function in parallel to BCK2, we expect disomes lacking this gene to exhibit an increased critical size. If aneuploidy delays G1 by disrupting BCK2 function, the effects of combining aneuploidy and a BCK2 deletion should not be additive.

Small, unbudded cells were isolated by centrifugal elutriation and budding was measured as a function of volume. In wild-type cells, deletion of BCK2 caused an increase in critical size of about 11% (Figure 10A; Supplemental Table 1). Deletion of BCK2 had a greater effect on the disomic strains than the wild-type strain, causing increases in critical size ranging from 29%-46% in the disomes tested (Figure 10B-E; Supplemental Table 1). A greater increase in critical size in the disomes upon deletion of BCK2 suggests that disomic strains depend more on the BCK2 pathway for cell cycle entry than do wild-type cells. This conclusion is consistent with our data suggesting that the Cln3-CDK/Whi5/SBF pathway is impaired in disomic cells. Together, our results indicate that aneuploidy affects entry into the cell cycle in at least two ways: aneuploidy elicits a growth defect and causes a cell cycle entry delay that is due to a delay in Cln3 accumulation.

Many progeny of triploid meioses exhibit a G1 delay

The strategy we employed to create the disomic yeast strains is limited in that only low complexity aneuploidies (gains of one or two chromosomes) can be obtained (Torres et al., 2007). To determine whether yeast strains with more complex aneuploidies also exhibit a G1 delay, we induced triploid strains to undergo meiosis. In such meioses progeny are generated with a range of karyotypes: haploid, diploid, and aneuploidies ranging from 1N+X to 2N-X. Nearly all aneuploid strains created in this manner are highly unstable (Pavelka et al., 2010; St Charles et al., 2010; Sheltzer et al., 2011), but, we reasoned, could be cultured for brief periods of time to determine whether they also exhibit a G1 delay.

The viability of spores obtained from triploid meioses was very low, as expected (Parry and Cox, 1970; St Charles et al., 2010). The few colonies that did grow up exhibited a wide range of proliferation rates. Based on our previous studies, we assume that progeny obtained from these meioses with wild-type or near wild-type proliferation rates are either euploid or harbor aneuploidies of chromosomes with little impact on cell proliferation, such as chromosome I (Torres et al., 2007), whereas slow-growing progeny harbor aneuploidies of (presumably multiple) chromosomes that cause significant growth impairment.

DNA content analysis revealed that many of the progeny of triploid meioses exhibit a G1 delay (Figure 11). This G1 delay correlated remarkably well with proliferation rates as observed by growth on solid medium (Figure 11). Spores 1-1, 1-2 and 1-3 obtained from the triploid parent 1 grew very well and exhibited a DNA content profile very similar to that of the parent (Figure 11B-E). Spores 1-4, 1-5, 1-6 and 1-7 grew poorly and showed an increase in G1 cells (Figure 11F-I). Similar results were obtained with progeny from triploid parent 2. The percentage of cells in G1 was very low in exponentially growing cultures of this strain, the reason for which is currently unknown. It was nevertheless evident that in the aneuploid progeny, the G1 delay
correlated with growth ability. Spore 2-1 grew well and exhibited a cell cycle distribution close to that of the parent (Figure 11K); cultures of spores that grew poorly (spores 2-2, 2-3, 2-4, 2-5 and 2-6) showed an increase in G1 cells (Figure 11L-P). We conclude that many aneuploid yeast strains obtained as offspring of triploid meioses also exhibit a G1 delay.

DISCUSSION

Here, we investigate the basis of the G1 delay observed in many disomic yeast strains to understand how aneuploidy alters the regulatory networks controlling the G1 – S phase transition. Two aspects of G1 regulation appear to be affected: cell growth and entry into the cell cycle. Thus, the disomic yeast strains are unique in that they grow slowly, yet, unlike most slow-growing mutants, they must reach a larger critical size to pass through START.

Most of the disomic strains show a growth defect during G1 as measured by cell volume accumulation (Figure 3; Supplemental Figure 3). This growth defect is not nearly as dramatic as that seen in mutants impaired in ribosome biogenesis or protein synthesis (Supplemental Figure 2; (Jorgensen et al., 2002; Hoose et al., 2012)), but nevertheless, the defect is readily observed by cell volume measurements. However, it should be noted that we did not detect any defects in global protein synthesis as judged by polysome profiles or 35S-methionine incorporation (Figure 5A; C.G. unpublished observations). These observations raise the possibility that the disomic cells are defective in cell volume accumulation but not protein synthesis, which would imply that these aneuploid cells must be denser than euploid cells. We favor the idea that the defect in cell volume accumulation reflects an overall defect in macromolecule biosynthesis, but the defect is too subtle to be detected by methods such as 35S-methionine incorporation and polysome profiling.

Our investigation of the basis for the cell volume accumulation defect of the disomic yeast strains indicates that it does not stem from diminished amino acid pools (Figure 4) or a constitutively active amino acid starvation response (Supplemental Figure 4), nor from shifts in translation efficiency (Figure 5A). Rather, we propose that macromolecule biosynthesis and/or other aspects of cell growth are hampered in aneuploid cells. One explanation for this growth defect is that particular genes present on the additional chromosome(s) in aneuploid cells may be causing slow growth. For instance, additional copies of key regulators of growth processes such as macromolecule biosynthesis, vesicle fusion, protein sorting and/or energy production could lead to a slowing of growth. In this scenario, the reason for the growth defect is different in each aneuploid strain. It is also possible that slow growth is caused by a feature common to all aneuploidies, such as an increase in the number of gene products due to the extra chromosome(s). Prior studies have shown the additional chromosomes to be actively transcribed and translated in the disomic strains (Torres et al., 2007; Torres et al., 2010); therefore, it is possible that an increased burden on the transcription and translation machinery hampers cell growth. This scenario predicts that the growth defect correlates with the amount of additional translation that occurs in the disomes. This is not what we observe, as the growth defect in the disomes does not correlate with the size of the additional chromosome (Supplemental Figure 3K). Therefore, we favor the idea that
imbalances in individual proteins critical for cell growth hamper cell volume accumulation in the disomic strains.

The delay in cell cycle entry (as judged by a delay in bud formation) observed in the disomes was more pronounced than the growth defect, which led us to investigate this phenotype in more detail. Of the 14 aneuploid strains studied, 10 showed increased critical size. The following observations points towards a common origin of the cell cycle entry defects observed in the disomes:

1. The cell cycle entry delay correlated with the size of the additional chromosome (Figure 1; Table 1).
2. All disomes analyzed, namely disomes IV, XI and XV, exhibit a delay in \textit{CLN2} mRNA accumulation (Figure 6A, B, D);
3. The increase in critical size of all disomes analyzed (disomes IV, VIII and XI) is suppressed by high levels of \textit{CLN2} (Figure 7).
4. Whi5 exit from the nucleus was delayed in all disomes studied (disomes IV, XI, and XV; Figure 8).
5. The accumulation of Cln3 is delayed in all disomes analyzed (disomes VIII and XI).

If the detailed characterization of a subset of disomic strains is representative of all disomic strains exhibiting a cell cycle entry defect, our data point towards Cln3 accumulation being affected by aneuploidy.

How does aneuploidy interfere with Cln3 accumulation and/or other aspects of cell cycle entry? Are these G1 processes affected by changes in copy number of specific proteins or features common to all disomies? Consistent with the former idea are the critical size increases observed in disomes VIII, XIV and VIII+XIV. Whereas the single disomes exhibit increases in critical size of 8 fL (disome VIII) and 5 fL (disome XIV), the double disome shows only a 3 fL increase in critical size compared to wild-type cells (compare Figures 1D, I, N). This finding suggests that the cell cycle entry delay in the individual disomes is caused by a defined number of imbalanced gene products that can be suppressed by providing a surplus of gene products located on another chromosome. In this model, the correlation between increased critical size and size of the additional chromosome would be explained by genes affecting cell cycle entry being distributed more or less evenly across the yeast genome. (Moriya et al., 2006) have identified all yeast genes whose increased dosage affects cell proliferation. It will be interesting to determine, which of these when present at two copies affect cell cycle entry.

Although single gene imbalances could contribute to the delay in cell cycle entry, our data also indicate that a response to a common feature of disomy is responsible for the observed cell cycle entry delay observed in many disomic strains. The critical size increase in the disomes correlates with the size of the additional chromosome (Figure 1O) and all disomes analyzed appear to show a similar defect in the molecular pathway governing START (see above). An increase in the amount of DNA is not responsible for the G1 delay in the disomes as YACs containing human DNA do not cause an increase in critical size. A previous study suggested that increasing the number of SBF binding sites delays cell cycle entry because they titrate Cln3 molecules (Wang et al., 2009). We analyzed the distribution of SBF binding sites in the yeast genome as published in (Ferrezuelo et al., 2010) and found them to be evenly distributed across the genome (data...
It is thus possible that Cln3 titration by SBF binding sites contributes to the G1 delay observed in the disomes. While an increase in SBF binding sites could contribute to the G1 delay in the disomes, our analyses point towards delayed Cln3 protein accumulation as the major defect in at least disomes VIII and XI. We do not yet know whether CLN3 translation and/or Cln3 stability is affected in the aneuploid strains. Protein synthesis rates affect CLN3 translation (Polymenis and Schmidt, 1997) and so aneuploidy could, via its effects on macromolecule biosynthesis, affect Cln3 synthesis. Aneuploidy could also affect Cln3 stability. Recently the two ubiquitin ligases SCF-Cdc4 and SCF-Grr1 have been identified as being responsible for regulating Cln3 stability (Landry et al., 2012). It will be interesting to determine whether aneuploidy affects Cln3 ubiquitination by these ligases.

Many stresses cause a G1 arrest or delay (Herrero et al., 2003). Aneuploidy has been suggested to cause proteotoxic and energy stress in yeast and mouse cells (Torres et al., 2007; Torres et al., 2010; Tang et al., 2011). These or other stresses elicited by aneuploidy could elicit a transient G1 delay by interfering with Cln3 accumulation. Indeed, proteotoxic stress induced by heat shock causes a G1 delay (Johnston and Singer, 1980), which, like the G1 delay seen in the disomic yeast strains, can be suppressed by CLN2 overexpression (Rowley et al., 1993). Whether and how aneuploidy and other cellular stresses antagonize Cln3 accumulation is an important question that remains to be addressed.

Is the G1 delay that we observe in many of the disomes also seen in other models of aneuploidy? Our study of the progeny of triploid meioses suggests that many of the poorly growing and thus presumably highly aneuploid yeast strains show a significant G1 delay that is detectable in exponentially growing cells. A study of aneuploid fission yeast cells created by triploid meioses showed that these cells also exhibit a delay in G1 during germination (Niwa et al., 2006), but the cause of the G1 delay is not known. Mouse embryonic fibroblasts trisomic for either chromosome 1, 13, 16 or 19 (Williams et al., 2008) and trisomy 21 human fibroblasts (Segal and McCoy, 1974) exhibit cell proliferation defects, but the basis of this proliferation defect has not been explored. Thus, whether a conserved effect on cell cycle entry exists in aneuploid cells of different species remains to be established. However, a common transcriptional response reminiscent of the budding yeast environmental stress response (indicative of cellular stress and slowed proliferation) is observed in aneuploid yeast, Arabidopsis, mouse and human cells (Sheltzer et al., 2012). It is tempting to speculate that aneuploidy, like many other cellular stresses, impacts cell proliferation by delaying cells in G1. Determining the molecular mechanisms underlying the effects of aneuploidy on progression through G1 will be an important aspect of understanding how aneuploidy impacts cell physiology.

**MATERIALS AND METHODS**

**Yeast strains, plasmids, and growth conditions**

All yeast strains are derivatives of W303 and are described in Supplemental Table 2. Yeast strains were generated and manipulated as described previously (Guthrie and Fink, 1991). The GCN4-LacZ construct (Hinnebusch, 1985) was integrated at the URA3 locus. Cells were grown at 30°C in YEP or in synthetic medium containing G418 at 0.2 mg/ml, supplemented with 2% raffinose, 2% raffinose + 2% galactose, or 2% glucose (YEPD).
Elutriation
Cells were grown in 1-2 L of synthetic medium supplemented with 2% raffinose at 30°C. Cells were collected by centrifugation and resuspended in 30 ml YEP without sugar. Cells were sonicated to break clumps and kept at 4°C for the duration of the elutriation. A Beckman elutriation rotor JE 5.0 (Beckman Coulter, Brea, CA) was chilled to 4°C and equilibrated with YEP (no sugar) at 2400 rpm. Cells were loaded into the rotor at a pump speed of approximately 14 ml/min and allowed to equilibrate for 15-20 minutes at a rotor speed of 2400 rpm and a pump speed of approximately 14 ml/min. The pump speed was increased until small, unbudded cells exited the elutriation chamber. One liter was collected at that pump speed (about 20 ml/min for most strains). Pump speed settings varied slightly among experiments as the disomic strains have different cell volumes. Elutriated cells were concentrated using a filtration apparatus and resuspended in YEP supplemented with 2% glucose at 30°C.

Cell volume determination
Cell volume was measured using a Beckman Coulter Multisizer 3 (Beckman Coulter). 40-100 µl of culture was diluted into Isoton II solution (Beckman Coulter) to give <10% saturation. 50,000 particles were sized for each sample. Particles smaller than 3 µm were excluded to decrease background signal and decrease the chances of counting dead cells. Cell volume distributions were smoothed with a rolling window of 3 bins. Modal population values were used for the plots. The critical size of a strain is defined as the modal cell volume at half-maximal budding.

Ribosome profiling and mRNA-seq
Ribosome profiling was performed as described (Ingolia et al., 2009), with subtraction of three prominent ribosomal rRNA species performed as in (Brar et al., 2012). In short, cells were treated with 100 µg/ml cycloheximide to freeze ribosomes, then flash-frozen, and subjected to mixermill lysis. Clarified extract was treated with RNAse I and subjected to sucrose gradient fractionation. The 80S/monosome fraction was harvested and RNA was extracted and size-selected for ribosome footprints (28-30 nt). These footprints were converted to a sequencing library, which was sequenced using an Illumina GAII machine (Illumina, San Diego, CA). Sequences were aligned to the W303 genome and quantified as described (Ingolia et al., 2009). For translation efficiency (TE) analyses, calculations were only done on genes with 128 summed counts among mRNA and footprint samples, with TE = (ribosome footprint RPKM)/(mRNA RPKM). RPKM = reads per kilobase million, a normalizing unit for gene length and total sequencing counts per lane. mRNA-seq was also performed as described (Ingolia et al., 2009). In short, flash-frozen cells were lysed in hot acid phenol and total RNA was extracted. polyA selection was performed to isolate mRNA, which was fragmented to 40-60 nt species, which were converted to a sequencing library, sequenced and analyzed in parallel with ribosome footprint samples.

Polyribosome profile analysis
Polysomes were prepared as described (Clarkson et al., 2010). Briefly, 250 ml cultures were grown in YEPD at 30°C to an OD<sub>600</sub> = 0.5. Cycloheximide was added to a final concentration of 0.1 mg/ml for 3 minutes. Cells were pelleted by centrifugation and lysed
by vortexing with zirconia/silica beads in 1x PLB (20 mM HEPES-KOH [pH 7.4], 2 mM magnesium acetate, 100 mM potassium acetate, 0.1 mg/ml cycloheximide, 3 mM DTT) and treated with RNAsin Plus RNase inhibitor (Promega, Fitchburg, WI). Lysates were clarified by centrifugation and 25 A260 units were resolved on 11 ml linear 10%-50% sucrose gradients in 1x PLB by centrifugation in a Beckman SW41 rotor (Beckman Coulter) for 3 h at 35,000 rpm.

**Metabolomic analysis**

All strains were grown in shake flasks at 30°C and 250 rpm in defined minimal medium containing per liter (adapted from (Verduyn et al., 1992)): 5 g KH2PO4, 0.5 g MgSO4*7H2O, 2 g monosodium glutamate, 1.5 mg EDTA, 4.5 mg ZnSO4*7H2O, 0.3 mg CoCl2, 1 mg MnCl2*4H2O, 0.3 mg CuSO4*5H2O, 4.5 mg CaCl2*2H2O, 3 mg FeSO4*7H2O, 0.4 mg NaMoO4*2H2O, 1 mg H3BO3, 0.1 mg KI. Filter-sterilized vitamins were added separately to a final concentration per liter of: 0.005 mg biotin, 0.1 mg Ca-pantothenate, 0.1 mg nicotinic acid, 2.5 mg inositol, 0.1 mg pyridoxine, 0.02 mg p-aminobenzoic acid, 0.1 mg thiamine. The medium was buffered with 100 mM KH-phthalate at a pH of 5. Due to the genetic markers in the strains, the medium was supplemented with the following compounds per liter: 0.031 g lysine, 0.02 g uracil, 0.24 g leucine, 0.08 g adenine, 0.02 g tryptophan, 0.021 g methionine, 0.2 g geneticin (G418). Cells were grown to an OD600 between 0.7 and 1.5 when metabolism of 1 ml culture was arrested by quenching in -40°C methanol + 10 mM ammonium acetate. After centrifugation at -9°C (3 min, 5000 rpm) the samples were stored at -80°C. Intracellular metabolites were extracted by incubation in 75% ethanol + 10 mM ammonium acetate for 3 min at 80°C. The supernatant was dried using a vacuum centrifuge. The dried extracts were derivatized with TBDMS (N-tert-butyldimethylsilyl-N-methyltrifluoroacetamide) for quantification. The metabolites were separated by gas chromatography and injected to a time-of-flight mass spectrometer as described in (Ewald et al., 2009).

**β-galactosidase activity assay**

Cell extracts and activity measurements were performed as described (Dever, 1997). Cells were grown in synthetic medium lacking histidine and supplemented with 2% glucose. Amino acid starvation was induced by the addition of 3-amino-1,2,4-triazole (3-AT), a competitive inhibitor of an intermediate step in histidine synthesis, to a final concentration of 100 mM.

**Northern blot analysis**

Cells were grown at 30°C in synthetic medium supplemented with 2% raffinose, elutriated as described and released into the cell cycle. Time points were taken until after the critical size had been reached. Total RNA was isolated as described (Cross and Tinkelenberg, 1991) and Northern blots were performed as described (Hochwagen et al., 2005). Blots were probed for CLN2 and the loading control ACT1.

**Real-time RT-PCR**

Approximately 2–4 OD600 units of culture were pelleted by centrifugation at 4°C (3000 rpm, 2 min). The pellet was resuspended in 1 ml cold DEPC-treated water, transferred to a 2 ml microfuge tube and repelleted by centrifugation at 4°C (3000 rpm, 2 min). The
pellet was flash-frozen in liquid nitrogen and stored at -80°C. To extract total RNA, ~200 µl of glass beads, 400 µl TES buffer (10 mM Tris [pH 7.5], 10 mM EDTA, 0.5% SDS) and 400 µl acid phenol:chloroform (pH 4.5) were added to the cell pellet and the tubes were vortexed for 45 minutes at 65°C. The phases were separated by centrifugation, and the top phase was transferred to a new tube containing 1 ml of 120 mM sodium acetate in ethanol to precipitate RNA at 4°C. Precipitates were collected by centrifugation and resuspended in 100 µl DEPC-treated water. Total RNA was further purified using the RNeasy Mini kit (Qiagen, Hilden, Germany), including DNase treatment, according to the manufacturer’s instructions. cDNA was synthesized from 750 ng total RNA using the SuperScript III First-Strand Synthesis SuperMix kit (Invitrogen, Grand Island, NY) with random hexamer primers according to the manufacturer’s instructions. Real-time qPCR reactions were run using the SYBR Premix Ex Taq Perfect Real Time kit (TaKaRa Bio, Otsu, Japan) and a Roche LightCycler 480 (Roche, Penzberg, Germany) according to manufacturer’s instructions. qPCR primers are listed in Supplemental Table 3.

Western blot analysis
Approximately 2-4 OD_{600} units of culture were pelleted by centrifugation at 4°C (3000 rpm, 2 min). Cells were incubated at 4°C in 5% trichloroacetic acid for at least 10 minutes. Cell pellets were washed once with acetone and dried. Cells were lysed with glass beads in 100 µL of lysis buffer (50 mM Tris-Cl at pH 7.5, 1 mM EDTA, 2.75 mM dithiothreitol [DTT], complete protease inhibitor cocktail [Roche, Mannheim, Germany]) with a bead mill and samples were boiled in 1X SDS loading buffer. Following sodium dodecyl sulfate-polyacrilamide gel electrophoresis (SDS-PAGE) and transfer of proteins to a nitrocellulose membrane, proteins were detected with the following antibodies. Whi5-GFP was detected using a mouse anti-GFP antibody (JL-8 epitope, Clontech, Mountain View, CA) at a dilution of 1:1000. Cln3-13Myc was detected using a mouse anti-Myc antibody (9e10 epitope; Covance, Princeton, NJ) at a dilution of 1:600. Pgg1 was detected using a mouse anti-Pgg1 antibody (Invitrogen, Carlsbad, CA) at a 1:10,000 dilution. Kar2 was detected using a rabbit anti-Kar2 antibody (kindly provided by Mark Rose) at a dilution of 1:400,000. Quantification was performed using ImageQuant software.

Fluorescence microscopy
For Whi5-GFP imaging, cells were fixed in 4% paraformaldehyde/3.4% sucrose for 15 minutes and then resuspended in 0.1 M potassium phosphate (KPi pH 6.4)/1.2 M sorbitol/1% Triton. Cells were then resuspended in 0.05 µg/ml DAPI in KPi/sorbitol and imaged using a Zeiss Axioplan 2 microscope (Carl Zeiss, Göttingen, Germany) and a Hamamatsu OCRA-ER digital camera (Hamamatsu, Hamamatsu, Japan).

Flow cytometric analysis
Analysis of DNA content by flow cytometry was performed as previously described (Hochwagen et al., 2005).

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Mai, B., and Breeden, L. (2000). CLN1 and its repression by Xbp1 are important for efficient sporulation in budding yeast. Mol Cell Biol 20, 478-487.


Table 1. Critical Sizes and Growth Constants of Disomic Cells

<table>
<thead>
<tr>
<th>Strain</th>
<th>Critical Size (fL)*</th>
<th>% of WT</th>
<th>Exponential Growth Constant, k (min⁻¹)*</th>
<th>% of WT</th>
</tr>
</thead>
<tbody>
<tr>
<td>Dis II</td>
<td>35 (38)</td>
<td>92%</td>
<td>0.006144 (0.005882)</td>
<td>104%</td>
</tr>
<tr>
<td>Dis IV</td>
<td>53 (39)</td>
<td>136%</td>
<td>0.004667 (0.006939)</td>
<td>67%</td>
</tr>
<tr>
<td>Dis V</td>
<td>33 (40)</td>
<td>83%</td>
<td>0.006101 (0.007391)</td>
<td>83%</td>
</tr>
<tr>
<td>Dis VIII</td>
<td>46 (38)</td>
<td>121%</td>
<td>0.006339 (0.007433)</td>
<td>85%</td>
</tr>
<tr>
<td>Dis X</td>
<td>40 (35)</td>
<td>114%</td>
<td>0.006894 (0.006931)</td>
<td>99%</td>
</tr>
<tr>
<td>Dis XI</td>
<td>55 (40)</td>
<td>138%</td>
<td>0.005967 (0.006944)</td>
<td>86%</td>
</tr>
<tr>
<td>Dis XII</td>
<td>53 (38)</td>
<td>139%</td>
<td>0.004160 (0.005239)</td>
<td>79%</td>
</tr>
<tr>
<td>Dis XIII</td>
<td>40 (40)</td>
<td>100%</td>
<td>0.004738 (0.005839)</td>
<td>81%</td>
</tr>
<tr>
<td>Dis XIV</td>
<td>41 (36)</td>
<td>114%</td>
<td>0.006343 (0.007126)</td>
<td>89%</td>
</tr>
<tr>
<td>Dis XV</td>
<td>52 (39)</td>
<td>133%</td>
<td>0.005213 (0.005098)</td>
<td>102%</td>
</tr>
<tr>
<td>Dis XVI</td>
<td>36 (38)</td>
<td>95%</td>
<td>0.005558 (0.007879)</td>
<td>71%</td>
</tr>
<tr>
<td>Dis XI + XV</td>
<td>60 (38)</td>
<td>158%</td>
<td>0.006480 (0.007471)</td>
<td>87%</td>
</tr>
<tr>
<td>Dis XI + XVI</td>
<td>54 (37)</td>
<td>146%</td>
<td>0.005835 (0.008001)</td>
<td>73%</td>
</tr>
<tr>
<td>Dis VIII + XIV</td>
<td>42 (39)</td>
<td>108%</td>
<td>0.006196 (0.006460)</td>
<td>96%</td>
</tr>
<tr>
<td>Diploid</td>
<td>70 (38)</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
</tbody>
</table>

* Values in parentheses are the haploid wild-type results for each experiment
Figure 1: Cells carrying an extra chromosome delay entry into the cell cycle by increasing the critical size for budding.

Wild-type ([A-N], A11311, filled circles) and disomes (open circles) II ([A], A6865), IV ([B], A12687), V ([C], A14479), VIII ([D], A13628), X ([E], A21986), XI ([F], A13771), XII ([G], A12694), XIII ([H], A21987), XIV ([I], A13979), XV ([J], A12697), XVI ([K], A12700), XI+XV ([L], A12691), XI+XVI ([M], A12699), and VIII+XIV ([N], A15615)
were grown at 30°C in selective medium supplemented with 2% raffinose. Small, unbudded cells were isolated by centrifugal elutriation and released into the cell cycle at 30°C in YEPD. Time points were taken every 15 minutes to measure cell volume and the percentage of budded cells. (O) Correlation between the critical size for budding and the size of the extra chromosome in the disome. The critical size (shown as percent of wild-type critical size for each experiment) is positively correlated with the size of the extra chromosome ($r^2 = 0.41$, $p = 0.019$). Disome XII was excluded from the correlation analysis due to variations in rDNA copy number, which preclude determination of the exact chromosome size.
**Figure 2:** Yeast artificial chromosomes carrying human DNA do not cause an increase in critical size.

Wild-type (A11311, filled circles), YAC-2 (A17392, 850 kb, open triangles), YAC-3 (A17393, 670 kb, open circles), and YAC-4 (A17394, 620 kb, open squares) cells were grown at 30°C in selective medium supplemented with 2% raffinose. Small unbudded cells were isolated by centrifugal elutriation and released into the cell cycle at 30°C in YEPD. Time points were taken every 15 minutes to measure cell volume and the percentage of budded cells.
Figure 3: Cells carrying an extra chromosome show a decrease in growth rate during G1.

(A-H) Wild-type ([A-H], A11311, black), disome IV ([A,B], A12687, red), disome XIV ([C,D], A13979, red), disome XV ([E,F], A12697, red), and disome XVI ([G,H], A12700, red) cells were grown at 30°C in selective medium supplemented with 2% raffinose. Small unbudded cells were isolated by centrifugal elutriation and released into the cell cycle at 30°C in YEPD. Time points were taken every 15 minutes to monitor cell
volume. G1 time points (less than 20% budded cells in the population) were fitted to an exponential function to determine growth parameters. (B, D, F, H) Cell size distributions for elutriated wild-type (black) and disomic cells (red) of the same volume and after 1 hour of growth in YEPD medium.

(I) Distribution of growth constants for disomes II (A6865), IV (A12687), V (A14479), VIII (A13628), X (A21986), XI (A13771), XII (A12694), XIII (A21987), XIV (A13979), XV (A12697), XVI (A12700), XI+XV (A12691), XI+XVI (A12699), and VIII+XIV (A15615), are shown as the percent of the wild-type (A11311) growth constant for each experiment. Disomes IV, XVI, and XI+XVI (red) show growth rates <75% of wild-type; disomes V, VIII, XI, XII, XIII, XIV, and XI+XV (yellow) show growth rates 75-90% of wild-type; disomes II, X, XV, and VIII+XIV (green) show growth rates 90-110% of wild-type.
Figure 4: Amino acids and organic acid levels are not altered in aneuploid cells.

(A-R) Metabolomic analysis of wild-type (A11311), sfp1Δ (A3009), and disomes IV (A12687), VIII (A13628), XI (A13771), XV (A12697), and XVI (A12700). Cells were grown in synthetic medium supplemented with the necessary nutrients for growth (see Materials and Methods). Metabolites were extracted, separated by gas chromatography and quantified by time-of-flight mass spectrometry. Error bars represent standard error of the mean (n≥5 for metabolites [A-Q]; n=4 for doubling time [R]).

(S) Overview of TCA cycle.
Figure 5: Translational efficiency is not altered in disome XVI.
(A) Polysome profiles of wild-type (A22361) and disome XVI (A12700) cells.
(B-D) Analysis of translation in wild-type (A11311) and disome XVI (A12700) cells.
(B) The log$_2$ ratio of disome XVI to wild-type mRNA RPKM values is shown with the x-axis representing genes in rank order by chromosomal location. Genes included have 128 raw counts between the wild-type and disomic strains.
(C) The log$_2$ ratio of disome XVI to wild-type ribosome footprint RPKM values is shown with the x-axis representing genes in rank order by chromosomal location. Genes included have 128 raw counts between the wild-type and disomic strains.
(D) Translation efficiencies [TE= log$_2$ (mRNA RPKM/footprint RPKM)] were calculated for each gene with at least 128 summed raw counts between mRNA and footprints in both the disomic and wild-type strains. These TE values are represented as a histogram.
Figure 6: Transcription of CLN2 and other SBF and MBF targets is delayed in disomes IV, XI and XV.

Disomic and wild-type cells were grown at 30°C in selective medium supplemented with 2% raffinose to mid-log phase. Small unbudded cells were isolated by centrifugal elutriation and released into the cell cycle at 30°C in YEPD. Samples were taken to measure RNA levels and cell volume.

(A, B) CLN2 RNA levels were examined in wild-type (A11311), disome IV (A12687) and disome XI (A13771) cells. ACT1 was used as a loading control. (B) Quantification of...
CLN2 levels in (A) normalized by ACT1.
(C-H) RNA levels of the SBF targets CLN1 (C), CLN2 (D), and PCL1 (E) and of the MBF targets POL1 (F) RNRI (G), and CLB5 (H) were determined by real-time RT-PCR in wild-type ([C-H], A11311, filled circles), disome XI ([C-H], A28266, open circles), and disome XV ([C-H], A27930, open triangles). Transcript levels were normalized to ACT1 levels. Dashed lines indicate the critical sizes (modal volume at half-maximum of budding) for each strain.
Figure 7: Overexpression of CLN2 suppresses the critical size increase in disomes IV, VIII and XI.

Wild-type ([B], A25475, filled circles), disome IV ([B], A25477, open circles), disome VIII ([B], A25476, open triangles), and disome XI ([B], A25474, open squares) cells carrying a GAL-CLN2 construct and wild-type ([A], A11311, filled circles), disome IV ([A], A12687, open circles), disome VIII ([A], A13628, open triangles), and disome XI ([A], A13771, open squares) cells lacking the construct were grown to mid-log phase in selective medium supplemented with 2% raffinose and induced for one hour with 2% galactose. Small, unbudded cells were isolated by centrifugal elutriation and released at 30°C in YEP medium containing 2% galactose + 2% raffinose. Samples were taken every 15 minutes to determine cell volume and the percentage of budded cells.
Figure 8: Nuclear exit of Whi5 is delayed in disomes IV, XI, XV and XVI, but deletion of WHI5 does not suppress the S phase entry delay of disomes IV, VIII and XI.

(A-E) Wild-type ([A], A25877), disome IV ([B], A25879), disome XI ([C], A31515), disome XV ([D], A31516), and disome XVI ([E], A25880) were grown at 30°C in selective medium supplemented with 2% raffinose. Small unbudded cells were isolated
by centrifugal elution and released into the cell cycle at 30°C in YEPD. Time points were taken every 15 minutes to monitor budding, Whi5-GFP localization and cell volume. Bud = budding; Cyt = cytoplasmic Whi5.

(F-I) Wild-type ([F], A25479, open circles), disome IV ([G], A25481, open circles), disome VIII ([H], A25480, open circles), and disome XI ([I], A25482, open circles) with WHI5 deleted and wild-type ([F], A11311, filled circles), disome IV ([G], A12687, filled circles), disome VIII ([H], A13628, filled circles), and disome XI ([I], A13771, filled circles) cells with functional WHI5 were grown in selective medium supplemented with 2% raffinose. Small unbudded cells were isolated by centrifugal elution and released at 30°C in YEPD. Time points were taken every 15 minutes to determine cell volume and the percentage of budded cells.

(J) Wild-type (A25877), disome IV (A25879), disome XI (A31515), disome XV (A31516), and disome XVI (A25880) cells were grown at 30°C in YEPD and Whi5-GFP protein levels were detected by Western blot analysis. Disome XV (A31516) contains one copy of Whi5-GFP and one untagged copy of Whi5, as WHI5 is located on chromosome XV. Wild-type cycling cells (A2587) were used as a no-tag control.
Figure 9: Cln3 accumulation is delayed in disome XI.

Wild-type ([A,C], A33692) and disome XI ([B,C], A33694) cells were grown at 30°C in selective medium supplemented with 2% raffinose. Small unbudded cells were isolated by centrifugal elutriation and released into the cell cycle at 30°C in YEPD. Time points were taken every 15 minutes to determine the percentage of budded cells, cell volume and Cln3 protein levels. In separate experiments, wild-type (A) and disome XI (B) were
elutriated as described above and samples were taken every 15 minutes to determine the percentage of budded cells, cell volume and \textit{CLN3} RNA levels by RT-PCR. \textit{CLN3} protein and RNA levels are presented as fold-change from the first time point and are plotted as a function of cell volume to demonstrate that Cln3 protein but not \textit{CLN3} RNA levels fluctuate significantly during G1. Cln3 protein levels were normalized to Pgi1 and \textit{CLN3} RNA levels were normalized to \textit{ACT1}. Wild-type cycling cells (A2587) were used as a no-tag control for the Western blots. The arrow underneath the blot marks the critical size (cs).

(C) Wild-type and disome XI Cln3 protein levels from the experiments in (A) and (B) were quantified and normalized to Pgi1 levels. The Western blot used for quantification is not shown. This analysis shows that overall Cln3 protein levels are comparable between wild-type and disome XI strains.
Figure 10: Deletion of $BCK2$ exacerbates the G1 delay in disomic yeast cells.
Wild-type ([A], A31519, open circles), disome VIII ([B], A26163, open circles), disome XI ([C], A26162, open circles), disome XV ([D], A31520, open circles), and disome XVI ([E], A26161, open circles) lacking $BCK2$ and wild-type ([A], A11311, filled circles), disome VIII ([B], A13628, filled circles), disome XI ([C], A13771, filled circles), disome XV ([D], A12697, filled circles), and disome XVI ([E], A12700, filled circles) with functional $BCK2$ were grown at 30°C in selective medium supplemented with 2% raffinose. Small unbudded cells were isolated by centrifugal elutriation and released into the cell cycle at 30°C in YEPD. Time points were taken every 15 minutes to determine cell volume and the percentage of budded cells. Experiments for strains containing $BCK2$ are the same experiments as in Figure 1 and are shown here for comparison; strains were not run in parallel with the $bck2\Delta$ strains.
Figure 11: Progeny of triploid meioses exhibit a G1 delay.
MATa/a/α ([B], triploid parent 1, A6424) and MATa/a/α ([J], triploid parent 2, A6425) cells were induced to sporulate and tetrads were dissected. Individual colonies were picked after 1 week and grown to exponential phase in YEPD medium to determine DNA content ([C-I], A6424 progeny; [K-P], A6425 progeny). FL2-H detection thresholds were adjusted for each strain so that the cell population was within the linear detection range of the instrument. Plus signs indicate proliferation rates on solid medium, with +++ representing proliferation rates close or equal to wild-type controls, and fewer plus signs indicating poorer proliferation rates. A haploid strain (A2587) is shown in (A).