Coding of Electric Pulse Trains Presented through Cochlear Implants in the Auditory Midbrain of Awake Rabbit: Comparison with Anesthetized Preparations

The MIT Faculty has made this article openly available. Please share how this access benefits you. Your story matters.

Citation

As Published
http://dx.doi.org/10.1523/jneurosci.2084-13.2014

Publisher
Society for Neuroscience

Version
Final published version

Accessed
Sun Dec 30 17:44:49 EST 2018

Citable Link
http://hdl.handle.net/1721.1/91535

Terms of Use
Article is made available in accordance with the publisher's policy and may be subject to US copyright law. Please refer to the publisher's site for terms of use.

Detailed Terms
Coding of Electric Pulse Trains Presented through Cochlear Implants in the Auditory Midbrain of Awake Rabbit: Comparison with Anesthetized Preparations

Yoojin Chung,1,3 Kenneth E. Hancock,1,2 Sung-Il Nam,1,3 and Bertrand Delgutte1,2,4
1Eaton-Peabody Laboratories, Massachusetts Eye and Ear Infirmary, Boston, Massachusetts 02114, 2Department of Otology and Laryngology, Harvard Medical School, Boston, Massachusetts 02115, 3Department of Otolaryngology, School of Medicine, Keimyung University, Daegu, South Korea 700-712, and 4Research Laboratory of Electronics, Massachusetts Institute of Technology, Cambridge, Massachusetts 02139

Cochlear implant (CI) listeners show limits at high frequencies in tasks involving temporal processing such as rate pitch and interaural time difference discrimination. Similar limits have been observed in neural responses to electric stimulation in animals with CI; however, the upper limit of temporal coding of electric pulse train stimuli in the inferior colliculus (IC) of anesthetized animals is lower than the perceptual limit. We hypothesize that the upper limit of temporal neural coding has been underestimated in previous studies due to the confound of anesthesia. To test this hypothesis, we developed a chronic, awake rabbit preparation for single-unit studies of IC neurons with electric stimulation through CI. Stimuli were periodic trains of biphasic pulses with rates varying from 20 to 1280 pulses per second. We found that IC neurons in awake rabbits showed higher spontaneous activity and greater sustained responses, both excitatory and inhibitory, at high pulse rates. Maximum pulse rates that elicited synchronized responses were approximately two times higher in awake rabbits than in earlier studies with anesthetized animals. Here, we demonstrate directly that anesthesia is a major factor underlying these differences by monitoring the responses of single units in one rabbit before and after injection of an ultra-short-acting barbiturate. In general, the physiological rate limits of IC neurons in the awake rabbit are more consistent with the psychophysical limits in human CI subjects compared with limits from anesthetized animals.

Key words: anesthesia; cochlear implant; inferior colliculus; temporal coding

Introduction

Although cochlear implants (CIs) restore open-set speech comprehension in a majority of deaf subjects, most CI users still have difficulty with pitch perception, speech reception in noise, and sound localization even with bilateral implantation. These difficulties may stem in part from limitations in temporal processing. A majority of CI users can discriminate the rate of electric pulse trains up to ~300 pulses per second (pps), with a few exceptional subjects showing limits as high as 1000 pps (Townshend et al., 1987; Kong and Carlyon, 2010; Moore and Carlyon, 2010). Responses of auditory neurons to pulse trains delivered through CIs also show an upper limit of temporal coding, but the physiological limits in anesthetized animals are lower than the perceptual limits in human CI users. For the most part, neurons in the inferior colliculus (IC) show sustained and pulse-locked responses to periodic pulse trains only up to 30–300 pps (Snyder et al., 1995; Vollmer et al., 1999, 2005; Middlebrooks and Snyder, 2010; Hancock et al., 2012, 2013). A similar pulse rate limit of 10–200 pps is observed for neural sensitivity to interaural time differences (ITDs) in IC neurons (Smith and Delgutte, 2007; Hancock et al., 2010, 2012), but this limit is lower than that the 250–600 pps limit for perceptual ITD sensitivity in CI listeners (Laback et al., 2007; van Hoesel, 2007). In anesthetized animals, the underlying reason for the limits on temporal coding and ITD sensitivity is that the majority of IC neurons only respond to the onset of pulse trains at rates >100–200 pps. This lack of sustained responses in IC of anesthetized animals is inconsistent with percepts lasting throughout the stimulus even at much higher pulse rates in most CI users, although some subjects experience loudness adaptation (Tang et al., 2006).

We hypothesize that temporal coding, ITD sensitivity, and sustained firing to CI stimulation at high pulse rates may be underestimated due to the effect of anesthesia. Barbiturate anesthesia has been shown to reduce both spontaneous and evoked neural activity in the IC of normal-hearing animals (Bock and Webster, 1974; Kuwada et al., 1989; Torterolo et al., 2002), which is consistent with an enhancement of inhibitory influences. Similar differences between anesthetized and awake conditions have been observed in primary cortex of deaf marmosets and guinea
pigs with CI stimulation (Johnson et al., 2011; Kirby and Middlebrooks, 2012). A degradation of temporal coding by anesthesia has been reported in the IC (Tollin et al., 2004; Song et al., 2011) and auditory cortex (Ter-Mikaelian et al., 2007) of normal-hearing animals.

We developed an awake rabbit model of CIs for single-unit recording from the IC to eliminate the confound of anesthesia. Although awake preparations are increasingly common in single-unit studies of normal-hearing animals, they have not yet been used to study responses of subcortical auditory neurons to CI stimulation in deaf animals. Our results show higher pulse rate limits of temporal coding in the awake rabbit compared with earlier results from anesthetized preparations. We also found unsynchronized, sustained responses at high pulse rates that are rarely observed in the IC of anesthetized animals with CI stimulation. We demonstrate that these differences are largely due to the effect of anesthesia by monitoring the responses of single units in one rabbit before and after injection of an ultra-short-acting barbiturate.

Materials and Methods

Animals

We measured responses of single units in the IC of awake rabbits to electric stimulation presented through CIs. Experiments were performed on four female Dutch-belted rabbits, two of which received unilateral cochlear implantation in the right ear and the other two bilateral cochlear implantations. All procedures were approved by the animal care and use committees of both Massachusetts Eye and Ear and the Massachusetts Institute of Technology. A preliminary report of some of these findings has been published previously (Chung et al., 2013).

Surgical procedures

Methods for recording from single units in the IC of awake rabbits were as described in previous reports with normal-hearing animals (Devore and Delgutte, 2010; Day et al., 2012). Methods for chronic cochlear implantation in adult rabbits were newly developed. Anesthesia for all survival surgeries was induced with xylazine (5–6 mg/kg, s.c.) and ketamine (35–44 mg/kg, i.m.). For prolonged surgeries such as cochlear implantation, a surgical level of anesthesia was maintained by isoflurane mixed with oxygen (2.5%) and delivered via a facemask (0.8 l/min).

Each rabbit underwent three surgical procedures separated by recovery and training periods: a first one to affix a headpost and stainless steel cylinder to the skull, a second one to implant intracochlear electrode arrays, and a third one to perform a craniotomy for accessing the IC. In the first surgery, a stainless steel cylinder and brass head bar were affixed to the skull using stainless steel screws and dental acrylic (Jet Denture Repair; Lang Dental). After the rabbit was fully recovered, surgery for cochlear implantation was performed. A postauricular incision was made to access the tympanic bulla and then part of the bulla was drilled to expose the round window. An eight-contact ring-type electrode array (Zeo0274; Cochlear Ltd) was inserted into the cochlea through the round window. Before inserting the electrode array, 5–10 μl of distilled water was injected into the cochlea through the round window to deafer the ear by causing hypotonic stress to the organ of Corti (Ebert et al., 2004). The round window was sealed with soft tissue harvested from nearby muscle and secured with dental cement (Durelon; 3M). The percutaneous connector terminating the lead wire of the electrode array was secured to the side of the headpost with dental acrylic. The superficial muscles and the skin incision were closed with sutures that were removed 10–14 d after the operation. Success of cochlear implantation was verified immediately after surgery by measuring the electrically evoked auditory brainstem response (EABR) to biphasic current pulses delivered between the most apical and the most basal electrodes in the intracochlear array. EABR thresholds were usually in the range of 16–212 dB re 1 mA. The ABR to 100 μs acoustic clicks at 100 dB peak SPL was also measured to verify the effectiveness of deafening in the implanted ears. No attempt was made to deafer the unimplanted left ear in the two unilaterally implanted rabbits.

After an ~1 week recovery from cochlear implantation surgery, rabbits were habituated to the experimental setup until they could sit quietly for 2–3 h with the head attached by the headpost while receiving electric stimulation through the cochlear implant. After habitation, the animals underwent a third aseptic procedure to make a small (~2 mm diameter) craniotomy 0.5 mm posterior and 3 mm lateral to bregma. In the two unilaterally implanted rabbits, the craniotomy was made contralateral to the implanted right ear. The exposed area was covered with a topical antibiotic (Bacitracin) and sealed with dental impression material (Reprosil). Single-unit recording sessions started after 2–3 d of recovery and lasted until 26–431 d after implantation. Rabbts were monitored on a closed-circuit video system throughout the recording sessions and only received auditory stimulation through their cochlear implant during recording sessions.

Electrophysiological methods

Stimuli

The main stimuli were 300 ms periodic trains of biphasic pulses (50 μs/phase) presented every 600 ms. Pulse trains were generated using 16-bit D/A converters (PXI-6221; National Instruments) and delivered to each cochlear implant through a pair of custom-built, high-bandwidth, isolated current sources. Stimulation was between the most apical and most basal intracochlear electrodes. This wide bipolar electrode configuration stimulates auditory nerve fibers over the entire length of the tonotopic axis while reducing stimulus artifact relative to monopolar stimulation (Litvak et al., 2001). Stimuli were presented dichotically in the bilaterally implanted rabbits and monaurally to the ear contralateral to the recorded IC in the unilaterally implanted rabbits.

The stimulus used for searching for single units in bilaterally implanted rabbits was a sequence of three biphasic pulses presented alternately to both ears (dichotically), the left ear, and the right ear. The interval between consecutive pulses in the triplet was 100 ms and the silent interval between triplets was 200 ms. In unilaterally implanted rabbits, the search stimulus was a pair of pulses separated by 100 ms with a 300 ms silent interval between pairs.

Single unit recordings

Single units were isolated using epoxy-insulated tungsten electrodes (A-M Systems) advanced with a remote-controlled microdrive. The electrode was advanced in a dorsoventral direction through theoccipital cortex up to the IC, which was identified by background entrainment to the search stimulus. Signals from the recording electrode were preamplified (Axoprobe 1A; Molecular Devices), bandpass filtered (1–3 kHz), and sampled at 100 kHz using a high-speed, 12-bit A/D converter (PXI-6123; National Instruments). The stimulus artifact was removed by gating out a short interval (0.3–0.5 ms) after each stimulus pulse. Spike times were detected by software triggering on level crossings. Only well isolated single units were studied, as assessed by the stability of spike waveforms and amplitudes, which were always at least three times above the noise floor and clearly distinct from the local field potential. Representative spike waveforms recorded from IC neurons are shown in Figure 1, A–C, for pulse-train stimulation. These examples include a range of spike quality, from excellent (Fig. 1C) to cases that are clearly single units but include a small fraction (<1%) of false triggers (Fig. 1A,B).

When a single unit was isolated, the threshold for a single biphasic pulse was determined by measuring responses to the search stimulus from just below threshold to at least 2 dB above threshold in 1 or 2 dB steps. Then responses to 300 ms pulse trains were measured for pulse rates ranging from 20 to 1280 pps in half-octave steps. Pulse rates were presented in random order and each rate was repeated 12–20 times. Pulse trains were presented at 2–4 dB above the single-pulse threshold.

Histological processing

In the last recording session from one unilaterally implanted rabbit, electrolytic lesions were made to mark the borders of the region showing activity evoked by CI stimulation while the animal was under deep anesthesia (xylazine 6 mg/kg, s.c., and ketamine 44 mg/kg, i.m.). The rabbit was then perfused intracardially using a solution of 1.5% paraformaldehyde and 2.5% gluteraldehyde in phosphate buffer. The brain was immersed in fixative for 24 h and then transferred to 25% sucrose solution for several days. Coronal sections (80 μm) were cut with
a freezing microtome and mounted on subdued slides. Borders of IC regions were identified by staining cell bodies with azure-thionin. All identified lesions were located within the central nucleus.

Injection of ultra-shorting-acting barbiturate
To assess the effect of anesthesia in individual neurons directly, a catheter was surgically inserted into the right jugular vein of one unilaterally implanted rabbit to allow the administration of an ultra-short-acting barbiturate while recording from single units (Kuwada et al., 1989). A median ventral neck incision was made to expose the bifurcation of the right jugular vein into the anterior and posterior facial veins. A silicon catheter (2 Fr/outer diameter 0.67 mm, 20 cm) filled with diluted heparin (10 U/ml) was inserted at ~6 cm, approximately to the level of the superior vena cava, into the posterior facial vein just above the bifurcation point. The catheter was secured to the vein with ligatures and led subdermally to an injection port on the nape of the neck. The catheter was flushed with diluted heparin (10 U/ml) every day and after each use to keep it patent (Kaplan and Timmons, 1979; Kuwada et al., 1989).

In recording sessions from this rabbit, the responses of a single unit to pulse trains were first characterized in the awake condition as a function of pulse rate using 12 repetitions as described above. Following this characterization, 5 mg/kg methohexital sodium (Brevital) was injected through the intravenous catheter over 10 s. The unit responses to pulse train stimuli were then monitored for 10–30 min after the injection, yielding 80–240 repetitions of responses to 12 different pulse rates. Up to two injections separated by at least 30 min were given over a 2 h recording session. A total of 13 injections were made in this rabbit while successfully maintaining contact with a single unit.

Single unit recordings from anesthetized cats
Because direct data on the effects of anesthesia were only available from a small number of neurons, we reanalyzed data from the IC of anesthetized cats that have been described in part previously (Hancock et al., 2010, 2012, 2013) to allow quantitative comparison with the awake rabbit data at the population level. These data came from six adult cats that were deafened with ototoxic drugs and received cochlear implants bilaterally at the beginning of the neurophysiological experiment.

In the first of two procedures, adult cats were deafened by injection of kanamycin (300 mg/kg, s.c.) followed after 30 min by ethacrynic acid (25 mg/kg, i.v.; Xu et al., 1993) while under ketamine anesthesia (33 mg/kg, i.m.). The effectiveness of deafening was verified by the absence of ABR response to 100 μs acoustic clicks up to 100 dB peak SPL (Hancock et al., 2010). The neurophysiological experiment took place either 1–2 weeks (n = 3) or 6 months (n = 3) after deafening. Cats were anesthetized with a combination of urethane (300 mg/kg, i.p.) and either diallyl barbituric acid (75 mg/kg, i.p.) or sodium pentobarbital (37 mg/kg, i.p.). The lateral and dorsal aspects of the skull were exposed. Tympanic bullae were opened bilaterally to allow access to the round window. A small cochleostomy was made and eight-contact electrode arrays (260274; Cochlear Ltd) were inserted bilaterally. An opening was made in the skull and the dorsal surface of the IC was exposed by aspirating the overlying cerebral cortex and removing part of the bony tentorium.

Aside from the use of anesthesia in cat, all stimulation protocols, recording procedures, and methods for data analysis were essentially the same in cat and in rabbit except for minor differences. Single-unit recordings in cat were made using 16-site multichannel electrodes (Neuropixel) advanced dorsally to ventrally into the exposed IC. Signals were amplified (RA16; TDT) and then filtered (300–3000 Hz) and amplified using a digital signal processor (Medusa; TDT). The recording was preamplified (RA16; TDT) and then filtered (300–3000 Hz) and amplified using a digital signal processor (Medusa; TDT). The recording was typically made differentially between adjacent electrodes in the array to minimize the amplitudes of stimulus artifacts and local field potentials. Criteria for defining single units were the same as in rabbit. Figure 1 D shows spike waveforms from an example neuron in anesthetized cat.

Data analysis
Spontaneous activity. In some units, spontaneous activity was recorded for 30 s without any stimulation. In cases when the spontaneous activity was not measured directly, we used responses to the search stimulus to estimate the spontaneous firing rate. Specifically, we counted the spikes occurring at subthreshold levels when available. If too few subthreshold responses were available, spikes were counted over the last 100 ms of the 200–300 ms silent interval between consecutive search stimuli and averaged over all tested levels.

Dependence of average firing rate on pulse rate. To compute the “sustained” firing rate in response to the 300 ms pulse trains, the onset response was removed by excluding the smallest integer number of
stimulus periods having a total duration of at least 30 ms. The firing rate was averaged over the remaining stimulus duration with the onset window excluded. In awake rabbits, both background activity and rebound responses after stimulus offset were observed frequently. To minimize the effect of rebound responses, the spike rate during the off period was calculated over a 200 ms window extending from 100 ms after stimulus offset to the onset of the next stimulus.

To test for stimulus-driven activity (excitatory or suppressive) for each pulse rate, we used receiver operating characteristic analysis to compare the sustained firing rates during the on and off periods on a trial-by-trial basis. A response was considered to be significantly excitatory if the firing rate during the on period exceeded the rate during the subsequent off period in at least 75% of stimulus trials. Conversely, the response was considered to be suppressive if the rate during the off period exceeded the rate during the on period in at least 75% of stimulus trials.

A spike will be undetected if the portion of its waveform exceeding the user-set threshold occurs entirely during the 0.3–0.5 ms stimulus artifact gating window. These missed spikes result in firing rates being underestimated, especially for high pulse rates, when the gating window covers a substantial fraction of the stimulus period. This underestimation of firing rates during the on period could lead to responses being incorrectly identified as suppressive when they are compared with the off period response, in which there is no artifact gating. To avoid this problem, all apparently suppressive responses were corrected by recomputing the on period firing rate using a stimulus duration excluding the gating windows. This correction assumes that spikes are equally likely to occur within and outside of the gating windows. No correction was applied to pulse-locked excitatory responses in order overestimating a firing rate in this case this assumption does not hold. This method ensures a conservative identification of both excitatory and suppressive responses.

**Cluster analysis of on and off period responses.** We observed considerable variability between neurons in both the dependence of responses on pulse rate and on the relative magnitudes of responses during and after the stimulus. To characterize this variability objectively, we used k-means clustering (MATLAB; MathWorks) to categorize neurons based on the joint pulse-rate dependence of on and off responses. For each neuron, the sustained firing rate versus pulse rate curves during the on and off periods were concatenated into a single vector and then normalized by the maximum value over the entire vector. The clustering algorithm iteratively partitions the set of vectors (one for each neuron) into a prespecified number of clusters to minimize the sum of the Euclidean distances of each vector to the cluster centroid. The number of clusters was chosen by monitoring the summed Euclidean distance between neural responses and the cluster centroid as a function of the number of clusters. **Pulse-locked responses.** Two different methods were used to characterize the synchronization of spikes to the stimulus pulses and to determine a upper rate limit of pulse locking for each unit. A method based on cross-correlation between stimulus and response (Hancock et al., 2013) and a more standard method based on vector strength, a.k.a. synchronization index (Vollmer et al., 2005; Middlebrooks and Snyder, 2010). The two methods differ in that the cross-correlation method requires spikes to occur within a relatively narrow latency range after each pulse to be considered pulse locked, whereas all spikes contribute to the vector strength regardless of when they occur over the stimulus period.

The following description of the cross-correlation analysis supersedes that in our earlier report (Hancock et al., 2013), which was oversimplified. For each pulse rate, the stimulus pulse train was cross-correlated with the spike train (excluding the onset response) and a histogram constructed using a 0.1 ms bin width. When properly normalized, this cross-correlogram represents the mean instantaneous firing rate after each stimulus pulse. A perfectly pulse-locked response would yield a single peak at the spike latency in the cross-correlogram. To assess the statistical significance of cross-correlogram peaks, we computed cross-correlograms for random spike trains containing the same number of spikes as the data, but uniformly distributed over the sustained response window excluding the artifact gating windows. This computation was repeated for 5000 random spike trains and a confidence bound was defined as the 99.5th percentile (p = 0.005) of these synthetic cross-correlograms (Fig. 3A,B, gray shading). A correlogram peak was regarded as significant when the average of two consecutive bins exceeded the confidence bound. The height of the largest peak in the cross-correlogram (after averaging consecutive bins) relative to the 99.5% confidence bound was used as a measure of the strength of pulse locking. The pulse-locking limit was computed by interpolating to find the pulse rate where the peak height intercepted the confidence bound (Fig. 3C,D). Spike latency was defined as the latency of the highest significant peak in the cross-correlogram. The latencies were averaged over all pulse rates up to 80 pps to get a single latency for each unit.

The cross-correlograms were also used to separate pulse-locked spikes from unsynchronized spikes and to define a pulse-locked firing rate for each pulse rate. For this purpose, we used the area of each significant cross-correlogram peak lying above the average firing rate (Fig. 3A,B, white filled areas). Some units fired at multiple latencies within an inter-pulse interval and therefore showed multiple peaks in their cross-correlogram. In such cases, the peak areas were summed over all significant peaks. The summed peak areas were converted to units of firing rate (spikes/s) for comparison with the overall firing rate.

**Results** We recorded from 136 single units in the IC of awake rabbit, 94 units from two unilaterally deafened and implanted rabbits and 42 from two bilaterally deafened and implanted rabbits. The single-unit recordings from unilaterally implanted rabbits were made 29–431 d after implantation, whereas the recordings from bilaterally implanted rabbits were made 26–172 d after implantation. Unless otherwise noted, responses to contralateral stimulation in unilaterally implanted rabbits and to diotic stimulation in bilaterally implanted rabbits were combined because the patterns of response were similar.

**Dependence of firing rates on pulse rate** Responses to periodic electric pulse trains were measured as a function of pulse rate in all neurons. We observed wide variability in both the pulse-rate dependence of responses and the relationship between responses during the on and off periods. Responses of three example units from awake rabbit and one unit from anesthetized cat are shown in Figure 1. The first neuron from awake rabbit (Fig. 1A) illustrates the suppressive responses (except for an onset response) that are commonly observed in this preparation. A very high level of background activity (~70 spikes/s) is observed during the interstimulus period; this activity is suppressed during presentation of the pulse train, with the greatest suppression occurring at 160 pps. In contrast, the second neuron from awake rabbit (Fig. 1B) shows excitatory responses for all pulse rates and lower background activity. The neuron fires approximately one spike per pulse up to 160 pps in a pulse-locked manner, as indicated by periodic patterns in the dot raster. Pulse-
locked firing at low rates transitions to unsynchronized firing at high rates, and sustained responses are observed up to the highest rate tested (1280 pps). The background activity is suppressed after each stimulus, especially at intermediate pulse rates that evoke the largest sustained response. Although the types of responses illustrated in Figure 1, A and B, are primarily observed in awake rabbit, responses such as those of the rabbit neuron in Figure 1C are observed in both awake and anesthetized preparations (Hancock et al., 2012). This neuron fires approximately two spikes per pulse at 20 pps; firing efficiency starts to decrease from 56 pps up and only a response to the first pulse is observed for rates >320 pps, resulting in a “band-pass” pattern for the sustained response. Finally, the pattern illustrated by the neuron from anesthetized cat in Figure 1D is rarely observed in awake rabbit. This neuron fires one spike per pulse at 20 pps, but firing efficiency starts to decrease from 40 pps up and there is only a response to the first pulse for rates >80 pps. The lack of background activity during the interstimulus period in Figure 1D contrasts with the strong spiking in neurons from awake rabbit in Figure 1, A and B.

The firing rates during the on period (excluding the onset response) and the off period (excluding the first 100 ms) were compared for each pulse rate to determine whether the pulse train elicited a significant excitatory or suppressive response (see Materials and Methods). Figure 2 shows the fraction of units that showed significant excitatory and suppressive responses as a function of pulse rate for all neurons from awake rabbit. The data were no significant differences in the distributions of either spontaneous firing rates (Kolmogorov–Smirnov, p = 0.18) or firing rates during the interstimulus off period (Kolmogorov–Smirnov, p = 0.41) between unilaterally and bilaterally implanted rabbits.

**Figure 2.** Fraction of IC units showing excitatory (positive ordinates) and suppressive (negative ordinates) responses to pulse-train stimulation as a function of pulse rate in unilaterally and bilaterally implanted rabbits.

Pulse-locked responses and response latency

The degree of synchronization (“pulse-locking”) between neural spikes and stimulus pulses was assessed using two complementary methods: one based on cross-correlation between spike trains and stimulus pulse trains (Hancock et al., 2013) and the traditional method based on vector strength (Vollmer et al., 2005; Middlebrooks and Snyder, 2010). The cross-correlation method should provide a more stringent criterion for pulse locking because it requires spikes to occur within a restricted range of latencies after each stimulus pulse, whereas the vector strength method counts all spikes regardless of their temporal position over the stimulus period (Hancock et al., 2013). Figure 3A, B shows example cross-correlograms for the same rabbit IC neurons as in Figure 1, C and B, respectively. Robust pulse locking was evoked by low-rate pulse trains in both neurons, as shown by prominent peaks at a latency of ~6 ms in the cross-correlograms. The peak amplitude dropped markedly (>112 pps) in the first neuron (Fig. 3A) and firing was minimal (>224 pps). In contrast, pulse-locked spikes were observed up to 320 pps in the other neuron (Fig. 3B). The multiple cross-correlogram peaks for pulse rates between 80 and 320 pps reflect the periodicity of the pulse train stimulus. Above 320 pps, no cross-correlogram peak exceeds the 99.5% confidence bound for a random spike train (gray shading), meaning that the responses are unsynchronized to the stimulus pulses.

For each neuron, the pulse-locking limit was defined as the highest pulse rate where a significant peak was observed in the cross-correlogram (see Materials and Methods). Figure 3, C and D, shows the normalized height of the correlation peaks relative to the 99.5% confidence bound for random spike trains as a function of pulse rate. The pulse-locking limit is the rate where the normalized peak height just falls below 1, meaning the correlation peak no longer exceeds the confidence bound. The pulse-locking limits were 156 and 356 pps for the two example neurons. In comparison, Figure 3, E and F, show the vector strength and its 99.5% confidence bound for a random spike train as a function of pulse rate for these two neurons. Pulse-locking limits determined by the vector strength method were 220 and 312 pps.

The area of the significant cross-correlogram peaks lying above the average firing rate (Fig. 3A, B, white filled area) was used to define a “pulse-locked firing rate” that was compared with the overall sustained firing rate during the on period (comprising both pulse-locked and unsynchronized spikes). Pulse-locked firing rates are shown as a function of pulse rate and compared with overall firing rates for the two example neurons in Figure 3, G and H. In the first neuron, the overall and pulse-locked firing rates approximately parallel each other, which is consistent with the decrease in firing rates near the pulse-locking limit (Figs. 1C, 3A). In contrast, in the other neuron, the pulse-locked firing rate falls sharply above 224 pps, whereas the overall firing rate remains substantial up to the highest rate tested, re-
Table 1

<table>
<thead>
<tr>
<th>Firing rate (spikes/s)</th>
<th>Pulse rate (pps)</th>
</tr>
</thead>
<tbody>
<tr>
<td>All spikes</td>
<td>20</td>
</tr>
<tr>
<td>Pulse-locked spikes</td>
<td>220</td>
</tr>
</tbody>
</table>

Figure 3. A, B, Cross-correlograms between stimulus pulse trains and neural spike trains for the same two neurons as in Figure 1, C and B, respectively. Each trace shows the cross-correlogram for one pulse rate. Gray shading indicates the 99.5% upper confidence bound for a random spike train; correlation peaks exceeding the confidence bound are filled in white. C, D, Normalized height of the main correlogram peak as a function of pulse rate. The peaks are normalized to the 99.5% confidence bound (dashed line). Filled marker indicates that the peak height exceeds the confidence bound. The pulse-locking limit is defined as the rate where the peak height intercepts the 99.5% upper confidence bound. E, F, Vector strength and 99.5% upper confidence bound for a random spike train as a function of pulse rate. The limit is defined as the rate where the vector strength intercepts the 99.5% upper confidence bound. G, H, Comparison of firing rates computed using all spike to firing rates computed using only pulse-locked spikes.

The distributions of pulse-locking limits obtained by both methods in awake rabbit are compared in Figure 4A and Table 1 (the cat data also included in Table 1 will be discussed later). The main difference between the two methods is the much higher fraction of neurons that did not pulse lock at any rate using the cross-correlation method (29%) compared with using vector strength (6%), which is consistent with the idea that the cross-correlation method provides a more stringent criterion for pulse locking. Differences between the two methods were minimal when only pulse-locking units were considered and the median pulse-locking limits were similar (148 pps for cross-correlation vs 158 pps for vector strength). Neither the differences in the shapes of the distributions (as assessed by the Kolomogorov–Smirnov test), nor the difference in median pulse-locking limits (Wilcoxon rank-sum test) reached statistical significance (Table 1).

Using the cross-correlation method, there was no significant difference in median pulse-locking limits for contralateral stimulation in unilaterally implanted rabbits compared with diotic stimulation in bilaterally implanted rabbits (Wilcoxon rank-sum test, p = 0.11). In addition, no significant difference in median pulse-locking limits between contralateral and bilateral stimulation was observed among 20 neurons from bilaterally implanted rabbits that were studied with both modes of stimulation (Wilcoxon signed-rank test, p = 0.08). Moreover, a linear regression showed no effect of duration of deafness (the time interval between implantation and recording) on pulse-locking limits (R² = 0.0002, p = 0.89). Finally, pulse-locking limits were not correlated with spontaneous firing rates across the sample of neurons (R² = 0.022, p = 0.15).

Previous studies of the IC using anesthetized preparations (Snyder et al., 1995; Vollmer et al., 1999, 2005) have reported an inverse correlation between pulse-locking limit and spike latency in response to pulsatile CI stimulation. To assess whether such a correlation also holds in awake rabbit, the spike latency in response to pulse trains was calculated from the largest peak in the cross-correlogram (which excludes the onset response). Only responses to low-rate pulse trains (< 80 pps) were used to avoid ambiguities arising when the latency becomes comparable to the interpulse interval. The mode of the latency distribution was centered at 6–7 ms, with a minority of units (23%) having latencies >10 ms (Fig. 4B). In contrast to previous reports from anesthetized preparations, there was no significant correlation between pulse-locking limit and spike latency in awake rabbit (R² = 0.006, p = 0.47; Fig. 4C). This lack of correlation occurs because some long-latency units had a pulse-locked response up to relatively high rates. Previous studies in anesthetized animals measured the first-spike latency in response to the entire pulse train rather than the latency in response to each individual pulse in the train (excluding the onset response). To determine whether this difference in the definition of latency might account for the contrasting findings, we reanalyzed data from previous experiments in anesthetized cat (Hancock et al., 2010, 2012, 2013) using cross-correlation to determine latency and pulse-locking limits. There was a significant negative correlation between the two metrics (R² = 0.36, p < 0.001; Fig. 4C) in anesthetized cat, suggesting that the failure to observe such a correlation in awake rabbit reflects genuine differences between the two preparations rather than differences in the definition of latency.

**Effect of barbiturate on spontaneous activity and response to pulse trains**

The response properties of IC neurons in awake rabbit possess unique features that have not been reported in previous studies of responses to CI stimulation that used anesthetized preparations (Snyder et al., 1991, 1995; Vollmer et al., 1999, 2005; Hancock et al., 2010, 2012, 2013). These features include prominent spontaneous activity, a high incidence of suppressive responses to pulse trains, and unsynchronized, sustained responses at high pulse rates. To assess directly whether these differences reflect a specific effect of anesthesia as opposed to other methodological differ-
ences with these earlier studies (none of which was conducted in rabbit), we monitored changes in the responses of 13 single units after intravenous injection of an ultra-short-acting barbiturate in one unilaterally implanted rabbit.

Illustrative results are presented in Figure 5 for one neuron. Before injection (Fig. 5A), the neuron showed background activity, clear pulse-locked spikes up to 320 pps, and unsynchronized sustained firing at higher pulse rates. One minute after injection (Fig. 5B), the background activity largely disappeared, pulse-locked responses were observed only up to 160 pps and there was no sustained activity at higher pulse rates. Ten minutes after injection (Fig. 5C), the response largely recovered back to the preinjection pattern. The firing rate was not uniformly attenuated across all pulse rates after injection, but rather the relative reduction was more pronounced at high pulse rates so that the “half-maximum pulse rate,” where the firing rate falls to 50% of its maximum value, shifted to a lower value (Fig. 5D).

This change in half-maximum rate is especially apparent when pulses were locked to low-rate pulse trains in awake rabbit. C, Scatter plot of pulse-locking limit against spike latency in awake rabbit and anesthetized cat.

Table 1. Pulse-locking limits by method and preparation

<table>
<thead>
<tr>
<th>Method</th>
<th>Cross-correlation</th>
<th>Vector strength</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Unsynchronized</td>
<td>Median (pps)</td>
</tr>
<tr>
<td>Awake rabbit</td>
<td>29%</td>
<td>148</td>
</tr>
<tr>
<td>Anesthetized cat</td>
<td>20%</td>
<td>80</td>
</tr>
<tr>
<td>Rabbit vs cat</td>
<td>p = 0.07^c</td>
<td>p &lt; 0.001^d</td>
</tr>
</tbody>
</table>

^aPulse-locked units only.

^bKolmogorov–Smirnov test.

^cχ2 test.

^dWilcoxon rank-sum test.
the pre- and postinjection curves are each normalized by their maximum (Fig. 5).

Two additional examples with different response patterns are presented in Figure 6. Before injection, the neuron in Figure 6, A and B, showed high background activity of ~70 spikes/s and firing was strongly suppressed during pulse train stimulation. One minute after injection, the background firing rate was reduced to half its preinjection value and the firing rate during the stimulus was also reduced, especially at low pulse rates. The unit in Figure 6, C and D, fired several spikes per pulse at low pulse rates before injection and showed pronounced rebound responses at high pulse rates. One minute after injection, the background activity was greatly reduced, pulse-locked responses were limited to 20 and 40 pps, and rebound responses were eliminated.

Three metrics were used to characterize these changes for the 13 single units studied with barbiturate injection: the background firing rate between stimulus presentations, the pulse-locking limit based on cross-correlograms, and the half-maximum pulse rate. Figure 7A compares the background firing rates before and after barbiturate injection across the unit sample. Background activity was decreased in all units and the median decrease in background rate (from 8.6 to 1.2 spikes/s) across the sample was statistically significant (Wilcoxon signed-rank test, \( p < 0.001 \)). Figure 7B compares the preinjection and postinjection half-maximum rates and pulse-locking limits. All of the data points lie below the main diagonal, meaning that both the half-maximum rate and the pulse-locking limit were decreased after injection in all 13 units studied. In five units, pulse locking was completely eliminated. The median changes in half-maximum rate (from 176 to 64 pps) and pulse-locking limit (from 224 to 95 pps) across the unit sample were both highly significant (Wilcoxon signed-rank tests, \( p < 0.001 \) for both). However, the median response latencies measured by cross-correlation were not significantly altered by anesthesia (Wilcoxon signed-rank test, \( p = 0.48 \)), which is consistent with the lack of correlation between latency and pulse-locking limit in the awake state (Fig. 4C).

**Comparison with results from anesthetized cat**

We have shown that injection of barbiturate anesthesia causes decreases in background activity, pulse-locking limits, and sustained responses to high rate stimulation in a small number of single units from rabbit IC. The response properties altered by anesthesia in rabbit are precisely those that distinguish results in...
the awake rabbit preparation from responses previously measured in anesthetized animals with CI stimulation, suggesting that anesthesia is a major factor underlying the differences with previous studies. However, the sample of units studied with barbiturate injection is too small \((n = 13)\) to allow a detailed, quantitative comparison with these earlier studies at the neural population level. Moreover, the level of anesthesia induced by the ultra-short-acting barbiturate in rabbit is not as deep as the level used in previous studies \(\text{(which were based on nonsurvival experiments)}\), so that the effects of anesthesia are likely to be underestimated in the rabbit.

To address these issues, we reanalyzed data from previous experiments in anesthetized cats \(\text{(Hancock et al., 2010, 2012, 2013)}\) that used the same intracochlear electrodes, pulse train stimuli, and recording methods to quantitatively compare key response properties of IC neurons in the awake rabbit with those in an anesthetized preparation. This anesthetized cat preparation has been the leading animal model for studies of neural coding with CI stimulation in both the IC \(\text{(Snyder et al., 1991, 1995; Vollmer et al., 1999, 2005; Shepherd et al., 2001; Smith and Delgutte, 2007; Middlebrooks and Snyder, 2010)}\) and auditory cortex \(\text{(Raggio and Schreiner, 1994, 1999, 2003; Kral et al., 2000, 2005, 2009; Fallon et al., 2009)}\). Specifically, data from 117 single units in six bilaterally implanted, anesthetized cats were analyzed. Unless otherwise noted, data from both short-term-deafened \(\text{(1 week)}\) and long-term-deafened \(\text{(6 months)}\) cats were combined because the differences between the two groups were minor for the response measures of interest.

### Spontaneous activity

There were obvious differences in spontaneous activity between anesthetized cats and awake rabbits. Figure 8A shows histograms of the distribution of spontaneous firing rates \(\text{(SRs)}\) across our neuron samples in the two preparations. Approximately \(94\%\) of units in awake rabbit had SRs \(>1\) spike/s with a median SR of 17.27 spikes/s. In contrast, a majority of units in anesthetized cats did not have any SRs and only \(25\%\) of the units had SRs \(>1\) spike/s. The difference in SR distributions between the two preparations was highly significant \(\text{(Kolmogorov–Smirnov test,} p < 0.001)\).

We reported previously that spontaneous firing rates are higher in long-term-deafened cats than in short-term-deafened cats \(\text{(Hancock et al., 2013)}\), which might confound the comparison with the rabbit data when both groups of cats are combined. However, the median spontaneous rates in awake rabbit \(\text{(17.27 spike/s)}\) were significantly higher than those in long-term-deafened cats \(\text{(0.10 spike/s; Wilcoxon rank-sum test,} p < 0.001)\).  

### Prevalence of excitatory and suppressive responses

There were also major differences in the prevalence of excitatory and suppressive responses between the two preparations. In anesthetized cats, the fraction of units showing significant excitatory responses dropped rapidly with increasing pulse rate so that only \(11–15\%\) of the units had an excitatory response for rates of 160 pps and above \(\text{(Fig. 8B)}\). Although a drop with increasing pulse rates is also observed in awake rabbit, the fraction of units with excitatory responses remains \(>30\%\) for all pulse rates and is higher than the cat fraction for all pulse rates except 20 pps. A two-way ANOVA on the arc-sine-transformed data showed significant effects of both pulse rate and preparation \(\text{(} p < 0.001 \text{for both)}\). There is an even clearer contrast in suppressive response between the two preparations. Only \(5–10\%\) of the units at in anesthetized cat show a suppressive response for any pulse rate, as opposed to \(20–50\%\) in awake rabbits. A two-way ANOVA showed a significant effect of preparation on the incidence of suppressive responses \(\text{(} p < 0.001 \text{) but no effect of pulse rate} (p = 0.48)\).

### Temporal coding

Using the cross-correlation method to determine pulse locking, there were fewer neurons that did not pulse lock at any rate in anesthetized cat \(\text{(20\%)}\) than in awake rabbit \(\text{(28\%)}\) \(\text{(Table 1)}\). This result is consistent with the high incidence of suppressive responses that did not give rise to a short-latency cross-correlogram peak in rabbit. Among the units that did show pulse locking, the distribution of pulse-locking limits based was strongly biased toward higher rates in awake rabbit compared with anesthetized cat and this held for both the cross-correlation method and the vector strength method. The difference is clearly apparent in the cumulative distributions based on the cross-correlation method \(\text{(Fig. 8C)}\), which include only pulse-locking units. The median pulse-locking limit was 1.9 times higher in awake rabbit \(\text{(148 pps)}\) than in anesthetized cat \(\text{(80 pps)}\). Approximately \(50\%\) of pulse-locking limits fell in the range \(45–150\ pps\) in anesthetized cat versus \(103–272\ pps\) in awake rabbit. The difference in median pulse-locking limits between the two preparations was highly significant when only pulse-locking units were compared \(\text{(Wilcoxon rank sum test,} p < 0.001)\) and remained significant \(\text{(} p = 0.03 \text{) when all units were included despite the higher proportion of unsynchronized units in rabbit.} \)

In summary, the three effects that were observed directly in a few rabbit single units after barbiturate injection (i.e., decreases in background activity, pulse-locking limits, and incidence of excitatory responses at high pulse rates) are also observed when response properties are compared quantitatively across much larger samples of IC neurons from awake rabbit and anesthetized cat, suggesting that anesthesia is a major factor underlying the differences between the two preparations.

Characterization of the variability in pulse rate dependence of on and off responses

The example neurons in Figures 1 illustrate the wide variability that was observed in both the pulse-rate dependence of responses to pulse trains and the degree to which responses are primarily excitatory or suppressive. To characterize this variability objectively, we used k-means clustering to categorize the various response patterns observed in awake rabbit and anesthetized cat (see Materials and Methods). This clustering analysis was applied to the combined data from both preparations so that the full range of variability would be represented and the distributions of the different clusters could be compared between the two preparations. We chose to split the neuron population into four clusters because this categorization accounted for >50% of the variance in the data and the clusters were easily interpretable. The pulse rate dependence of responses for the four cluster centroids is shown in Figure 9, A–D. The first cluster (Fig. 9A) contains neurons with predominantly suppressive responses, in that the off period response is greater than the on period response at most pulse rates. The other three clusters (Fig. 9B–D) contain predominantly excitatory responses but differ in the pulse rate yielding the largest response. Cluster 2 (Fig. 9B) contains “all-pass” neurons that have an excitatory response over a broad range of pulse rates, often with the maximum firing rate occurring at higher pulse rates (> 320 pps). Cluster 3 (Fig. 9C) contains “band-pass” neurons that have a maximum in firing rate near 80 pps, with a clear drop on either side. Finally, Cluster 4 (Fig. 9D) contains “low-pass” neurons in which the firing rate decreases rapidly with increasing pulse rate and there is little or no sustained response > 80 pps. The example neurons in Figure 1, A–D, belong to the suppressive, all-pass, band-pass, and low-pass clusters, respectively.

The relative incidence of the four response types differed markedly between the two preparations ($\chi^2$ test, $p < 0.001$; Fig. 9E). In awake rabbit, the most common response pattern was the suppressive type (43%), followed by band-pass and all-pass responses in similar proportions (25–30%). Very few neurons in awake rabbit showed the low-pass response pattern. In contrast, the most common response pattern in anesthetized cats was low-pass followed by band-pass. <14% of units in anesthetized cats showed all-pass or suppressive responses. There was no significant difference in the distribution of neurons across the four clusters between unilaterally deafened rabbits and bilaterally deafened rabbits ($\chi^2$ test, $p = 0.92$).

The four clusters defined based on average firing rates also differ in pulse-locked responses. Figure 10 shows both the pulse-locked firing rate (as presented in Fig. 3G,H) and the overall firing rate as a function of pulse rate for each of the four cluster centroids defined in Figure 9. In suppressive neurons (Fig. 10A), the pulse-locked firing rate is much lower than the overall firing rate for all pulse rates, reflecting the lack of a sharp peak in the cross-correlogram for these neurons, which are mostly recorded from awake rabbit. All-pass neurons (Fig. 10B) show a similar dependence of overall and pulse-locked firing rates at low pulse rates, but the pulse-locked rate begins to fall markedly for pulse rates > 224 pps. At high pulse rates (> 320 pps), the overall firing rate is much larger than the pulse-locked rate, which is characteristic of the sustained, unsynchronized responses only observed significantly in awake rabbit. In band-pass neurons (Fig. 10C), the pulse-locked firing rate approximately parallels the overall firing rate with some attenuation. Finally, in low-pass
neurons (Fig. 10D), there is little difference between pulse-locked and overall firing rates, consistent with the lack of background activity in these neurons, which overwhelmingly come from anesthetized cats.

Discussion

We measured the responses of single units in IC of awake rabbit to electric pulse trains delivered through intracochlear electrode arrays. Compared with results from anesthetized preparations, IC neurons in rabbit showed higher spontaneous activity and greater sustained responses (excitatory or suppressive) to high-rate pulse trains. Temporal coding of pulse trains was also enhanced at high pulse rates. Decreases in background activity, pulse-locking limits, and sustained responses at pulse high rates were observed directly in a small number of single units from one rabbit after injection of an ultra-short-acting barbiturate. These changes were consistent with quantitative differences between much larger samples of neurons between the awake rabbit preparation and earlier data from anesthetized cats. Suppression of spontaneous activity may be a neural substrate for the suppression of tinnitus by electric stimulation reported in some CI users (Rubinstein et al., 2003; Van de Heyning et al., 2008; Chang and Zeng, 2012).

Mechanisms of anesthesia effects

Higher evoked firing rates in awake animals have been observed in the IC of normal-hearing rabbits (Kuwada et al., 1989) and guinea pigs (Torterolo et al., 2002) and in the primary auditory cortex of deaf marmosets and guinea pigs with CI stimulation (Johnson et al., 2011; Kirby and Middlebrooks, 2012). Using “precedence effect” stimuli, Tollin et al. (2004) found shorter suppression of the response to the lagging click by the leading click in the IC of awake, behaving cat compared with barbiturate-anesthetized cat (Litovsky and Yin, 1998). A similar effect of barbiturates on recovery times has also been observed in rat IC (Song et al., 2011). Shorter suppression may lead to improved temporal coding of acoustic stimuli at high stimulation rates, which is consistent with our finding of higher pulse-locking limits in awake rabbit.

The reduction in spontaneous and evoked firing rates and the prolongation of recovery times by anesthesia are consistent with an enhancement of inhibition mediated by GABA receptors, as suggested by Kuwada et al. (1989). However, in auditory thalamic neurons, the suppressive effect of pentobarbital on evoked firing rates was not eliminated by iontophoresis of GABA receptor antagonists, suggesting that additional mechanisms are involved (Wan and Puil, 2002). The urethane anesthesia used in our cat preparation acts via a wide spectrum of neurotransmitter-gated ion channels (Hara and Harris, 2002), so the effects of anesthesia are likely to be even more complex.

Earlier reports (Colburn et al., 2009; Hancock et al., 2012) suggested that the upper rate limit on pulse-locked responses to electric pulse trains may be mediated by the low-threshold, voltage-activated potassium currents (I_{K,LVA}) present in many brain-
stem auditory neurons, including medial superior olivary neurons (Svirskis et al., 2004). Neurons expressing \( I_{\text{K,VA}} \) show a cumulative increase in membrane conductance after each stimulus pulse that increases the spiking threshold (Manis and Marx, 1991). Barbiturates decrease excitatory neurotransmitter release by reducing voltage-dependent calcium conductances (Werz and Macdonald, 1985). The resulting reduction of excitative drive may further limit the ability of neurons to reach threshold at high pulse rates under anesthesia.

Perceptual implications of higher coding of high-rate pulse trains

The temporal coding of constant-amplitude pulse trains by IC neurons has been studied extensively in anesthetized preparations, with a focus on characterizing the maximum pulse rate for which responses are synchronized (“pulse-locked”) to the stimulus (Snyder et al., 1995; Vollmer et al., 1999, 2005; Middlebrooks and Snyder, 2010). Using the Rayleigh test for vector strength as a pulse-locking criterion, median pulse-locking limits of IC neurons for intracochlear stimulation range from 95 pps (Vollmer et al., 2005) to 120 pps (Middlebrooks and Snyder, 2010). (Higher limits were obtained with intraneural stimulation of the auditory nerve, which facilitates the stimulation of low-frequency tonotopic regions.) Despite differences among these studies in timing of deafness, stimulation regimens, pulse-locking criteria, and unit selection criteria, temporal coding limits in awake rabbit (median 158 pps using the vector strength method) are clearly higher than those from these anesthetized preparations.

Our finding of higher pulse-locking limits in awake rabbit contrasts with those of Ter-Mikaelian et al. (2007), who observed no difference in the phase-locking limits of IC neurons to sinusoidally amplitude-modulated (SAM) tones between anesthetized and unanesthetized gerbils, even though these investigators found clear effects of anesthesia in the auditory cortex. A transition from sustained to onset response was clearly observed at high pulse rates for individual neurons in a unilaterally deaf rabbit after administration of barbiturate (Fig. 7A–C). Such transitions may be less common in normal-hearing animals, in which both SAM tones and pulse trains can already produce sustained responses at high rates even under anesthetized conditions (Krishna and Semple, 2000; Nelson and Carney, 2007; Krebs et al., 2008).

The higher temporal coding limits of IC neurons in awake rabbits better agree with human performance in temporal processing tasks such as rate pitch discrimination. Most CI listeners can discriminate the rate of periodic pulse trains and rank them based on perceived pitch up to ~300 pps (Moore and Carlyon, 2010). A few “star subjects” show an ability to detect changes in pulse rate perhaps as high as 1000 pps (Townshend et al., 1987; Kong et al., 2009; Kong and Carlyon, 2010). Temporal coding in awake rabbit, where >20% of the neurons had temporal pulse-locking limits ≥224 pps, seems sufficient to account for the rate discrimination abilities of the “typical” CI user. However, the performance of the star subjects still seems hard to account for from the awake IC data.

Pulse rates >300 pps might be coded by the firing rates of IC neurons giving unsynchronized, sustained responses to high-rate pulse trains, which were common in awake rabbit. Such sustained, unsynchronized responses are rarely observed in anesthetized animals. Unsynchronized activity to high-rate stimulation was first described in the primary auditory cortex (Lu et al., 2001; Wang et al., 2008) and thalamus (Bartlett and Wang, 2007) of normal-hearing, awake marmosets, but were rarely observed in the auditory cortex of anesthetized cats (Lu and Wang, 2000).

Subsequently, unsynchronized activity to electric stimulation through CI was also observed in the primary auditory cortex of awake, deaf animals (Johnson et al., 2011; Kirby and Middlebrooks, 2012). The crossover pulse rate where unsynchronized responses become more prevalent than pulse-locked responses was 224 pps in deaf rabbits (data not shown), which is much higher than the ~50 pps crossover rate in the auditory cortex of normal-hearing marmosets (Liu et al., 2001; Fig. 3) and still higher than the ~120 pps crossover in marmoset thalamus (Bartlett and Wang, 2007; Fig. 3). The presence of sustained responses at high pulse rates in the IC and cortex of awake animals is consistent with auditory percepts lasting throughout the stimulus duration in most CI users (Tang et al., 2006).

In the IC of anesthetized animals, sensitivity to ITDs for periodic electric pulse trains is reduced or eliminated at high pulse rates (Smith and Delgutte, 2007; Hancock et al., 2012). Perceptual sensitivity to ITDs also degrades at high stimulation rates in bilateral CI users (Laback et al., 2007; van Hoesel, 2007), but the perceptual limits (250–600 pps) are higher than the limits for most IC neurons in anesthetized cat. It has been suggested that the limits on rate discrimination and ITD sensitivity with CI stimulation may share a common mechanism (van Hoesel, 2007; Carlyon et al., 2008). Whether the enhanced pulse locking at high rates in awake rabbits also translates to improved ITD sensitivity in bilaterally implanted animals will be of interest in future studies and will shed light on the nature of the perceptual limitations.

References


Hancock KE, Noel V, Ryugo DK, Delgutte B (2010) Neural coding of inter-

CrossRef Medline

Coding of Electric Pulse Trains in Awake IC


Coding of Electric Pulse Trains in Awake IC

Chung et al.

Coding of Electric Pulse Trains in Awake IC

CrossRef Medline

CrossRef Medline

CrossRef Medline

CrossRef Medline

CrossRef Medline

CrossRef Medline

CrossRef Medline

CrossRef Medline

CrossRef Medline

CrossRef Medline

CrossRef Medline

CrossRef Medline

CrossRef Medline

CrossRef Medline

CrossRef Medline

CrossRef Medline

CrossRef Medline

CrossRef Medline

CrossRef Medline

CrossRef Medline

CrossRef Medline

CrossRef Medline

CrossRef Medline

CrossRef Medline

CrossRef Medline

CrossRef Medline

CrossRef Medline

CrossRef Medline

CrossRef Medline

CrossRef Medline

CrossRef Medline

CrossRef Medline

CrossRef Medline