The Highly Conserved Bacterial RNase YbeY Is Essential in Vibrio cholerae, Playing a Critical Role in Virulence, Stress Regulation, and RNA Processing

Citation

As Published
http://dx.doi.org/10.1371/journal.ppat.1004175

Publisher
Public Library of Science

Version
Final published version

Accessed
Fri Jun 23 15:07:08 EDT 2017

Citable Link
http://hdl.handle.net/1721.1/88174

Terms of Use
Creative Commons Attribution

Detailed Terms
http://creativecommons.org/licenses/by/4.0/
The Highly Conserved Bacterial RNase YbeY Is Essential in *Vibrio cholerae*, Playing a Critical Role in Virulence, Stress Regulation, and RNA Processing

Maarten Vercruysse1, Caroline Köhrer1, Bryan W. Davies2, Markus F. F. Arnold1, John J. Mekalanos3, Uttam L. RajBhandary1, Graham C. Walker1*

1 Department of Biology, Massachusetts Institute of Technology, Cambridge, Massachusetts, United States of America, 2 Department of Molecular Biosciences, The University of Texas at Austin, Austin, Texas, United States of America, 3 Department of Microbiology and Immunobiology, Harvard Medical School, Boston, Massachusetts, United States of America

Abstract

YbeY, a highly conserved protein, is an RNase in *E. coli* and plays key roles in both processing of the critical 3’ end of 16 S rRNA and in 70 S ribosome quality control under stress. These central roles account for YbeY’s inclusion in the postulated minimal bacterial genome. However, YbeY is not essential in *E. coli* although loss of *ybeY* severely sensitizes it to multiple physiological stresses. Here, we show that YbeY is an essential endoribonuclease in *Vibrio cholerae* and is crucial for virulence, stress regulation, RNA processing and ribosome quality control, and is part of a core set of RNases essential in most representative pathogens. To understand its function, we analyzed the RNA and ribosome profiles of a *V. cholerae* strain partially depleted for YbeY and other RNase mutants associated with 16 S rRNA processing; our results demonstrate that YbeY is also crucial for 16 S rRNA 3’ end maturation in *V. cholerae* and that its depletion impedes subunit assembly into 70 S ribosomes. YbeY’s importance to *V. cholerae* pathogenesis was demonstrated by the complete loss of mice colonization and biofilm formation, reduced cholera toxin production, and altered expression levels of virulence-associated small RNAs of a *V. cholerae* strain partially depleted for YbeY. Notably, the ybeY gene is conserved in several distinctantly related pathogens can fully complement an *E. coli* *ybeY* strain under various stress conditions, demonstrating the high conservation of YbeY’s activity in stress regulation. Taken together, this work provides the first comprehensive exploration of YbeY’s physiological role in a human pathogen, showing its conserved function across species in essential cellular processes.


Copyright: © 2014 Vercruysse et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This study was supported by grants from the National Institute of Health to GCW (GM31030) and ULR (GM17151), the MIT Deshpande Center to GCW, the National Institute of Allergy and Infectious Diseases (AI-018045) to JJM, and P30 ES002109 to the MIT Center for Environmental Health Sciences. GCW is an American Cancer Society Professor. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: gwalker@mit.edu

Introduction

To thrive in diverse and changing environments, bacteria have to quickly sense and respond to a broad range of stimuli and signals. The ability to adapt to environmental changes is particularly important for pathogens during invasion of the host. Bacterial adaptation is often mediated through modulation of gene expression, to quickly sense and respond to a broad range of stimuli and signals. The ability to adapt to environmental changes is particularly important for pathogens during invasion of the host. Bacterial adaptation is often mediated through modulation of gene expression, including small RNAs (sRNAs) that control gene expression. These regulatory molecules are perfectly suited to initiate rapid regulatory circuits, as they do not require translation and can be turned over quickly by various RNases [1–6].

RNases, which can be divided into endo- and exonucleases according to their substrate specificities, play crucial roles in bacterial pathogenesis by regulating the expression of many virulence factors. In uropathogenic *Escherichia coli*, the endoribonuclease RNase E controls expression of Pap pili required for attachment to the kidney by selectively processing the bicistronic *papA2I* mRNA [7]. In *Staphylococcus aureus*, the endoribonuclease RNase III mediates the degradation of duplexes of RNAIIH with mRNAs coding for early virulence factors, whereas the exoribonuclease PNPase is involved in the cold stress response and global mRNA turnover [3,9]. In Gram-positive bacteria, RNase Y is the functional equivalent of RNase E in *E. coli* and affects virulence of *S. aureus* and *Streptococcus pyogenes* in silkworm and mouse models [3].

Small RNAs (sRNAs) that control gene expression are perfectly suited to initiate rapid regulatory circuits, as they do not require translation and can be turned over quickly by various RNases [5,9]. For example, in *Vibrio cholerae*, several sRNAs have been identified with a role in pathogenesis, such as the quorum-sensing sRNAs *Qyr1*-4 and the ToxT activated sRNAs *TarA* and *TarB* [10,11]. In *S. aureus*, RNAIIH acts as both an activator and repressor of early virulence factors, and 6S RNA of *Legionella pneumophila* is crucial for replication in host cells [12,13]. The function of regulatory sRNAs often relies on their interactions with accessory proteins, e.g. the RNA chaperone Hfq for stabilization of the sRNA/target mRNA interaction and various RNases for specific degradation of sRNAs and their target mRNAs [2,5].

One of the most recently identified RNases is YbeY, a highly conserved member of the UPF0054 protein family. It is found in...
Author Summary

Bacteria adapt and survive unfavorable environments by quickly changing their gene expression and physiology, for example as pathogens do during infection of host cells. Gene expression is often determined by RNA turnover, a balance between transcription and RNA decay carried out by multiple RNases. The recently identified RNase YbeY was shown in *E. coli* to participate in rRNA maturation and 70 S ribosome quality control, however YbeY’s roles in other organisms and the extent of functional conservation is unknown. Here, we show that YbeY is an essential RNase in the pathogen *Vibrio cholerae*, critical for cell fitness and general stress tolerance. We demonstrate that YbeY is crucial for 16 S rRNA 3’ end maturation, assembly of functional 70 S ribosomes and ribosome quality control. Moreover, YbeY regulates virulence-associated small RNAs and its depletion leads to an overall reduction in pathogenesis, exemplified by significantly decreased biofilm formation, mouse colonization and cholera toxin production. We also show that YbeY belongs to a minimal core set of RNases essential in most representative pathogenic bacteria, and its depletion leads to an overall reduction in pathogenesis, exemplified by significantly decreased biofilm formation, mouse colonization and cholera toxin production. YbeY acts together with RNase R can remove defective 70 S ribosomes, 16 S rRNA precursors and free ribosomal subunits. Compared to the 16 S rRNA processing RNase G [29] and the exonuclease RNase R, also shown to be involved in 16 S rRNA processing [16,30], only the loss of YbeY led to severe defects in 16 S rRNA 3’ end maturation and 70 S ribosome assembly. In the case of the multifunctional RNase E [31], the gene could not be deleted because it is essential, but a strain encoding RNase E ΔCTD, in which the carboxy-terminal domain is disrupted, had no effect on 16 S processing. YbeY is also crucial for the virulence of *V. cholerae*, since a ΔybeY deletion strain that expresses reduced levels of YbeY fails to colonize infant mice intestines and its biofilm and CT production are significantly reduced compared to wild-type (Wt). In addition, we experimentally identified several YbeY- and RNase E ΔCTD-dependent sRNAs associated with *V. cholerae* virulence. Strikingly, ybeY genes from four distantly related pathogens can fully complement an *E. coli* ΔybeY strain, demonstrating the high conservation of YbeY activity across species.

Results

YbeY is part of a core set of RNases essential in most representative pathogens

To date, YbeY’s physiological roles have been studied in *E. coli* [16–19,32] and *S. meliloti* [14,21]. Its multifaceted functions in rRNA and sRNA metabolism make YbeY a key player among RNases. To gain further insight into the role of YbeY within the network of bacterial RNases, we carried out a comprehensive phylogenetic analysis of RNases in a selection of bacteria with a focus on human pathogens, assessing (i) the presence or absence of major RNases and (ii) their essentiality.

Figure 1 summarizes the results of our analysis for the main endo- and exoribonucleases in eleven intensively studied bacterial species, using available transposon collections and genome-wide gene-deletion studies [33–50]. As described in detail in Materials and Methods, this summary represents our best effort to analyze publically available databases, taking into account possible misclassification of the essential nature of individual RNases (e.g., transposon mutagenesis can miss mutants that grow very slowly or tolerate certain insertions; gene inactivation cannot detect essential functions covered by multiple genes). Bacterial species were sorted according to phylogeny based on the amino acids of YbeY.
acid sequence of each YbeY homolog; RNases were sorted according to their distribution, the most ubiquitous RNases being listed first.

This comparative analysis of RNases (Figure 1) reveals numerous patterns: a) YbeY is more often essential than not, indicating that the model organism *E. coli* is not always the most representative; b) the RNases of the postulated minimal bacterial genome [15] overlap well with the most ubiquitous RNases of both Gram-negative and Gram-positive bacteria, e.g. YbeY, RNase P, RNase III and PNPase; c) the majority of essential RNases are endonucleases, whereas bacterial exonucleases are functionally more redundant than endonucleases and are hence rarely

**Table 1.** Endonucleases and Exonucleases Present in Human Pathogens

<table>
<thead>
<tr>
<th>Substrate</th>
<th>YbeY</th>
<th>RNase P</th>
<th>RNase III</th>
<th>RNase HII</th>
<th>RNase L-PSP</th>
<th>RNase E</th>
<th>RNase Z</th>
<th>RNase J1</th>
<th>RNase H1</th>
<th>RNase G</th>
<th>RNase M5</th>
<th>RNase HIII</th>
<th>RNase J2</th>
<th>RNase Mini III</th>
<th>RNase I</th>
</tr>
</thead>
<tbody>
<tr>
<td>m</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>r</td>
<td>+</td>
<td>+</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>N</td>
</tr>
<tr>
<td>t</td>
<td>+</td>
<td>+</td>
<td>E</td>
<td>E</td>
<td>E</td>
<td>E</td>
<td>E</td>
<td>E</td>
<td>E</td>
<td>E</td>
<td>E</td>
<td>E</td>
<td>E</td>
<td>E</td>
<td>E</td>
</tr>
<tr>
<td>s</td>
<td>+</td>
<td>+</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>N</td>
</tr>
<tr>
<td>o</td>
<td>+</td>
<td>+</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>N</td>
<td>N</td>
</tr>
</tbody>
</table>

Exonucleases

- PNPase
- RNase R
- RNase PH
- Oligoribonuclease
- RNase Z
- RNase J1
- RNase T
- RNase D
- RNase II
- RNase J2

Number of essential RNase genes: 5-6, 3, 4, 6-7, 6-7, 5-6, 6, 7, 5, 4-5, 6, 4

Total number of RNase genes: 15, 17, 17, 15, 14, 15, 14, 14, 12, 10, 9, 4

Figure 1. Essential RNases in human pathogens. Compilation of major RNases in *V. cholerae* (Vc), *E. coli* (Ec), *S. typhi* (St), *H. influenzae* (Hi), *P. aeruginosa* (Pa), *B. subtilis* (Bs), *S. aureus* (Sa), *S. pneumoniae* (Sp), *M. tuberculosis* (Mt), *H. pylori* (Hp), and *M. genitalium* (Mg). The asterisk indicates RNases present in the minimal bacterial genome set [15]. RNases with dual endo- and exonuclease functions are listed twice, but are counted only once to assess the total number of genes. Blank squares indicate that no homolog for the respective RNase could be identified. The designation of RNases as putatively essential (E) and non-essential (N) is based on the availability of deletion or transposon mutants according to Materials and Methods. Current databases list these RNases as non-essential; follow-up experiments suggest these RNases could by essential (personal communication; BW Davies, LA Simmons, Y Furuta). Only mutants with transposon insertions in the last 10% of the gene are available. The key roles of RNases in mRNA (m), rRNA (r), tRNA (t) and sRNA (s) metabolism are indicated; other functions (o). The phylogenetic tree on top is based on alignment of various YbeY protein sequences using Geneious software.

doi:10.1371/journal.ppat.1004175.g001

YbeY’s Functional Role in *V. cholerae*
YbeY’s Functional Role in V. cholerae

Figure 2. Generation of a ΔybeY strain in V. cholerae. A) Genomic organization of the ybeZ (VC0961)-ybeY (VC0960) locus in the V. cholerae strain C6706. An in-frame deletion of ybeY was carried out in C6706; the position of the chromosomal ybeY deletion is indicated (△). B) Growth analysis of the C6706 wild-type strain (WT pY) and ΔybeY pY mutant strain (△ybeY pY) transformed with a plasmid containing ybeY under control of an arabinose-inducible promoter. Cells were first grown in LB medium supplemented with arabinose and then subcultured in medium containing arabinose or glucose to obtain full induction or maximum depletion of YbeY, respectively. “pY” indicates that ybeY is expressed from a plasmid. Ara+, cells were grown in LB in the presence of arabinose. Gluc+, cells were grown in LB in the presence of glucose. C) RT-qPCR analysis of ybeY mRNA in the C6706 wild-type strain (WT) and the ΔybeY pY strain. Samples were taken from log phase cultures (OD600 0.5), as shown above. The levels of ybeY mRNA were normalized to 5 S rRNA. The WT strain grown in the absence of any carbon source and without the maintenance plasmid was set to 1.

DOI:10.1371/journal.ppat.1004175.g002

YbeY is essential for growth of V. cholerae

A large-scale, saturating transposon library of V. cholerae showed no hits for the ybeY homolog, designated VC0960 [36]. To experimentally verify the essentiality of ybeY in V. cholerae, we attempted to create a non-polar deletion of ybeY, which is part of the ybeZ operon (Figure 2A), in V. cholerae C6706 using standard allelic exchange [52]. However, chromosomal ybeY could be deleted only in a strain carrying a copy of ybeY on the arabinose inducible vector pBAD18 (henceforth referred to as ΔybeY pY) in the presence of 0.1% arabinose. In the absence of arabinose, no such non-polar deletion of ybeY could be obtained.

The growth rate of the ΔybeY pY strain is identical to its parental strain in LB medium in the presence of 0.1% arabinose (denoted as Ara+) (Figure 2B). When the ΔybeY pY strain was transferred to arabinose-free medium (denoted as Ara-) (Figure S1), cells showed a reduction in growth rate indicating YbeY depletion. If these cells were subcultured a second time into medium lacking arabinose (denoted as Ara-/Ara-) (Figure S1), cell growth was diminished to such an extent that we could not obtain enough cells for experimental measurements. An experimentally more convenient protocol for decreasing the expression of YbeY from pBAD18 is to shift the Ara+ cells to medium lacking arabinose but containing glucose, which decreases the 3’,5’-cyclic AMP levels, lowering expression from the catabolite-repressed PBAD promoter. Thus, maximal depletion of YbeY was obtained by subculturing overnight cultures of the ΔybeY pY strain grown in the presence of arabinose into medium that lacked arabinose but was supplemented with 0.2% glucose (denoted as Gluc+) (Figure 2B). After being switched to medium with glucose, ΔybeY pY cells initially grow similar to the wild-type (WT) but grow slower after mid-exponential phase and stop growing at a significantly lower optical density (OD600) in stationary phase. No cell death or loss of viability was observed in late-stationary phase after YbeY depletion when cells were plated on arabinose-containing solid media to determine colony-forming units (data not shown). Since simply inoculating overnight cultures of cells grown with arabinose into medium containing glucose (Gluc+) gave robust YbeY depletion, we have chosen these conditions for most experiments described in this study, as they allowed us to maximally deplete essential; and d) the genome size of each organism correlates well with the number of major RNases, although the total number of essential RNases remains relatively constant with an average of five to six RNases. E. coli and S. typhi are extremes and show a remarkable redundancy of RNase genes with the highest number of total RNase genes and the lowest number of essential RNase genes, suggesting that many RNases share overlapping functions.

Overall, our comparative analysis of the essential nature of RNases in eleven different bacteria from a wide phylogenetic range allowed us to put YbeY firmly within a minimal core set of RNases, which overlaps with the set of RNases found in the minimal bacterial genome. Thus, it is not surprising that YbeY is essential in most pathogens that were included in our analysis. We chose to study YbeY in the model pathogen V. cholerae strain C6706, which is a clinical isolate of the seventh pandemic belonging to the O1 El Tor biotype and highly similar to the sequenced strain N16961 [51]. Of particular interest, its ybeY homolog is proposed to be essential [36].
YbeY levels in a controlled fashion (see also Figure 2C) without affecting cell viability.

The levels of ybeY mRNA in V. cholerae wild-type and the plasmid-containing mutant strain were determined by RT-qPCR (Figure 2C). Notably, the presence of the pBAD18 derivative containing the extrachromosomal copy of ybeY (pY) led to a moderate increase in ybeY mRNA levels due to leaky transcription in the absence of arabinose or glucose. Expression of ybeY is strongly induced by arabinose (Ara+). In contrast, ybeY expression was reduced to 20% of the endogenous wild-type ybeY levels when arabinose was removed from the media and glucose was added (Gluc+).

Our ability to generate a ybeY deletion in V. cholerae C6706 only in the presence of a complementing plasmid and the striking decrease in growth rate upon YbeY depletion strongly support the conclusion that YbeY is essential for growth of V. cholerae in rich media.

Depletion of YbeY in V. cholerae affects 16 S rRNA maturation and 70 S ribosome assembly

Functional 70 S ribosomes are composed of 16 S, 23 S, and 5 S rRNAs that are processed from a larger RNA precursor. In E. coli, YbeY has been shown to be primarily involved in the maturation of both termini of 16 S rRNA and to also play a minor role in the 5’ maturation of 23 S and 5 S rRNAs [16]. Moreover, YbeY is essential for 16 S rRNA maturation at 45°C [18]. Very little is known about rRNA maturation in V. cholerae to date. To determine the role of V. cholerae YbeY in rRNA maturation, we isolated total RNA from the wild-type and the ΔybeY pY strain (Figure 3A). ΔybeY pY cells grown in the presence of arabinose (Ara+) exhibited a wild-type like rRNA profile. However, maximum depletion of YbeY in cells grown in the presence of glucose (Gluc+) showed strong accumulation of 17 S rRNA. We were able to detect an intermediate decrease of 16 S RNA and a concomitant intermediate increase of 17 S precursor levels compared to the parental strain by switching the carbon source of the ΔybeY pY strain (Figure 3A). Mapping of the 5’ and 3’ termini of all rRNAs confirmed the strong defect of 16 S rRNA maturation upon YbeY depletion, however no 23 S and 5 S rRNA processing deficiencies were detected (Figure 3B, S2). The latter shows that although E. coli and V. cholerae are considered to be fairly closely related species, the substrate specificity of individual RNases may be different as the distribution of RNase genes and their essentiality also differ (Table S1). Mapping of the 16 S rRNA 5’ end showed, as expected, two precursors since 4 of the 8 16 S rRNA genes have a ~20-nucleotide deletion within the 5’ terminal precursor sequence in V. cholerae C6706.

The 16 S rRNA maturation defect also affects the ribosome pool as indicated by sucrose gradient analysis of ribosomes from V. cholerae wild-type and the ΔybeY pY strain. Upon gradual depletion of YbeY, the level of intact 70 S ribosomes decreased accordingly and the level of individual 50 S and 30 S subunits increased compared to the parental strain (Figure 3C). Compared to an E. coli ΔybeY strain in which the ybeY gene was completely deleted, we observed a more pronounced accumulation of individual ribosomal subunits in the V. cholerae ΔybeY pY strain (Figure 4A), when ybeY mRNA levels are reduced to 20% of wild-type levels (Figure 2C, maximum YbeY depletion).

Mapping of both ends of 16 S rRNA in total RNA, 70 S ribosomes and 30 S subunits isolated from the V. cholerae ΔybeY pY strain upon maximum YbeY depletion and from the E. coli ΔybeY strain demonstrated that 30 S subunits contain mostly immature 16 S rRNA, whereas rRNA isolated from assembled 70 S ribosomes is mostly mature (Figure 4B). Total RNA samples, representing a mixture of free rRNA and rRNA from 30 S and 70 S ribosomes, also show strong maturation defects. These effects are similar in V. cholerae and E. coli. Observations made here are consistent with the assembly of immature or only partially matured 16 S rRNA into 30 S subunits. However, if such subunits are not processed further, as is the case in cells with reduced or no YbeY activity, they cannot be assembled efficiently into 70 S ribosomes. As a consequence, although the 70 S ribosome peaks in both the V. cholerae YbeY-depleted strain and the E. coli ΔybeY strain (Figures 3C and 4A) are drastically reduced, they contain mostly mature 16 S rRNA (Figure 4B). Notably, the level of 17 S precursor in 70 S ribosomes isolated from E. coli ΔybeY shown in this work (Figure 4B, 4C) is lower compared to our previous work [16,18], though 16 S rRNA with mature termini is consistently the dominant species in 70 S ribosomes in contrast to 30 S ribosomal subunits. In the course of our work with ΔybeY strains, we have observed that the absolute amount of precursor species can vary, depending on slight differences in culturing conditions, such as the level of aeration, source of LB medium etc. (data not shown).

YbeY of E. coli can degrade rRNA, mRNA and oligoribonucleotides in vitro [18]. Its active site has a highly conserved metal-coordinating histidine H3XH5XH motif that is characteristic to the UFP0054 protein family. The RNase activity of V. cholerae YbeY was confirmed by our demonstration that it was capable of degrading total V. cholerae rRNA, while its metal-dependency was confirmed by showing that the addition of EDTA inhibited this degradation (Figure S3A). Using an RNA hairpin and single-strand oligoribonucleotide substrates, we further demonstrated that V. cholerae YbeY is an endoribonuclease, whose substrate specificity is very similar to E. coli YbeY (Figure S3B, S3C). To improve in vitro activity and to prevent aggregation of purified V. cholerae YbeY, we used a plasmid that encodes both GroES and GroEL to increase chaperone activity. The RNase activity of purified V. cholerae YbeY was still slightly lower than that of purified E. coli YbeY, which could be due to the fact that YbeY was purified from E. coli BL21, a non-native host strain. In addition, we showed that YbeY together with RNase R of both V. cholerae and E. coli can degrade 70 S ribosomes isolated from the V. cholerae ΔybeY pY strain grown under maximum YbeY depletion, but does not degrade V. cholerae wild-type ribosomes (Figure 4C, S3D). This is consistent with analogous experiments in E. coli [18], thereby demonstrating the conservation of the role of V. cholerae YbeY in ribosome quality control and the cross-species conservation of the YbeY-dependent ribosome quality control mechanism because E. coli enzymes recognize and degrade defective V. cholerae ribosomes.

16 S rRNA maturation in V. cholerae mutant strains carrying transposon insertions in genes encoding RNase E, RNase G, or RNase R.

To analyze the involvement of V. cholerae YbeY in maturation of 16 S rRNA in comparison to other RNases that are functionally associated with 16 S rRNA maturation in E. coli, we compared the 5’ and 3’ termini of 16 S rRNA from V. cholerae wild-type, the mectD::Tn [36], rng::Tn [36] and mrc::Tn mutant strains [36], and the ΔybeY pY strain depleted of YbeY (Gluc+) (Figure 5). In E. coli, RNase E (rne) and RNase G (rng) participate in the two-step, sequential maturation of the 5′ end of 16 S RNA [29]. Although RNase E is essential for growth in E. coli, several rne::Tn mutants were found in V. cholerae transposon libraries [36,53]. However, all transposons were inserted downstream of the N-terminal domain (NTD) of RNase E that specifies the endonuclease activity,
Figure 3. Analysis of rRNA and ribosome profiles in V. cholerae ΔybeY. 

A) Total RNA was isolated from C6706 Wt pY and the ΔybeY pY strain grown in the presence of arabinose or glucose as specified and analyzed by agarose gel electrophoresis. The positions of 23 S, 17 S, and 16 S rRNAs are indicated based on their mobility.

B) The 5' and 3' termini of 16 S rRNA from C6706 Wt pY and the ΔybeY pY strain were mapped as described in detail in Materials and Methods. "P" and "M" specify the positions of bands derived from the precursor and mature forms of 16 S rRNA.

C) Ribosome profiles for C6706 Wt pY and the ΔybeY pY strain (top); quantitation of polysomes, 70 S, 50 S and 30 S ribosomes (bottom pie charts). "pY" indicates that ybeY is expressed from a plasmid. Ara+, cells were grown in LB in the presence of arabinose. Gluc+, cells were grown in LB in the presence of glucose. Ara+/Gluc+, intermediate YbeY depletion by switching the carbon source of the ΔybeY pY strain in early exponential phase from arabinose to glucose (for details see Materials and Methods).

doi:10.1371/journal.ppat.1004175.g003
confirming the essentiality of its nuclease activity in *V. cholerae*. The C-terminal domain (CTD) functions as a scaffold for various interacting proteins that make up the degradosome and was recently shown to be required for binding to 30 S subunits of *E. coli*.[31,54]. Since multiple mutants with transposon insertions within the CTD of RNase E are available in *V. cholerae*[36,53], a fully functional degradosome may not be essential for the viability of *V. cholerae*. The transposon of the representative *rne::Tn* mutant we chose to investigate further is located between segment A and the first RNA binding domain of RNase E.[31]. Therefore, this mutant, designated as *rne*-CTD::Tn, lacks the full C-terminal scaffolding domain as confirmed by RT-qPCR (data not shown).

**Figure 4.** Comparison of ribosome profiles and 16 S rRNA maturation in *V. cholerae* ΔybεY and *E. coli* ΔybεY. **A**) Ribosome profiles for *V. cholerae* C6706 Wt pY and the ΔybεY pY strain, and *E. coli* MC4100 Wt and the ΔybεY strain (top); quantitation of polysomes, 70 S, 50 S and 30 S ribosomes (bottom pie charts). “pY” indicates that ybeY is expressed from a plasmid. *V. cholerae* Wt pY was grown in LB in the presence of arabinose (Ara+) and *V. cholerae* ΔybεY pY was grown in LB in the presence of glucose for maximum depletion (Gluc-). *E. coli* cells were grown in LB medium without additional carbon source. **B**) Mapping of 5’ and 3’ termini of 16 S rRNA in total RNA, 30 S ribosomal subunits and 70 S ribosomes from C6706 Wt pY (W) and the ΔybεY pY strain (Δ), and *E. coli* MC4100 Wt (W) and the ΔybεY strain (Δ). rRNA was isolated from samples shown in A). “P” and “M” specify the positions of bands derived from the precursor and mature forms of 16 S rRNA. **C**) *In vitro* ribosome quality control by *V. cholerae* (Vc) and *E. coli* (Ec) YbeY together with RNase R. 70 S ribosomes of a *V. cholerae* ΔybεY pY strain upon maximum YbeY depletion were incubated with YbeY, RNase R, or a mixture of YbeY and RNase R from *E. coli* or *V. cholerae* as indicated. The positions of 23 S and 16 S rRNAs are indicated based on their mobility. doi:10.1371/journal.ppat.1004175.g004
YbeY’s Functional Role in *V. cholerae*

The efficiency of 16 S rRNA 5’ end processing is reduced considerably upon depletion of YbeY as well as in the absence of RNase G, but is unaffected in both *rne*-CTD::Tn and *rne*:Tn mutants (Figure 5, top panel). Notably, no mature 5’ end was detected in total RNA samples from the *V. cholerae rng*:Tn mutant, in contrast to *E. coli*, which still contains a substantial amount of the mature form in the absence of RNase G [29]. These results were confirmed by agarose gel electrophoresis, showing a 17 S-like precursor in total RNA of the *rng*:Tn mutant (Figure 5, bottom panel). Maturation of the 3’ end of 16 S rRNA is defective upon maximum depletion of YbeY in *V. cholerae* but not in the *rne*-CTD::Tn, *rng*:Tn and *rne*:Tn mutants, demonstrating that YbeY is a key RNase required for processing of the critical 16 S rRNA 3’ end (Figure 5, middle panel).

In both the 30 S subunits and 70 S ribosomes of the *V. cholerae rng*:Tn mutant, additional, although still incomplete, processing of the 5’ end was observed, showing that low-level maturation occurs in the ribosomal context. The 70 S ribosome of the *V. cholerae* *rng*:Tn mutant contains immature 5’ ends, which is different from the *V. cholerae* YbeY-depleted ΔybeY pY strain. This is consistent with the fact that the *V. cholerae* *rng*:Tn has no apparent growth phenotype (data not shown) and ribosomes with immature 16 S rRNA 5’ termini can function in contrast to ribosomes with immature 3’ termini [53–57].

Sucrose gradient analysis of *V. cholerae* ribosomes shows no defects in the absence of RNase G, RNase E, ΔCTD, and RNase R compared to the wild-type (Figure S4), possibly due to correct 16 S rRNA 3’ maturation. Therefore, among the RNases whose function we assessed, YbeY is the only RNase crucial for 3’ processing of 16 S rRNA and its assembly into 70 S ribosomes in *V. cholerae*.

**YbeY affects regulation of virulence associated small RNAs**

Expression of critical virulence genes in various pathogens is often regulated by small RNAs (sRNAs). In many cases, regulation by sRNA depends on the RNA chaperone Hfq with a subsequent involvement of RNase E [58]. In *S. meliloti*, YbeY function clearly affected the regulation of certain sRNAs and their target mRNAs [21]. To investigate a possible role of *V. cholerae* YbeY in sRNA regulation, we selected five sRNAs thought to be important for pathogenicity: MicX and VrrA, which regulate the expression of outer membrane proteins [59,60]; TarB, which targets the colonization factor TcpF [61] and the *Vibrio* seventh pandemic regulator VspR [62]; Qrr1–4 sRNAs, which control quorum sensing and those virulence genes that are regulated by quorum sensing [63]; and the regulator 6 S RNA, which binds to and thereby affects transcription of RNA polymerase σ70-driven genes [12].

Expression patterns of MicX, VrrA, TarB, Qrr1–4 and 6 S RNA were monitored by Northern blot analysis of total RNA isolated from ΔybeY pY cells upon maximum YbeY depletion and from *rne*-CTD::Tn cells (Figure 6). Steady state levels of MicX and TarB sRNAs were significantly reduced upon YbeY depletion compared to the parental strain, while levels of Qrr1–4 sRNAs were elevated (Figure 6A). In contrast, the *rne*-CTD::Tn mutant, which harbors an RNase E mutant lacking the docking domain for Hfq and other components of the degradosome [31], showed no changes in MicX and TarB sRNA levels but strongly increased VrrA and Qrr1–4 sRNA levels (Figure 6B). 6 S RNA levels did not change in either of the mutant strains (Figure 6A and B). Quantitation of sRNA levels was carried out using 3 S rRNA as a loading control (Figure 6C and D). Taken together, these results

---

**Figure 5.** 16 S rRNA maturation in *V. cholerae* mutant strains with transposon insertions in genes encoding RNase E (*rne*-CTD::Tn), RNase G (*rng*:Tn), or RNase R (*rne*:Tn). Mapping of 5’ and 3’ termini of 16 S rRNA in total RNA, 30 S ribosomal subunits and 70 S ribosomes from C6706 Wt pY (W), ΔybeY pY (Δ), *rne*-CTD::Tn (E), *rng*:Tn (G) and *rne*:Tn (R); “P” and “M” specify the positions of bands derived from the precursor and mature forms of 16 S rRNA (top and middle panel). Analysis of total RNA by agarose gel electrophoresis; the positions of 23 S, 17 S and 16 S rRNAs are indicated based on their mobility (bottom panel). “P” indicates that ybeY is expressed from a plasmid. *V. cholerae* Wt pY was grown in LB in the presence of arabinose (Ara+) and *V. cholerae* ΔybeY pY was grown in LB in the presence of glucose for maximum depletion (Gluc+); all other strains were grown in LB medium without additional carbon source. doi:10.1371/journal.ppat.1004175.g005
show a clear role of *V. cholerae* YbeY in sRNA regulation, different from that of the traditional Hfq/RNase E-mediated system.

**YbeY plays an important role in *V. cholerae* pathogenesis**

Certain RNases are important for pathogens to survive the fluctuating conditions during host infection and to control the expression of many virulence factors [64]. We have shown YbeY to be an RNase in *V. cholerae* that plays a role in regulating the levels of several virulence-associated sRNAs. To study YbeY’s role in *V. cholerae* pathogenesis, we investigated the effect of YbeY’s depletion on a number of specific virulence factors of *V. cholerae*.

Initially, we observed an apparent loss of pigmentation in the ΔybeY strain upon maximum YbeY depletion to 20% of wild-type YbeY levels (Figure 7A). Also, overnight biofilm formation in static LB cultures was dramatically reduced upon maximum YbeY depletion, compared to wild-type and ∆ybeY pY cultures grown in the presence of arabinose (Figure 7B). Additionally, depletion of YbeY significantly reduced CT production of ΔybeY pY cells (Figure 7C). Exposure to bile salts resulted in a two to four order of magnitude drop in survival of the ΔybeY pY strain upon YbeY depletion, 1 to 3 hours after cells were shifted to media lacking arabinose (Figure 7D). Finally, as depletion of YbeY clearly affects multiple virulence factors, we assessed YbeY’s overall ability to colonize and persist in an infant-mouse colonization model (Figure 7E). Using a competition assay, we grew the wild-type and ΔybeY pY strain in the absence of arabinose prior to inoculation (Figure 7E). Only a minor colonization defect was observed with strains grown in the presence of glucose.
The presence of arabinose prior to inoculation. This lack of complete complementation is probably due to the absence of arabinose in the intestinal tract of mice, allowing for some/slight YbeY depletion. Taken together, these results suggest that YbeY plays a critical role during multiple stages of the V. cholerae’s infection cycle.

YbeY plays a central role in stress regulation of V. cholerae

The phenotype of ΔybeY deletion mutants in S. meliloti and E. coli is extremely pleiotropic, as they exhibit a strong sensitivity to a wide range of physiological stresses [14,16]. To analyze the importance of YbeY function in protecting V. cholerae against stress, we tested the sensitivity of the V. cholerae ΔybeY pY strain to a broad variety of stresses targeting key cellular processes after YbeY had been depleted by growth in glucose. These stresses include antibiotics targeting protein synthesis, transcription, and cell wall synthesis, the oxidative stress \( \text{H}_2\text{O}_2 \), the DNA-damaging agent ultraviolet radiation and heat (Figure 8).

Cell growth was tested on plates by serial dilution of overnight cultures of V. cholerae wild-type and ΔybeY pY cells grown in the presence of arabinose; to deplete YbeY, glucose was present in the plating medium as indicated. Under non-stress conditions (37°C), ΔybeY pY showed similar growth upon YbeY depletion compared to wild-type, although mutant colonies at the highest dilutions were smaller than wild-type colonies due to their reduced growth rate (Figure 8A). Although ybeY is essential, no cell death was observed under these conditions, most likely because ybeY’s leaky expression still reaches approximately 20% of the endogenous wild-type ybeY levels (Figure 2B).

In general, cell growth of the V. cholerae ΔybeT pY strain on plates supplemented with glucose and one of several stress agents was drastically reduced compared to the parental strain (Figure 8A). Since YbeY has a major role in 16 S rRNA maturation, we tested three different antibiotics that target the 30 S ribosomal subunit, tetracycline, gentamycin and kasugamycin. The ΔybeT pY strain showed a severe increase in sensitivity upon YbeY depletion to all of these antibiotics. A similar effect was observed with antibiotics targeting the 50 S ribosomal subunit, chloramphenicol and erythromycin. Additionally, we found that the ΔybeT pY strain also exhibited increased sensitivity to inhibition of RNA synthesis by rifampicin and cell wall synthesis.
by both ampicillin and cefotaxime. As *V. cholerae* is exposed to oxidative stress during host infection and UV radiation outside of the host, both stresses were tested as well, showing a consistent decrease in protection against oxidative and UV stress upon YbeY depletion (Figure 8A).

In *E. coli*, YbeY is an important factor for rRNA maturation even at 37 °C but becomes essential upon temperature shift to 45 °C as shown by a complete loss of mature 16 S rRNA and cell viability [16,18,19]. YbeY depletion in a *V. cholerae ΔybeY* strain grown in glucose also causes a growth defect at 45 °C after 3 hours compared to wild-type (Figure 8B), however only a two to three orders of magnitude drop in survival was consistently observed, in contrast to *E. coli*’s overall loss of viability at elevated temperatures (see Figure 9A). This difference might in part be *V. cholerae* specific but may also be due to the low background levels of YbeY still present in the *V. cholerae* depletion strain. Overexpression of *ybeY* does not provide additional heat protection to *V. cholerae* because the wild-type strain with and without the *ybeY* overexpression plasmid showed a similar level of heat tolerance (data not shown). Interestingly, a cold-sensitive phenotype was found upon depletion of YbeY in a *V. cholerae ΔybeY* strain at 18 °C (Figure 8C). A similar cold-sensitivity was not observed in *E. coli*, further illustrating the disparity in response to changes in temperature between *V. cholerae* and *E. coli* depleted of YbeY.

YbeY’s activity in stress regulation and rRNA maturation is highly conserved among evolutionary distant pathogens

YbeY is a highly conserved protein found in almost all sequenced bacteria. To determine the level of functional conservation, we complemented the *E. coli ΔybeY* strain – grown under a variety of stress conditions – using constitutively expressed *ybeY* of four distantly related human pathogens, *V. cholerae*, *P. aeruginosa*, *Klebsiella pneumoniae*, and *A. aerogenes*.

Figure 8. Sensitivity of *V. cholerae ΔybeY* to various stresses. A) C6706 Wt pY and ΔybeY pY cells were grown overnight in LB medium supplemented with arabinose, subsequently diluted to an OD₆₀₀₀.1 and then spotted as a dilution series (1:10) onto LB plates supplemented with various antibiotics or H₂O₂, or irradiated with UV as indicated. Cells were grown in the presence of glucose (Gluc+) or arabinose (Ara+) as indicated. B) To determine heat-sensitivity of C6706 Wt pY and ΔybeY pY cells, early exponential cultures supplemented with glucose were incubated for 3 h at 37 °C and 45 °C before spotting as a dilution series (1:10) onto LB plates supplemented with glucose (Gluc+). Cells were then grown for 18 h at 37 °C. 

C) To determine cold-sensitivity of C6706 Wt pY and ΔybeY pY cells, cultures were spotted as a dilution series (1:10) onto LB plates supplemented with glucose (Gluc+). Cells were then grown for 2 days at 37 °C and 18 °C as indicated.

doi:10.1371/journal.ppat.1004175.g008
The function of YbeY in stress regulation and rRNA maturation is well conserved among distantly related pathogens.

**Discussion**

We have shown that YbeY is an essential RNAse in the pathogen *V. cholerae*. Depletion of YbeY: i) results in severe defects in 16 S rRNA maturation, ribosome assembly, and stress regulation, ii) affects regulation of virulence-associated sRNAs, and iii) reduces overall pathogenesis (Figure 10). Moreover, we found that key functions of YbeY’s multifaceted activity are highly conserved among pathogens.

YbeY is an essential RNAse in *V. cholerae* playing a central role in RNA metabolism

YbeY is the first endoribonuclease of *E. coli* shown to play a direct or indirect role in processing of the 16 S rRNA 3’ terminus [16,18]. However, most of what we know about rRNA processing comes primarily from studies in *E. coli* [31,63], while relatively little information is available in other organisms. Here, we have shown that *V. cholerae* YbeY is also a metal-dependent endoribonuclease, although in contrast to *E. coli*, ybeY is essential in *V. cholerae* (Figure 2). Gradual depletion of YbeY in *V. cholerae* results in gradual accumulation of 16 S RNA precursors that are defective in maturation of both ends, along with a gradual decrease in the efficiency of ribosome assembly (Figure 3). Similar to *E. coli*, where YbeY plays a minor role in 5’ maturation of 23 S and 5 S rRNAs [16], YbeY does not appear to be required for the maturation of 23 S and 5 S rRNAs in *V. cholerae*. We also examined 16 S rRNA processing in *V. cholerae* mutants of RNases associated with 16 S rRNA maturation in *E. coli*, the 5’ end processing RNases E and G and RNase R. The latter endonuclease plays an important role in 70 S ribosome quality control together with YbeY [18] and has been shown to largely bind to the ribosome close to the 5’ end of 16 S rRNA [66]. Using an *mc*::Tn mutant (*mc*-CTD::Tn) that contains the essential N-terminal nucleic domain but lacks the C-terminal degradosome scaffold [31], we have shown that an intact RNase E nucleic domain is sufficient for 5’ processing of 16 S rRNA (Figure 5). As expected, a 3’ processing defect was observed in the *rng*:Tn mutant. 3’ maturation was not affected in the *mc*-CTD::Tn and *rng*:Tn mutants, and lack of RNase R had no effect on 5’ or 3’ processing.

Similar to *E. coli* [16], depletion of YbeY leads to processing defects at both the 5’ and 3’ ends of 16 S rRNA (Figure 4). The role of YbeY in 5’ maturation of 16 S rRNA is likely indirect. It is possible that 5’ processing of 16 S rRNA is dependent on 3’ processing, which is in accord with a recent report showing that in *E. coli* 5’ processing proceeded much less efficiently in the absence of four exoribonucleases thought to be involved in 3’ maturation of 16 S rRNA [30]. Moreover, the 30 S particles in the YbeY depleted samples contained mostly 16 S rRNA precursors, indicating that correct 16 S rRNA maturation may be required for efficient 70 S ribosome assembly and that the final maturation of 16 S rRNA occurs on the ribosome.

We also demonstrated that 70 S ribosome quality control mediated by YbeY and RNase R is not limited to *E. coli*. Recent *V. cholerae* 70 S ribosomes were degraded by the combined action of *V. cholerae* YbeY and RNase R (Figure 4 and S3). Similar to YbeY, RNase R is also present in most representative pathogens (Figure 1), suggesting the possibility of a universal ribosome quality control mechanism executed jointly by an endonuclease and an exonuclease. Since 70 S ribosomes isolated from a *V. cholerae* YbeY strain containing 16 S rRNA with mostly mature termini are completely degraded by YbeY and RNase R, we can assume that this quality control mechanism is not solely triggered by unprocessed RNA. In agreement with our previously proposed model [18], YbeY alongside RNase R may act as a sensor of rRNA perturbations caused by misfolding or lack of certain base modifications, rRNA damage by environmental factors, or an imbalance in ribosomal proteins or ribosome maturation factors in defective 70 S ribosomes. In addition, it is quite possible that some of the factors that trigger the YbeY-dependent ribosome quality control described above may also contribute to the defects in ribosome assembly observed in YbeY-depleted strains.

YbeY’s function in RNA metabolism also extends to sRNA regulation as reported earlier in *S. meliloti* [21]. YbeY proteins share structural similarities with the MID domain of Argonaute proteins, illustrating a conceptual parallel between prokaryote and eukaryote sRNA pathways [21]. Regulatory RNAs are a powerful means of regulating gene expression at multiple levels, allowing bacteria to respond quickly to environmental changes. Often, sRNAs depend on the RNA chaperone Hfq and RNA degradation by RNases, such as RNase E or PNPase [5,58,67]. Hfq was shown to be important for stress tolerance and virulence in many pathogens [68]. Currently, around 20 sRNAs have been experimentally confirmed in *V. cholerae* [69,70]. Here, we show for the first time, that YbeY is also involved in the regulation of several virulence-associated sRNAs in *V. cholerae*, namely MicX, TarB and Qrr1–4 (Figure 6A). It is currently not known, whether YbeY requires Hfq to do so. In *V. cholerae*, regulation of quorum sensing sRNAs and TarB is mediated by Hfq [62,71]. One could easily envision that YbeY functions alongside Hfq and replaces the function of the commonly used RNase E for a certain subset of sRNAs. Conversely, YbeY may also target certain sRNAs in an Hfq-independent manner as it has been shown for PNPase in *E. coli* [72]. Moreover, consistent with current knowledge of sRNA regulation we observed differential sRNA levels of VrrA and TarB in *V. cholerae* YbeY mutants (Figure 6B), but saw no changes for sRNAs MicX and TarB that are targeted by YbeY. Although the essential nucleic activity of RNase E is maintained in *mc*-CTD::Tn, the docking site for various degradosome components such as Hfq and PNPase is absent, impeding the RNase E-mediated recruitment of interacting partners that are required for certain sRNAs. These data hence establish YbeY as an important player in sRNA regulation alongside Hfq, RNase E and PNPase, and add to its key role in bacterial RNA metabolism.
YbeY affects multiple virulence factors and the overall stress response of V. cholerae

Virulence factors are gene products that improve bacterial survival during host infection. We demonstrate that depletion of YbeY impedes V. cholerae's virulence by affecting virulence factors at multiple stages along the pathogenesis cycle (Figure 7). First, YbeY depletion reduces the level of pigmentation. Pigments like melanin protect cells against UV radiation, which is a major source of stress for pathogens outside the host and was previously shown to regulate expression of virulence factors [73]. Second, biofilm formation is drastically reduced upon YbeY depletion. Biofilms are highly organized microbial communities that are characterized by the ability to resist diverse stresses like antibiotics and host immune systems [74]. Biofilms are central to pathogenesis of many bacterial infections [73]. Third, YbeY depletion significantly reduces cholera toxin (CT) production. Colonization of the small bowel by V. cholerae must occur before CT is secreted into the intestinal epithelial cells and causes severe diarrhea. Therefore, YbeY-depleted cells that colonize the intestine are substantially less toxic than the parental strain. Fourth, lack of YbeY highly sensitizes the V. cholerae ybeY mutant to bile salts, which are antimicrobial substances present in the small intestines of mammals and one of the first stress factors encountered by the bacteria after passage through the acidic stomach. Lastly, intestinal colonization by the V. cholerae ybeY mutant is almost completely lost upon YbeY depletion in competition with the wild-type strain in infant mice, illustrating the overall reduced in vivo fitness or reduced ability to establish an infection. This is consistent with YbeY being one of the 507 genes that are highly expressed in both stool samples from cholera patients and during mid-exponential growth in rabbit ileal loops compared to growth in LB [76,77].

Virulence defects observed upon YbeY depletion in V. cholerae could be partly indirect, since YbeY affects ribosome biogenesis and thereby globally affects protein synthesis.

---

**Figure 9. Complementation of E. coli ΔybeY by ybeY of various pathogens.**

A) E. coli MC4100 Wt pY and the ΔybeY pY strain grown in LB medium were serially (1:10) diluted and spotted onto LB plates containing various antibiotics or H2O2 treated with heat (45 °C) or UV (60 J/m2) as indicated. “pY EC” refers to a plasmid expressing ybeY of E. coli, similarly V. cholerae N16961 (pY_Vc), P. aeruginosa PAO1 (pY_Pa), M. tuberculosis H37Rv (pY_Mt), and S. pneumoniae R6 (pY_Sp). B) Total RNA was isolated from E. coli MC4100 transformed with the empty vector, and the E. coli ΔybeY strain transformed with plasmids containing ybeY from various bacteria as indicated. RNA was analyzed by agarose gel electrophoresis. The positions of 23 S, 17 S, 16 S and 16 S* rRNAs are indicated based on their mobility.

doi:10.1371/journal.ppat.1004175.g009
The reduced virulence of the ybeY mutant is, however, opposite to that expected from the relative fall in TarB levels and rise in Qrr1–4 levels upon YbeY depletion; low TarB and elevated Qrr1–4 levels generally correlate with increased expression of specific virulence genes [61,62,71,78]. Virulence, as defined by successful penetration of the host’s defense mechanisms, colonization and toxigenesis, is nonetheless an intricate phenomenon and represents the net result of the coordinated expression of hundreds of genes including sRNAs, that are subject to multiple regulatory circuits [10,79,80]. While YbeY’s exact role in the regulation of virulence-associated sRNAs and their effect on various virulence factors is unclear, YbeY evidently plays a crucial role in V. cholerae pathogenesis.

Stress adaptation is key to establishing a successful infection. YbeY was originally discovered in S. meliloti as a stress regulator crucial to establish a chronic intracellular infection required for the nitrogen-fixing symbiosis within its host plant Medicago sativa [14]. In E. coli, deletion of ybeY causes a pleiotropic stress phenotype, especially to heat [16]. Hence, YbeY was initially identified as a heat shock protein [20]. In V. cholerae, we showed that YbeY depletion drastically impairs the stress response to a wide variety of stress agents. Its remarkably pleiotropic phenotype (Figure 8A) distinguishes it from most RNase mutants studied so far, which typically show fewer phenotypic traits. However, YbeY of V. cholerae does not act like a heat shock protein. Depletion of YbeY results only in a modest decrease of cell survival after heat treatment compared to the wild-type (Figure 8B). On the other hand, unlike in E. coli, a modest cold-sensitive phenotype was found upon YbeY depletion in V. cholerae ybeY at 18°C (Figure 8C). Cold-sensitivity is a characteristic of ribosome assembly defects, since lower temperatures can stabilize incorrect ribosome intermediates [81], and is a well-established phenotype of RNase mutants lacking PNPase, RNase PH or RNase R [82,83]. Hence, cold-sensitivity is in agreement with YbeY’s key role in RNA metabolism.

YbeY is part of a core set of bacterial RNases representing an attractive antimicrobial drug target

Based on high sequence conservation, YbeY is found in virtually all sequenced bacteria. Orthologs of ybeY are much less conserved in eukaryotes. So far, no orthologs could be identified in sequenced Fungi and Archaea although they are present in some higher eukaryotes, including humans. We have demonstrated here that YbeY’s function in stress regulation and rRNA maturation is
highly conserved in bacteria as ybeY orthologs of four distantly related pathogens, S. pneumoniae, M. tuberculosis, P. aeruginosa, and V. cholerae, could fully complement an E. coli ΔybeY mutant subjected to a variety of stress agents and could correctly mature 16 S rRNA of E. coli during heat treatment (Figure 9). This is notable as conserved proteins do not always retain their specific characteristics, e.g. MazG of M. tuberculosis. This conserved NTP-pyrophosphohydrolase was shown to have a distinct function in M. tuberculosis compared to E. coli in response to starvation and oxidative stress [84].

Using a comprehensive phylogenetic analysis of major RNases in a selection of important bacterial pathogens (Figure 1), we showed that YbeY is more often essential than not and is part of a minimal core set of RNases, consistent with its remarkable level of functional conservation. While the overarching mechanisms by which organisms process RNA are conserved, our analysis also shows the disparity of RNases and their essentiality among different organisms. Thus, RNA maturation and degradation machineries necessitate the implementation of a set of RNases, which may vary from organism to organism, to fulfill all required functions. Altogether, the relatively low number of essential RNases underlines functional redundancy within groups of RNases and highlights the fundamental role of YbeY as a bacterial RNase.

Novel drug targets are crucial to combat the rapid rise of antibiotic resistance. The majority of today’s antibiotics have been developed against essential components of central cellular pathways. New potential targets worth evaluating include bacteria-specific proteases, aminoacyl-tRNA synthetases, RNA-modification enzymes, and RNases. For example, the essential process of RNA turnover via RNases is considered rich in candidate antimicrobial targets [63]. The RNase YbeY represents an important new promising drug target. We have demonstrated that YbeY in V. cholerae plays an essential role in various aspects of RNA metabolism and overall pathogenesis (Figure 10). Depletion of YbeY in V. cholerae limits growth and increases the organism’s sensitivity towards stress, making YbeY well suited for the development of target-specific antibiotics in whole-cell screening assays based on differential sensitivity of target-depleted and complemented strains. Such an approach was used successfully to discover platensimycin, a new class of broad-spectrum antibiotics for Gram-positive bacteria [85]. The availability of a suitable model host is also crucial for early antibiotic development [86]. Since depletion of YbeY in V. cholerae significantly reduces colonization of the pathogen in infant mice as well as expression of numerous virulence factors, a mouse model can be adapted for such a purpose. Although functionally associated with a well-established antibiotic target, the ribosome (Figure 10), YbeY is so far unexploited as a drug target and its use as a drug target might lead to the discovery of completely novel antibiotic scaffolds. A YbeY-specific inhibitor could also be used in combination with other antibiotics for enhanced antimicrobial activity, similar to combining silver and vancomycin [87] or rifampicin and acyldepsipeptide that activates the CbpP protease [88]. Considering YbeY’s high level of conservation, its essential nature in many pathogens and its ability to sensitize pathogens by disrupting stress tolerance and virulence, a YbeY-specific antibiotic could have broad-spectrum antimicrobial activity.

Materials and Methods

Ethics statement

The animal experiments were performed with protocols approved by Harvard Medical School Office for Research Protection Standing Committee on Animals. The Harvard Medical School animal management program is accredited by the Association for the Assessment and Accreditation of Laboratory Animal Care, International (AAALAC), and meets National Institutes of Health standards as set forth in the Guide for the Care and Use of Laboratory Animals (DHHS Publication No. (NIH) 85-23 Revised 1996). The institution also accepts as mandatory the PHS Policy on Humane Care and Use of Laboratory Animals by Awardee Institutions and NIH Principles for the Utilization and Care of Vertebrate Animals Used in Testing, Research, and Training. There is on file with the Office of Laboratory Animal Welfare (OLAW) an approved Assurance of Compliance (A3431-01).

General

DNA and RNA-DNA chimeric oligonucleotides were obtained from IDT or Eurofins MWG Operon. Sequences of oligonucleotides used in this study are summarized in Table S1.

Genome analysis of essential RNases in human pathogens

RNase genes were identified as putatively essential based on comparison of various deletion or transposon libraries in V. cholerae C6706 [36,53]; this work, E. coli MG1655 [39,40], S. typhi Ty2 [37,45], H. influenzae Rd KW20 [33,34], P. aeruginosa PA14 ([46]; http://ausubellab.mgh.harvard.edu/cgi-bin/pa14/home.cgi), B. subtilis 168 [44], S. aureus NCTC 8325 [38], S. pneumoniae TIGR4 [48], M. tuberculosis H37v [42,49,50], H. pylori G27 [47], and M. genitalium G37 [41]. RNases were designated as essential if no deletion or transposon mutant was available for the respective open reading frame. For RNase E, several libraries contained mutants with transposon insertions in the second half of the gene corresponding to the C-terminal docking domain of RNase E, but no transposon insertions in the first half of the gene which corresponds to the essential RNase domain of RNase E [51]; in these cases, RNase E was termed essential. RNases were designated as non-essential when multiple deletion or transposon mutants were available, with transposon insertions randomly distributed within the respective open reading frame. The identity of individual RNases was confirmed by BLAST searches (NCBI).

Strain construction and DNA manipulations

To generate a non-polar deletion of VC0960, we first constructed a V. cholerae C6706 strain carrying a copy of VC0960 on the arabinose inducible vector pBAD18 [89], named pY in the main text. The deletion of VC0960 (chromosomal location 1,024,609–1,024,573) was performed in the presence of arabinose as previously described [52].

The complementation plasmids expressing YbeY of various organisms were constructed by PCR amplification of each ybeY homolog from chromosomal DNA (i.e. b0659 of E. coli MG1655, VC0960 of V. cholerae C6706, PA3982 of P. aeruginosa PA01, SPR0809 of S. pneumoniae R6, and Rv2367c of M. tuberculosis H37v), and subsequently cloned into pBR322 under control of the constitutive tetracycline promoter.

DNA manipulations were performed according to the methods of Sambrook [90] and cloning protocols were sequence-verified.

Bacterial strains and growth conditions

The bacterial strains used for this work are V. cholerae wild-type C6706 [91], V. cholerae C6706 ΔybeY pY (this work), V. cholerae C6706 mc-CTD::Tn [36], Mutant ID EC2413, Genomic locus 2.183.265), V. cholerae C6706 mg::Tn [36], Mutant ID EC135315, Genomic locus 445.329), V. cholerae C6706 no::Tn [36], Mutant...
ID EC9412, Genomic locus 2.768.302, E. coli wild-type MC4100 [92], and E. coli MC4100 AybeF [16]. V. cholerae and E. coli were cultured in LB medium (1% tryptone, 0.5% yeast extract, 0.5% NaCl) at 37°C when used for RNA isolation, ribosome isolation, protein purification or stress tests. V. cholerae strains were maintained in LB medium supplemented with 0.1% arabinose (Ara+; YbeY induction) or 0.2% glucose (Gluc+; YbeY depletion) as indicated. For depletion of YbeY (Gluc+), cells were pre-cultured in LB containing arabinose, pelleted, washed twice with 0.85% saline solution, diluted into fresh LB containing glucose to an OD600 less than 0.001 and then cultured until mid exponential phase (OD600 0.5). For intermediate depletion of YbeY (indicated as Ara+/Gluc+), cells were grown in LB containing arabinose until early exponential phase (OD600 0.1), pelleted, washed twice and subsequently grown in LB containing glucose until mid exponential phase.

To analyze growth, C6706 WT pY and AybeF pY strains were cultured as described above and growth was monitored for 8–10 h until saturation. Additionally, the AybeF pY mutant strain was grown in the absence of arabinose (Ara−) for depletion of YbeY and subsequently subcultured a second time in LB medium lacking arabinose (Ara−/Ara+) to further deplete YbeY.

Antibiotics were used at the following concentrations: kanamycin (Kan: 40 μg/ml), tetracycline (Tet: 0.05 μg/ml), gentamycin (Gen: 2.0 μg/ml for V. cholerae and 0.4 μg/ml for E. coli), kasugamycin (Kas: 100 μg/ml), chloramphenicol (Cm: 0.25 μg/ml), V. cholerae and 2.5 μg/ml E. coli), crythromycin (Ery: 0.5 μg/ml), rifampicin (Rif: 0.1 μg/ml), ampicillin (Amp: 2 μg/ml), and cefotaxime (Cef: 0.0002 μg/ml for V. cholerae and 0.2 μg/ml for E. coli).

Ribosome isolation and profiles

70 S ribosomes, and 50 S and 30 S subunits from V. cholerae C6706, V. cholerae AybeF pY mutant, V. cholerae rnc::Tn mutant, V. cholerae mc-CTD::Tn mutant, V. cholerae mc::Tn mutant, E. coli MC4100, and E. coli AybeF mutant were isolated as described previously [93] with minor modifications. Briefly, log phase cultures (OD600 0.5) grown in LB at 37°C were harvested, resuspended in ice-cold buffer A (20 mM HEPES pH 7.5, 50 mM NH4Cl, 10 mM MgCl2, 5 mM β-mercaptoethanol, 0.1 mM PMSF), and lysed by French press (two passes at 11,000 psi). All subsequent steps were carried out at 4°C. Cell debris was removed by low speed centrifugation. The 30 S supernatant was obtained by two spins at 30,000 g for 30 min and was subsequently subjected to centrifugation at 100,000 g for 3 h. The final pellet containing the ribosomes was resuspended in buffer A and loaded onto a 10–40% sucrose gradient made in buffer A. After centrifugation at 150,000 g for 7 h, the 70 S ribosomes and ribosomal subunits were collected by monitoring the fractions spectro-photometrically (λ260).

Isolation and analysis of RNA

Total RNA from log phase cultures (OD600 0.5) grown in LB at 37°C and rRNA from purified ribosome fractions were extracted using TRIzol (Ambion) according to the manufacturer’s protocol. Samples used for RT-qPCR and sRNA analysis were treated with TURBO DNase (Ambion) to remove any DNA contamination. Total rRNA profiles were analyzed by Synergy/Gagarose gel electrophoresis as described [94].

cDNA synthesis and RT-qPCR
cDNA was synthesized using the iScript cDNA Synthesis Kit (BioRad) according to the manufacturer’s protocol, and 1 μg of total RNA per 20 μl reaction. Relative expression levels were determined by RT-qPCR using the LightCycler 480 Real-time PCR system. Primers were designed using Primer Express 3.0 (optimal primer length of 20 bases, optimal amplicon length of 100 bases, GC-content of 50% and Tm of 50–60°C; see Table S1). 10 ng of cDNA and 0.2 μM of each primer was mixed with LightCycler 480 Cybr Green I master mix in a 20 μl reaction.

PCR conditions were: a pre-incubation of 5 min at 95°C, an amplification stage of 45 cycles of 20 s at 95°C, 30 s at 57°C and 20 s at 72°C, and a melting curve stage of 5 s at 95°C, 1 min at 65°C increased to 97°C with steps of 2.2°C/s. All reactions were performed in triplicate and samples without reverse transcriptase were used as a control to assess genomic DNA contamination. The raw data was analyzed using LightCycler 480 Software (1.5.0 SP2), normalized against 5 S rRNA and the wild-type strain C6706 (without the pY maintenance plasmid; without additional carbon source) was used as calibrator condition. Relative gene expression was calculated using the Livak method [95].

Northern blot analysis of RNA

RNA was separated by denaturing polyacrylamide gel electrophoresis (6% polyacrylamide/7 M urea). The transfer of RNA onto Nytran membrane and Northern blot hybridization were as described in [96] with minor modifications. DNA oligonucleotides were end labeled with 32P using T4 polynucleotide kinase (New England Biolabs). Prehybridizations were carried out at 42°C and hybridizations were carried out at 40°C–45°C. Following hybridization, membranes were washed at room temperature with 6×–2× SSC. Autoradiograms were analyzed by phosphorimager using a Typhoon scanner and ImageQuant software.

Mapping of 5′ and 3′ termini of rRNA

To map the 5′ termini of rRNA, primer extension assays were performed using Superscript II reverse transcriptase (Invitrogen). Typically, a 10 μl reaction contained 100 ng of total RNA or 25–50 ng of rRNA isolated from ribosomes, 100,000 cpm of the 32P-labeled DNA oligonucleotide and 2 μl of the 5′ reaction buffer (Invitrogen). After denaturation (65°C, 5 min), samples were chilled on ice for 5 min and then brought to room temperature for 5 min. 0.5 μl of DTT solution (Invitrogen), 0.25 μl of a 10 mM dNTP solution and 50 μl of Superscript II (Invitrogen) were added. The reverse transcription reaction was carried out at 45°C for 20 min. The reaction was stopped by addition of loading dye containing formamide and heat denaturation. Reaction products were separated on denaturing polyacrylamide gels (6–10% polyacrylamide/7 M urea).

The 3′ termini of rRNA was mapped by site-specific RNase H cleavage assay as described by Deutscher and colleagues [29,97,98] with minor modifications, followed by Northern hybridization using probes specific for the mature 3′ termini of 16 S rRNA, 23 S rRNA or 5 S rRNA, respectively. Briefly, 100 ng of total RNA or 50 ng of rRNA isolated from ribosomes were mixed with a chimeric RNA/DNA oligonucleotide and 1 μl of 10× RNase H buffer (New England Biolabs) in a 10 μl reaction. After denaturation (95°C, 1 min), samples were brought to 65°C for 5 min, then 55°C for 15 min, 35°C for 15 min and then to room temperature for another 15 min to allow for formation of the DNA:RNA hybrid. 2.5 U of RNase H (New England Biolabs) was added. Samples were incubated for 60 min at 37°C. The reaction was stopped by addition of loading dye containing formamide and heat denaturation. Reaction products were separated on denaturing polyacrylamide gels (6–10% polyacrylamide/7 M urea) and analyzed by Northern blot hybridization.
Overexpression and purification of YbeY and RNase R

To construct the overexpression plasmid, ybeY of E. coli (b0659) and V. cholerae (VC0960) was cloned into pET28a under control of a T7 promoter with an N-terminal His-MBP-TEV (site)-tag. Purification of YbeY without RNase R contamination was accomplished by expression in BL21 (DE3) Arna [18]. To improve activity and yield, pET28a_YbeY was co-transformed with pGro7, a chaperone plasmid that encodes both GroES and GroEL under an arabinosin inducible promoter [99]. MBP-YbeY was purified using amylose resin (New England Biolabs), the MBP-tag was cleaved by Turbo TEV protease (Eton Bioscience Inc.) and removed by size exclusion chromatography as described previously [18]. The purified YbeY protein was subjected to mass spectrometry to assess its purity.

For generating RNase R, VC0960 of V. cholerae was cloned into pET15b under control of a T7 promoter with an N-terminal His-tag. After overexpression in BL21 (DE3) Arna [18], RNase R was purified by affinity chromatography using Talon resin (Clontech) following the manufacturer’s protocol for batch-gravity-flow purification of proteins.

In vitro RNase assays

All YbeY RNase assays were carried out as described previously in 50 mM HEPES pH 7.5 in a 20 μl volume [18]. Total RNA (250 ng) was incubated with E. coli or V. cholerae YbeY (5–10 μM) for 1 h at 37°C. Digestion products were separated on Syngergel agarose gels. Short synthetic RNA substrates were labeled at their 5’ terminus using γ-32P-ATP and T4 polynucleotide kinase (New England Biolabs). For each reaction, 0.03 μM short RNA substrate was incubated with V. cholerae or E. coli YbeY (1-5 μM) for 1 h at 37°C. Products were separated on a 10% denaturing polyacrylamide gel containing 7 M urea.

70 S ribosomes (0.05 μM) isolated from V. cholerae ΔybeY pY (grown under maximum YbeY depletion) and V. cholerae C6706 Wt pY pY (grown in the presence of arabinose) were incubated with YbeY and RNase R of V. cholerae (0.1 μM) or E. coli (0.05 μM), either alone or together in 50 mM HEPES pH 7.5 and 10 mM MgCl2 for 1 h at 37°C. Ribosomal RNA was subsequently isolated using TRIzol, separated by Syngergel/agarose gel electrophoresis and stained with ethidium bromide.

Mouse infections

Infant mice were inoculated with V. cholerae ΔybeY pY cells grown in the absence or presence of 0.2% arabinose. We assessed their ability to colonize the mouse intestine in competition with the parental strain V. cholerae C6706 as previously described [100].

Western blot analysis of Cholera toxin

The levels of cholera toxin produced by V. cholerae C6706 Wt pY and ΔybeY pY mutant were analyzed by western blotting. Supernatants of cultures (10 mL) grown at 37°C in LB medium in the presence of 0.1% arabinose or 0.2% glucose as indicated, were collected after 8 h by centrifugation and subsequently passed through a 0.2 μm cellulose acetate filter (VWR). The total protein content of the samples was estimated by Bradford using BSA as reference. 5 μg of supernatant protein mixed with 5 μg of BSA were applied to a 12% SDS-PAGE and transferred onto an Immobilon-P membrane (PVDF) following standard procedures [87]. Non-specific sites were blocked using 5% milk powder in TBS-T (0.05% Tween20 in 20 mM Tris-HCl, 150 mM NaCl, pH 7.5) by incubation at room temperature for 4 h. The primary Rabbit polyclonal anti-Cholera toxin antibody (Sigma-Aldrich) was dissolved in 5% milk powder/TBS-T at a concentration of 0.05 μg/ml and incubated with the membrane overnight at 4°C. After 5 washes with TBS-T for 5 min, the secondary antibody (Anti-Rabbit IgG HRP; Thermo Scientific) was dissolved in 5% milk/TBS-T (0.002 μg/ml) and incubated with the membrane for 2 h at room temperature. After 5 washes with TBS-T for 5 min, 1 ml of SuperSignal West Dura Chemiluminescent Substrate (Thermo Scientific) was added for 5 min before exposing the X-ray film.

Stress tests and complementation assays

Stress sensitivity of the V. cholerae ΔybeY pY mutant compared to the C6706 Wt pY was determined by spotting serially diluted (1:10, starting at OD600 0.1) cultures, grown overnight in LB medium and subsequently washed with 0.85% saline solution, onto LB plates containing various antibiotics or 0.1 mM H2O2 along with either 0.1% arabinose or 0.2% glucose as indicated. For UV survival, cultures of the V. cholerae strains were spotted onto LB plates and irradiated with a UV dose of 20 J/m2 using a G15T8 UV lamp (GE) at 254 nm, then incubated in the dark. To determine heat sensitivity of C6706 Wt pY and ΔybeY pY, cells from an early exponential culture supplemented with 0.2% glucose were incubated for 3 h at 37°C and 45°C before spotting as a dilution series onto LB plates supplemented with 0.2% glucose; plates were subsequently incubated for 18 h at 37°C. Cold sensitivity of C6706 Wt pY and ΔybeY pY mutant cells was resolved by spotting serially diluted cultures of a late exponential culture, grown in the presence of 0.1% arabinose, onto LB plates containing 0.2% glucose; plates were incubated for 2 days at 37°C and 18°C as indicated.

Similarly, complementation of the E. coli ΔybeY mutant by ybeY of various pathogens was determined by spotting serially diluted cultures (1:10, starting at OD600 0.1), grown overnight in LB medium, onto LB plates containing various antibiotics or 0.75 mM H2O2. A higher UV dose of 60 J/m2 was used to measure UV resistance of the E. coli strains. Heat sensitivity of E. coli cells was determined by spotting serially diluted cultures of a late exponential culture onto LB plates; plates were incubated for 18 h at 45°C as indicated. The complementation vectors constitutively express ybeY of E. coli MC4100 (pY_Ec), V. cholerae N16961 (pY_Vc), P. aeruginosa H37Rv (pY_Pa), M. tuberculosis H37v (pY Mt), or S. pneumoniae R6 (pY Sp).

Bile-salt sensitivity of C6706 Wt pY and ΔybeY pY mutant was determined by growth in LB medium supplemented with 2.0% bile salt and 0.1% arabinose as specified. The number of colony forming units (CFU) was determined after 0, 1, 2, and 3 h of bile-salt treatment.

Pigment formation of C6706 Wt pY and ΔybeY pY mutant was determined by spinning down a similar number of cells grown to mid-log phase in LB medium containing 0.1% arabinose or 0.2% glucose as indicated.

Supporting Information

Figure S1 Growth analysis of V. cholerae ΔybeY. Growth curve of C6706 Wt pY and the ΔybeY pY strain in LB medium at 37°C. The Wt and mutant strains were grown in medium supplemented with arabinose (Ara+). The ΔybeY pY strain was diluted into medium lacking arabinose (Ara-) for depletion of YbeY, grown to saturation and then subcultured a second time into fresh medium lacking arabinose (Ara-/Ara-).

Figure S2 Analysis of 23 S rRNA and 5 S rRNA in V. cholerae ΔybeY. Mapping of 5’ and 3’ termini of A) 23S RNA...
and B) 5 S rRNA from C6706 Wt pY and the ΔybeY pY strain. “P” and “M” specify the positions of bands derived from the precursor and mature forms of 23 S rRNA and 5 S rRNA. “pY” indicates that ybeY is expressed from a plasmid. Ara+, cells were grown in LB in the presence of arabinose. Gluc+, cells were grown in LB in the presence of glucose. Ara+/Gluc+, intermediate YbeY depletion by switching the carbon source of the ΔybeY pY strain in early exponential phase from arabinose to glucose (for details see Materials and Methods).

Figure S3 Analysis of V. cholerae YbeY’s in vitro RNase activity. A) Total RNA (250 ng) isolated from V. cholerae can be degraded by purified YbeY of V. cholerae (10 μM), similar to E. coli YbeY (5 μM), EDTA (50 mM) inhibits YbeY’s RNase activity. RNA was analyzed by agarose gel electrophoresis. The positions of the 23 S and 16 S rRNA are indicated. B–C) Purified YbeY of V. cholerae (1–5 μM) shows endoribonuclease activity on 32P-labeled oligoribonucleotides (0.03 μM); hairpin substrate in B) and ssRNA in C). Digestion products were analyzed by polyacrylamide gel electrophoresis. D) In vitro ribosome quality control by V. cholerae (Vc) and E. coli (Ec) YbeY together with RNase R. 70 S ribosomes of C6706 Wt pY grown in the presence of arabinose (Ara+) were incubated with YbeY, RNase R, or a mixture of YbeY and RNase R from E. coli or V. cholerae as indicated. The positions of 23 S and 16 S rRNAs are indicated based on their mobility.

Figure S4 Ribosome profiles of V. cholerae mutant strains with transposon insertions in genes encoding RNase E (rne::CTD::Tn), RNase G (rng::Tn) or RNase R (rnr::Tn). Analysis of ribosome profiles in C6706 Wt pY (W), ΔybeY pY (Δ, rne::CTD::Tn: E), rng::Tn (G) and rnr::Tn (R). Wt pY and ΔybeY pY cells were first grown in LB medium supplemented with arabinose and then subcultured into glucose-containing medium for depletion of YbeY; all other strains were grown in LB medium without additional carbon source. Polysomes, 70 S, 50 S and 30 S ribosomes are indicated.

Figure S5 YbeY is highly conserved among bacteria. Sequence alignment of YbeY proteins from E. coli MG1655 (Ec), V. cholerae N16961 (Vc), P. aeruginosa PAO1 (Pa), M. tuberculosis H37Rv (Mt), and S. pneumoniae R6 (Sp). The highly conserved H3XH5X motive in the catalytic pocket is indicated.

Table S1 Oligonucleotides used in this study.

Acknowledgments

We thank Richard Goldstein for his generous gift of S. pneumoniae DNA and Deborah Hung for her generous gift of M. tuberculosis DNA. We also thank all members of the Walker lab for discussion and input.

Author Contributions

Conceived and designed the experiments: MV CK BWD MFFA JJM ULR GCW. Performed the experiments: MV CK BWD MFFA. Analyzed the data: MV CK BWD MFFA. Contributed reagents/materials/analysis tools: MV CK BWD MFFA JJM ULR GCW. Wrote the paper: MV CK ULR GCW.

References

Haemophilus influenzae Bacillus subtilis essential gene set with hidden Markov virulence factors. Infect Escherichia coli MazG is a in vitro


genes using in vivo is essential for growth at low temperature and plays a novel role in the 3'


10: 291.


PLOS Pathogens | www.plospathogens.org 19 June 2014 | Volume 10 | Issue 6 | e1004175