Movement of Replicating DNA through a Stationary Replisome

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Movement of Replicating DNA through a Stationary Replisome

Katherine P. Lemon and Alan D. Grossman*
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Summary

We found that DNA is replicated at a central stationary polymerase, and each replicated region moves away from the replisome. In Bacillus subtilis, DNA polymerase is predominantly located at or near midcell. When replication was blocked in a specific chromosomal region, that region was centrally located with DNA polymerase. Upon release of the block, each copy of the duplicated region was located toward opposite cell poles, away from the central replisome. In a roughly synchronous population of cells, a region of chromosome between origin and terminus moved to the replisome prior to duplication. Thus, the polymerase at the replication forks is stationary, and the template is pulled in and released outward during duplication. We propose that B. subtilis, and probably many bacteria, harness energy released during nucleotide condensation by a stationary replisome to facilitate chromosome partitioning.

Introduction

The production of viable progeny depends on the faithful replication and partitioning of genomic DNA. Bacteria do not appear to have a eukaryotic-like mitotic apparatus. Like many bacteria, B. subtilis has a single circular chromosome (4200 kb; Kunst et al., 1997) with a single origin (oriC) from which bidirectional DNA replication initiates (Kornberg and Baker, 1992). Separation of newly duplicated chromosome regions occurs while DNA replication of the rest of the chromosome proceeds (Webb et al., 1997; Teleman et al., 1998; Webb et al., 1998).

Recent work with B. subtilis, E. coli, and C. crescentus demonstrates that the chromosome is oriented such that for most of a cell cycle the two sister origins reside on opposite sides of the cell, while the unduplicated terminus region is centrally located (Glaser et al., 1997; Gordon et al., 1997; Lewis and Errington, 1997; Lin et al., 1997; Mohl and Gober, 1997; Webb et al., 1997; Niki and Hiraga, 1998; Sharpe and Errington, 1998; Teleman et al., 1998; Webb et al., 1998; Jensen and Shapiro, 1999; Niki et al., 2000). Regions located between the origin and terminus on the circular chromosome appear to be positioned between them in the cell (Teleman et al., 1998; Niki et al., 2000). Efforts to associate chromosome movement with the act of DNA replication are confounded by two factors. First, the rate of bacterial DNA polymerization is estimated at 500–1000 nucleotides per second (Baker and Bell, 1998). This is equivalent to moving of a single base pair a distance of ~170–340 nanometers each second. Second, DNA inside cells is highly compacted (reviewed in Trun and Marko, 1998). The ~4200 kilobase pair (kbp) circular genome of B. subtilis (Kunst et al., 1997) is neatly organized in a rod-shaped cell two to four microns in length and about half a micron in diameter.

Here, we describe experiments that demonstrate that DNA is duplicated at a centrally located, stationary replisome. Previously, we visualized the B. subtilis replicative DNA polymerase using a functional fusion of the catalytic subunit, PolC, to green fluorescent protein (GFP) (Lemon and Grossman, 1998). PolC–GFP localizes to the duplication arrest in regions ~100–130 kbp to the left (LSTer or left stringent terminus region) and ~150–200 kbp to the right (RSTer) of oriC (Lemon and Grossman, 1998; Autret et al., 1999). DNA polymerase resumes chromosome synthesis in the STer regions when amino acids are added back to the cells. Starvation for an amino acid activates the production of the alarmone (p)ppGpp by the relA gene product, which induces the pleiotropic stringent response in many bacteria (Cashel et al., 1996; Wendrich and Marahiel, 1997), though stringent arrest of DNA replication at sites away from the origin has only been characterized in B. subtilis.

We visualized the more precisely mapped LSTer region (Autret et al., 1999). During stringent arrest, the LSTer region was located at or near midcell. Under these conditions, the newly duplicated sister origins are already separated to opposite sides of the cell (Sharpe and Errington, 1998). After release from replication arrest, the LSTer region was duplicated, and the two copies were positioned away from midcell in opposite halves of the cell.

We also visualized a region of the chromosome (270°) approximately midway between origin and terminus. Just prior to a burst of duplication of the 270° region in a roughly synchronous population of cells, we observed an increase in the occurrence of coincidence of the 270° region and DNA polymerase. Taken together, our results indicate that the stationary, centrally positioned DNA polymerase does indeed represent the location of active replication forks and suggest that the DNA is threaded through the polymerase during duplication and then extruded from the polymerase after duplication.

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Table 1. Number of Foci of the LSTer Region in Cells before, during, and after Stringent Arrest

<table>
<thead>
<tr>
<th>Conditions* (Number of Cells)</th>
<th>1</th>
<th>2</th>
</tr>
</thead>
<tbody>
<tr>
<td>Asynchronous growth (728)</td>
<td>42</td>
<td>56</td>
</tr>
<tr>
<td>Stringent arrest (572)</td>
<td>77</td>
<td>22</td>
</tr>
<tr>
<td>Release from arrest (504)</td>
<td>28</td>
<td>64</td>
</tr>
</tbody>
</table>

*Strain KPL549 [dnaB134(ts), hutM−lacO, lacI-gfp] was grown in defined minimal medium at 30°C, and the number of foci of the LSTer region per cell was determined. Samples were taken (1) during asynchronous exponential growth, (2) 30 min after induction of stringent arrest in a synchronized population, and (3) 30 or 35 min after release from stringent arrest.

Table 2. Number of Foci of the LSTer Region in Cells during Growth after Synchronization

<table>
<thead>
<tr>
<th>Timea</th>
<th>Number of Cells</th>
<th>Percentage of Cells with Indicated Number of Focib</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>1</td>
<td>2</td>
</tr>
<tr>
<td>0</td>
<td>406</td>
<td>62.3</td>
</tr>
<tr>
<td>15</td>
<td>442</td>
<td>64.3</td>
</tr>
<tr>
<td>30</td>
<td>348</td>
<td>52</td>
</tr>
<tr>
<td>45</td>
<td>449</td>
<td>39.7</td>
</tr>
<tr>
<td>60</td>
<td>394</td>
<td>31.2</td>
</tr>
<tr>
<td>75</td>
<td>566</td>
<td>27.2</td>
</tr>
</tbody>
</table>

*The replication cycle was synchronized by shifting cells to nonpermissive temperature (45°C) to block initiation of replication and to allow most ongoing rounds of replication to finish. Samples of strain KPL549 [dnaB134(ts), hutM−lacO, lacI-gfp] were taken at the indicated times (min) after shift back to permissive temperature, and the number of foci of the LSTer region per cell was determined.

Results

Visualization of the LSTer Region

We chose the more precisely defined LSTer region over the RSTer region for analysis (Autret et al., 1999). The LSTer region contains two approximately equivalent arrest sites, LSTer1 and LSTer2, separated by about 27 kbp (Autret et al., 1999). Stalling of DNA polymerase in both the left and right STer regions is about 80% effective in a synchronized population of cells treated with arginine hydroxymate (Levine et al., 1991, 1995; Autret et al., 1999). We marked the LSTer region by inserting a tandem array of E. coli lac operator sites adjacent to hutM on the B. subtilis chromosome, which is ~40 kbp from LSTer2 and ~67 kbp from LSTer1. We chose this position for convenience and to minimize the possibility that the array would disrupt function of the LSTer region. The hutM-lacO array was visualized with a carboxy-terminal fusion of Lac repressor to a variant of GFP (LacI-GFP) (Cormack et al., 1996).

During exponential growth, most cells have a partly duplicated chromosome. The LSTer region is within 130 kbp of the origin and is duplicated relatively early (Autret et al., 1999). Indeed, during asynchronous exponential growth, most cells had two well-separate foci of the LSTer region (Table 1). The pattern of localization of the sister LSTer regions appeared to be similar to that of the origin region (Webb et al., 1997; Telemam et al., 1998; Webb et al., 1998) and the chromosome partitioning protein Spo0J (Glaser et al., 1997; Lewis and Errington, 1997; Lin et al., 1997; Sharpe and Errington, 1998; Telemam et al., 1998), which binds to sites (parS) in the origin-proximal 20% of the chromosome (Lin and Grossman, 1998).

Synchronization of DNA Replication in a Population of B. subtilis

In order for most of the cells to undergo replication arrest at a similar time, the population was first synchronized for DNA replication (Levine et al., 1991, 1995; Autret et al., 1999). A mutant that is temperature sensitive for initiation of DNA replication, dnaB134 (Mendelson and Gross, 1987; Karamata and Gross, 1970), was shifted to nonpermissive temperature to allow most ongoing rounds of replication to finish while preventing initiation of new rounds of replication. In B. subtilis, the replication initiation protein DnaB (not a helicase as is E. coli DnaB) is membrane associated and is required for origin enrichment in membrane fractions (Winston and Sueoka, 1980). Immediately after shiftdown to permissive temperature, ~62% of the population was synchronized as judged by having a single focus of the LSTer chromosome (Table 2), indicating a single chromosome per cell. This method gives at least one roughly synchronous round of replication in a population (Dworkin et al., 1972).

The LSTer Region Is Centrally Located during Stringent Response–Induced Replication Arrest

Stringent response was induced in a dnaB134(ts)-synchronized population of cells by addition of the amino acid analog arginine hydroxymate (250–500 μg/ml) (Levine et al., 1991, 1995; Autret et al., 1999). Thirty minutes after induction of stringent DNA replication arrest, 77% of cells had a single, unduplicated focus of the LSTer region (Table 1). In contrast, in the absence of stringent arrest, the percent of cells (52%) (Table 2) with a single focus 30 min after synchronization was similar to that in a culture of asynchronously growing cells (45%) (Table 2). During stringent arrest, the single focus of LSTer was located at or near midcell (Figure 1A).

DNA Polymerase Is Centrally Located during Stringent Response–Induced Replication Arrest

To be sure that the manipulations to synchronize and induce stringent arrest did not affect DNA polymerase position, we observed DNA polymerase in live cells using a functional fusion of the Tau subunit to GFP. A dimer of Tau is the scaffolding that holds the leadsing and lagging strand polymerases together (Kelman and O’Donnell, 1995; Baker and Bell, 1998). The Tau-GFP fusion produced a slightly stronger signal than the PolC-GFP fusion, and previous work indicated that during exponential growth, the position of Tau-GFP is similar.
Movement of DNA through a Stationary Replication Complex

**Figure 1. Position of the LSTer Region and DNA Polymerase during Replication Arrest Induced by the Stringent Response**

During stringent arrest, most cells have a single centrally located focus of the LSTer region. After release, most cells have two well-separated foci of the LSTer region. (A) shows the position of the LSTer region (hutM-lacO) in cells with a single focus during stringent arrest (341 cells). In (B), the position of DNA polymerase in cells with a single focus of Tau-GFP during stringent arrest (402 cells) is shown. (C) shows the position of the LSTer region (hutM-lacO) in cells with two foci after release from stringent arrest by nutrient addback (114 cells). Position was determined by measuring the distance from midfocus to the cell pole. In cells with two foci, distance was measured from the same cell pole. (A) and (C), strain KPL549 [dnaB134(ts), hutM1-lacO, lacI-gfp]; (B), strain KPL502 [dnaB134(ts), dnaX-gfp].

**After Release from Replication Arrest, Most Cells Had Two Well-Separated Foci of the LSTer Region**

DNA replication arrest was released by washing cells (to remove arginine hydroximate) and resuspending them in a nutrient-rich medium. Within 30–35 min after release, most cells (64%) had two foci of the LSTer region (Table 1). Thus, a majority (67%) of the cells with a single focus during arrest appeared to replicate the LSTer region after release. Duplication of the LSTer region correlated with a resumption in growth as measured by an increase in the culture's optical density. The two newly duplicated copies of the LSTer regions were well separated from each other and located at ~35% and ~65% of the cell length (Figure 1C). In addition to the shift from one to two foci, after release there was an increase in the proportion of cells with three and four foci (from 1% to 8%).

**Visualization of DNA Polymerase and LSTer in the Same Cells**

The experiments described above were done in live cells from two different strains; one contained the LacI-GFP fusion and the lac operator array near the LSTer region, and the other contained a functional Tau-GFP fusion. When DNA replication was stalled at the LSTer region, the majority of cells had a single focus of both the LSTer region and DNA polymerase at or near midcell (Table 1; Figures 1A and 1B). To directly show that the LSTer region and the polymerase colocalize during replication arrest, we visualized the LSTer region and the replisome in the same cells with and without the stringent arrest.

The development of variants of GFP with sufficiently separable excitation and emission spectra permits the observation of two separate fusion proteins in the same cell (Tsien, 1989; Miller et al., 1999). DNA polymerase was visualized using a functional fusion of Tau to a yellow variant of GFP (YFP) (Ormo et al., 1996). The lacO array near the LSTer region (hutM-lacO) was visualized using a fusion of LacI to a cyan variant of GFP (CFP) (Heim and Tsien, 1996).

During asynchronous growth, the majority of cells (83%) had two foci of the LSTer region (Table 3, line A). Immediately after synchronization of the replication cycle, ~43% of the cells had two foci of the LSTer region and ~55% had a single focus (Table 3, line B). One hour later after growth at permissive temperature, most of the cells (83%) had two foci of the LSTer region (Table 3, line C). In marked contrast, when stringent arrest was induced, only ~47% of the cells had two foci and 49% had a single focus of the LSTer region (Table 3, line D). When stringent arrest was released, cells began to grow, and by thirty minutes a majority had duplicated and separated the LSTer region (Table 3, line E). The increase in cells with three and four foci after release (Table 3) indicates the occurrence of multiform fork replication under these growth conditions (see below).

A clear pattern emerged when we observed cells that had a centrally located polymerase (either a single focus or two closely juxtaposed foci [Lemon and Grossman, 1998]) and one or two foci of LSTer. During exponential growth in an asynchronous culture, 86% of the cells had two well-separated foci of the LSTer region, one on either side of the central DNA polymerase (Figure 2). This same pattern was observed in the majority of cells (79%) that were growing without arrest 60 min after synchronization (Figures 2, 3A–3C, and 4). In contrast, when DNA replication arrest was induced (stringent response), there was a marked increase in the proportion of cells in which a single focus of LacI-CFP bound near the LSTer was either coincident with (31%) or touching (18.5%) a focus of DNA polymerase (Tau-YFP) (Figures 2, 3D–3F, and 4). Likewise, there were many fewer cells, only ~36%, with the typical exponential growth pattern (Figures 2 and 4). The lacO array is inserted ~67 kbp from the first arrest site (LSTer1) and ~40 kbp from the second site (LSTer2). LSTer1 and 2 are approximately equivalent (Aurzet et al., 1999), so in about half the cells the DNA replication fork is blocked ~67 kbp away from the lacO array that we visualized. It is likely that in cells in which the LacI-CFP focus touches the Tau-YFP focus, LSTer is coincident with the replication fork. Therefore, we have combined these two classes in Figures 2 and 4.

When cells were released from stringent arrest, within 30 min the pattern shifted back toward that seen for cells growing without arrest. Approximately 53% of cells had two well-separated LSTer foci flanking a central...
Table 3. Number of Foci of the LSTer Region in Cells Containing Tau-YFP

<table>
<thead>
<tr>
<th>Conditions* (Number of Cells Counted)</th>
<th>Percentage of Cells with Indicated Number of Foci</th>
</tr>
</thead>
<tbody>
<tr>
<td>(A) Asynchronous (623)</td>
<td>5.6 83.3 4.7 6.4</td>
</tr>
<tr>
<td>(B) Synchronous, 0 min (706)</td>
<td>54.8 42.5 2 0.7</td>
</tr>
<tr>
<td>(C) Synchronous, no arrest (600)</td>
<td>12.7 83 2.7 1.7</td>
</tr>
<tr>
<td>(D) Synchronous, stringent arrest (457)</td>
<td>49 47.3 2.8 0.9</td>
</tr>
<tr>
<td>(E) Synchronous, release from arrest (696)</td>
<td>20 66.4 9 4.6</td>
</tr>
</tbody>
</table>

*Samples of strain KPL730 [dnaB134(ts), hutM-1acO, lacI-cfp, dnaX-yfp] were taken under the indicated conditions, and the number of foci of the LSTer region per cell was determined. (A), asynchronous growth; (B), synchronous cells immediately after shift to permissive temperature after 45 min at 42°C (to prevent initiation of new rounds of replication and to allow most ongoing rounds to finish); (C), synchronous cells 60 min after shift to permissive temperature with no stringent arrest; (D), synchronous cells 60 min after shift to permissive temperature and during stringent arrest; and (E), synchronous cells 30 min after addition of nutrients to release stringent arrest. Nutrients were added 60 min after stringent arrest and shift to permissive temperature.

DNA polymerase (Figures 2, 3G–3I, and 4). The pattern of localization of DNA polymerase and the LSTer region described here strongly supports the model that the DNA template is duplicated by a stationary, centrally located DNA polymerase. After duplication, the two sister chromosome regions then move in opposite directions apart from each other and away from the replisome.

**Multifork Replication**

*B. subtilis*, like many other bacteria, is capable of doubling, under optimal growth conditions, in about a third of the time it takes to duplicate the genome. This is accomplished by reinitiating DNA replication from the sister origins before cell division so that there are multiple bidirectional replication forks per cell, essentially giving the replication cycle a headstart on the division cycle. When cells were released from stringent arrest by resuspension in a nutrient-rich medium, multifork replication began to occur in about 20% of cells, as indicated by cells with more than two LSTer foci (Table 3). After release, we observed cells that had a focus of LSTer out toward each pole, both of which were either adjacent to or coincident with a focus of DNA polymerase (Figures 3J–3O). In these cells, DNA replication has probably reinitiated or is about to initiate at the two sister origins prior to cell division. We speculate that initiation occurs at the cell quarters, which are nascent...
midcell positions. Some of these cells also had a centrally located DNA polymerase that probably represents an initial round of replication that was not yet complete (Figures 3J–3L).

Visualization of DNA Polymerase and the 270° Region of the Chromosome in Synchronized Cells

It appears that in B. subtilis, the origin region is located at approximately midcell or a position that will become midcell, prior to duplication (Lewis and Errington, 1997; Telemam et al., 1998) (D. C.-H. Lin, P. S. Lee, and A. D. G., unpublished data). The LSTer region is close to midcell, prior to duplication (Lewis and Errington, 1997; Anderson et al., 1998). The 270° region is halfway between oriC and the terminus. Sister origin regions are localized at approximately the cell quarter positions (D. C.-H. Lin, P. S. Lee, and A. D. G., unpublished data), while the terminus is at approximately midcell (Webb et al., 1997; Telemam et al., 1998; Webb et al., 1998). Thus, the 270° region is usually positioned between the quarter and midcell (Telemam et al., 1998). We visualized the 270° region using a tandem array of lac operators adjacent to ctsS (Telemam et al., 1998) combined with our LacI-CFP fusion. DNA polymerase was again visualized with Tau-YFP.

Immediately after synchronization (t = 0), most of the cells (82%) had a single focus of the 270° region (Figure 5A). Between 60 and 75 min after synchronization, there was a marked increase in the number of cells with two foci of the 270° region (Figure 5A), indicating that in many cells the 270° region was duplicated during this interval. We observed three general patterns when we compared the position of the 270° region to that of DNA polymerase over the time course in cells with a single focus of 270°. (1) At all time points, 30%–35% of the cells had the 270° region adjacent to a focus of DNA polymerase. (2) The percentage of cells with the 270° region coincident with (squares) or separate from (circles) a focus of DNA polymerase (Tau-YFP) was taken at the indicated times after shiftdown to permissive temperature, and the foci of DNA polymerase (Tau-YFP) and the 270° region (LacI-CFP) were analyzed. (3) At all time points, 30%–35% of the cells had the 270° region adjacent to a focus of DNA polymerase. This is not surprising, given that 270° is usually positioned between the cell quarter and midcell (Telemam et al., 1998), while the replisome is at or near midcell (Lemon and Grossman, 1998). (2) We also observed cells in which the 270° region was clearly separated from DNA polymerase and (3) cells in which the 270° region was coincident with a focus of DNA polymerase. The percent of cells in which the 270° region was coincident with DNA polymerase increased between 45 and 60 min after synchronization (Figure 5B). This increase occurred just prior to the shift from permissively to 270° per cell (Figure 5A). The timing of this position change is consistent with movement of the 270° region to the stationary DNA polymerase just prior to duplication.

Discussion

A Centrally Located, Stationary Bacterial Replisome

Our results indicate that during replication in B. subtilis, DNA moves through a stationary replisome located at or near midcell. After a region of DNA is replicated, the two duplicated regions move away from the replisome
E. coli

of SeqA (Hiraga et al., 1998) also indicates that newly

B. subtilis

most cells (Koppes et al., 1999). Previously, we specu-

oriC

croscopy indicates that the bulk of SeqA does not colo-

GFP system, the origin is centrally located in cells with

mass of the chromosome dwarfs that of the replisome. A mbb/ruva.html). When a model of head-to-head heli-

Goldman, 1998; Cook, 1999; Mehta et al., 1999; Vale

helicase for each fork is loaded onto the origin bubble

them at midcell, they are captured and maintained at ap-

approximately cell quarter positions until DNA replication

reinitiates.

DNA and RNA Polymerases Are Motor Proteins

For years, people have thought of DNA polymerase as

propelling itself along the DNA template during replica-

tion. A stationary DNA polymerase would instead spool

the template through itself during replication. Measure-

ments of the force generated by a single, stationary

RNA polymerase during transcription (25 picoNewtons)

(Gelles and Landick, 1998; Wang et al., 1998) indicate

that RNA polymerase is a more powerful motor than

either myosin or kinesin (for reviews see Block, 1998;

Goldman, 1998; Cook, 1999; Mehta et al., 1999; Vale

and Milligan, 2000). A replisome contains two types of

nucleoside triphosphate-dependent molecular motors,

the DNA polymerase and helicase. Given that the rate

of DNA synthesis is approximately ten times faster than

that of RNA synthesis, it seems likely that the replisome

is at least as powerful a motor as RNA polymerase.

Although the replisome has a mass of ~1,000 kDa, the

mass of the chromosome dwarfs that of the replisome. A

stationary replisome would bind and release and move

the chromosome in single base pair increments of ~660

Da and ~0.34 nm (Kornberg and Baker, 1992; Cook,

1999). During chromosome duplication, the same amount

of energy is used whether the polymerase is stationary

or moving. Given the limits of our understanding of the

viscosity and crowding of the intracellular environment,

it has been difficult to estimate if spooling the chromo-

somal through the replisome would be more energeti-

cally cost effective than moving the replisome along the

polymerase. We speculate that an added benefit of the

stationary polymerase, combined with rapid recompa-

tion of the DNA after duplication, is the ability to rapidly

separate newly duplicated regions, thus limiting the pos-

sibility of sister chromosomes tangling in the crowded

nucleoid mass.

E. coli Probably Has a Stationary Replisome

There is increasing evidence that the gram-negative bacterium E. coli also has a centrally located, stationary replisome. In E. coli, newly replicated DNA detected by pulse-labeling with [3H]thymidine, is centrally located in most cells (Koppes et al., 1999). Previously, we specu-
lated (Lemon and Grossman, 1998) that the location of SeqA (Hiraga et al., 1998) also indicates that newly

duplicated DNA is found at or near midcell in E. coli.

SeqA binds both the origin and newly replicated, hemi-
methylated DNA (Slater et al., 1995). Fluorescence mi-

roscopy indicates that the bulk of SeqA does not colo-

calize with oriC. Rather, SeqA localizes as discrete foci

predominantly at midcell or the cell quarters (Hiraga et

al., 1998; Onogi et al., 1999) in a pattern similar to that

of the B. subtilis replisome (Lemon and Grossman,

1998). The SeqA foci probably represent the majority of

SeqA bound to newly duplicated, hemimethylated DNA

adjacent to the replication forks. We suspect that the

extraction-capture model for how DNA replication might

power, at least in part, chromosome partitioning will

apply to other bacteria.

Is the Replisome Anchored?

While there is no evidence to suggest that the replisome is

membrane bound, it seems reasonable to expect it

might be anchored in order to prevent movement during

DNA replication. In B. subtilis, one of the proteins (DnaB)

needed to initiate replication is a membrane protein and

might play a role in helping to anchor the replisome.

The bidirectional replication forks might also be main-

tained in place by the viscosity inside the cell. Recent

findings indicate that large protein complexes may not

diffuse freely throughout the cell (Elowitz et al., 1999).

Head-to-Head Replication Forks

In vitro data from E. coli replication indicates that the

helicase for each fork is loaded onto the origin bubble in

a such a way that they either must pass each other

or, if stationary, remain head-to-head as each spools

and unwinds DNA (Fang et al., 1999). Head-to-head sta-

tionary helicases spooling DNA might be envisioned in

a manner similar to models for how RuvA and RuvB

accompilish Holliday junction branch migration (Rafferty

et al., 1996) (see also http://www.sdsc.edu/journals/

mbb/ruva.html). When a model of head-to-head heli-

cases is drawn (Fang et al., 1999), it suggests that the

leading and lagging strand DNA polymerase at each fork

might use the same template strand (Figure 6). In other

words, we suggest that each replication fork duplicates

one entire template strand (one fork for Watson and one

for Crick) instead of each fork duplicating both strands

from half of the chromosome. Head-to-head bidirec-

tional forks might help to facilitate chromosome separa-

tion and also help to limit movement of the replisome.

The Replisome Is Probably Positioned

during Initiation of Replication

The simplest model is that DNA replication initiates cen-

trally or at sites that will become central after cell divi-

sion, and this establishes the position of the replisome.

This model is supported by the pattern of origin region

localization in both B. subtilis and E. coli which indicates

that replication initiates at or near midcell or at positions

that will become midcell. In E. coli, the origin region as

detected by fluorescence in situ hybridization (FISH)

appears to be centrally located in cells just prior to onset

of DNA replication (Roos et al., 1999; Niki et al., 2000).

B. subtilis origin-region DNA can be labeled with BrdU

when spores of a thymine auxotroph are germinated.

Initially, the BrdU-labeled origin regions form a single

focus near midcell (Lewis and Errington, 1997). Later,

cells have two well-separated origin regions. When the

B. subtilis origin region is labeled with the lacO-Lacl-

GFP system, the origin is centrally located in cells with

a single focus (D. C.-H. Lin and A. D. G., unpublished

data). Spo0J, which is an excellent marker of origin re-

region position in B. subtilis (Lewis and Errington, 1997;
Smc may be loaded onto newly replicated DNA to establish these constrained (+) supercoils and compact the DNA. In addition to Smc, Spo0J, a member of the ParB protein family, is required for efficient partitioning (Treton et al., 1994), probably by functioning to organize the origin region into a higher order structure (Glaser et al., 1997; Lin et al., 1997; Britton et al., 1998; Lin and Grossman, 1998; Britton and Grossman, 1999).

The Common Theme of Stationary Replisomes
A diverse array of organisms including prokaryotes, archaea, and eukaryotes may have stationary replisomes. Many of the components required for DNA replication are functionally conserved and homologous in all organisms (Baker and Bell, 1998). The term “replication factory” was first applied to the multiple foci of replication proteins observed in the eukaryotic nucleus using immunofluorescence microscopy (for review see Newport and Yan, 1996; Cook, 1999). The prevailing view is that these factories represent a clustering of many replicons that probably initiate replication synchronously during S phase (Newport and Yan, 1996). Recently, a fusion of GFP to the eukaryotic sliding clamp, GFP-PCNA, has been observed using time-lapse microscopy in a stable tissue culture cell line (Leonhardt et al., 2000). Replication foci (GFP-PCNA) assembled and disassembled during S phase but did not show directional motion (Leonhardt et al., 2000). This observation is consistent with earlier models suggesting that these eukaryotic “replication factories” might represent a conglomerate of stationary replisomes through which the DNA template moves (Hozak et al., 1993).

Most eukaryotes use a mitotic spindle and mitotic motors for the separation of sister chromosomes. These components involved in chromosome partitioning have not been found in prokaryotes. Perhaps the differences in the distances needed to move chromosomes during partitioning partly explain the dramatically different mechanisms used by eukaryotes and prokaryotes.

Experimental Procedures

Growth Media and Antibiotics
Rich medium was Luria-Bertani broth (LB), and defined minimal medium was S7 medium with 1% glucose and 0.1% glutamate (Vasantha and Freese, 1980; Jaacks et al., 1989) and required amino acids (phenylalanine [40 μg/ml], tryptophan [40 μg/ml], and threonine [240 μg/ml]). Where needed, antibiotics were used at the indicated concentrations: chloramphenicol (cat), 5 μg/ml; erythromycin, 0.5 μg/ml and lincomycin, 12.5 μg/ml together (mls); and spectinomycin (spc), 100 μg/ml.

Strains and Alleles
All strains are derivatives of JH642, which is topC2, pheA1 (Peregonot alone. It is likely that proteins involved in chromosome organization, compaction, and supercoiling contribute to the orderly movement of sister DNA regions away from the central replisome. We speculate that Smc, which is clearly important for chromosome partitioning (Britton et al., 1998; Graumann et al., 1998; Moriya et al., 1998; Britton and Grossman, 1999; Hirano, 1999), may play such a role. It has recently been proposed that Smc and the E. coli analog MukB generate higher order compaction of bacterial DNA by introducing constrained positive supercoils (Holmes and Cozzarelli, 2000; Sawitzke and Austin, 2000).

Figure 6. A Model of Two Head-to-Head, Opposing, Stationary Hexameric Helicases Spooling the DNA Template through as They Unwind the Double Helix
Adapted from Fang et al. (1999).

Lin et al., 1997; Teleman et al., 1998), binds to eight identified parS sites located in the origin-proximal 20% of the chromosome (Lin and Grossman, 1998). During exponential growth, most cells have two well-separated foci of Spo0J. In cells with a single focus, Spo0J, and hence the origin region, is predominantly at midcell (Teleman et al., 1998; P. S. Lee and A. D. G., unpublished data). Most cells in which the initiation of DNA replication is inhibited and old rounds have finished have a single focus of Spo0J-GFP located at approximately midcell (our unpublished data). Finally, our results with replication upon release from stringent arrest (Figures 3J–3O) indicate that during multifork replication initiation occurs at the cell quarters, positions that will become midcell upon division.

Other Proteins Are Required for Efficient Chromosome Partitioning
Although we have proposed that the replisome plays an important role in chromosome partitioning, it is certainly not alone. It is likely that proteins involved in chromosome organization, compaction, and supercoiling contribute to the orderly movement of sister DNA regions away from the central replisome. We speculate that Smc, which is clearly important for chromosome partitioning (Britton et al., 1998; Graumann et al., 1998; Moriya et al., 1998; Britton and Grossman, 1999; Hirano, 1999), may play such a role. It has recently been proposed that Smc and the E. coli analog MukB generate higher order compaction of bacterial DNA by introducing constrained positive supercoils (Holmes and Cozzarelli, 2000; Sawitzke and Austin, 2000). Smc may be loaded onto newly replicated DNA to establish these constrained (+) supercoils and compact the DNA. In addition to Smc, Spo0J, a member of the ParB protein family, is required for efficient partitioning (Treton et al., 1994), probably by functioning to organize the origin region into a higher order structure (Glaser et al., 1997; Lin et al., 1997; Britton et al., 1998; Lin and Grossman, 1998; Britton and Grossman, 1999).
lacI Operator/Lac Repressor System for Visualizing Regions of Chromosomal DNA

The Lac repressor/lac operator system for visualizing chromosome regions was first developed for budding yeast and Chinese hamster ovary cells (Robinett et al., 1996; Straight et al., 1996; Belmont and To

To release cells from stringent response, cells were removed from medium with arginine hydroxymate by pouring the culture onto a sterile 0.2 μm Naigene bottle top filter. Cells were washed with 2–3 volumes of medium and resuspended in fresh growth medium plus the following additions: arginine (600 μg/ml), a standard mix of all 20 amino acids (Harwood and Cutting, 1990), adenosine (20 μg/ml), guanosine (20 μg/ml), thymidine (50 μg/ml), and cytidine (50 μg/ml).

All data presented in the tables and figures are cumulative from at least two experiments, with each experiment giving similar results.

Live Cell Microscopy

Slides of live cells were prepared essentially as described (Lemon and Grossman, 1998). Briefly, an aliquot of cells was removed from an exponentially growing culture and stained with the vital membrane stain FM4-64 (140–280 ng/ml) (Molecular Probes) prior to placement on a pad of 0.5%–1% agarose in a solution of 1 mM MgSO4 (Harwood and Cutting, 1990). The following filter sets were used: a Cy3 filter set for FM4-64, Chroma filter set 41029 for YFP, and Chroma filter set 31044 for CFP. Images were acquired using a Nikon E800 microscope equipped with a CCD camera (Harmamatsu, model C4742-95) and Improvion LiveLab software. Images were colorized and merged using Improvion LiveLab software and cropped in Adobe Photoshop 3.05 or 5.5.

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References


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...ized mutants of the green fluorescent protein (GFP). Gene 173, 33–38.
the cell cycle of *Escherichia coli* as analyzed by fluorescent in situ hybridization. Biochimie 81, 797–802.


