Siderophore-Mediated Cargo Delivery to the Cytoplasm of Escherichia coli and Pseudomonas aeruginosa: Syntheses of Monofunctionalized Enterobactin Scaffolds

The MIT Faculty has made this article openly available. Please share how this access benefits you. Your story matters.

| As Published | http://dx.doi.org/10.1021/ja3077268 |
| Publisher | American Chemical Society (ACS) |
| Version | Author's final manuscript |
| Citable Link | http://hdl.handle.net/1721.1/83505 |
| Terms of Use | Article is made available in accordance with the publisher's policy and may be subject to US copyright law. Please refer to the publisher's site for terms of use. |
| Detailed Terms | |
Siderophore-Mediated Cargo Delivery to the Cytoplasm of *Escherichia coli* and *Pseudomonas aeruginosa*: Syntheses of Monofunctionalized Enterobactin Scaffolds and Evaluation of Enterobactin-Cargo Conjugate Uptake

Tengfei Zheng, Justin L. Bullock, and Elizabeth M. Nolan*

Department of Chemistry, Massachusetts Institute of Technology, Cambridge, MA 02139

*Corresponding author: lnolan@mit.edu

Phone: 617-452-2495

Fax: 617-324-0505
Abstract

The design and syntheses of monofunctionalized enterobactin (Ent, L- and D-isomers) scaffolds where one catecholate moiety of enterobactin houses an alkene, aldehyde, or carboxylic acid moiety at the C5 position are described. These molecules are key precursors to a family of ten enterobactin-cargo conjugates presented in this work, which were designed to probe the extent to which the Gram-negative ferric enterobactin uptake and processing machinery recognizes, transports, and utilizes derivatized enterobactin scaffolds. A series of growth recovery assays employing enterobactin-deficient E. coli ATCC 33475 (ent-) revealed that six conjugates based on L-Ent having relatively small cargos promoted E. coli growth under iron-limiting conditions whereas negligible-to-no growth recovery was observed for four conjugates with relatively large cargos. No growth recovery was observed for the enterobactin receptor deficient strain of E. coli H1187 (fepA-) or the enterobactin esterase-deficient derivative of E. coli K-12 JW0576 (fes-), or when the D-isomer of enterobactin was employed. These results demonstrate that the E. coli ferric enterobactin transport machinery identifies and delivers select cargo-modified scaffolds to the E. coli cytoplasm. Pseudomonas aeruginosa PAO1 K648 (pvd-, pch-) exhibited greater promiscuity than that of E. coli for the uptake and utilization of the enterobactin-cargo conjugates, and growth promotion was observed for eight conjugates under iron-limiting conditions. Enterobactin may be utilized for delivering molecular cargos via its transport machinery to the cytoplasm of E. coli and P. aeruginosa thereby providing a means to overcome the Gram-negative outer membrane permeability barrier.
Introduction

Siderophores are low-molecular-weight high-affinity Fe(III) chelators that are biosynthesized and exported by bacteria, fungi, and plants during periods of nutrient limitation for acquiring this essential metal ion from the extracellular milieu.1,2 Both naturally-occurring and synthetic siderophore mimics are useful for bioremediation,3 iron chelation therapies,4,5 antibiotic drug-delivery strategies6-14 Fe(III) detection,15-18 protein identification,19 and pathogen capture.20,21 These types of applications benefit from or require siderophores amenable to facile and site-specific synthetic modification. In this work, we expand the current toolkit of site-specifically modifiable siderophore scaffolds to include triscatecholate enterobactin, and we report that various synthetic enterobactin-cargo conjugates are actively transported to the cytoplasm of the Gram-negative bacterial species Escherichia coli and Pseudomonas aeruginosa by the enterobactin uptake machinery.

Enterobactin (Ent, 1, Figure 1A) is a canonical siderophore biosynthesized by Gram-negative species of Enterobacteriaceae that include Escherichia coli, Salmonella, and Klebsiella.22 Decades of exploration pertaining to enterobactin biosynthesis and coordination chemistry, in addition to investigations of the proteins involved in its cellular transport and processing, provide a detailed molecular and physiological understanding of how this chelate contributes to bacterial iron homeostasis and colonization.22 The enterobactin synthetase is comprised of four proteins, EntBDEF, and is responsible for the production of enterobactin from L-serine and 2,3-dihydroxybenzoic acid (DHB).23 Following biosynthesis, Ent is exported into the extracellular space where it scavenges Fe(III). Enterobactin coordinates Fe(III) by its three catecholate groups with $K_a \sim 10^{49}$ M$^{-1}$.24 In E. coli, the outer membrane transporter FepA (and to a lesser extent Cir and Fiu) recognizes and binds ferric enterobactin with sub-nanomolar affinity,25,26 and provides periplasmic entry where the siderophore forms a complex with the periplasmic binding protein FepB.27 Subsequently, [Fe(Ent)]$^{3-}$ is transported into the cytosol, which requires the action of ExbBD, TonB, and FepCDG, the latter of which constitute the inner-membrane ATP-binding cassette (ABC) transporter system (Figure 1B).28-32 Fes, the cytosolic
enterobactin esterase, catalyzes the hydrolysis of the \([\text{Fe(Ent)}]^3\)-macrolactone,\textsuperscript{33} and the ferric reductase YgjH may subsequently assist in Fe(III) release such that the metal ion can be used metabolically.\textsuperscript{34} Several pathogenic Gram-negative species harbor gene clusters (e.g. iroA, MccE492) responsible for post-assembly line modifications of the enterobactin scaffold to provide the salmochelins.\textsuperscript{33,35-38} Salmochelins are a family of glucosylated enterobactin derivatives where the sugar moieties are attached to the C5 position of one or more catecholate rings (e.g. MGE 2 and DGE 3, Figure 1A).\textsuperscript{39}

![Figure 1](image.png)

**Figure 1.** Siderophores and siderophore transport machinery relevant to this work. (A) Structures of enterobactin 1 and the salmochelins MGE 2 and DGE 3. (B) Cartoon depiction of the enterobactin transport and processing machinery in *E. coli*.

Gram-negative bacteria have an outer membrane that serves as a permeability barrier and prevents cellular entry of many molecules, including antibiotics (e.g. vancomycin). Siderophore uptake machinery provides one route to overcome this permeability barrier,\textsuperscript{6-14} and enterobactin and its transporter FepA have been identified as a desirable siderophore/receptor pair for cargo delivery to Gram-negative bacterial species.\textsuperscript{13,37} FepA-mediated uptake of the ribosomal peptide antibiotics colicin B\textsuperscript{40} and MccE492m,\textsuperscript{41} in addition to bacteriophage,\textsuperscript{42}
indicates that this receptor has the capacity to transport large molecules. Moreover, the catecholate siderophore transporters of *E. coli* (e.g. Fiu, Cir) recognize synthetic catechol-modified \(\beta\)-lactam antibiotics;\(^\text{43-46}\) these serendipitous observations motivated early “Trojan horse” delivery strategies. Indeed, small-molecule antibiotics appended to siderophore-inspired di- and tricatecholate platforms have been evaluated for antibacterial activity with mixed results.\(^\text{47-51}\) Most recently, amoxicillin and ampicillin, \(\beta\)-lactam antibiotics that act in the periplasm and target bacterial cell wall biosynthesis, were covalently linked to a tripodal catecholate platform and remarkably afforded ca. \(10^2\)- to \(10^3\)-fold enhanced activity against *P. aeruginosa* PAO1 compared to the free drug.\(^\text{49}\)

The ability of FepABCDG and the TonB-ExbB-ExbD system of *E. coli*, as well as the enterobactin transport machinery of other bacterial species, to recognize and provide cytosolic transport of unnatural cargo appended to the native ligand remains unexplored. Enterobactin exhibits \(C_3\) symmetry and houses no unique functional group for site-specific synthetic modification. Total syntheses of enterobactin,\(^\text{52-56}\) hydrolytically stable enterobactin analogs,\(^\text{57-60}\) and salmochelins\(^\text{61}\) have been reported. To the best of our knowledge, no enterobactin scaffold housing a site-specific synthetic handle has been presented. Such scaffolds are a pre-requisite for employing enterobactin in a variety of paradigms that include cargo delivery, iron and siderophore detection, and bacterial capture.

Herein we present a family of ten enterobactin-cargo conjugates that are based on a monofunctionalized enterobactin scaffold. Inspired by the salmochelins, we have derivatized enterobactin at the C5 position of the catecholate, which provides a point for site-specific modification without compromising the Fe(III)-binding groups or the macrolactone (Figure 2). Moreover, we report that the ferric enterobactin uptake machineries of *Escherichia coli* and *Pseudomonas aeruginosa* PAO1 deliver enterobactin-derivatized cargo to the cytoplasm of both species under iron deficient conditions, and that cargo size is an important and species/strain-specific parameter to evaluate in enterobactin conjugate design.
**Experimental**

**Reagents.** Dimethylformamide (DMF) and dichloromethane (CH$_2$Cl$_2$) were dried over 4 Å molecular sieves or by using a VAC solvent purification system (Vacuum Atmospheres). Anhydrous dimethyl sulfoxide (DMSO) was purchased from Sigma-Aldrich and used as received. The triserine lactone 4 and its D-isomer 5 were synthesized according to a literature procedure.$^{55}$ 2,3-Bis(benzyloxy)benzoic acid 6,$^{62}$ vancomycin-alkyne 7,$^{63}$ and tert-butyl (2-oxo-2-(prop-2-yn-1-ylamino)ethyl)carbamate 8,$^{63}$ were synthesized according to literature procedures. L-Ent 1 and its D-isomer 9 were synthesized as reported elsewhere.$^{55,56}$ Tert-butyl 3-(2-(2-(aminoethoxy)ethoxy)ethoxy)propanoate 10 was purchased from BOC Sciences (Shirley, NY), 11-azido-3,6,9-trioxaundecan-1-amine 11 was purchased from Fluka, 6-((tert-butoxycarbonyl)amino)hexanoic acid 12 was purchased from Advanced Chem Tech, and Fmoc-PEG-CO$_2$H 13 was purchased from Chem-Impex International, Inc. The syntheses of the PEG-derivatized cargos 14-18 are provided as Supporting Information. Methyl-5-allyl-3-methoxysalicylate 19 was obtained from Sigma Aldrich. All other chemicals were purchased from Sigma-Aldrich, Alfa Aesar, or TCI in the highest available purity and used as received.

**General Synthetic Materials and Methods.** EMD TLC silica gel 60 F$_{254}$ plates were used for analytical thin-layer chromatography. EMD PLC silica gel 60 F$_{254}$ plates of 1-mm thickness were used for preparative TLC. Zeoprep 60HYD silica gel (40-63 μm) obtained from Zeochem was used for flash chromatography. $^1$H, $^{19}$F, and $^{13}$C NMR spectra were collected on a Varian
300 or 500 MHz spectrophotometer, which were operated at ambient probe temperature (283 K) and housed in the Department of Chemistry Instrumentation Facility. The $^1$H and $^{13}$C NMR spectra were referenced to internal standards and $^{19}$F spectra were referenced to an external CF$_3$Cl standard. An Avatar FTIR instrument was used to acquire IR spectra. Optical absorption spectra were recorded on an Agilent 8453 diode array spectrophotometer (1-cm quartz cuvettes, Starna). General methods for high performance liquid chromatography and mass spectrometry, $^1$H and $^{13}$C NMR spectra, and IR spectroscopic data are provided as Supporting Information.

**Methyl-5-allyl-2,3-dihydroxybenzoate (20).** Methyl-5-allyl-3-methoxysalicylate (19, 2.22 g, 10.0 mmol) and anhydrous $N,N$-diisopropylethylamine (DIPEA, 1.94 g, 15.0 mmol) were dissolved in 125 mL of dry CH$_2$Cl$_2$ and stirred at rt for five min. The solution was cooled to −78 °C in an acetone/dry ice bath, and boron tribromide (BBr$_3$, 1M solution in CH$_2$Cl$_2$, 30 mL, 30 mmol) was added slowly over ca. 10 min via a syringe to afford a yellow solution. The reaction was stirred at −78 °C for 1 h, warmed to −30 °C over the course of 1 h, and subsequently warmed to rt and stirred for another 4.5 h. Water (200 mL) was added slowly to quench the reaction, and the organic phase was washed with saturated aqueous potassium bicarbonate (K$_2$CO$_3$, 3 x 100 mL). The organic phase was dried over sodium sulfate (Na$_2$SO$_4$), and the solvent was removed under reduced pressure to afford a brown oil. Flash chromatography on silica gel with a solvent gradient (100% hexanes to 20% EtOAc/hexanes) gave the product as a white solid (1.09 g, 53%). TLC $R_f$=0.5 (silica, CH$_2$Cl$_2$); mp = 55-56 °C. $^1$H NMR (CDCl$_3$, 500 MHz), δ 3.29 (2H, d, $J$ = 7.0 Hz), 3.95 (3H, s), 5.05-5.10 (2H, m), 5.80 (1H, s), 5.91 (1H, m), 6.97 (1H, s), 7.18 (1H, s), 10.76 (1H, s). $^{13}$C NMR (CDCl$_3$, 125 MHz), δ 39.4, 52.3, 111.9, 116.0, 119.8, 120.4, 131.1, 137.0, 144.8, 147.2, 170.7. HRMS (DART): [M+Na]$^+$ m/z calcd., 231.0628; found, 231.0637.

**5-Allyl-2,3-bis(benzyloxy)benzoic acid (21).** Alkene 20 (2.18 g, 10.5 mmol), benzyl bromide (10.8 g, 60.3 mmol), and K$_2$CO$_3$ (24.5 g, 17.8 mmol) were combined in 200 mL of acetone at rt. The reaction was refluxed under N$_2$ for 16 h, which provided a yellow solution with
white solids, and the mixture was cooled to rt and filtered. The filtrate was concentrated under reduced pressure to afford a yellow oil. The oil was dissolved in a 375-mL mixture of 4:1 MeOH / 5 M NaOH (aq). The resulting solution was refluxed for 3.5 h and concentrated under reduced pressure to afford a white-yellow oil. Water (300 mL) was added to the oil, and the aqueous phase was washed with hexanes (4 x 100 mL). The pH of the aqueous phase was adjusted to ca. 1 by addition of 12 M HCl and the product precipitated as a white solid. A 100-mL portion of CH₂Cl₂ was added, and the resulting mixture was partitioned. The aqueous phase was extracted with additional CH₂Cl₂ (2 x 100 mL) and the combined organic layers were dried over Na₂SO₄ and concentrated under reduced pressure to yield 21 as a white solid (3.91 g, 99%). TLC Rf = 0.55 (silica, 100% CH₂Cl₂); mp = 135-136 °C. ¹H NMR (CDCl₃, 300 MHz), δ 3.38 (2H, d, J = 6.6 Hz), 5.06-5.14 (2H, m), 5.17 (2H, s), 5.22 (2H, s), 5.92 (1H, m), 7.09 (1H, d, J = 2.1 Hz), 7.31-7.50 (10H, m), 7.58 (1H, m). ¹³C NMR (CDCl₃, 125 MHz), δ 39.6, 71.4, 76.9, 116.7, 119.3, 122.6, 123.9, 127.8, 128.4, 128.7, 128.7, 129.1, 129.2, 134.8, 135.8, 136.2, 137.2, 145.5, 151.2, 165.6. HRMS (DART): [M-H]⁻ m/z calcd., 373.1445; found, 373.1439.

(E)-2,3-Bis(benzyloxy)-5-(prop-1-en-1-yl)benzoic acid (22). A 30-mL portion of methanol (MeOH) was degassed with N₂ for 4 h at rt and 21 (750 mg, 2.00 mmol) was subsequently added. The mixture was stirred at rt until 21 dissolved and PdCl₂ (58 mg, 0.32 mmol) was added to give a cloudy brown solution. The reaction was stirred at rt for 24 h and filtered. The filtrate was concentrated and purified by column chromatography using silica gel (1:4:5 EtOAc/hexanes/CH₂Cl₂) to yield 22 as a light yellow solid (666 mg, 89%). TLC Rf = 0.4 (40% EtOAc/hexanes); mp = 140-142 °C. ¹H NMR (CDCl₃, 300 MHz), δ 1.88-1.90 (3H, m), 5.19 (2H, s), 5.23 (2H, s), 6.25 (1H, dq, J = 15.9, 6.0 Hz), 6.32-6.38 (1H, m), 7.22 (1H, d, J = 2.1 Hz), 7.32-7.51 (10H, m), 7.69 (1H, d, J = 2.1 Hz). ¹³C NMR (CDCl₃, 125 MHz), δ 18.3, 71.4, 77.0, 115.8, 121.6, 122.7, 127.4, 127.7, 128.4, 128.7, 129.1, 129.2, 134.7, 135.7, 135.9, 145.7, 151.3, 165.5. HRMS (DART): [M-H]⁻ m/z calcd., 373.1445; found, 373.1457.

N,N′-((3S,7S,11S)-11-(2,3-Bis(benzyloxy)-5-((E)-prop-1-en-1-yl)benzamido)-2,6,10-trioxo-1,5,9-trioxacyclododecane-3,7-diyl)bis(2,3-bis(benzyloxy)benzamide) (23). Trilactone 4
(740 mg, 2.00 mmol) and DIPEA (2.58 g, 20 mmol) were mixed in dry DMSO (8 mL) and stirred for 10 min at rt to give a clear solution. PyAOP (3.13 g, 6.07 mmol), 22 (748 mg, 2.00 mmol) and 6 (1.00 g, 2.99 mmol) were dissolved in dry DMSO (10 mL) and added to the solution containing 4, and the reaction turned yellow and became orange after stirring for 2 h at rt. The orange solution was mixed with EtOAc (50 mL) and water (50 mL) and partitioned. The organic phase was washed with brine (3 x 50 mL), dried over Na₂SO₄, and concentrated to afford a yellow oil. Flash chromatography on silica gel with a solvent gradient (10% EtOAc/hexanes to 55% EtOAc/hexanes) yielded the product as a white foam (931 mg, 37%). TLC Rf = 0.3 (50% EtOAc/hexanes); mp = 100-102 °C (decomp). 1H NMR (CDCl₃, 300 MHz), δ 1.88-1.91 (3H, m), 4.01-4.11 (3H, m), 4.16-4.22 (3H, m), 4.91-4.98 (3H, m), 5.03-5.19 (12H, m), 6.17-6.40 (2H, m), 7.10-7.47 (32H, m), 7.66-7.71 (3H, m), 8.51-8.53 (3H, m). 13C NMR (CDCl₃, 125 MHz), δ 18.2, 40.6, 51.2, 63.9, 70.9, 76.0, 76.1, 114.2, 117.3, 120.4, 122.8, 124.1, 125.7, 126.1, 126.3, 127.4, 127.5, 127.9, 128.0, 128.2, 128.4, 128.4, 128.7, 128.7, 129.6, 134.1, 135.8, 135.8, 136.0, 136.0, 145.5, 146.7, 151.4, 151.4, 164.7, 168.8, 168.8. HRMS (DART): [M+H]⁺ m/z calcd., 1250.4645; found, 1250.4653.

N,N’-((3S,7S,11S)-11-(2,3-Bis(benzyloxy)-5-formylbenzamido)-2,6,10-trioxo-1,5,9-trioxacyclododecane-3,7-diyl)bis(2,3-bis(benzyloxy)benzamide) (24). A portion of compound 23 (285 mg, 0.228 mmol) was dissolved in 1,4-dioxane (9 mL) at rt, and water (3 mL) was added to give a colorless solution. Osmium tetroxide (OsO₄, 68 µL of 2.5% wt solution in 2-methyl-2 propanol, 6.7 µmol) was added and the reaction was stirred for 0.5 h at rt, which afforded a light brown solution. Sodium periodate (NaIO₄, 76.5 mg, 0.570 mmol) was then added and the reaction was stirred for another 2 h at rt. The suspension was partitioned in water (20 mL) and EtOAc (50 mL). The organic phase was washed with 0.1 M sodium thiosulfate (Na₂S₂O₃, 3 x 20 mL) and brine (2 x 20 mL), and dried over Na₂SO₄. Flash chromatography on silica gel with a solvent gradient (20% EtOAc/hexanes to 65% EtOAc/hexanes) yielded the product as white solid (165 mg, 58%). TLC Rf = 0.6 (70% EtOAc/hexanes); mp = 74 °C (decomp). 1H NMR (CDCl₃, 300 MHz), δ 4.03-4.11 (3H, m), 4.18-4.26 (3H, m), 4.90-4.96 (3H,
m), 5.05-5.28 (12H, m), 7.09-7.44 (31H, m), 7.65-7.67 (2H, m), 8.14-8.15 (1H, m), 8.46-8.52 (3H, m), 9.86 (1H, s). $^{13}$C NMR (CDCl$_3$, 125 MHz), $\delta$ 51.4, 51.4, 51.7, 64.1, 64.2, 71.0, 71.2, 76.2, 76.2, 76.5, 113.1, 117.3, 117.4, 122.9, 123.0, 124.2, 126.2, 126.3, 126.5, 127.5, 127.6, 127.8, 128.1, 128.3, 128.4, 128.4, 128.5, 128.5, 128.5, 128.6, 128.8, 128.9, 132.1, 135.2, 135.3, 135.9, 135.9, 136.0, 146.7, 146.8, 151.5, 151.5, 151.7, 152.2, 163.7, 164.9, 164.9, 168.7, 168.9, 169.1, 190.6. HRMS (DART): [M+H]$^+$ m/z calcd., 1238.4287; found, 1238.4279.

3,4-Bis(benzyloxy)-5-(((3S,7S,11S)-7,11-bis(2,3-bis(benzyloxy)benzamido)-2,6,10-trioxo-1,5,9-trioxacyclocodcan-3-yl)carbamoyl)benzoic acid (25). A portion of 24 (112 mg, 0.0903 mmol) was dissolved in 1,4-dioxane (3 mL) at rt. Sulfamic acid (NH$_3$SO$_3$, 15.8 mg, 0.162 mmol) was dissolved in water (0.75 mL) and added to the dioxane solution. Sodium chlorite (NaClO$_2$, 14.7 mg, 0.163 mmol) dissolved in 0.2 mL of water and the resulting solution was added to the reaction over the course of 10 min, and the reaction turned yellow. After stirring for 0.5 h at rt, the reaction was partitioned in water (10 mL) and EtOAc (20 mL), the aqueous phase was extracted with EtOAc (2 x 10 mL), and the combined organic phases were dried over Na$_2$SO$_4$. Flash chromatography on silica gel with a solvent gradient (CH$_2$Cl$_2$ to 10% MeOH/CH$_2$Cl$_2$) yielded the product as white solid (87 mg, 76%). TLC $R_f$ = 0.5 (10% MeOH/CH$_2$Cl$_2$); mp = 128-129 °C (decomp). $^1$H NMR (CDCl$_3$, 500 MHz), $\delta$ 4.05-4.08 (3H, m), 4.22-4.25 (3H, m), 4.93-4.98 (3H, m), 5.06-5.25 (12H, m), 7.06-7.47 (31H, m), 7.67-7.69 (2H, m), 7.86 (1H, s), 8.44-8.47 (2H, m), 8.54-8.57 (2H, m). $^{13}$C NMR (CDCl$_3$, 125 MHz), $\delta$ 51.4, 51.5, 51.6, 64.1, 71.1, 71.2, 76.2, 76.4, 117.5, 117.6, 123.0, 124.2, 125.4, 125.6, 126.2, 127.5, 127.6, 127.8, 128.1, 128.3, 128.4, 128.4, 128.5, 128.6, 128.7, 128.8, 128.9, 135.4, 135.6, 135.9, 136.1, 146.8, 150.7, 151.4, 151.5, 164.1, 165.0, 168.8, 168.9, 169.0, 169.3. HRMS (DART): [M+H]$^+$ m/z calcd., 1254.4230; found, 1254.4204.

Enantiomers 26-28. The D-isomers of the enterobactin alkene 23, aldehyde 24, and acid 25 were synthesized as described for the L-isomers except that triserine lactone 5 was employed instead of 4. The synthetic procedures and characterization are provided as Supporting Information.
Tert-butyl(1-3-(((3S,7S,11S)-7,11-bis(2,3-dihydroxybenzamido)-2,6,10-trioxo-1,5,9-trioxacyclododecan-3-yl)carbamoyl)-4,5-dihydroxyphenyl)-1-oxo-5,8,11-trioxo-2-azatridecan-13-yl)carbamate (29). Compound 25 (50 mg, 40 \text{ mol}), PyAOP (34 mg, 60 \text{ mol}) and DIPEA (15.2 \text{ mL}, 160 \text{ mol}) were mixed in 2 mL of dry CH$_2$Cl$_2$ at rt. A portion of 7 (15 mg, 48 \text{ mol}) was then added and the resulting yellow solution was stirred for 4 h at rt. The crude reaction was washed with 0.01N HCl (2 x 10 mL), dried over Na$_2$SO$_4$, and concentrated. The benzyl-protected product was purified by preparative TLC (10% MeOH/CH$_2$Cl$_2$) and obtained as a white viscous solid (46 mg, 75%). TLC $R_f = 0.7$ (10% MeOH/CH$_2$Cl$_2$). $^1$H NMR (CDCl$_3$, 500 MHz), $\delta$ 1.42 (9H, s), 3.27-3.28 (2H, m), 3.50-3.52 (2H, m), 3.59-3.66 (12H, m), 4.02-4.07 (3H, m), 4.15-4.18 (3H, m), 4.90-4.94 (3H, m), 5.03-5.20 (12H, m), 7.10-7.45 (36H, m), 7.65-7.67 (2H, m), 7.85-7.88 (1H, m), 7.99 (1H, bs), 8.49-8.54 (3H, m). $^{13}$C NMR (CDCl$_3$, 125 MHz), $\delta$ 28.3, 39.9, 40.2, 51.3, 51.4, 63.9, 64.1, 69.7, 70.0, 70.2, 70.3, 70.4, 71.1, 71.2, 76.2, 76.3, 79.0, 116.7, 117.5, 120.3, 123.0, 124.2, 125.4, 126.1, 126.2, 127.6, 127.6, 127.8, 128.2, 128.3, 128.4, 128.5, 128.6, 128.6, 128.7, 128.8, 128.9, 130.2, 135.4, 135.7, 135.9, 135.9, 136.1, 146.8, 146.9, 149.0, 151.5, 151.8, 155.9, 164.2, 164.8, 164.9, 165.8, 168.9, 169.0, 169.1. HRMS (ESI): [M+Na]$^+$ m/z calcd., 1550.5942; found, 1550.5977.

This benzyl-protected product was dissolved in 2 mL of 1:1 EtOAc/EtOH, the reaction flask was purged with N$_2$, and 45 mg Pd/C (10% wt) was added. The reaction was stirred under H$_2$ (1 atm) for 6 h at rt, and the Pd/C was removed by centrifugation (13,000 rpm, 10 min). The clear supernatant was decanted, concentrated, and re-dissolved in a 4:2:1 mixture of 1,4-dioxane/H$_2$O/MeOH, and purified by semi-preparative HPLC (20% B for 5 min followed by 20-70% B over 15 min, 4 mL/min). The product eluted at 15.8 min and was lyophilized to give 29 as white solid (15 mg, 50%). The analytical HPLC trace of the purified product is reported as Supporting Information. HRMS (ESI): [M+Na]$^+$ m/z calcd., 1010.3125; found, 1010.3173.

$N^1$-((3S,7S,11S)-7,11-Bis(2,3-dihydroxybenzamido)-2,6,10-trioxo-1,5,9-trioxacyclododecan-3-yl)-$N^2$-(1-cyclohexyl-1-oxo-5,8,11-trioxo-2-azatridecan-13-yl)-4,5-dihydroxyisophthalalimide (30). Compound 30 was synthesized as described for 29 except
that 14 (13.6 mg, 45.0 \text{ mol}) was used instead of 7. After purification by preparative TLC (10% MeOH/CH$_2$Cl$_2$), the benzyl-protected precursor of 30 was obtained as a white viscous solid (37 mg, 60%). TLC $R_f = 0.6$ (10% MeOH/CH$_2$Cl$_2$). $^1$H NMR (CDCl$_3$, 500 MHz), $\delta$ 1.17-1.21 (3H, m), 1.37-1.43 (2H, m), 1.62-1.63 (1H, m), 1.72-1.74 (2H, m), 1.78-1.81 (2H, m), 2.00-2.06 (1H, m), 3.39-3.42 (2H, m), 3.51-3.53 (2H, m), 3.59-3.61 (2H, m), 3.64-3.65 (10H, m), 4.01-4.06 (3H, m), 4.13-4.17 (3H, m), 4.88-4.93 (3H, m), 5.04-5.21 (12H, m), 6.23-6.25 (1H, m), 7.09-7.45 (35H, m), 7.64-7.66 (2H, m), 7.86 (1H, d, $J = 2.0$ Hz), 8.02 (1H, d, $J = 2.0$ Hz), 8.49-8.54 (3H, m). $^{13}$C NMR (CDCl$_3$, 125 MHz), $\delta$ 25.6, 29.5, 38.8, 40.0, 45.3, 51.3, 51.4, 63.9, 64.1, 69.8, 69.8, 70.0, 70.3, 70.4, 70.4, 71.2, 71.2, 76.2, 76.3, 116.8, 117.5, 120.4, 123.0, 124.3, 125.4, 126.1, 126.2, 127.6, 127.6, 127.9, 128.2, 128.3, 128.4, 128.4, 128.5, 128.5, 128.6, 128.6, 128.8, 128.8, 128.9, 129.0, 130.1, 135.4, 135.7, 135.9, 136.0, 136.1, 146.8, 146.9, 149.1, 151.6, 151.8, 164.3, 164.9, 164.9, 165.8, 168.9, 169.0, 169.1, 176.2. HRMS (ESI): [M+Na]$^+$ m/z calcd., 1560.6150; found, 1560.6269. Compound 30 was purified by semi-preparative HPLC (20% B for 5 min followed by 20-70% B over 15 min, 4 mL/min). The product eluted at 15.1 min and was obtained as white solid (20 mg, 58%). The analytical HPLC trace of the purified product is reported as Supporting Information. HRMS (ESI): [M+Na]$^+$ m/z calcd., 1020.3333; found, 1020.3346.

$N^2$-([3R,7R,11R]-7,11-Bis(2,3-dihydroxybenzamido)-2,6,10-trioxo-1,5,9-trioxacyclododecan-3-yl)-$N^1$-1(cyclohexyl-1-oxo-5,8,11-trioxa-2-azatridecan-13-yl)-4,5-dihydroxyiso-phthalamide (31). Compound 31 was synthesized as described for 30 except that 28 (36 mg, 29 \text{ mol}) was used instead of 25. After purification by preparative TLC (10% MeOH/CH$_2$Cl$_2$), the benzyl-protected precursor of 31 was obtained as a white oily solid (29 mg, 65%). TLC $R_f = 0.6$ (10% MeOH/CH$_2$Cl$_2$). $^1$H NMR (CDCl$_3$, 500 MHz), $\delta$ 1.17-1.25 (3H, m), 1.38-1.44 (2H, m), 1.63 (1H, m), 1.72-1.81 (4H, m), 2.01-2.06 (1H, m), 3.40-3.41 (2H, m), 3.39-3.42 (2H, m), 3.51-3.53 (2H, m), 3.58-3.65 (12H, m), 4.01-4.06 (3H, m), 4.13-4.16 (3H, m), 4.87-4.95 (3H, m), 5.03-5.21 (12H, m), 6.22-6.23 (1H, m), 7.09-7.45 (35H, m), 7.65-7.66 (2H, m), 7.86 (1H, s), 8.02 (1H, s), 8.49-8.54 (3H, m). $^{13}$C NMR (CDCl$_3$, 125 MHz), $\delta$ 25.6, 29.5, 38.8, 40.0, 45.3, 51.3, 51.4, 63.9, 64.1, 69.8, 69.8, 70.0, 70.3, 70.4, 70.4, 71.2, 71.2, 76.2, 76.3, 116.8, 117.5, 120.4, 123.0, 124.5, 126.3, 127.6, 127.9, 128.2, 128.3, 128.4, 128.4, 128.5, 128.6, 128.6, 128.8, 128.8, 128.9, 129.0, 130.1, 135.4, 135.7, 135.9, 136.0, 136.1, 146.8, 146.9, 149.1, 151.6, 151.8, 164.3, 164.9, 164.9, 165.8, 168.9, 169.0, 169.1, 176.2.
125.4, 126.1, 126.2, 127.6, 127.9, 128.2, 128.3, 128.4, 128.5, 128.6, 128.6, 128.8, 128.8, 128.9, 129.0, 130.1, 135.4, 135.7, 135.9, 136.0, 136.1, 146.8, 146.9, 149.1, 151.6, 151.8, 164.3, 164.9, 165.8, 168.9, 169.0, 169.1, 176.2. HRMS (ESI): [M+Na]^+ m/z calcd., 1560.6150; found, 1560.6141. Compound 31 was purified by semi-preparative HPLC (20% B for 5 min followed by 20-70% B over 15 min, 4 mL/min). The product eluted at 14.8 min and was obtained as white solid (5.1 mg, 27% yield). The analytical HPLC trace of the purified product is reported as Supporting Information. HRMS (ESI): [M+Na]^+ m/z calcd., 1020.3333; found, 1020.3328.

N^2-((3S,7S,11S)-7,11-Bis(2,3-dihydroxybenzamido)-2,6,10-trioxo-1,5,9-trioxacyclododecan-3-yl)-4,5-dihydroxy-N^1-(1-(naphthalen-2-yl)-1-oxo-5,8,11-trioxa-2-azatridecan-13-yl)isophthalamide (32). Compound 32 was synthesized as described for 29 except that 15 (20 mg, 44 [mol] was used instead of 7. After purification by preparative TLC (5% MeOH/CH$_2$Cl$_2$), the benzyl-protected precursor of 32 was obtained as a white-yellow oily solid (37 mg, 59%). TLC $R_f = 0.6$ (10% MeOH/CH$_2$Cl$_2$). $^1$H NMR (CDCl$_3$, 500 MHz), $\delta$ 3.44-3.74 (16H, m), 3.94-4.08 (4H, m), 4.12-4.16 (2H, m), 4.78-4.82 (1H, m), 4.87-4.92 (2H, m), 5.02-5.17 (12H, m), 7.01-7.52 (39H, m), 7.58-7.59 (1H, m), 7.64-7.66 (2H, m), 7.79-7.84 (3H, m), 7.94-7.94 (1H, m), 8.29-8.31 (1H, m), 8.47-8.50 (3H, m). $^{13}$C NMR (CDCl$_3$, 125 MHz), $\delta$ 39.6, 39.9, 51.4, 51.4, 63.9, 64.1, 69.6, 69.7, 70.2, 70.4, 71.1, 71.2, 71.2, 76.2, 76.3, 76.3, 116.7, 117.5, 120.3, 123.1, 124.3, 124.6, 125.0, 125.2, 125.4, 126.1, 126.2, 126.2, 126.9, 127.6, 127.6, 127.6, 127.9, 128.1, 128.2, 128.4, 128.4, 128.5, 128.6, 128.6, 128.8, 128.9, 128.9, 129.0, 130.0, 130.1, 130.3, 133.5, 134.5, 135.4, 135.7, 135.9, 136.0, 136.2, 146.9, 146.9, 149.0, 151.6, 151.7, 164.2, 164.9, 164.9, 165.7, 168.9, 169.0, 169.1, 169.6. HRMS (ESI): [M+Na]^+ m/z calcd., 1604.5837; found, 1604.5964. Compound 32 was purified by semi-preparative HPLC (20% B for 5 min followed by 30-55% B over 10 min, 4 mL/min) and eluted at 12.7 min. The isolated product was lyophilized and obtained as a white solid (4.4 mg, 18%). The analytical HPLC trace of the purified product is provided as Supporting Information. HRMS (ESI): [M+Na]^+ m/z calcd., 1064.3020; found, 1064.3084. Mass spectrometric analysis of the crude reaction indicated M+4 in addition to the
desired product 32 and suggested partial reduction of the naphthalene cargo under the deprotection conditions. From analysis of HPLC peak areas, the ratio between 32 and the partial reduction product is ca. 4:1.

\[ N^1-(1-(3-Benzylphenyl)-1-oxo-5,8,11-trioxo-2,6,10-trioxa-1,5,9-trioacyclocodecan-3-yl)-N^2-(3S,7S,11S)-7,11-bis(2,3-dihydroxybenzamido)-2,6,10-trioxo-1,5,9-trioacyclocodecan-3-yl)-4,5-dihydroxyisophthalamide \] (33). Compound 33 was synthesized as described for 29 except that 16 (24 mg, 62 \( \mu \)mol) was used instead of 7. Partial purification by preparative TLC (10% MeOH/CH\(_2\)Cl\(_2\)) afforded the benzyl-protected precursor of 33 as a white-yellow solid with a grease contamination (43 mg, 67%). TLC \( R_f = 0.6 \) (10% MeOH/CH\(_2\)Cl\(_2\)). \(^1\)H NMR (CDCl\(_3\), 500 MHz), \( \delta \) 3.57-3.61 (12H, m), 3.94-3.95 (2H, d, \( J = 6.0 \)) 3.97-4.05 (3H, m), 4.07-4.15 (3H, m), 4.85-4.90 (3H, m), 5.01-5.17 (12H, m), 7.01-7.40 (30H, m) 7.62-7.70 (3H, m), 7.82 (1H, d, \( J=2.0 \)), 7.99-8.00 (1H, d, \( J = 2.0 \)), 8.47-8.51 (3H, m) HRMS (ESI): [M+Na\(^+\) \( m/z \) calcd., 1644.6150; found, 1644.6105. A portion (32.5 mg, 20.0 \( \mu \)mol) of this material was carried without further purification or characterization. Compound 33 was purified by semi-preparative HPLC (20% B for 5 min followed by 20-70% B over 15 min, 4 mL/min). The product eluted at 15.8 min and was obtained as white solid (13.5 mg, 62%). The analytical HPLC trace of the purified product is provided as Supporting Information. HRMS (ESI): [M+Na\(^+\) \( m/z \) calcd., 1104.3333; found, 1104.3305.

\[ N^2-(3S,7S,11S)-7,11-Bis(2,3-dihydroxybenzamido)-2,6,10-trioxo-1,5,9-trioacyclocodecan-3-yl)-4,5-dihydroxy-N^1-(1-oxo-1-(11-oxo-2,3,5,6,7,11-hexahydro-1H-pyrido[2,3-f]pyrido[3,2,1-ij]quinolin-10-yl)-5,8,11-trioxa-2-azatridecan-13-yl)isophthalamide \] (34). Compound 34 was synthesized as described for 29 except that 17 (18 mg, 39 \( \mu \)mol) was used instead of 7. After purification by preparative TLC (10% MeOH/CH\(_2\)Cl\(_2\)) the benzyl-protected precursor of 34 was obtained as an orange oily solid (18 mg, 26%). TLC \( R_f = 0.7 \) (10% MeOH/CH\(_2\)Cl\(_2\)). \(^1\)H NMR (CDCl\(_3\), 500 MHz), \( \delta \) 1.93-1.95 (4H, m), 2.71-2.83 (4H, m), 3.26-3.32 (4H, m), 3.56-3.69 (16H, m), 3.99-4.18 (6H, m), 4.88-4.94 (3H, m), 5.01-5.18 (12H, m), 6.94 (1H, s), 7.06-7.43 (35H, m), 7.62-7.66 (2H, m), 7.80-7.80 (1H, m), 7.97-7.97 (1H, m),
8.47-8.53 (4H, m), 9.02-9.03 (1H, m). $^{13}$C NMR (CDCl$_3$, 125 MHz), δ 19.9, 20.0, 21.0, 27.3, 39.4, 40.1, 49.7, 50.2, 51.5, 64.1, 69.9, 71.1, 71.2, 76.3, 105.4, 108.1, 115.9, 117.5, 119.8, 123.0, 124.3, 125.7, 126.3, 127.2, 127.6, 127.6, 127.8, 128.1, 128.2, 128.5, 128.5, 128.6, 128.9, 128.9, 130.0, 135.7, 136.0, 136.2, 146.9, 148.2, 148.3, 149.0, 151.6, 151.7, 152.6, 162.9, 164.4, 165.0, 166.9, 169.1. HRMS (ESI): [M+Na]$^+$ m/z calcd., 1717.6313; found, 1717.6287. Compound 34 was purified by semi-preparative HPLC (20% B for 5 min followed by 20-70% B over 15 min, 4 mL/min). The product eluted at 17.1 min and was obtained as an orange solid (4.5 mg, 48%). The analytical HPLC trace of the purified product is provided as Supporting Information. HRMS (ESI): [M+Na]$^+$ m/z calcd., 1177.3496; found, 1177.3540.

7-(4-(1-(3-(((3S,7S,11S)-7,11-Bis(2,3-dihydroxybenzamido)-2,6,10-trioxo-1,5,9-trioxacyclododecan-3-yl)carbamoyl)-4,5-dihydroxyphenyl)-1-oxo-5,8,11-trioxo-2-azatetradecan-14-oyl)piperazin-1-yl)-1-cyclopropyl-6-fluoro-4-oxo-1,4-dihydroquinoline-3-carboxylic acid (35). Compound 35 was synthesized as described for 29 except that 18 (26 mg, 48 mol) was used instead of 7, and TMSCI (10 |L, 79 mol) and DIPEA (15 µL, 160 µmol) was mixed with 18 before addition to the solution containing 25. After purification by preparative TLC (10% MeOH/CH$_2$Cl$_2$), the benzyl-protected precursor of 35 was obtained as a yellow oily solid (46 mg, 65%). TLC R$_f$ = 0.65 (10% MeOH/CH$_2$Cl$_2$). $^1$H NMR (CDCl$_3$, 500 MHz), δ 1.13 (2H, bs), 1.33 (2H, bs), 2.64 (2H, bs), 3.23-3.30 (4H, m), 3.51 (1H, bs), 3.63 (14H, bs), 3.79 (4H, bs), 3.99-4.04 (3H, m), 4.11-4.14 (3H, m), 4.86-4.91 (3H, m), 5.01-5.19 (12H, m), 7.06-7.43 (39H, m), 7.59-7.61 (2H, m), 7.83 (1H, s), 7.97-7.99 (2H, m), 8.45-8.49 (3H, m), 8.69 (1H, s). $^{13}$C NMR (CDCl$_3$, 125 MHz), δ 8.2, 33.4, 35.4, 40.0, 41.1, 45.3, 49.3, 50.0, 51.3, 51.4, 51.4, 63.9, 64.1, 67.1, 69.7, 70.2, 70.3, 70.4, 70.5, 71.2, 71.3, 76.2, 76.3, 105.2, 108.0, 112.3, 112.4, 116.7, 117.5, 120.0, 120.0, 120.5, 123.0, 124.3, 125.6, 126.1, 126.1, 127.6, 127.6, 127.8, 128.2, 128.3, 128.4, 128.5, 128.6, 128.6, 128.8, 128.8, 128.8, 129.0, 130.2, 135.5, 135.7, 135.9, 136.0, 136.1, 138.9, 145.2, 145.3, 146.8, 146.8, 147.4, 149.0, 151.6, 151.6, 151.8, 152.4, 154.4, 164.2, 164.9, 164.9, 165.8, 166.9, 168.9, 169.0, 169.1, 169.7, 176.9. $^{19}$F NMR (CDCl$_3$, 282 MHz) δ -121.3. HRMS (ESI): [M+Na]$^+$ m/z calcd., 1792.6434; found, 1792.6337. Compound 35 was
purified by semi-preparative HPLC (20% B for 5 min followed by 20-70% B over 10 min, 4 mL/min) and eluted at 15.2 min. The isolated product was lyophilized and obtained as a white solid (2.5 mg, 9%). The HPLC trace of the purified product is provided as Supporting Information. HRMS (ESI): [M+Na]+ m/z calcd., 1252.3617; found, 1252.3633.

7-(4-(6-Aminohexanoyl)piperazin-1-yl)-1-cyclopropyl-6-fluoro-4-oxo-1,4-dihydroquinoline-3-carboxylic acid (36). Ciprofloxacin (37, 331 mg, 1.00 mmol) and DIPEA (1.0 mL, 5.7 mmol) were mixed in 6 mL of dry CH2Cl2, and TMSCl (370 µL, 2.91 mmol) was added to give a clear yellow solution. 6-((Tert-butoxycarbonyl)amino)hexanoic acid (12, 346 mg, 1.50 mmol), PyAOP (834 mg, 1.60 mmol), and DIPEA (700 µL, 4.02 mmol) were dissolved in 4 mL of dry CH2Cl2, and the two solutions were combined and stirred overnight at rt. The reaction was quenched with MeOH (10 mL), and the resulting solution was concentrated to dryness, and the crude product was redissolved in 40 mL of EtOAc. The organic phase was washed with 10 mM HCl (2 x 40 mL) and saturated aqueous NaHCO3 (2 x 40 mL), dried over Na2SO4, and purified by flash chromatography on silica gel (3% MeOH/CH2Cl2) to give 38 as yellow solid (243 mg, 45%). TLC Rf = 0.7 (5% MeOH/CH2Cl2). 1H NMR (CDCl3, 300 MHz), δ 1.14-1.20 (2H, m), 1.32-1.53 (13H, m), 1.59-1.69 (2H, m), 2.36 (2H, t, J = 6.0 Hz), 3.08 (2H, dt, J = 6.3, 6.3 Hz), 3.26-3.56 (4H, m), 3.51-3.59 (1H, m), 3.69-3.82 (4H, m), 4.68 (1H, bs), 7.32 (1H, d, J = 7.2 Hz), 7.82 (1H, d, J = 12.9 Hz), 8.60 (1H, s), 14.9 (1H, bs). 13C NMR (CDCl3, 125 MHz), δ 8.1, 24.7, 26.4, 28.3, 29.8, 32.9, 35.3, 40.2, 41.0, 45.1, 49.3, 49.9, 78.9, 105.0, 107.7, 111.9, 112.1, 119.6, 119.7, 138.8, 145.2, 145.3, 147.3, 152.4, 154.4, 155.9, 166.6, 171.4, 176.7. 19F NMR (CDCl3, 282 MHz), δ -121.1. HRMS (ESI): [M+H]+ m/z calcd., 545.2775; found, 545.2768.

The TFA salt of 36 (202 mg, 98%) was obtained as a yellow solid from 38 (201 mg, 0.369 mmol) by stirring 38 in 40% TFA/CH2Cl2 at rt for 3 h and removing the solvent. TLC Rf = 0.1 (10% MeOH/CH2Cl2). 1H NMR (CD3OD, 300 MHz), δ 1.41-1.52 (4H, m), 1.65-1.77 (4H, m), 2.52 (2H, t, J = 7.2 Hz), 2.96 (2H, t, J = 7.2 Hz), 3.34-3.43 (4H, m), 3.82 (5H, m), 7.57 (1H, d, J = 7.5 Hz), 7.85 (1H, d, J = 13.2 Hz), 8.76 (1H, s). 13C NMR (CDCl3, 125 MHz), δ 7.8, 23.8, 25.4, 26.5, 26.6, 32.2, 35.4, 39.0, 39.1, 41.0, 45.0, 48.1, 48.3, 48.5, 48.6, 48.8, 49.0, 49.1, 49.5, 105.0,
17 F NMR (CDCl$_3$, 282 MHz), δ -76.0, -120.9. HRMS (ESI): [M+H]$^+$ m/z calcd., 445.2251; found, 445.2255.

7-(4-(6-(3-(((3S,7S,11S)-7,11-bis(2,3-dihydroxybenzamido)-2,6,10-trioxo-1,5,9-trioxacyclododecan-3-yl)carbamoyl)-4,5-dihydroxybenzamido)hexanoyl)piperazin-1-yl)-1-cyclopropyl-6-fluoro-4-oxo-1,4-dihydroquinoline-3-carboxylic acid (40). Compound 40 was synthesized as described for 35 except that DMSO (1.5 mL) was used as the solvent and compound 36 (19.4 mg, 34.8 µmol) was used instead of 18. After preparative TLC purification (10% MeOH/CH$_2$Cl$_2$), 39 was obtained as white viscous solid (17 mg, 74%). TLC $R_f$ = 0.6 (10% MeOH/CH$_2$Cl$_2$). $^1$H NMR (CDCl$_3$, 300 MHz) δ 1.17-1.83 (10H, m), 2.40 (2H, bs), 3.29-3.44 (6H, m), 3.70-3.86 (5H, m), 4.02-4.17 (6H, m), 4.86-4.93 (3H, m), 5.04-5.21 (12H, m), 7.07-7.42 (33H, m), 7.60-7.64 (2H, m), 7.85-8.05 (3H, m), 8.47-8.50 (3H, m), 8.74 (1H, bs), 15.0 (1H, bs).

$^{13}$C NMR (CDCl$_3$, 125 MHz), δ 8.1, 12.3, 17.2, 18.6, 24.4, 26.3, 26.4, 26.5, 29.0, 32.8, 34.7, 39.7, 41.2, 45.3, 46.2, 46.3, 51.4, 51.5, 51.5, 52.0, 54.8, 63.9, 64.1, 64.2, 71.1, 71.2, 71.2, 76.2, 76.3, 76.3, 105.2, 109.5, 113.0, 113.2, 116.6, 117.5, 120.1, 123.0, 124.3, 125.5, 126.1, 127.6, 127.8, 128.2, 128.3, 128.4, 128.4, 128.5, 128.6, 128.6, 128.6, 128.8, 128.9, 128.9, 129.0, 130.3, 135.5, 135.8, 135.9, 136.0, 136.1, 138.1, 145.4, 146.8, 149.0, 151.6, 151.8, 152.3, 152.4, 154.1, 164.9, 165.0, 165.8, 166.1, 166.8, 169.0, 169.1, 171.5. HRMS (ESI): [M+H]$^+$ m/z calcd., 1680.6303; found, 1680.6352. Compound 40 was purified by semi-preparative HPLC (20% B for 5 min followed by 20-70% B over 15 min, 4 mL/min) and eluted at 16.1 min. The isolated product was lyophilized and obtained as a white-yellow solid (6.7 mg, 59%). The HPLC trace of the purified product is provided as Supporting Information. HRMS (ESI): [M+H]$^+$ m/z calcd., 1140.3486; found, 1140.3482.

$N_1$-(2-(2-(2-Azidoethoxy)ethoxy)ethoxy)ethyl)-4,5-bis(benzyloxy)-$N_3$-((3S,7S,11S)-7,11-bis(2,3-bis(benzyloxy)benzamido)-2,6,10-trioxo-1,5,9-trioxacyclododecan-3-yl)isophthalamide (41). 11-Azido-3,6,9-trioxaundecan-1-amine (11, 8.2 µL, 42 µmol) and 25 (40 mg, 32 µmol) were dissolved in 1 mL of dry CH$_2$Cl$_2$. PyAOP (33.2 mg, 63.8 µmol) and
DIPEA (22.2 mL, 128 mol) were added to give a light yellow solution. The reaction was stirred for 4 h at rt and concentrated, and the crude product was purified by preparative TLC (50% EtOAc/CH₂Cl₂) to afford 41 as a light yellow oil (31 mg, 68%). TLC Rᵣ = 0.3 (50% EtOAc/CH₂Cl₂). ¹H NMR (CDCl₃, 300 MHz) δ 3.34 (2H, t, J = 5.1 Hz), 3.61-3.69 (14H, m), 3.97-4.18 (6H, m), 4.88-4.94 (3H, m), 5.02-5.22 (12H, m), 7.08-7.46 (34H, m), 7.64-7.67 (2H, m), 7.85 (1H, d, J = 1.8 Hz), 7.99 (1H, d, J = 2.1 Hz), 8.48-8.52 (3H, m). ¹³C NMR (CDCl₃, 125 MHz) δ 40.0, 50.6, 51.4, 64.0, 64.1, 69.7, 69.9, 70.3, 70.6, 71.2, 71.2, 76.3, 116.7, 117.5, 120.4, 123.1, 124.3, 125.5, 126.2, 126.2, 127.6, 127.6, 127.9, 128.3, 128.4, 128.4, 128.5, 128.6, 128.7, 128.8, 128.9, 129.0, 130.2, 135.5, 135.8, 136.0, 136.0, 136.2, 146.9, 146.9, 149.1, 151.6, 151.8, 164.2, 164.9, 165.9, 168.9, 169.1, 169.1. HRMS (ESI): [M+Na]⁺ m/z calcd., 1476.5323; found, 1476.5345.

**Vancomycin-PEG-Ent (42).** A DMSO solution of 41 (19 mg/mL, 1.3 mM, 250 µL), an aqueous solution of 8 (20 mg/mL, 1.3 mM, 250 µL), a DMF solution of benzoic acid (49 mg/mL, 450 mM, 50 µL), and an aqueous solution of CuSO₄ (10 mg/mL, 45 mM, 50 µL) were mixed together, and an additional 400 µL of DMSO was added to yield a clear light blue solution. An aqueous solution of sodium ascorbate (NaAsc, 18 mg/mL, 90 mM, 50 µL) was subsequently added. The reaction become colorless to yellow and was stirred at rt for 15 min, at which time another 50 µL of aqueous NaAsc was added. After stirring for 15 min, the crude reaction was frozen and lyophilized to give a brown oil. The oil was dissolved in a 2:1:1 ratio of dioxane/MeOH/H₂O and purified by semi-preparative HPLC (50% B for 5 min followed by 50-100% B over 11 min, 4 mL/min). The benzyl-protected precursor of 42 eluted at 13 min and was obtained as white solid after lyophilization (3.5 mg, 36%). HRMS (ESI): [M+2Na]²⁺/2 m/z calcd., 1520.5030; found, 1520.5171.

A portion of this precursor (14 mg, 4.7 mol; obtained from four 250-µL scale Click reactions) was dissolved in 30% H₂O/MeOH, the flask was purged with N₂, and 16 mg Pd/C (10% wt) was added. The reaction was stirred under H₂ (1 atm) for 24 h at rt, and the Pd/C was removed by centrifugation (13,000 rpm, 10 min). The supernatant was concentrated by
lyophilization and the resulting residue was dissolved in a 2:1:1 mixture of dioxane/MeOH/H₂O. HPLC purification (20% B for 5 min followed by 20-46% B over 8 min, 4 mL/min) gave 43 as white solid (6.3 mg, 55%). The HPLC trace of the purified product is reported in Supporting Information. HRMS (ESI): [M+2H]^2+/2 m/z calcd., 1228.37960; found, 1228.37961.

**tert-Butyl(2-(((1-(1-(((3S,7S,11S)-7,11-bis(2,3-dihydroxybenzamido)-2,6,10-trioxo-1,5,9-trioxacyclododecan-3-yl)carbamoyl)-4,5-dihydroxyphenyl)-1-oxo-5,8,11-trioxa-2-azatridecan-13-yl)-1H-1,2,3-triazol-4-yl)methyl)amino)-2-oxoethyl)carbamate (43).** Compound 43 was synthesized as described for 42 except that a DMSO solution of 7 (2.8 mg/mL, 13 mM, 25 µL) was used instead of 8. HPLC purification gave 3.3 mg of the benzyl-protected precursor of 43 as a white solid (58%). HRMS (ESI): [M+H]^+ m/z calcd., 1688.6489; found, 1688.6421. Compound 43 (3.3 mg, 33%) was obtained from the precursor (13.3 mg, 7.88 µmol; obtained from four 25-µL scale Click reactions) following the same procedure as synthesizing 42. HPLC purification (0% B for 5 min followed by 0-45% B over 8 min, 4 mL/min) afforded 43 as a white solid with a retention time of 12.8 min. The HPLC trace of the purified product is reported as Supporting Information. HRMS (ESI): [M+H]^+ m/z calcd., 1126.3853; found, 1126.3832.

**Growth Recovery Assays.** General microbiology methods are included as Supporting Information. Overnight cultures were prepared by inoculating 5 mL of LB (E. coli) or LB base supplemented with 2.5 g/L NaCl (P. aeruginosa) with the appropriate freezer stocks and the cultures were incubated at 37 °C in a tabletop incubator shaker set at 150 rpm. The overnight culture was diluted 1:100 into 5 mL of fresh media with or without 200 µM 2,2'-dipyridyl (DP) and incubated at 37 °C with shaking at 150 rpm until the optical density at 600 nm (OD₆₀₀) reached 0.6. The cultures were diluted to an OD₆₀₀ value of 0.001 in 50% reduced MHB medium (10.5 g/L) with or without 200 µM (E. coli) or 600 µM DP (P. aeruginosa). A 90-µL aliquot of the diluted culture was mixed with a 10-µL aliquot of a 10x solution of the siderophore or siderophore-cargo conjugate in a 96-well plate, which was wrapped in parafilm and incubated at 30 °C with shaking at 150 rpm for 19 h. Bacterial growth was assayed by measuring OD₆₀₀.
using a BioTek Synergy HT plate reader. Each well condition was prepared in duplicate and three independent replicates of each assay were conducted on different days. The resulting mean OD$_{600}$ are reported and the error bars are the standard deviation of the mean obtained from the three independent replicates.

**Results and Discussion**

**Design and Synthesis of Monofunctionalized Enterobactin Platforms.** We present a stepwise synthesis to monofunctionalized enterobactin scaffolds in Scheme 1. Guided by the structures of MGE and DGE (Figure 1A), we chose to install functional groups amenable to synthetic modification at the C5 position of one enterobactin catechol ring. This position is remote from the iron-binding hydroxyl groups in addition to the macrolactone (Figure 2). Prior studies of the salmochelins indicate that modification at this site compromises neither Fe(III) complexation nor the esterase-catalyzed hydrolysis of the macrolactone.$^{33,64}$ The structure of the antibiotic-siderophore conjugate MccE492m (Figure S1),$^{65}$ which exhibits a monoglucosylated enterobactin derivative attached to a ribosomal peptide, also influenced our decision to prepare monofunctionalized enterobactin platforms. We selected methyl-5-allyl-3-methoxysalicylate 19 as a starting material because of its commercial availability. This precursor was demethylated using BBr$_3$ in the presence of DIPEA to prevent HBr addition to the alkene moiety, and 20 was obtained in 53% yield as a white powder. Benzyl protection of 20 and subsequent hydrolysis of the methyl ester in refluxing sodium hydroxide was performed following a literature protocol$^{62}$ for catecholate protection of 2,3-dihydroxybenzoic acid and 21 was obtained in 99% yield as a white powder. Palladium-catalyzed isomerization of the alkene was achieved by using PdCl$_2$ in degassed methanol and 22 was obtained in 89% yield as a light yellow solid following workup. Next, a one-pot coupling reaction between the enterobactin trilactone 4, 6 and 22 was performed with a 1:1.5:1 ratio and PyAOP as the coupling reagent. This reaction provided a mixture of 23, its di- and tri-substituted analogs, and unmodified Ent. These products were separated by flash chromatography and afforded 23 in 37% yield as a
white foam. The 1:1.5:1 ratio of 4/6/22 was chosen based on several optimization trails and this ratio provided the highest yield of the desired monosubstituted product. Oxidation of alkene 23 by using OsO₄ and NaIO₄ in mixed 1,4-dioxane/water afforded 24 as a white foam in 58% yield. Further oxidation of 24 under mild conditions provided carboxylic acid 25 in 76% yield as a white powder. This step-wise synthesis provides gram quantities of 23-25 (L-isomers) in high purity, and these molecules are stable when stored as dry solids at 4 °C. The stepwise coupling and oxidations were also performed using triserine lactone 5 to afford the D-enantiomers alkene 26, aldehyde 27, and acid 28 (Supporting Information). The D-enantiomer of Ent is transported into E. coli by FepA, but it is not a substrate for the enterbactin esterase Fes. We therefore reasoned that conjugates based on D-Ent would provide useful controls for conjugate uptake studies, and that this enantiomer may also be advantageous in antibacterial drug delivery applications because it provides an iron-starvation effect.

Scheme 1. Syntheses of 22 (top) and monofunctionalized enterobactin scaffolds (bottom).

This synthesis provides a family of enterobactin scaffolds amenable to functionalization. For instance, alkene 23 may be employed in olefin cross metathesis, aldehyde 24 in reductive amination, and acid 25 in peptide coupling reactions. Moreover, other organic transformations of
22 or 23 may provide additional versatile functional groups (e.g. hydroxyl), affording more synthetic possibilities for enterobactin derivatives that can be utilized in various applications.

**Design and Synthesis of Enterobactin-Cargo Conjugates.** We selected carboxylic acid 25 as a key intermediate for the preparation of enterobactin-cargo conjugates, and evaluated two strategies for appending cargo to the enterobactin scaffold. In one thrust, standard peptide coupling chemistry was employed to link cargo to the enterobactin acid via an amide bond (Schemes 2 and 3). In the second approach, enterobactin-azide 41 was prepared and “Click” chemistry utilized for cargo attachment (Scheme 4). In both cases, benzyl deprotection unmasked the enterobactin catecholates in the final step.

**Scheme 2.** Syntheses of enterobactin-cargo conjugates 29-35. The syntheses of 14-18 are provided as Supporting Information.

We selected a variety of commercially available molecules housing carboxylic acids as cargos. The cargos presented in Scheme 2 vary in size and shape and include a simple Boc
protecting group, cyclohexane, naphthalene, phenylmethylbenzene, ciprofloxacin, and coumarin. This selection includes cargo expected to be non-toxic (e.g. Boc, cyclohexane) in addition to an antibiotic (ciprofloxacin) and a fluorophore (coumarin 343). Next, we selected PEG₃ as a stable and water-compatible linker. It provides ca. 14-Å separation between enterobactin and the cargo. The conjugates depicted in Scheme 2 were prepared by coupling the PEG-derivatized cargo 10,14-18 to 25 using PyAOP as the coupling reagent. The resulting benzyl-protected conjugates were purified by preparative TLC and obtained in yields ranging from 26% (Bn-34) to 75% (Bn-29). Benzyl deprotection reactions were performed by hydrogenation over Pd/C and the resulting enterobactin-cargo conjugates were purified by reverse-phase semi-preparative HPLC. Conjugates 29-35 were obtained in milligram quantities and high purity judging by analytical HPLC (Figures S2-S11) and LC/MS analysis (Table S1). Conjugate 31 houses D-Ent and was prepared to probe the role of Fes-mediated hydrolysis in the bacterial growth recovery assays (vide infra).

Scheme 3 exhibits the synthesis of enterobactin-ciprofloxacin 40 where the PEG linker is substituted by a C₅ alkyl chain to probe the consequences of variable linker composition and hydrophilicity. The synthesis of 40 was carried out by reacting ciprofloxacin with 6-Boc-aminohexanoic acid 12 followed by Boc deprotection, coupling of the resulting free amine to 25, and benzyl deprotection. The carboxylic acid of ciprofloxacin was protected in situ by using trimethylsilyl chloride (TMSCl) to prevent self-coupling in the syntheses of both 35 and 40. In this general approach of attaching a carboxylic acid cargo, the linkers were first coupled to the cargo rather than to the Ent scaffold because the Ent macrolactone degrades in the presence of primary amines and under highly acidic conditions such as those required to remove Boc protecting groups.

In Scheme 4, we present the synthesis of 43, an enterobactin-vancomycin conjugate assembled via Click chemistry. Vancomycin is a nonribosomal peptide antibiotic active against Gram-positive organisms that inhibits cell wall biosynthesis by binding to the D-Ala-D-Ala of lipid II and blocking peptidoglycan cross-linking. It exhibits poor activity against Gram-negative bacteria because it is too large to cross the outer membrane. Because modification of the C-terminal carboxylic acid with a PEG chain did not perturb its antibacterial activity, we selected this site as a point of attachment. Moreover, we employed Click chemistry for the conjugate assembly to avoid complications with the various functional groups exhibited by vancomycin. First, the azide-functionalized PEG linker 11 was coupled to 25 to afford the enterobactin-azide 41 in 68% yield. Copper(I)-catalyzed azide-alkyne cycloaddition of 41 with alkyne 8 subsequently afforded enterobactin-vancomycin 42 in 55% yield following hydrogenation and purification. This synthetic approach was extended to 43, a small analog of 42 that houses a tert-butyl cargo, and the strategy is also applicable to other alkyne-substituted cargos that are compatible with the benzyl deprotection conditions.
Scheme 4. Syntheses of enterobactin-cargo conjugates by Click chemistry.

**Enterobactin-Cargo Conjugates Coordinate Fe(III).** The optical absorption spectrum of each enterobactin-cargo conjugate exhibited catecholate absorption at ca. 316 nm (MeOH, rt). With the exception of 34, which afforded a yellow solution because of the coumarin moiety, methanolic solutions of each conjugate turned from colorless to wine-colored following the addition of ca. one equiv of aqueous Fe(III), and the expected ligand-to-metal charge transfer (LMCT) bands were observed, indicating Fe(III) coordination to the enterobactin catecholates (Figure S12-S14).

**Enterobactin-Cargo Conjugate Delivery to the E. coli Cytoplasm.** We employed three non-pathogenic E. coli strains that are defective in enterobactin synthesis, enterobactin transport, or ferric enterobactin utilization in growth recovery assays (Table S2). E. coli ATCC 33475 (ent-) cannot biosynthesize enterobactin, but retains the capacity to import and metabolize the siderophore. E. coli H1187 (fepA-) lacks the outer membrane enterobactin
receptor. *E. coli* K-12 JW0576 (fes-) can accumulate ferric enterobactin, but cannot release the iron because it is deficient in the enterobactin esterase Fes. As a result of these defects in iron metabolism, all three strains grow poorly under conditions of iron limitation.\textsuperscript{71} The iron chelator dipyridyl (DP) was used to generate iron-deficient conditions and promote expression of siderophore transport machinery in the growth recovery assays.

We first evaluated whether the enterobactin conjugates afforded growth recovery of *E. coli* (ent-) cultured under iron-deficient conditions (50% MHB, 200 \mu M DP). *E. coli* (ent-) grew to \( \text{OD}_{600} \sim 0.35 \) in 50\% MHB medium (30 \( ^\circ \)C, \( t = 19 \) h), and this value decreased to <0.05 when 200 \( \mu \)M DP was added to the media. Low-micromolar concentrations of L-Ent restored growth, as expected,\textsuperscript{70} and the *E. coli* cultures reached \( \text{OD}_{600} \sim 0.2 \) in the presence of 10 \( \mu \)M Ent (Figure 3). Likewise, low-micromolar concentrations of the enterobactin-cargo conjugates 29-33 and 43 exhibiting Boc (29, 43), cyclohexyl (30), napthyl (32), and phenylmethylbenzyl (33) cargos afforded growth recovery to similar levels (Figures 3 and S15). No growth restoration was observed when *E. coli* (fepA-) or *E. coli* (fes-) were cultured with 29 or 30 (Figures 5 and S16), which supports the notion that the growth recovery of *E. coli* (ent-) results from FepA-mediated cytoplasmic transport and Fes-catalyzed hydrolysis of the enterobactin moiety to release iron.

Moreover, the D-enantiomer of enterobactin, D-Ent 9, is not a substrate for Fes and does not provide growth recovery (Figures 4 and S17).\textsuperscript{66} Indeed, no growth promotion occurred when *E. coli* (ent-) was treated with conjugate 31, the D-enantiomer of 30 (Figures 4 and S15). Taken together, these results demonstrate that the enterobactin transport machinery has the capacity to recognize and transport cargo-derivatized enterobactin scaffolds to the *E. coli* cytoplasm, and that these molecules are substrates for the cytoplasmic esterase Fes.
Figure 3. E. coli ATCC 33475 (ent-) and P. aeruginosa PAO1 (pvd-, pch-) growth recovery assays employing L-Ent and select enterobactin-cargo conjugates (50% MHB, ± 200 or 600 µM DP, t = 19 h, 30 °C). Grey bars: OD\textsubscript{600} of bacteria cultured in the absence of DP. Black bars: OD\textsubscript{600} of bacteria cultured in the presence of 200 (E. coli) or 600 (P. aeruginosa) µM DP. (A) L-Ent promotes growth recovery of E. coli. (B) Enterobactin conjugate 30 housing a cyclohexyl cargo affords growth recovery of E. coli. (C) Enterobactin conjugate 34 housing a coumarin moiety affords little-to-no growth recovery of E. coli. (D) L-Ent promotes growth recovery of P. aeruginosa. (E) Enterobactin conjugate 30 housing a cyclohexyl cargo affords growth recovery of P. aeruginosa. (F) Enterobactin conjugate 34 housing a coumarin moiety affords growth recovery of P. aeruginosa. Figures 4 and S15 contain the assay results for the other conjugates. Each bar indicates the average of three independent replicates (two wells per replicate) and the error bars are the standard deviation of the mean.
Figure 4. The comparative effects of enterobactin-cargo conjugates on bacterial cell growth. *E. coli* and *P. aeruginosa* were cultured in the presence of 10 µM of L-Ent 1, D-Ent 9 and the enterobactin-cargo conjugates 29-35, 40, 42, 43 in the absence (grey bars) and presence (black bars) of DP (50% MHB, T = 30 °C, t = 19 h). (A) *E. coli* ATCC 33475 (*ent*) and the DP concentration was 200 µM. (B) *P. aeruginosa* PAO1 (*pvd-, pch-*) and the DP concentration was 600 µM. NC refers to a no-conjugate control.

We observed no convincing evidence for marked uptake of larger cargos by *E. coli* ATCC 33475, which suggests that FepA of this *E. coli* strain has a cargo size limit. For instance, under iron limitation, negligible *E. coli* growth recovery and no toxicity was observed following treatment with the enterobactin-coumarin conjugate 34 (Figure 3), suggesting that *E. coli* (*ent*) may not readily import 34. Moreover, no growth recovery occurred following treatment of *E. coli* with either ciprofloxacin 35 or 40 (Figures 4 and S15). In the absence of DP, these conjugates afforded a concentration-dependent inhibition of *E. coli* growth. Likewise, 10 µM vancomycin 42
inhibited the growth of *E. coli* (± DP, Figures 4 and S15). This behavior contrasts that of unmodified vancomycin, which is inactive against *E. coli* over the concentration range employed in this study. Two possible origins for inhibitory activity of the ciprofloxacin and vancomycin conjugates are (i) enterobactin-antibiotic uptake and resulting antibacterial action or (ii) a lack of active transport into *E. coli*, resulting in extracellular iron chelation and hence nutrient deprivation. Taking all observations into account, including those for *P. aeruginosa* described below, we contend that the latter option is the most probable explanation.

**Figure 5.** Comparison of growth recovery for *E. coli* (ent-), *E. coli* (fepA-), and *E. coli* (fes-) with conjugate 29 in the presence of 200 µM DP. Black bars: *E. coli* (fes-) cultured with conjugate 29; white bars: *E. coli* (fepA-) cultured with conjugate 29; grey bars, *E. coli* (ent-) cultured with conjugate 29 in the presence of 200 µM DP. See Figures 4 and S16 for additional data.

**Enterobactin-Cargo Conjugate Delivery to the *P. aeruginosa* Cytoplasm.** We next sought to determine whether the enterobactin-cargo conjugates provided growth recovery for *Pseudomonas aeruginosa* PAO1. This Gram-negative opportunistic human pathogen synthesizes and exports two siderophores, pyoverdine (pvd) and pyochelin (pch), and employs
multiple additional mechanisms for iron acquisition.\textsuperscript{72,73} \textit{P. aeruginosa} utilizes enterobactin as a xenosiderophore, and the genes pfeA\textsuperscript{74,75} and pirA\textsuperscript{76} encode outer membrane enterobactin transporters. Similar to the \textit{E. coli} experiments, we focused on using \textit{P. aeruginosa} strains deficient in siderophore production or utilization in growth recovery assays. \textit{P. aeruginosa} K648 (pvd-, pch-) is deficient in both pyoverdine and pyochelin biosynthesis, and shows attenuated growth in iron-deficient conditions, whereas \textit{P. aeruginosa} K407 (pvd-, pFr-) is deficient in pyoverdine biosynthesis and lacks the enterobactin transporter PfeA.\textsuperscript{74}

In 50% MHB medium, \textit{P. aeruginosa} (pvd-, pch-) grew to OD\textsubscript{600} \~ 0.45 (30 °C, t = 19 h) and this value diminished to ca. 0.25 in the presence of 600 µM DP. Supplementation of the iron-limiting growth medium with low-micromolar concentrations of L-Ent resulted in the restoration of \textit{P. aeruginosa} growth to OD\textsubscript{600} \~ 0.40 (Figure 3). Comparable growth recovery was observed for cultures treated with eight of the nine conjugates based on L-Ent (Figures 3, 4 and S18). Vancomycin 42, which exhibits the largest cargo, afforded a growth inhibitory effect (±DP) as observed for \textit{E. coli} (ent-). In contrast to its L-Ent analog 30, conjugate 31 based on D-Ent was growth inhibitory as was D-Ent (Figures 4 and S18). This result demonstrates that \textit{P. aeruginosa} also requires the L-isomer for iron utilization. Lastly, no growth enhancement of \textit{P. aeruginosa} (pFr-) was observed in the presence of L-Ent or conjugate 30 (600 µM DP); instead, these siderophores caused growth inhibition at micromolar concentrations (Figure S19). These results demonstrate that PfeA is necessary for conjugate-mediated growth recovery, supporting its role as a transporter for the enterobactin conjugates. In total, these assays demonstrate that the enterobactin transport machinery of \textit{P. aeruginosa}, and PfeA in particular, recognizes and delivers various cargo-modified enterobactin scaffolds to the cytoplasm.

Ciprofloxacin is a fluoroquinolone antibiotic that acts in the cytoplasm and inhibits DNA gyrase.\textsuperscript{77} The fact that ciprofloxacin conjugates 35 and 40 each restored \textit{P. aeruginosa} growth demonstrated that the cargo was successfully delivered to the cytoplasm of this microbe with negligible impact of the variable linker composition, and that conjugation of ciprofloxacin to enterobactin attenuated its antibacterial activity. This observation is in general agreement with
reports of pyoverdine-fluoroquinolone\textsuperscript{78} and pyochelin-fluoroquinolone\textsuperscript{79,80} conjugates where the antibiotic was covalently attached to the siderophore and point to the need for appropriate linker design for fluoroquinolone delivery and release after cellular entry.\textsuperscript{81} These pyoverdine/pyochelin-antibiotic conjugates afforded no antipseudomonal activity or diminished activity relative to the unmodified drug, and the pyoverdine-fluoroquinolone antibiotic exhibited decreased \textit{E. coli} gyrase inhibitory activity \textit{in vitro}.\textsuperscript{78}

A comparison of the enterobactin-cargo growth recovery profiles for \textit{E. coli} and \textit{P. aeruginosa} (Figures 4, S15, S18) reveals that these particular microbes have different capacities for internalizing enterobactin-cargo conjugates, and that cargo size is an important factor. Vancomycin has a rigid dome-like structure and a molecular weight of ca. 1.4 KDa, and the assays presented in this work suggest that this molecule is too big for enterobactin-mediated transport into \textit{E. coli} or \textit{P. aeruginosa}. In contrast, small and malleable cargos such as a Boc protecting group and cyclohexane afforded growth recovery comparable to that of L-Ent for both strains. A comparison of OD\textsubscript{600} values for bacterial cultures treated with such conjugates (e.g. 29, 30, 32, 34) shows that growth recovery to levels comparable to that of L-Ent occurs at a conjugate concentration of 1 \textmu M for \textit{P. aeruginosa} whereas 10 \textmu M is required for \textit{E. coli}. \textit{P. aeruginosa} responds to lower Ent concentrations than \textit{E. coli}, which indicates a higher uptake efficiency. Coumarin 343 is an example of a cargo that exhibits no signs of toxicity over the concentration range tested and affords markedly different results on microbial growth promotion for these two species. A comparison of the ciprofloxacin conjugate data for \textit{E. coli} and \textit{P. aeruginosa} also suggests differential uptake. For both the ciprofloxacin and coumarin cargo, the growth recovery assays indicate that the enterobactin transport machinery of \textit{P. aeruginosa} imports these cargos whereas the \textit{E. coli} system does not. These observations suggest that species-selective targeting may be possible with strategic cargo choice even when a siderophore is utilized by multiple microbial species.
Summary and Perspectives

We have designed and prepared a family of monofunctionalized enterobactin derivatives, and utilized these scaffolds for the preparation of enterobactin-cargo conjugates bearing cargos of varying size and complexity. Growth recovery assays employing E. coli and P. aeruginosa revealed that the enterobactin uptake machineries of these Gram-negative species recognize and transport enterobactin-cargo conjugates to the Gram-negative cytoplasm. These studies are significant in several respects. First, the notion of using siderophores for antibiotic delivery across the Gram-negative outer membrane, which serves as a permeability barrier, has achieved long-term interest. Such “Trojan horse” antibiotics are largely inspired by the sideromycins, a family of siderophore-antibiotic conjugates produced by the soil bacterium Streptomyces, and by early observations that catechol-modified β-lactams were recognized by the iron-uptake machinery of Gram-negative microbes. Significant efforts have been made to prepare and characterize synthetic siderophore-antibiotic conjugates with the goal of targeting drug-resistant Gram-negative pathogens. Timely examples of siderophore-antibiotic conjugates with antimicrobial activity include a mycobactin-artemisinin conjugate that kills Mycobacterium tuberculosis and Plasmodium falciparum, and amoxicillin/ampicillin-appended tripodal triscatecholates that exhibit potent antipseudomonal activity relative to the parent β-lactam antibiotics. One bottleneck with this general approach, and using siderophores in other applications, is that few synthetically tractable and modifiable native siderophores are available. DFO B and pyoverdine, which are readily obtained commercially (DFO B) or from bacterial cultures (pyoverdines), provide free amino groups useful for conjugation and are most commonly derivatized for application-based work. Syntheses of modified pyochelin, petrobactin, and mycobactin platforms that house functional groups amenable to site-specific elaboration have been reported, and these scaffolds are important contributions to the toolkit of siderophores that can be modified without compromising Fe(III) coordination in addition to recognition by siderophore-binding proteins. The syntheses described in this work provide enterobactin with a functional handle for versatile chemical modifications,
and will allow strategic use of this canonical siderophore in a multitude of chemical biology and biotechnology initiatives.

Unanswered questions regarding the antibacterial activity and fate of reported synthetic siderophore-antibiotic conjugates exist. Whether a given conjugate is actively transported into the bacterial cell is oftentimes unclear. Because FepA recognizes relatively large biomolecules including MccE492m (84-aa) and colicin B (324-aa), it is tempting to predict that FepA may accommodate almost any cargo appended to an enterobactin or catecholate platform. The results presented in this work challenge this notion and indicate that cargo size is an important and species-specific parameter. Our assays indicate that P. aeruginosa PAO1 has a greater capacity to import enterobactin-cargo conjugates than E. coli ATCC 33475. It will be interesting to determine the cargo scope of other E. coli strains and bacterial species that utilize enterobactin for iron acquisition, and understand the molecular and physiological basis for such variations. Colicins are largely α-helical and MccE492m shares some sequence homology with colicins. It is likely that some enterobactin receptors have decreased propensity to transport synthetic small molecules or natural products with less structural malleability (i.e. vancomycin) than an α-helical peptide.

The mechanisms of iron release from siderophores, which vary tremendously for the myriad of siderophores produced by different bacterial species, are another important consideration in siderophore-cargo conjugate design. Guided by studies of chiral recognition in enterobactin transport, which demonstrated that D-Ent is transported into E. coli but cannot be hydrolyzed by Fes, we designed the monofunctionalized D-Ent scaffolds to probe cytosolic delivery. This design feature prevents esterase-catalyzed iron release from enterobactin-based conjugates in the cytoplasm and may have practical utility. From the standpoint of drug delivery, a tug-of-war may result from utilizing an iron-supplying siderophore that confers a growth advantage for delivering a toxic payload to a bacterial cell, and preventing iron release may be beneficial. In other applications, siderophore-fluorophore conjugates are of interest for bacterial
detection and diagnostics, and Fe(III) binding to and release from the siderophore will likely influence the photophysical properties of such molecules.

In summary, these investigations reveal that the enterobactin transport machineries of *E. coli* (e.g. FepABCDG and TonB-ExbB-ExbD) and *P. aeruginosa* will deliver enterobactin-modified cargo to the Gram-negative cytoplasm. Moreover, the preparative work affords a new siderophore platform amenable to synthetic elaboration and an entry route for employing the native enterobactin scaffold in a multitude of application-based initiatives that include intracellular cargo delivery, iron sensing, siderophore labeling, protein and pathogen detection, and therapeutic development.

**Acknowledgements.** The Searle Scholars Program (Kinship Foundation), the Department of Chemistry and the Undergraduate Research Opportunities Program (UROP) at MIT, and the Amgen Scholars Program (J.L.B) are gratefully acknowledged for financial support. We thank Professor Keith Poole for providing the *Pseudomonas aeruginosa* strains employed in this work, Professor Klaus Hantke for providing the *E. coli* H1178 (fepA-) strain, Professor Stephen J. Lippard for use an IR spectrophotometer and a melting point apparatus, and Dr. Andrew Wommack for carefully proof-reading the manuscript. *E. coli* K-12 JW0576 was obtained from the Keio Collection. NMR instrumentation maintained by the MIT DCIF is supported by NSF grants CHE-9808061 and DBI-9729592.

**Supporting Information.** Syntheses and characterization of 14-18 and 26-28, general liquid chromatography, mass spectrometry and microbiology methods, summary of enterobactin-cargo conjugate characterization (Table S1), summary of bacterial strains and sources (Table S2), structure of MccE492m (Figure S1), HPLC traces for the purified conjugates (Figures S2-S11), optical absorption spectra (Figures S12-S14), growth recovery assays (Figures S15-19), H and C NMR spectra, and IR spectroscopic data. This material is available free of charge via the Internet at http://pubs.acs.org.
References


Supporting Information for
Siderophore-Mediated Cargo Delivery to the Cytoplasm of Escherichia coli and Pseudomonas aeruginosa: Syntheses of Monofunctionalized Enterobactin Scaffolds and Evaluation of Enterobactin-Cargo Conjugate Uptake

Tengfei Zheng, Justin L. Bullock, and Elizabeth M. Nolan*

Department of Chemistry, Massachusetts Institute of Technology, Cambridge, MA 02139

*Corresponding author: inolan@mit.edu

Phone: 617-452-2495
Fax: 617-324-0505
This Supporting Information includes:

**Supplementary Experimental Section** ................................................................. S3
Synthesis of 14 ........................................................................................................... S3
Synthesis of 15 ........................................................................................................... S3
Synthesis of 16 ........................................................................................................... S4
Synthesis of 17 ........................................................................................................... S4
Synthesis of 18 ........................................................................................................... S5
Synthesis of 26 ........................................................................................................... S5
Synthesis of 27 ........................................................................................................... S6
Synthesis of 28 ........................................................................................................... S7
General Liquid Chromatography and Mass Spectrometry Methods ..................... S8
General Microbiology Methods ............................................................................. S9

Scheme S1. Syntheses of 14-18 .................................................................................. S11

Table S1. Characterization of enterobactin-conjugates .............................................. S12

Table S2. Bacterial strains employed in this study ..................................................... S13

Figure S1. Structure of MccE492m ........................................................................... S14

Figures S2-S11. Analytical HPLC of purified enterobactin-cargo conjugates ............. S15

Figures S12-S14. Optical absorption spectra of enterobactin-cargo conjugates ........ S20

Figure S15. Growth recovery assays with *E. coli* ATCC 33475 (ent-) ..................... S23

Figure S16. Growth recovery assays with *E. coli* H1187 (fepA-) and JW0576 (fes-) ... S24

Figure S17. Growth recovery assays with D-Ent ........................................................ S25

Figure S18. Growth recovery assay with *P. aeruginosa* PAO1 K648 (pvd-, pch-) .... S26

Figure S19. Growth recovery assay with *P. aeruginosa* PAO1 K407 (pvd-, pFr-) ... S27

Spectroscopic data ..................................................................................................... S28
Supplementary Experimental Section

*N-(2-(2-(2-(2-Aminoethoxy)ethoxy)ethoxy)ethyl)cyclohexanecarboxamide* (14).

Cyclohexanecarboxylic acid (64 mg, 0.50 mmol) and 7 (192 mg, 0.599 mmol) were combined in 5 mL of dry CH₂Cl₂, and 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide (EDC, 143 mg, 0.751 mmol), 4-dimethylaminopyridine (DMAP, 30 mg, 0.25 mmol), and DIPEA (435 μL, 2.52 mmol) were added. The reaction was stirred for 4 h at rt, and the organic phase was washed with 50 mM HCl (3 x 20 mL) and brine (1 x 20 mL). The organic phase was dried over Na₂SO₄ and concentrated. Flash chromatography on silica gel with a solvent gradient (CH₂Cl₂ to 10% MeOH/CH₂Cl₂) afforded the Boc-protected product as a colorless oil (190 mg, 94%). TLC R_f = 0.65 (10% MeOH/CH₂Cl₂). ¹H NMR (CDCl₃, 300 MHz), δ 1.13-1.37 (14H, m), 1.58-1.60 (1H, m), 1.68-1.78 (4H, m), 1.97-2.05 (1H, m), 3.23-3.24 (2H, m), 3.34-3.40 (2H, m), 3.45-3.50 (4H, m), 3.53-3.59 (8H, m), 5.09 (1H, bs), 6.11 (1H, bs). ¹³C NMR (CDCl₃, 125 MHz), δ 25.5, 28.2, 29.4, 38.7, 40.1, 45.2, 69.7, 69.9, 70.0, 70.2, 78.9, 155.8, 176.0. HRMS (ESI): [M+Na⁺]⁺ m/z calcd., 425.2622; found, 425.2654.

A portion of this Boc-protected product (118 mg, 0.278 mmol) was dissolved in 2.5 mL of 40% TFA/CH₂Cl₂ and the light red solution was stirred at rt for 2.5 h. The reaction was concentrated to give 14 as a light-yellow oil in quantitative yield. TLC R_f = 0.2 (10% MeOH/CH₂Cl₂). ¹H NMR (CDCl₃, 500 MHz), δ 1.13-1.37 (5H, m), 1.62-1.64 (1H, m), 1.71-1.77 (4H, m), 2.10-2.15 (1H, m), 3.16 (2H, bs), 3.36 (2H, bs), 3.48-3.65 (10H, m), 3.76 (2H, bs), 6.93 (1H, bs), 7.88 (2H, 3s). ¹³C NMR (CDCl₃, 125 MHz), δ 25.4, 25.5, 29.4, 39.1, 39.7, 45.1, 66.6, 69.6, 69.8, 69.9, 145.8. HRMS (ESI): [M+Na⁺]⁺ m/z calcd., 325.2098; found, 325.2119.

*N-(2-(2-(2-(2-Aminoethoxy)ethoxy)ethoxy)ethyl)-2-naphthamide* (15). Compound 15 was synthesized as described for 14 except that 2-naphthoic acid (86 mg, 0.50 mmol) was used instead of cyclohexanecarboxylic acid. The Boc-protected product was obtained as light yellow oil (178 mg, 80%). TLC R_f = 0.7 (10% MeOH/CH₂Cl₂). ¹H NMR (CDCl₃, 500 MHz), δ 1.36 (9H, s), 3.11-3.12 (2H, m), 3.28 (2H, t, J = 5.2 Hz), 3.34-3.35 (2H, m), 3.45 (2H, t, J = 4.5 Hz), 3.54-3.60 (4H, m), 3.65 (4H, bs), 4.99 (1H, bs), 6.82 (1H, bs), 7.37 (1H, dd, J = 7.8, 7.8 Hz), 7.43-
7.49 (2H, m), 7.55 (1H, d, J = 7.0 Hz), 7.79-7.83 (2H, m), 8.26 (1H, d, J = 8.0 Hz). $^{13}$C NMR (CDCl$_3$, 125 MHz), δ 28.2, 39.5, 39.9, 69.5, 69.7, 70.0, 70.1, 70.2, 78.9, 124.5, 124.9, 125.2, 126.1, 126.7, 128.0, 129.9, 130.2, 133.4, 134.3, 155.7, 169.4. HRMS (ESI): [M+Na$^+$] $m/z$ calcd., 469.2309; found, 469.2335.

Compound 15 was obtained as light orange oil (quantitative yield from 91.6 mg, 0.205 mmol of the Boc-protected precursor). TLC $R_f$ = 0.15 (10% MeOH/CH$_2$Cl$_2$). $^1$H NMR (CDCl$_3$, 300 MHz), δ 3.06 (2H, bs), 3.56-3.66 (14H, m), 7.15 (1H, bs), 7.40-7.56 (6H, m), 7.85-7.94 (2H, m), 8.08-8.11 (1H, m). $^{13}$C NMR (CDCl$_3$, 125 MHz), δ 39.3, 39.6, 66.4, 69.6, 69.6, 69.7, 69.9, 124.7, 125.0, 125.1, 126.3, 127.0, 128.3, 129.8, 130.6, 133.5, 133.7, 170.7. δ HRMS (ESI): [M+Na$^+$] $m/z$ calcd., 369.1785; found, 369.1806.

$N$-(2-(2-(2-(2-Aminoethoxy)ethoxy)ethoxy)ethyl)-4-benzylbenzamide (16). Compound 16 was synthesized as described for 14 except that 4-benzylbenzoic acid (106 mg, 0.50 mmol) was used instead of cyclohexanecarboxylic acid. The Boc-protected product was obtained as light yellow oil (220 mg, 90%). TLC $R_f$ = 0.7 (10% MeOH/CH$_2$Cl$_2$). $^1$H NMR (CDCl$_3$, 500 MHz), δ 1.48 (9H, s), 3.32 (2H, s), 3.51-3.54 (1H, m), 3.61-3.71(11H, m), 4.07 (2H, s), 5.10 (1H, s), 6.82 (1H, s), 7.21-7.34 (6H, m), 7.78-7.81 (2H, m). $^{13}$C NMR (CDCl$_3$, 125 MHz), δ 28.3, 39.6, 40.2, 41.7, 69.8, 70.1, 70.1, 70.2, 70.4, 70.4, 79.1, 94.0, 126.2, 127.2, 128.5, 128.8, 128.9, 132.4, 140.3, 144.7, 145.8, 145.8, 155.9, 167.3. HRMS (ESI): [M+Na$^+$] $m/z$ calcd., 509.2622; found, 509.2628.

Compound 16 was obtained as brown oil (quantitative yield from 220 mg of the Boc-protected precursor). TLC $R_f$ = 0.4 (10% MeOH/CH$_2$Cl$_2$). $^1$H NMR (CDCl$_3$, 500 MHz), δ 3.12 (2H, s), 3.55-3.62 (12H, m), 3.63-3.71 (2H, m), 4.00 (2H, s), 7.15-7.30 (8H, m), 7.68-7.73 (5H, m), 8.49 (3H, m). $^{13}$C (CDCl$_3$, 125 MHz), δ 39.2, 39.5, 41.6, 67.1, 66.8, 69.6, 70.09, 126.2, 127.4, 128.5, 128.7, 128.9, 131.5, 140.3, 145.1. [M+Na$^+$] $m/z$ calcd., 409.2098; found, 409.2093.

$N$-(2-(2-(2-(2-Aminoethoxy)ethoxy)ethoxy)ethyl)-11-oxo-2,3,5,6,7,11-hexahydro-1H-pyrano[2,3-f]pyrido[3,2,1-ij]quinoline-10-carboxamide (17). Coumarin 343 (142 mg, 0.50
mmol), EDC (143 mg, 0.751 mmol), DMAP (30 mg, 0.25 mmol), and DIPEA (435 μL, 2.52 mmol) were mixed in 15 mL of CH₂Cl₂. A portion (280 μL, 1.50 mmol) of 2,2'-(oxybis(ethane-2,1-diyl))bis(oxy))diethanamine was added and the reaction mixture was stirred overnight at rt. The solvent was removed under reduced pressure and 17 was purified by preparative TLC (15% MeOH/CH₂Cl₂ with 1% TEA) and obtained as an orange oil (86 mg, 38%). TLC Rᵋ = 0.8 (10% MeOH/CH₂Cl₂). ¹H NMR (CDCl₃, 300 MHz), δ 1.95 (4H, bs), 2.75 (2H, t, J = 6.0 Hz), 2.85 (2H, t, J = 6.3 Hz), 3.22 (2H, bs), 3.29-3.34 (4H, m), 3.56-3.70 (14H, m), 3.89 (2H, bs), 7.03 (1H, s), 7.65 (2H, bs), 8.55 (1H, s), 9.10 (1H, s). ¹³C NMR (CDCl₃, 125 MHz), δ 19.8, 19.9, 20.8, 27.2, 39.3, 39.9, 49.6, 50.0, 66.5, 69.8, 69.9, 70.0, 105.3, 107.8, 108.0, 119.6, 127.0, 148.1, 148.2, 152.4, 162.8, 164.1. HRMS (ESI): [M+H]^+ m/z calcd., 460.2442; found, 460.2435.

7-(4-(3-(2-(2-Aminoethoxy)ethoxy)ethoxy)propanoyl)piperazin-1-yl)-1-cyclopropyl-6-fluoro-4-oxo-1,4-dihydroquinoline-3-carboxylic acid (18). Ciprofloxacin (115 mg, 0.50 mmol) and DIPEA (0.5 mL, 2.8 mmol) were mixed in 5 mL of CH₂Cl₂, and TMSCl (135 μL, 1.45 mmol) was added to give a clear yellow solution. Fmoc-PEG-CO₂H 13 (333 mg, 1.50 mmol), EDC (144 mg, 1.50 mmol), DMAP (30 mg, 0.050 mmol), and DIPEA (0.35 mL, 2 mmol) were dissolved in 2 mL of dry CH₂Cl₂, and the two solutions were combined and stirred at rt overnight. The resulting solution was washed with water (1x10 mL), 0.1M HCl (2x20 mL), and brine (1x20 mL), dried over Na₂SO₄, and purified by flash chromatography on silica gel (3% isopropanol/CH₂Cl₂) to give the product as yellow solid (206 mg, 54%). TLC Rᵋ = 0.6 (10% MeOH/CH₂Cl₂); mp = 83 °C (decomp). ¹H NMR (CDCl₃, 300 MHz), δ 1.11-1.16 (2H, m), 1.30-1.34 (2H, m), 2.64 (2H, t, J = 6.6 Hz), 3.23-3.69 (19H, m), 3.78-3.83 (4H, m), 4.17 (1H, t, J = 6.9 Hz), 4.35 (2H, d, J = 6.9 Hz), 5.64-5.66 (1H, m), 7.26-7.31 (3H, m), 7.34-7.39 (2H, m), 7.57-7.59 (2H, m), 7.70-7.73 (2H, m), 7.90-7.94 (1H, m), 8.66 (1H, s), 14.9 (1H, s). ¹³C NMR (CDCl₃, 125 MHz), δ 7.7, 33.0, 35.0, 40.5, 40.7, 44.9, 46.8, 49.5, 66.1, 66.8, 69.7, 69.8, 70.0, 70.0, 104.7, 170.2, 111.3, 111.6, 119.0, 119.1, 119.5, 124.7, 126.7, 127.3, 138.5, 140.7, 143.5, 144.8, 145.0, 146.9, 151.3, 154.6, 156.2, 166.4, 169.3, 176.2, 176.3. ¹⁹F NMR (CDCl₃, 282 MHz) δ -121.2. HRMS (ESI): [M+Na]^+ m/z calcd., 779.3063; found, 779.3052.
A portion of this product (182 mg, 0.240 mmol) was dissolved in CH₂Cl₂ (2 mL) and diethylamine (2 mL, 19.3 mmol) was added. The solution was stirred for 2 h and concentrated under reduced pressure, and this procedure was repeated. A portion of the crude yellow product was dissolved in 3:7 H₂O/DMSO and purified by semi-preparative HPLC (20% B for 5 min followed by 20%-70% B over 20 min, 4 mL/min). Compound 18 eluted at 11 min and a yellow powder was obtained after lyophilization (38 mg). TLC Rf = 0.15 (10% MeOH/CH₂Cl₂). ¹H NMR (CDCl₃, 500 MHz), δ 1.15 (2H, bs), 1.41 (2H, bs), 2.70 (2H, bs), 3.19 (2H, bs), 3.30-3.37 (4H, m), 3.62-3.82 (17 H, m), 7.37 (1H, bs), 7.83-7.85 (1H, m), 8.16 (3H, bs), 8.65 (1H, s). ¹³C NMR (CDCl₃, 125 MHz), δ 8.1, 33.1, 35.5, 40.0, 41.3, 45.2, 49.0, 49.6, 66.4, 67.1, 69.9, 70.1, 70.2, 105.4, 107.7, 111.9, 112.1, 119.8, 119.9, 139.0, 145.2, 145.3, 147.4, 152.4, 152.4, 167.0, 170.4, 176.8. ¹⁹F NMR (CDCl₃, 282 MHz) δ -121.4, -75.6. HRMS (ESI): [M+H]+ m/z calcd., 535.2563; found, 535.2578.

N,N'-(3S,7S,11S)-11-(2,3-bis(benzyloxy)-5-((E)-prop-1-en-1-yl)benzamido)-2,6,10-trioxo-1,5,9-trioxacyclododecane-3,7-diyl)bis(2,3-bis(benzyloxy)benzamide) (26). Compound 5 (0.441 g, 1.19 mmol) was dissolved in 8 mL of DMSO and DIPEA (2.28 mL, 13.1 mmol) was added. In a separate flask, compounds 6 (0.591 g, 1.78 mmol), 22 (0.669 g, 1.78 mmol), and PyAOP(2.48 g, 4.76 mmol) were combined in 15 mL of DMSO. This mixture was added drop wise to the solution of 5, and the resulting solution was stirred for 2 h at rt during which time it turned dark red-brown. The reaction was diluted with 50 mL of EtOAc and 25 mL of water. The layers were partitioned, and the organic phase was washed with saturated brine (2 x 25 mL). The organic phase was dried over Na₂SO₄ and concentrated under reduced pressure, which yielded a red-orange oil. Flash chromatography on silica gel with a solvent gradient (10% EtOAc/hexanes to 50% EtOAc/hexanes) afforded 26 as a white-yellow solid (207 mg, 15%). TLC Rf = 0.4 (50% EtOAc/Hexanes). ¹H NMR (CDCl₃, 500 MHz), δ 1.88-1.89 (3H, m) 4.01-4.04 (3H, m) 4.14-4.16 (3H, m) 4.91-4.92 (3H, m) 5.02-5.27 (12H, m) 6.18-6.22 (1H, m) 6.35-6.40 (1H, m) 7.12-7.68 (36H, m) 8.48-8.49 (3H, d, J=7.5) ¹³C NMR (CDCl₃, 500 MHz) δ 18.4, 51.4, 51.4, 64.1, 71.2, 76.3, 76.4, 76.7, 117.6, 120.7, 123.2, 124.3, 126.0, 126.4, 126.5, 127.6, 127.7,
127.8, 128.2, 128.4, 128.6, 128.6, 128.8, 128.8, 128.9, 128.9, 129.3, 129.8, 134.3, 136.0, 136.3, 147.0, 151.6, 164.9, 169.1. HRMS (ESI): [M+Na]+ m/z calcd., 1272.4464; found, 1272.4434.

$N,N'-(\text{(3S,7S,11S)-11-(2,3-bis(benzyloxy)-5-formylbenzamido)-2,6,10-trioxo-1,5,9-trioxacyclododecane-3,7-diyl)bis(2,3-bis(benzyloxy)benzamide)}}$ (27): Compound 26 (159 mg, 0.127 mmol) was dissolved in 6 mL of 1,4-dioxane. Water (2 mL) was slowly added drop wise to the solution. With each drop of water, a white cloudy precipitate appeared and then disappeared. OsO$_4$ (39 µL of a 2.5 wt % solution in 2-methyl-2-propanol, 3.8 µmol) was then added to the solution, and the reaction was stirred for 0.5 h, which yielded a brown solution. NaIO$_4$ (67 mg, 0.42 mmol) was added to the reaction, which was stirred for 2 h and a white precipitate formed. The reaction was diluted with EtOAc (25 mL) and water (10 mL). The organic phase was washed with 0.1 M NaS$_2$O$_3$ (3 x 20 mL) and brine (1 x 20 mL). The organic phase gained a yellow tint with the addition of NaS$_2$O$_3$. The organic phase was dried over Na$_2$SO$_4$ and concentrated in vacuo. Flash chromatography on silica gel using a solvent gradient (20% EtOAc/hexanes to 70% EtOAc/hexanes) yielded the product as a white foam (101 mg, 64%). TLC $R_f$ = 0.7 (10% MeOH/CH$_2$Cl$_2$). $^1$H NMR (CDCl$_3$, 500 MHz), $\delta$ 4.00-4.08 (3H, m), 4.15-4.22 (3H, m), 4.88-4.93 (3H, m), 5.03-5.26 (12H, m), 7.08-7.43 (31H, m) 7.65-7.65 (2H, m), 8.14 (1H, s), 8.41-8.47 (3H, m), 9.88 (1H, s). $^{13}$C NMR (CDCl$_3$, 125 MHz) $\delta$ 51.4, 51.5, 51.5, 51.7, 64.1, 64.3, 71.2, 71.3, 76.3, 76.6, 113.2, 117.5, 117.5, 123.1, 123.2, 124.3, 126.3, 126.3, 126.6, 127.6, 127.6, 127.8, 128.2, 128.2, 128.4, 128.4, 128.5, 128.6, 128.6, 128.7, 128.9, 128.9, 129.0, 132.2, 135.3, 135.4, 136.0, 136.0, 136.2, 146.8, 146.9, 151.6, 151.8, 152.3, 163.8, 165.0, 165.0, 168.8, 169.0, 169.2, 190.7. HRMS (ESI): [M+Na]$^+$ m/z calcd., 1260.4101; found, 1260.4094.

$3,4$-Bis(benzyloxy)-5-(((3S,7S,11S)-7,11-bis(2,3-bis(benzyloxy)benzamido)-2,6,10-trioxo-1,5,9-trioxacyclododecan-3-yl)carbamoyl)benzoic acid (28): Aldehyde 27 (0.092 g, 0.074 mmol) was dissolved in 2 mL of 1,4-dioxane. In a separate flask, NH$_3$SO$_3$ (12.8 mg, 0.132 mmol) was dissolved in 0.5 mL of H$_2$O. The sulfamic acid solution was added to the 1,4-dioxane solution. The reaction turned cloudy shortly after the addition of NH$_3$SO$_3$. NaClO$_2$ (12
mg, 0.13 mmol) was dissolved in 0.4 mL of H₂O and added drop wise over ten minutes to the dioxane solution. The reaction was stirred for 30 min and diluted with 10mL of H₂O and 10 mL of EtOAc. The aqueous phase was back extracted with EtOAc (2 x 10 mL), and the combined organic phases were dried over Na₂SO₄ and concentrated under reduced pressure. Flash chromatography on silica gel with a solvent gradient (CH₂Cl₂ to 10% MeOH/CH₂Cl₂) yielded a white solid (83 mg, 89%). TLC \( R_f = 0.6 \) (10% MeOH/CH₂Cl₂). \(^1\)H NMR (CDCl₃, 500 MHz), \( \delta \) 4.08-4.11 (3H, m), 4.25-4.28 (3H, m), 4.96-5.00 (3H, m), 5.08-5.29 (12H, m), 7.14-7.51 (30H, m), 7.70-7.72 (2H, m), 7.89 (1H, s), 8.45-8.48 (2H, m), 8.53-8.59 (2H, m). \(^{13}\)C NMR (CDCl₃, 125 MHz), \( \delta \) 51.5, 51.5, 51.6, 53.6, 64.2, 71.2, 71.3, 76.3, 76.5, 117.5, 117.7, 123.1, 124.3, 125.7, 126.3, 127.6, 127.9, 128.2, 128.4, 128.5, 128.6, 128.7, 128.8, 128.9, 129.0, 135.5, 135.6, 136.0, 136.2, 145.9, 146.9, 150.8, 151.4, 151.6, 164.1, 165.1, 168.9, 169.0, 169.1, 169.6. HRMS (DART): [M+H]+ m/z calc., 1254.4230; found, 1254.4225.

**General Liquid Chromatography and Mass Spectrometry Methods.** HPLC-grade acetonitrile (MeCN) and trifluoroacetic acid (TFA) were purchased from EMD. LC-MS grade MeCN containing 0.1% formic acid and water containing 0.1% formic acid were obtained from J. T. Baker. Semi-preparative and analytical high-performance liquid chromatography (HPLC) were performed by using an Agilent 1200 series HPLC system outfitted with an Agilent Zorbax reverse-phase C18 column (5 \( \mu \)m pore size, 9.4 x 250 mm) at a flow rate of 4 mL/min and a Clipeus reverse-phase C18 column (5 \( \mu \)m pore size, 4.6 x 250 mm; Higgins Analytical, Inc.) at a flow rate of 1 mL/min, respectively. The multi-wavelength detector was set to read the absorption at 220, 280, and 316 (catecholate absorption) nm using a reference wavelength of 500 nm. For all HPLC runs, solvent A was 0.1% TFA/H₂O and solvent B was 0.1% TFA/MeCN. Each semi-preparative or analytical run began with a five-minute equilibration at the %B used at the start of the gradient followed by a gradient of increasing %B. The HPLC solvents were prepared with HPLC-grade MeCN and TFA, and Milli-Q water (18.2 mΩcm⁻¹), and filtered through a 0.2-\( \mu \)m filter before use. For analytical HPLC to evaluate conjugate purity, the entire portion of each HPLC-purified compound was dissolved in a mixture of 1:1:1 1,4-
dioxane/methanol/water and an aliquot was taken for HPLC analysis, and the solution was subsequently lyophilized. Conjugate 40 was an exception, and this molecule was dissolved in DMSO prior to analytical HPLC. Most high-resolution mass spectrometry was performed by using an Agilent LC-MS system comprised of an Agilent 1260 series LC system outfitted with an Agilent Poroshell 120 EC-C18 column (2.7 μm pore size) and an Agilent 6230 TOF system housing an Agilent Jetstream ESI source. For all LC-MS analyses, solvent A was 0.1% formic acid / H₂O and solvent B was 0.1% formic acid / MeCN. The samples were run using a gradient of 5-95% B over five min with a flow rate of 0.4 mL/min. In some instances, high-resolution mass spectrometry was performed by staff at the MIT Department of Chemistry Instrumentation Facility, which houses a Bruker Daltonics APEXIV 4.7 Tesla Fourier Transform Ion Cyclotron Resonance Mass Spectrometer (FT-ICR-MS) with a direct analysis in real time (DART) ionization source.

**General Microbiology Materials and Methods.** *E. coli* 33475 (ent-) was purchased from American Type Culture Collection (ATCC). *E. coli* K-12 JW0576 (fes-) was obtained from the Keio Collection (Japan). *Pseudomonas aeruginosa* K648 (pvd’/pch’) and K407 (pvd-/pFr-) were gifts from Professor Keith Poole (Department of Biomedical and Molecular Sciences, Queen's University, Canada). Freezer stocks of all *E. coli* strains were prepared in 25% glycerol/Luria Broth (LB) medium. Freezer stocks of all *Pseudomonas aeruginosa* strains were prepared in 7.5% DMSO/LB base medium supplemented with 2.5 g/L NaCl. Luria Broth (tryptone 10 g/L, yeast extract 5 g/L, NaCl 10 g/L), Luria Broth base (pancreatic digest of casein 10 g/L, yeast extract 5 g/L, NaCl 0.5 g/L), Mueller Hinton Broth (MHB, beef extract powder 2.0 g/L, acid digest of casein 17.5 g/L, soluble starch 1.5 g/L), and agar were purchased from BD. All growth media and Milli-Q water used for bacterial cultures or for preparing solutions of the enterobactin-cargo conjugates were sterilized by using an autoclave. The iron chelator 2,2’-dipyridyl (DP) was purchased from Sigma-Aldrich. A 200 mM DP stock solution was prepared in DMSO and used in the bacteria growth assays. All enterobactin-cargo conjugates, L-Ent and D-Ent were stored as DMSO stock solutions at -20 °C. With the exception of the coumarin 343 conjugate, the stock
solution concentrations were determined by using the reported extinction coefficient for enterobactin (316 nm, 9,500 M\(^{-1}\)cm\(^{-1}\))\(^{S1}\) with the assumption that the cargo had no effect on catecholate absorption. Working dilutions of the Ent conjugates, L-Ent, and D-Ent were prepared in 20% DMSO/H\(_2\)O, and the growth recovery assay cultures all contained 2% v/v DMSO. Sterile polypropylene culture tubes and sterile polystyrene 96-well plates used for culturing were manufactured by VWR and Corning Incorporated, respectively. OD\(_{600}\) values were recorded on an Agilent 8453 diode array spectrophotometer or by using a BioTek Synergy HT plate reader.

References
Scheme S1. Syntheses of PEG-derivatized cargos 14-18.
<table>
<thead>
<tr>
<th>No.</th>
<th>Cargo</th>
<th>HPLC retention time (min)</th>
<th>m/z obs</th>
<th>m/z cald.</th>
</tr>
</thead>
<tbody>
<tr>
<td>29</td>
<td>Boc</td>
<td>25.3</td>
<td>1010.3173</td>
<td>1010.3125</td>
</tr>
<tr>
<td>30</td>
<td>Cyclohexyl</td>
<td>24.6</td>
<td>1020.3346</td>
<td>1020.3333</td>
</tr>
<tr>
<td>31</td>
<td>Cyclohexyl (D-Ent)</td>
<td>24.6</td>
<td>1020.3328</td>
<td>1020.3333</td>
</tr>
<tr>
<td>32</td>
<td>Naphthal</td>
<td>25.3</td>
<td>1064.3086</td>
<td>1064.3020</td>
</tr>
<tr>
<td>33</td>
<td>Phenylmethylbenzyl</td>
<td>25.2</td>
<td>1104.3305</td>
<td>1104.3333</td>
</tr>
<tr>
<td>34</td>
<td>Coumarin 343</td>
<td>27.2</td>
<td>1177.3570</td>
<td>1177.3496</td>
</tr>
<tr>
<td>35</td>
<td>Ciprofloxacin (PEG)</td>
<td>24.9</td>
<td>1252.3633</td>
<td>1252.3617</td>
</tr>
<tr>
<td>40</td>
<td>Ciprofloxacin (alkyl)</td>
<td>26.7</td>
<td>1140.3482</td>
<td>1140.3486</td>
</tr>
<tr>
<td>42</td>
<td>Vancomycin (triazole)</td>
<td>20.4</td>
<td>1228.3796</td>
<td>1228.3796</td>
</tr>
<tr>
<td>43</td>
<td>Boc-glycine (triazole)</td>
<td>23.8</td>
<td>1126.3775</td>
<td>1126.3832</td>
</tr>
</tbody>
</table>

\(^a\) HPLC gradient used for all compounds is 0% B for 5 min followed by 0-100% B over 30 min, 1 mL/min. 
\(^b\) All m/z values correspond to [M+Na]^+ unless specified otherwise. 
\(^c\) The m/z value corresponds to [M+2Na]^{2+}/2. 
\(^d\) The m/z value corresponds to [M+H]^+. 
<table>
<thead>
<tr>
<th>Bacterial strain</th>
<th>Description</th>
<th>Source</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>E. coli</em> ATCC 33475</td>
<td>ent-</td>
<td>ATCC</td>
</tr>
<tr>
<td><em>E. coli</em> JW0576</td>
<td>fes-</td>
<td>Kieo Collection</td>
</tr>
<tr>
<td><em>E. coli</em> H1187</td>
<td>fepA-</td>
<td>Professor Klaus Hantke</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Universität Tübingen</td>
</tr>
<tr>
<td><em>P. aeruginosa</em> K648</td>
<td>pvd-, pch-</td>
<td>Professor Keith Poole</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Queen’s University, Canada</td>
</tr>
<tr>
<td><em>P. aeruginosa</em> K407</td>
<td>pvd-, pFr-</td>
<td>Professor Keith Poole</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Queen’s University Canada</td>
</tr>
</tbody>
</table>
Figure S1. Structure of MccE492m. The C-terminal amino acids are depicted by the cartoon.
Figure S2. Analytical HPLC traces of purified 29 (0% B for 5 min following by 0-100% B over 30 min, 1 mL/min). The minor peak at ca. 24.6 min is attributed to loss of the Boc group resulting from the acidic HPLC conditions.

Figure S3. Analytical HPLC traces of purified 30 (0% B for 5 min followed by 0-100% B over 30 min, 1 mL/min).
**Figure S4.** Analytical HPLC traces of purified **31** (0% B for 5 min followed by 0-100% B over 30 min, 1 mL/min).

**Figure S5.** Analytical HPLC traces of purified **32** (0% B for 5 min followed by 0-100% B over 30 min, 1 mL/min).
**Figure S6.** Analytical HPLC traces of purified 33 (0% B for 5 min followed by 0-100% B over 30 min, 1 mL/min).

**Figure S7.** Analytical HPLC traces of purified 34 (0% B for 5 min followed by 0-100% B over 30 min, 1 mL/min).
Figure S8. Analytical HPLC traces of purified 35 (0% B for 5 min followed by 0-100% B over 30 min, 1 mL/min).

Figure S9. Analytical HPLC traces of purified 40 (0% B for 5 min followed by 0-100% B over 30 min, 1 mL/min). The peak at ca. 5 min in the 220 nm trace is because of the DMSO solvent.
Figure S10. Analytical HPLC traces of purified 42 (0% B for 5 min followed by 0-100% B over 30 min, 1 mL/min).

Figure S11. Analytical HPLC traces of purified 43 (0% B for 5 min followed by 0-100% B over 30 min, 1 mL/min). The minor peak at ca. 22 min is attributed to loss of the Boc group resulting from the acidic HPLC conditions.
Figure S12. Optical absorption spectra of L-Ent and enterobactin-cargo conjugates 29-31 in the absence (grey) and presence (black) of Fe(III). (MeOH, rt). The concentration of ligand was ca. 40 µM and ca. one equiv of Fe(III) was added. (A) L-Ent; (B) conjugate 29, Boc cargo; (C) conjugate 30, cyclohexyl cargo; (D) conjugate 31, cyclohexyl cargo and D-Ent.
Figure S13. Optical absorption spectra of enterobactin-cargo conjugates 32-34 in the absence (grey) and presence (black) of Fe(III). (MeOH, rt). The concentration of ligand was ca. 40 µM and ca. one equiv of Fe(III) was added. (A) Conjugate 32, napthyl cargo; (B) conjugate 33, phenylmethylbenzyl cargo; (C) conjugate 34, coumarin 343 cargo. The absorption feature at ca. 440 nm is from the coumarin moiety.
Figure S14. Optical absorption spectra of enterobactin-cargo conjugates 35, 40, 42 and 43 in the absence (grey) and presence (black) of Fe(III). (MeOH, rt). The concentration of ligand was ca. 40 µM and ca. one equiv of Fe(III) was added. (A) Conjugate 35, ciprofloxacin cargo; (B) conjugate 40, ciprofloxacin cargo; (C) conjugate 42, vancomycin cargo. (D) conjugate 43, Boc-glycine-triazole cargo.
Figure S15. *E. coli* ATCC 33475 (*ent*) growth recovery assays employing enterobactin-cargo conjugates (50% MHB, ± 200 µM DP, t = 19 h, T = 30 °C). Grey bars: *E. coli* cultured in the absence of DP. Black bars: *E. coli* cultured in the presence of 200 µM DP. (A) L-Ent; (B) conjugate 29; (C) conjugate 30; (D) conjugate 31; (E) conjugate 32; (F) conjugate 33; (G) conjugate 34; (H) conjugate 35; (I) conjugate 40; (J) conjugate 42; (K) conjugate 43.
**Figure S16.** *E. coli* H1187 (*fepA*- ) and *E. coli* K-12 JW0576 (*fes*- ) growth recovery assays employing L-Ent and enterobactin-cargo conjugates 29 and 30 (50% MHB, 200 µM DP, t = 19 h, T = 30 °C). (A) *E. coli* H1187 (*fepA*- ). This strain grows to an OD$_{600}$ of ~0.2 in the absence of DP (data not shown). (B) *E. coli* JW0576 (*fes*- ). This strain grows to an OD$_{600}$ of ~0.2 in the absence of DP (data not shown). Grey bars: L-Ent; black bars, conjugate 29; white bars, conjugate 30.
Figure S17. *E. coli* ATCC 33475 (ent-) and *P. aeruginosa* PAO1 K648 (pvd-, pch-) growth assays with the D-isomer of enterobactin (50% MHB, ± 200 or 600 µM DP, t = 19 h, T = 30 °C). Grey bars: bacteria cultured in the absence of DP. Black bars: bacteria cultured in the presence of 200 (*E. coli*) or 600 (*P. aeruginosa*) µM DP. (A) *E. coli*. (B) *P. aeruginosa*. 
Figure S18. *P. aeruginosa* PAO1 K648 (pvd-, pch-) growth recovery assays employing enterobactin-cargo conjugates (50% MHB, ± 600 µM DP, t = 19 h, T = 30 °C). Grey bars: In the absence of DP. Black bars: In the presence of 600 µM DP. (A) L-Ent; (B) conjugate 29; (C) conjugate 30; (D) conjugate 31; (E) conjugate 32; (F) conjugate 33; (G) conjugate 34; (H) conjugate 35; (I) conjugate 40; (J) conjugate 42; (K) conjugate 43.
Figure S19. *P. aeruginosa* PAO1 K407 (*pvd*, *pFr*) growth recovery assays employing L-Ent and enterobactin-cargo conjugates 29 and 30 (50% MHB, ± 600 µM DP, *t* = 19 h, *T* = 30 °C). Grey bars: In the absence of DP. Black bars: In the presence of 600 µM DP. (A) L-Ent; (B) conjugate 29; (C) conjugate 30.
19F OBSERVE
STANDARD PARAMETERS

exp3 spul

date Jul 12 2012
solvent DCE
file /data/molan/Hr
PEG-NH2-SF1 ftd
ACQUISITION

frq 396.157
at 91
pp 1000
fz 0
fn 1
hs 0
bm 4
pw 11.0
ai 4.000
lof 29967.2
nt 10
ct 16
alock n

flags not used

11 n
in n
hp y

display

vp 9999.9
vs 56
sc 0
wz 256
z 256
ls 500.00
rf 4588.1
fp 0
th 10
ins 100.000

18 TFA

HO
O
F

18

NH2
### Standard Proton Parameters

**SAMPLE**: s2pul

**Acq**: acq 129.672

**Solvent**: D2O

**Fil**: exp dpwr 30

**Acq**: exp dpwr 30

**T1**: 4.01 mm

**T2**: 4.01 ms

**T3**: 1.0 s

**Gain**: not used homo n

**Flags**: not used 1.0

**Hs**: n homo2 DEC3

**Display**: dpwr3 0

**Sm**: dpwr3 0

**W**: dpwr3 0

**Vs**: dpwr3 0

**Sc**: dpwr3 0

**Wc**: dpwr3 0

**Hzw**: dpwr3 0

**F**: PROCESSING

**Rf**: 1024.5 wft 162144

**Tm**: 3.060 proc

**Wt**: wft

---

**1H NMR Spectrum**

- **Chemical Shifts**:
  - 0.77 ppm
  - 2.54 ppm
  - 2.22 ppm
  - 3.19 ppm
  - 4.0 ppm
  - 3.6 ppm
  - 11.48 ppm
  - 3.22 ppm

---

**Structural Formula**

- Compound 24

**Chemical Structure**

- Benzyl (Bn)
- Acetyl (OAc)
- Oxalyl (O=C=O)
- Amid (NH)
- Phenyl (C6H5)
- Benzyl (Bn)
- Oxalyl (O=C=O)

---
Bn_Int_aldehyde_1_H

exp2: 22:00
SAMPLE: DEC. & VT

date: Aug 1 2012
seq: 500.176
solvent: CDCl3
file: exp
dwpr: 32
acq: 500.176
dm: mnn
t: 2.048
tr: 8770
np: 32768
sw: 8000.0
drez: 1.0
bu: 4000
hs: 4
temp: 23.0
vs: 2
PROCCESSING
pu: 5.0
r: proc
of: 0
nt: 2
ct: 2

block: n
gain: not used
flg: wwr
li: n
hi: n
sp: y
hs: nn
sp: -250.1

pp: 4000.0
vs: 3.0
sc: 0
wc: 250
hmm: 44.01
ls: 88.64
rt: 5130.3
ref: 3651.3
sh: 
hrs: 3.000

ph

11 10 9 8 7 6 5 4 3 2 1 ppm

7.75 7.75 3.19 2.61 17.46 13.83 3.00 3.57 3.54

27
07182012_coumarin_PEG_BnEnt_I3C

Sample data

Solvent: CHCl3

File: /data/nolan/N=dpwr_34

Movie: /data/nolan/N=dpwr_34

Acquisition: 1H 10400

Freq: 125.722

Gain not used

Display:

Select: 1H

Display:

Select: 1H
<table>
<thead>
<tr>
<th>Sample</th>
<th>DEC. &amp; VT</th>
</tr>
</thead>
<tbody>
<tr>
<td>Date</td>
<td>Aug 5 2011</td>
</tr>
<tr>
<td>Solvent</td>
<td>DMSO</td>
</tr>
<tr>
<td>File</td>
<td>TFA-36</td>
</tr>
</tbody>
</table>

**Acquisition**
- SFreq: 300.1105
- PROCESSING: wfile
- Proc: 4.063
- FM: 3602.9
- FB: not used
- BS: 4
- TPW: 5.9
- DT: 8.99
- TOF: 887.7
- CT: 28
- GAIN: not used

**Display**
- SP: -156.1
- WP: 3081.6
- SC: 151
- WC: 290
- Hz mw: 12.66
- IS: 277.66
- RF1: 1825.3
- RFp: 553.4
- TH: 20
- INT: 2.0

**Diagram**
- Chemical structure of compound 36 with peaks at ppm values: 0.99, 1.00, 1.92, 4.92, 4.99, 5.01, 2.00, 2.03, 4.00, 4.09, 2.03.
STANDARD CARBON PARAMETERS

SAMPLE

DEC. & VT

solvent

CDCl3
dn
H2N

TFA•

S81

36

H2N

TFA•

S81

36
<table>
<thead>
<tr>
<th>SAMPLE</th>
<th>DEC. &amp; VT</th>
</tr>
</thead>
<tbody>
<tr>
<td>Date</td>
<td>Aug 1 2011</td>
</tr>
<tr>
<td>Solvent</td>
<td>CDCl3</td>
</tr>
<tr>
<td>Notes</td>
<td></td>
</tr>
</tbody>
</table>

**Acquisition Details**

- **Field Strength (MHz):** 300.168
- **Processing:**
  - **t1:** 131071.3
  - **t2:** 8.0
  - **t3:** 0.050
  - **t4:** 4.053
  - **t5:** 48002.9
  - **t6:** 6092.4
  - **t7:** 48002.9
  - **t8:**

**Display Settings**

- **Display:** y
- **Time:** 16
- **Gain:** n
- **Display Settings:**
  - **sp:** 350.1
  - **wp:** 4800.7
  - **V5:** 151
  - **ac:** 0
  - **wc:** 250
  - **hzone:** 18.21
  - **ls:** 181.82
  - **r1:** 2612.5
  - **r2:** 2178.8
  - **th:** 0.97
  - **ins:** 2.000

**Chemical Shifts**

- **Peak 1:** 0.97
- **Peak 2:** 1.03
- **Peak 3:** 1.02
- **Peak 4:** 2.68
- **Peak 5:** 15.71

**Diagram**

- **Chemical Structure**
- **Label:** BocHN
- **Molecular Formula:** 38

**Note:** The chemical structure and spectrum indicate the presence of a complex molecule with multiple resonance peaks.
<table>
<thead>
<tr>
<th>Sample</th>
<th>DEC. &amp; VT</th>
<th>Solvent</th>
<th>Compound</th>
<th>Structure</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>10.00</td>
<td>3.00</td>
<td>DMSO</td>
<td><img src="image" alt="Structure" /></td>
</tr>
<tr>
<td>2</td>
<td>5.80</td>
<td>1.50</td>
<td>DMSO</td>
<td><img src="image" alt="Structure" /></td>
</tr>
</tbody>
</table>

**Table:**

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>ppm</td>
<td>10</td>
</tr>
</tbody>
</table>

**Legend:**

- **HO2C**: Carboxylic acid group
- **O**: Oxygen atom
- **Bn**: Benzyl group
- **F**: Fluorine atom
- **N**: Nitrogen atom
- **39**: Identified compound number

**Notes:**

- **Sample Identification:**
  - **Sample 1:**
    - Date: Aug 1 2012
    - Temperature: 25.67°C
    - Solvent: DMSO
    - Compound: DMSO
    - Structure: ![Structure](image)
  - **Sample 2:**
    - Date: Aug 1 2012
    - Temperature: 25.67°C
    - Solvent: DMSO
    - Compound: DMSO
    - Structure: ![Structure](image)
FT-IR (KBr pellet or NaCl disk, cm⁻¹)

Boc-14
3443 (m), 3325 (s), 3074 (w), 2976 (s), 2930 (s), 2858 (s), 1698 (s), 1652 (s), 1535 (s),
1451 (s), 1391 (m), 1366 (s), 1351 (m), 1329 (w), 1273 (s), 1253 (s), 1217 (w), 1173 (s),
1126 (s), 1041 (w), 969 (w), 945 (w), 895 (w), 865 (w), 780 (w), 756 (m).

14
3307 (m), 3084 (m), 2932 (s), 2855 (m), 1680 (s), 1644 (s), 1542 (m), 1453 (w), 1433 (w),
1351 (w), 1321 (w), 1308 (w), 1273 (w), 1203 (s), 1178 (s), 1137 (s), 932 (w), 897 (w),
836 (w), 799 (w), 722 (w), 706 (w).

Boc-15
3438 (w), 3326 (m), 3050 (w), 2975 (m), 2933 (m), 2870 (m), 1708 (s), 1648 (s), 1592 (w),
1580 (w), 1530 (s), 1455 (w), 1391 (w), 1366 (m), 1351 (w), 1303 (m), 1254 (m), 1172 (s),
1124 (s), 1041 (w), 970 (w), 866 (w), 807 (w), 785 (m), 756 (m), 655 (w).

15
3270 (w), 3075 (m), 2918 (m), 2872 (m), 1680 (s), 1638 (s), 1592 (w), 1539 (m), 1480 (w),
1455 (w), 1429 (w), 1350 (w), 1307 (w), 1259 (w), 1203 (s), 1174 (s), 1135 (s), 945 (w),
880 (w), 834 (w), 798 (m), 785 (m), 721 (w), 706 (w).

Boc-16
3346 (s), 3092 (w), 3071 (w), 3027 (m), 2977 (s), 2929 (s), 2863 (s), 2708 (w), 2498 (w),
1956 (w), 1693 (s), 1649 (s), 1543 (s), 1505 (s), 1454 (s), 1392 (m), 1366 (s), 1301 (s), 1251
(s), 1251 (s), 1201 (s), 1165 (s), 1131 (s), 1030 (m), 943 (w), 863 (m), 831 (m), 801 (m),
753 (s), 720 (m), 700 (s).

16
3426 (m), 3027 (m), 2918 (m), 1948 (w), 1683 (m), 1632 (m), 1548 (m), 1505 (m), 1454
(m), 1420 (m), 1351 (m), 1309 (m), 1203 (m), 1134 (m), 1022 (w), 940 (w), 862 (w), 836
(w), 801 (m), 743 (m), 722 (m), 700 (m).

17
3412 (s), 2976 (m), 2919 (m), 2742 (m), 2673 (m), 2535 (m), 2497 (m), 1694 (s), 1616 (s),
1583 (s), 1521 (s), 1485 (s), 1445 (s), 1367 (s), 1398 (m), 1367 (s), 1310 (s), 1243 (m),
1212 (s), 1175 (s), 1098 (s), 1037 (m), 962 (w), 932 (w), 895 (w), 851 (w), 832 (w), 793
(m), 751 (m), 664 (w).

Fmoc-18
3309 (m), 3049 (w), 3010 (w), 2924 (m), 2868 (m), 1719 (s), 1627 (s), 1508 (m), 1465 (s),
<table>
<thead>
<tr>
<th>Page</th>
<th>Numbers</th>
</tr>
</thead>
<tbody>
<tr>
<td>18</td>
<td>1390 (w), 1337 (w), 1295 (w), 1260 (m), 1148 (w), 1112 (w), 1024 (w), 949 (w), 884 (w), 833 (w), 807 (w), 745 (m).</td>
</tr>
<tr>
<td>20</td>
<td>2918 (m), 2876 (m), 1718 (m), 1692 (s), 1628 (s), 1506 (m), 1470 (m), 1385 (w), 1340 (w), 1295 (w), 1251 (w), 1201 (m), 1178 (w), 1125 (m), 1025 (w), 947 (w), 886 (w), 831 (w), 799 (w), 776 (w), 748 (w), 720 (w).</td>
</tr>
<tr>
<td>21</td>
<td>3467 (s), 3177 (m), 3079 (m), 3005 (m), 2978 (m), 2955 (m), 2905 (m), 2854 (w), 1678 (s), 1640 (w), 1622 (m), 1485 (s), 1442 (s), 1411 (w), 1370 (m), 1319 (s), 1282 (s), 1235 (m), 1195 (s), 1168 (s), 1130 (m), 916 (m), 867 (w), 791 (s), 772 (w), 681 (m), 630 (w).</td>
</tr>
<tr>
<td>22</td>
<td>3062 (m), 3032 (m), 2976 (m), 2937 (m), 2912 (m), 2889 (m), 1692 (s), 1639 (w), 1603 (w), 1578 (w), 1482 (m), 1453 (w), 1414 (w), 1378 (w), 1331 (m), 1268 (m), 1219 (w), 1146 (w), 1049 (m), 1028 (w), 968 (w), 914 (w), 858 (w), 791 (w), 758 (w), 697 (m).</td>
</tr>
<tr>
<td>23</td>
<td>3084 (w), 3063 (m), 3031 (m), 2955 (m), 2941 (m), 2876 (m), 2846 (m), 1692 (s), 1599 (m), 1574 (m), 1498 (m), 1483 (m), 1454 (m), 1414 (m), 1377 (m), 1336 (m), 1272 (m), 1256 (m), 1218 (m), 1153 (m), 1081 (w), 1051 (s), 1028 (w), 958 (s), 940 (m), 914 (w), 887 (w), 859 (m), 843 (w), 782 (m), 757 (s), 744 (m), 727 (m), 695 (s), 650 (w).</td>
</tr>
<tr>
<td>24</td>
<td>3356 (m), 3065 (w), 3031 (w), 2954 (w), 2927 (w), 2876 (w), 1752 (s), 1660 (s), 1597 (w), 1576 (m), 1514 (s), 1455 (m), 1424 (w), 1376 (w), 1346 (m), 1312 (m), 1266 (s), 1205 (s), 1133 (w), 1082 (w), 1054 (w), 1029 (w), 962 (w), 914 (w), 852 (w), 806 (w), 754 (s), 698 (s), 668 (w).</td>
</tr>
<tr>
<td>25</td>
<td>3654 (w), 3628 (w), 3359 (m), 3088 (w), 3064 (w), 3032 (w), 2946 (w), 2933 (w), 2874 (w), 1761 (s), 1660 (m), 1599 (w), 1576 (m), 1519 (m), 1490 (m), 1454 (m), 1427 (w), 1374 (m), 1345 (m), 1267 (m), 1203 (m), 1134 (w), 1081 (w), 1048 (w), 1028 (w), 954 (w), 912 (w), 849 (w), 807 (w), 754 (s), 734 (s), 697 (s).</td>
</tr>
</tbody>
</table>
26
3356 (m), 3058 (w), 3032 (w), 2929 (w), 2881 (w), 1750 (s), 1660 (s), 1597 (w), 1576 (m), 1514 (s), 1455 (m), 1375 (w), 1346 (w), 1312 (w), 1266 (s), 1204 (s), 1133 (w), 1082 (w), 1053 (w), 1027 (w), 962 (w), 914 (w), 851 (w), 802 (w), 754 (s), 698 (m).

27
3358 (m), 3062 (w), 3031 (w), 2957 (w), 2928 (w), 2876 (w), 1751 (s), 1697 (m), 1660 (s), 1577 (m), 1517 (s), 1455 (m), 1375 (w), 1346 (w), 1303 (w), 1266 (s), 1206 (m), 1134 (w), 1082 (w), 1054 (w), 1022 (w), 957 (w), 914 (w), 851 (w), 806 (w), 754 (s), 698 (m).

28
3356 (m), 3088 (w), 3064 (w), 3032 (w), 2957 (w), 2924 (w), 2872 (w), 1753 (s), 1718 (m), 1662 (s), 1599 (w), 1577 (s), 1517 (s), 1454 (s), 1427 (w), 1375 (m), 1346 (m), 1267 (s), 1205 (s), 1133 (w), 1112 (w), 1082 (m), 1052 (m), 1029 (m), 957 (m), 914 (m), 851 (w), 808 (w), 755 (s), 736 (s), 699 (s).

Bn₆-29
3356 (m), 3064 (w), 3033 (w), 2931 (m), 2872 (m), 1752 (s), 1710 (m), 1661 (s), 1577 (m), 1516 (s), 1455 (s), 1367 (m), 1346 (m), 1266 (s), 1205 (s), 1130 (m), 1083 (m), 1047 (m), 958 (w), 915 (w), 851 (w), 808 (w), 755 (m), 699 (m).

Bn₆-30
3353 (m), 3065 (w), 3032 (w), 3010 (w), 2930 (m), 2855 (m), 1752 (s), 1660 (s), 1593 (w), 1577 (m), 1518 (s), 1454 (m), 1424 (w), 1374 (w), 1346 (m), 1301 (m), 1265 (s), 1206 (s), 1130 (m), 1083 (m), 1050 (m), 1022 (w), 957 (w), 915 (w), 850 (w), 809 (w), 755 (s), 699 (m), 659 (w).

Bn₆-31
3354 (m), 3062 (w), 3032 (w), 3006 (w), 2929 (m), 2855 (m), 1752 (s), 1660 (s), 1577 (m), 1518 (s), 1454 (m), 1374 (w), 1346 (w), 1303 (w), 1265 (m), 1206 (m), 1130 (w), 1083 (w), 1061 (w), 1031 (w), 957 (w), 915 (w), 850 (w), 811 (w), 755 (s), 698 (m).

Bn₆-32
3353 (m), 3067 (w), 3028 (w), 2997 (w), 2924 (m), 2859 (m), 1751 (s), 1659 (s), 1577 (m), 1519 (s), 1455 (m), 1372 (w), 1346 (w), 1295 (m), 1205 (m), 1122 (w), 1083 (w), 1053 (w), 1027 (w), 958 (w), 914 (w), 845 (w), 806 (w), 785 (w), 754 (m), 698 (m).

Bn₆-33
3356 (m), 3071 (w), 3031 (w), 2924 (m), 2868 (m), 1751 (s), 1660 (s), 1576 (m), 1519 (s), 1454 (m), 1374 (w), 1346 (m), 1301 (m), 1265 (s), 1205 (m), 1130 (w), 1083 (w), 1057 (w), 1022 (w), 957 (w), 915 (w), 851 (w), 806 (w), 754 (s), 698 (m).
**Bn₆-34**
3350 (m), 3088 (w), 3065 (w), 3033 (w), 2931 (m), 2868 (m), 1752 (s), 1660 (s), 1616 (m), 1577 (s), 1518 (s), 1455 (s), 1369 (m), 1348 (m), 1309 (s), 1266 (s), 1211 (s), 1152 (w), 1129 (m), 1083 (m), 1053 (w), 1027 (w), 958 (w), 912 (m), 844 (s), 794 (w), 734 (s), 699 (m).

**Bn₆-35**
3369 (m), 3075 (w), 3036 (m), 3015 (m), 2950 (m), 2907 (m), 2855 (m), 1750 (s), 1662 (s), 1627 (s), 1577 (m), 1512 (s), 1464 (s), 1375 (m), 1339 (m), 1302 (m), 1263 (s), 1208 (s), 1121 (m), 1082 (m), 1053 (m), 1027 (m), 962 (m), 910 (w), 884 (w), 867 (w), 849 (w), 806 (w), 754 (s), 699 (m), 664 (w).

36
2954 (m), 2922 (m), 2859 (m), 1718 (m), 1682 (s), 1616 (s), 1469 (s), 1385 (w), 1333 (w), 1255 (w), 1199 (m), 1165 (m), 1139 (m), 1022 (w), 828 (w), 799 (w), 720 (w).

38
3345 (m), 3019 (m), 2967 (m), 2931 (m), 2863 (m), 1707 (s), 1628 (s), 1509 (s), 1467 (s), 1389 (m), 1365 (w), 1338 (m), 1301 (w), 1260 (s), 1170 (m), 1109 (w), 1026 (m), 988 (w), 940 (w), 888 (w), 834 (w), 807 (w), 751 (m), 711 (w), 665 (w).

39
3354 (m), 3065 (w), 3028 (m), 3009 (m), 2933 (m), 2859 (w), 1751 (s), 1658 (s), 1619 (s), 1577 (m), 1499 (s), 1455 (s), 1374 (m), 1346 (s), 1308 (s), 1262 (s), 1208 (s), 1128 (w), 1083 (m), 1028 (m), 979 (w), 957 (w), 914 (w), 851 (w), 806 (w), 754 (s), 699 (s), 665 (m).

41
3409 (s), 3002 (w), 2963 (m), 2918 (m), 2855 (m), 1748 (m), 1660 (s), 1576 (w), 1541 (w), 1437 (m), 1403 (m), 1342 (w), 1313 (m), 1260 (m), 1204 (w), 1025 (s), 953 (m), 845 (w), 798 (w), 703 (w).