Mechanism of assembly of the dimanganese-tyrosyl radical cofactor of class Ib ribonucleotide reductase: Enzymatic generation of superoxide is required for tyrosine oxidation via a Mn(III)Mn(IV) intermediate

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Abstract

Ribonucleotide reductases (RNRs) utilize radical chemistry to reduce nucleotides to deoxynucleotides in all organisms. In the class Ia and Ib RNRs, this reaction requires a stable tyrosyl radical (Y•) generated by oxidation of a reduced dinuclear metal cluster. The Fe³⁺₂-Y• cofactor in the NrdB subunit of the class Ia RNRs can be generated by self-assembly from Fe²⁺₂-NrdB, O₂, and a reducing equivalent. By contrast, the structurally homologous class Ib enzymes require a Mn³⁺₂-Y• cofactor in their NrdF subunit. Mn²⁺₂-NrdF does not react with O₂, but it binds the reduced form of a conserved flavodoxin-like protein, NrdIhq, which, in the presence of O₂, reacts to form the Mn³⁺Mn⁴⁺ intermediate. Here we investigate the mechanism of assembly of the Mn³⁺₂-Y• cofactor in Bacillus subtilis NrdF. Cluster assembly from Mn²⁺₂-NrdF, NrdIhq, and O₂ has been studied by stopped flow absorption and rapid freeze quench EPR spectroscopy. The results support a mechanism in which NrdIhq reduces O₂ to O₂⁻ (40-48 s⁻¹, 0.6 mM O₂), the O₂⁻ channels to and reacts with Mn²⁺₂-NrdF to form a Mn³⁺Mn⁴⁺ intermediate (2.2 ± 0.4 s⁻¹), and the Mn³⁺Mn⁴⁺ species oxidizes tyrosine to Y• (0.08-0.15 s⁻¹). Controlled production of O₂⁻ by NrdIhq during class Ib RNR cofactor assembly both circumvents the unreactivity of the Mn²⁺₂ cluster with O₂ and satisfies the requirement for an “extra” reducing equivalent in Y• generation.

1. INTRODUCTION

Ribonucleotide reductases (RNRs) catalyze the reduction of nucleotides to their corresponding deoxynucleotides and serve as the only de novo source of the deoxynucleotides required for DNA replication and repair for all organisms.¹ RNRs are classified² on the basis of the stable metallocofactor required for transient generation of a cysteine thyl radical³⁴ that initiates nucleotide reduction. In the cases of the structurally

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Supporting information. Experimental details of the determination of the UV-vis spectra of the Mn³⁺₂ cluster and Y•; NrdI-NrdF Kᵣ determination; expression, purification, and characterization of Y105F NrdF; EPR simulations; and attempts at cluster assembly using KO₂ as well as Equations S1-S9, Tables S1-S3, and Figures S1-S20. This material is available free of charge via the Internet at http://pubs.acs.org.
homologous class Ia and Ib RNRs, the oxidizing equivalent necessary for reversible thiol radical generation is stored as a stable tyrosyl radical (Y•) in the enzymes’ β2 subunits. The essential Y• is generated by reaction of a reduced, dinuclear metal cofactor with an oxidant. In class Ia RNRs, the active cofactor is a diferrocium-Y• (FeIII2-Y•), which can be assembled in vitro and in vivo using O2 as oxidant.6 7 Although class Ib RNRs can also assemble an active FeIII2-Y• cofactor in vitro in their β2 subunits (NrdFs), we recently discovered that an active dimanganese(III)-Y• (MnIII2-Y•) cofactor can be generated as well.7 The relevance of the MnIII2-Y• cofactor in vivo has been demonstrated recently by purification of the NrdFs of Corynebacterium ammoniagenes,8 Escherichia coli,3 and Bacillus subtilis9,10 from their native organisms; this result is likely extendable to most or all class Ib RNRs.9 Unlike the FeIII2 forms of the class Ia and Ib RNRs, the MnIII2 form of NrdF is unreactive with O2.7,11 and MnIII2-Y• assembly in vitro requires a flavodoxin-like protein, NrdI, conserved in class Ib systems, in addition to O2.7 Here we report our efforts to elucidate the mechanism of, and essential role of NrdI in, MnIII2-Y• cofactor assembly in the B. subtilis class Ib RNR using stopped flow (SF) absorption and rapid freeze quench (RFQ) EPR spectroscopies.

Extensive studies of the mechanism of FeIII2-Y• cofactor assembly12 in class Ia RNRs (Scheme 1) have provided a framework for thinking about the mechanism of MnIII2-Y• cofactor assembly. The FeIII2-Y• cofactor can be self-assembled in vitro from apo-β2, FeII, O2, and a reducing equivalent (Scheme 2A). The diferrous form of the protein reacts with O2 to generate a μ-peroxodiferrous intermediate.13,14 This intermediate is reduced by a neighboring tryptophan residue to form a FeIIIFeIV intermediate, termed X.15-19 and a tryptophan cation radical (W•+).15,20,21 X is the species responsible for oxidation of the catalytically essential tyrosine (Y122 in E. coli class Ia RNR). In the presence of excess reducing equivalents (FeII, ascorbate, or thiols), this W•+ does not accumulate.15 A protein factor, the ferredoxin YfaE in E. coli, is proposed to act as the donor of the extra electron in vivo.22

Our previous results have provided the first and, to date, only insight into the mechanism of MnIII2-Y• cofactor assembly by demonstrating that this cofactor’s reconstitution in vitro requires the presence of MnII, O2, and the reduced (hydroquinone,hq) form of NrdI.7 Our studies of MnIII2-Y• assembly in E. coli suggested that NrdI reacts with O2 to generate an oxidant competent to oxidize the MnII2 cluster, and that this oxidant channels within a NrdI•NrdF complex from its site of production at the FMNH− cofactor of NrdI to the metal site in NrdF.7 The channeling proposal is supported by the crystal structure of the E. coli NrdI•MnII2-NrdF complex.23 NrdI can conceivably generate either HOO(H) (represented as H2O2 in Scheme 2B) or O2•− (Scheme 2C) as oxidant; previous experiments were unable to distinguish between these options.2,7

MnIII2-Y• cluster is assembled in vitro with the highest yields to date in B. subtilis NrdF (0.6 Y•/β2),10 providing an opportunity to monitor cluster assembly in this system by SF absorption and RFQ-EPR spectroscopies. The results of these studies, presented in this manuscript, strongly suggest that the oxidant is O2•−, produced by reaction of NrdI•hq with O2, oxidizing NrdI to its neutral semiquinone form, NrdI•sq. The first metal-centered intermediate observed is a MnIII•MnIV species, which is kinetically competent to oxidize tyrosine Y105 to Y•. This is the first catalytically relevant MnIII•MnIV dimer in biology, and the analogue to X in FeIII2-Y• cofactor assembly. With O2•− as oxidant, the exact number of oxidizing equivalents necessary for tyrosine oxidation is provided, and the neighboring W residue does not appear to be oxidized during this process. Thus, O2•− is an elegant solution to both the unreactivity of the MnII2 cluster with O2 and the need for three oxidizing equivalents for MnIII2 formation and tyrosine oxidation.
2. EXPERIMENTAL SECTION

2.1. General considerations

Chemical reagents and CuZn superoxide dismutase from bovine erythrocytes (SOD, 4000 U/mg) were obtained from Sigma-Aldrich at the highest purity available. Manganese concentrations were determined using a Perkin-Elmer AAnalyst 600 atomic absorption (AA) spectrometer and a Mn standard solution (Fluka). Iron quantification was carried out using the ferrozine method. SF experiments were carried out using an Applied Photophysics DX 17MV instrument with the Pro-Data upgrade, using a PMT detector. RFQ experiments were performed using an Update Instruments 1019 syringe ram unit and a model 715 syringe ram controller. In both cases, the temperature was maintained at 25 °C using a Lauda circulating water bath. The temperature of the isopentane bath for RFQ was maintained using a liquid N

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2.2. Protein purification

N-terminally His\textsubscript{6}-tagged apoNrdF (tag: MGSSH\textsubscript{6}SSGLVPRGSH) was purified as previously described, with 1,10-phenanthroline added to the culture medium at 100 μM 20 min prior to induction. An additional chromatographic step was added to the published procedures to increase purity and remove minor DNA contaminants. Purifications were typically carried out starting from ~24 g cell paste (16-18 L growth). The eluent following Ni-NTA chromatography (10 mL column, 2.5 × 2 cm) was diluted 4-fold in 50 mM Tris, 5% glycerol, pH 7.6 (Buffer A) and loaded onto a Q Sepharose column (30 mL, 2.5 × 6.5 cm) equilibrated in Buffer A containing 150 mM NaCl, washed with 2 column volumes of the same buffer, and eluted with a 100 × 100 mL gradient of Buffer A containing 150-450 mM NaCl, and 2.5-3 mL fractions were collected. ApoNrdF eluted at 280-380 mM NaCl. The pooled fractions were concentrated and exchanged into 50 mM HEPES, 5% glycerol, pH 7.6 (Buffer B) using an Amicon Ultra 30 kDa MWCO centrifugal filtration device, yielding 7-8 mg/g cell paste. ApoNrdF concentrations (expressed per β2) were assessed using ε\textsubscript{280} = 110 mM\textsuperscript{−1} cm\textsuperscript{−1}. ApoNrdF contained <0.01 Mn/β2 and 0.02 Fe/β2 as purified.

N-terminally His\textsubscript{6}-tagged NrdI (tag: MGSSH\textsubscript{6}SSGLVPRGSH) was purified as described with minor modifications. Following Ni-NTA chromatography, the eluent was diluted 4-fold in 50 mM sodium phosphate, 5% glycerol, pH 7.6 and loaded onto an SP Sepharose column (10 mL, 2.5 × 2 cm), which was washed with 4 column volumes of Buffer B and eluted with Buffer B containing 200 mM NaCl. The eluted protein was concentrated and exchanged into Buffer B using an Amicon Ultra 10 kDa MWCO centrifugal concentrator.

2.3. Determination of the UV-visible spectra of NrdI in the oxidized (ox), sq, and hq forms

The extinction coefficient of oxidized NrdI at 449 nm in Buffer B was determined to be 12.3 mM\textsuperscript{−1} cm\textsuperscript{−1} by trichloroacetic acid precipitation as described. From this value, the spectra of the hq and sq forms were determined as described for E. coli NrdI.

2.4. Preparation of NrdI\textsubscript{hq}, Mn\textsuperscript{II}-loaded NrdF, and O\textsubscript{2}\textsuperscript{−}-saturated buffer

Anaerobic solutions of NrdI (350-450 μM) were reduced by titration with a solution of sodium dithionite (5-6 mM in Buffer B), in a septum-sealed anaerobic cuvette fitted with a gastight syringe with repeating dispenser. Sodium dithionite was added in 1 μL aliquots.
and monitored spectrophotometrically (300-800 nm) until no further change occurred. There was <5% excess dithionite in the resulting NrdI\textsubscript{hq} solutions.

To a solution of ~450 \( \mu \)M apoNrdF, a solution of 3-10 mM MnCl\textsubscript{2} in Buffer B (Mn concentration determined by AA spectroscopy) was added to a final concentration of 3.5 Mn\textsuperscript{II}/β2. The protein was incubated >5 min before use. For SF experiments, the solutions were aerobic, while for RFQ-EPR experiments, the procedure was carried out in an anaerobic chamber. We denote this protein “Mn\textsuperscript{II}-loaded NrdF” rather than “Mn\textsuperscript{II2}-NrdF” because titrations of apoNrdF with Mn\textsuperscript{II} monitored by EPR spectroscopy (Figure S3A) show that not all of the added Mn\textsuperscript{II} is bound under these conditions.

\( \text{O}_2 \)-saturated Buffer B (nominally 1.3 mM \( \text{O}_2 \)) was prepared immediately prior to use at 23 °C by sparging with 100% \( \text{O}_2 \) in a vented container for 0.5-1 h. Where noted, SOD (section 2.1) was added to the buffer at a final concentration of 100-500 U/mL.

### 2.5. Fluorometric determination of the \( K_d \) for NrdI\textsubscript{hq} binding to Mn\textsuperscript{II2}-NrdF

Fluorescence titration studies were carried out using a Photon Technology International QM-4-SE spectrofluorometer equipped with FELIX software and 0.5 mm excitation and 0.75 mm emission bandwidth slits. The excitation wavelength was 380 nm and the emission data were acquired at 475-625 nm, with 1 nm steps and 0.5 s integration time.

All solutions were prepared in the anaerobic chamber. A typical experiment contained in a final volume of 700 \( \mu \)L: 1 \( \mu \)M apo-NrdF, 4 \( \mu \)M MnCl\textsubscript{2}, and 100 \( \mu \)M dithionite in Buffer B. Excess dithionite was added to ensure anaerobicity throughout the duration of the titration. This solution was placed in a semi-micro quartz fluorometer cell (10 mm pathlength, Starna Cells), which was sealed with a septum and screw cap. An airtight 50 \( \mu \)L Hamilton syringe containing 240 \( \mu \)M NrdI\textsubscript{hq} and 100 \( \mu \)M dithionite in Buffer B, fitted to a repeat dispenser, was inserted into the cuvette. The apparatus was removed from the glovebox and equilibrated at 23 °C for 5 min, and a baseline spectrum was recorded. NrdI\textsubscript{hq} was then added in 1 or 2 \( \mu \)L aliquots, the sample was mixed and equilibrated for 1 min, and the spectrum was recorded. The shutter was opened just before each scan and closed immediately after to minimize photobleaching. The final concentration of NrdI\textsubscript{hq} was 15 \( \mu \)M. Data were analyzed by the method of Eftink,\textsuperscript{29} described in detail in the Supporting Information (SI, section S1.1), and provided the stoichiometry of NrdI binding (\( n \)) and the \( K_d \) for its interaction with NrdF.

### 2.6. Kinetics of Mn\textsuperscript{III2}-Y• cofactor assembly monitored by SF absorption spectroscopy

SF kinetics experiments were carried out at 25 ± 1 °C, maintained using a Lauda circulating water bath. The SF apparatus is in the open air; to minimize \( \text{O}_2 \) contamination, the connections of the syringes were purged continuously before and during the experiment with N\textsubscript{2}, and prior to the experiment, the SF lines were rinsed with 10 mL 300 mM dithionite followed by 25-30 mL anaerobic Buffer B. In a typical experiment, 20 \( \mu \)M NrdI\textsubscript{hq} in Buffer B, prepared anaerobically in one gastight Hamilton syringe, was mixed in a 1:1 ratio with \( \text{O}_2 \)-saturated Buffer B (1.3 mM \( \text{O}_2 \)) and 500 U/mL (~0.1 mg/mL) SOD (section 2.1), or \( \text{O}_2 \)-saturated Buffer B and 50 \( \mu \)M Mn\textsuperscript{II}-loaded NrdF (3.5 Mn\textsuperscript{II}/NrdF), in a second syringe. The reaction was monitored at single wavelengths (340, 410, or 610 nm), 4-5 shots were collected and averaged, and the experiments were repeated 2 or 3 times. Reactions were also monitored from 310 to 700 nm in 10 nm intervals (one shot per wavelength, performed on five separate occasions and each data set analyzed independently). After blanking the instrument at each wavelength, a zero timepoint spectrum was obtained by mixing 20 \( \mu \)M NrdI\textsubscript{hq} 1:1 with anaerobic Buffer B. Global analysis of the multiwavelength SF data was carried out in KinTek Explorer v 3.0 with SpectraFit.\textsuperscript{30,31}
For studies of the $O_2$ dependence of the cluster assembly reaction, 25%, 50%, and 75% $O_2$-saturated buffer (0.3, 0.6, and 0.9 mM $O_2$) was prepared by drawing the appropriate amount of $O_2$-saturated buffer into a gastight syringe containing anaerobic buffer. The contents were mixed and loaded immediately to the SF apparatus.

### 2.7. Kinetics of Mn$^{III_2}$ Y• cofactor assembly monitored by RFQ-EPR spectroscopy

In a typical experiment, Mn$^{II_2}$-NrdF (150 μM, 3.5 Mn$^{II}/β_2$) and NrdI$_{sq}$ (100 μM) in Buffer B in one syringe, prepared in the anaerobic box, was mixed with $O_2$-saturated Buffer B in the second syringe in a 1:1 ratio at 25 °C and aged for a pre-determined time period (6 ms – 60 s) in the reaction loop. The reaction mixture (350-400 μL) was sprayed, using a drive ram velocity of 1.25-3.2 cm/s, into liquid isopentane at −140 ± 5 °C in a glass funnel attached to an EPR tube. The samples were packed into the EPR tubes using a stainless steel rod and stored in liquid N$_2$ until analysis. Under these conditions, no decay of NrdI$_{sq}$ and the Mn$^{III}$Mn$^{IV}$ intermediate was observed during storage for 1 month, but ~20-30% decay of both signals was observed over 7 months. The quench times stated in the Results (12 ms – 60 s) include the time required to pass through the reaction loop after mixing plus 6 ms for quenching (estimated as described in ref. 35). The packing factor for NrdF was determined to be 0.55 ± 0.03, using Fe$^{III_2}$-Y• NrdF in Buffer B, prepared as described.

### 2.8. Determination of the UV-visible absorption spectra of the Mn$^{III_2}$ cluster and the Y•

#### 2.8.1. Preparation of Mn$^{III_2}$-Y• NrdF

Protein solutions from 12-13 RFQ reactions quenched at ≥48 ms (section 2.7, 7 mL total volume) were recovered from the reaction loops, pooled, and incubated on ice for 30 min with 5 mM EDTA. The mixture was then loaded onto a Q Sepharose column (3 mL, 1 × 3.5 cm) preequilibrated in Buffer B containing 100 mM NaCl, and the column washed with 9 mL of the same buffer. NrdI eluted in the loading and wash fractions. NrdF eluted with 4 mL Buffer B containing 500 mM NaCl. Protein-containing fractions were pooled/concentrated and repeatedly diluted with Buffer B so that the concentration of NaCl was <10 mM. The isolated Mn$^{III_2}$-Y• NrdF contained 1.0 Mn/$β_2$ and 0.36 ± 0.01 Y•/$β_2$. Thus, for a reaction mixture containing 75 μM Mn$^{II}$-loaded NrdF and 50 μM NrdI, 75 μM Mn is oxidized, 27 μM Y• is generated, and 21 μM Mn$^{III}$ is not associated with Y•.

#### 2.8.2. Reduction of Y• using hydroxylamine and extinction coefficient determination of Mn$^{III_2}$ cluster

Hydroxylamine and hydroxyurea reduce Y• and, albeit more slowly, the Mn$^{III_2}$ cluster. Thus, determination of the individual spectra of the Mn$^{III_2}$ cluster and Y• requires measurement of the UV-vis spectra of NrdF before and after hydroxylamine treatment, accompanied by determination of the Y• concentration by EPR spectroscopy and the Mn concentration by AA spectroscopy. The detailed procedure is given in section S1.2.

### 2.9. EPR spectroscopy

The concentrations of Y• and NrdI$_{sq}$ were determined on a Bruker EMX X-band spectrometer at 77 K using a quartz finger dewar. Analysis of Y• has been described. Analysis of the sq was carried out at 77 K using the following parameters: 9.34 GHz frequency, 5 μW power, 5 × 10$^4$ gain, 100 kHz modulation frequency, 0.15 mT modulation amplitude, 5.12 ms time constant, 20.48 ms conversion time. Spin quantitation of both Y• and sq was carried out using an E. coli Fe$^{III_2}$-Y• NrdF standard calibrated against a Cu$^{II}$ perchlorate standard.

Other EPR spectra were measured at the CalEPR Center at the University of California, Davis. Continuous-wave (CW) X-band spectra were acquired with an ECS106 or E-500 spectrometer (Bruker, Billerica, MA) under non-saturating slow-passage conditions using a
Super-High Q resonator (ER 4122SHQE). Cryogenic temperatures were achieved and maintained using an Oxford Instruments ESR900 liquid helium cryostat in conjunction with an Oxford Instruments ITC503 temperature and gas flow controller. Spectral simulations were performed with Matlab using the EasySpin 4.0 toolbox.\textsuperscript{37}

Quantitation of the unpaired spin concentration within a sample was achieved by comparison of the double integral of the EPR intensity to that of a 308 μM Cu\textsuperscript{II} in 100 mM EDTA, pH 6.2. Due to the presence of multiple paramagnetic species in the RFQ samples, double integration of each of the individual components was not possible. In the case of the transiently generated Mn\textsuperscript{III}Mn\textsuperscript{IV} species, a simulated spectrum was scaled until the integrated intensity of peaks 11-14 (ca. 349, 358, 365, 373 mT, Figure 6) matched that of the experimental spectrum. The entirety of the scaled simulated spectrum was then doubly integrated and compared to the spin standard.

2.10. Determination of rate constants of NrdI comproportionation and disproportionation by SF absorption spectroscopy

NrdI was reduced by anaerobic titration with sodium dithionite as described\textsuperscript{26} and the SF apparatus was prepared as described in section 2.6. In a typical experiment, one syringe contained 20 μM NrdI\textsubscript{hq} in Buffer B and the second contained 20 μM NrdI\textsubscript{ox} either alone or with 80 μM apoNrdF. The reactions were monitored at 25 °C at 610 nm from 1.5 ms – 15 s or 1.5 ms – 200 s, respectively. At least three replicate traces were collected and averaged in three separate experiments and analyzed using KinTek Explorer v. 3.0 according to eq. 4 (section 3.5).

2.11. Analysis of kinetics data

Nonlinear least-squares fitting of single wavelength data from SF and RFQ-EPR experiments to sums of single exponentials was carried out using Origin (Microcal) or KaleidaGraph (Synergy Software). All other kinetic analysis used KinTek Explorer v. 3.0 with SpectraFit.\textsuperscript{30,31}

3. RESULTS

3.1. Proposed model for Mn\textsuperscript{III}\textsubscript{2}-Y• assembly

Here, we describe rapid kinetics studies using SF absorption and RFQ-EPR spectroscopies, to determine the mechanism by which NrdI is involved in Mn\textsuperscript{III}\textsubscript{2}-Y• cofactor assembly in the \textit{B. subtilis} class Ib RNR. Our studies have led to the working model in Scheme 3. Specifically, our evidence described subsequently supports 1) rapid one-electron reduction of O\textsubscript{2} by NrdI\textsubscript{hq} to generate O\textsuperscript{2−} (40-48 s\textsuperscript{-1}); 2) slower formation of a Mn\textsuperscript{III}Mn\textsuperscript{IV} intermediate (2.2 ± 0.4 s\textsuperscript{-1}) subsequent to O\textsuperscript{2−} production; and 3) decay of the Mn\textsuperscript{III}Mn\textsuperscript{IV} intermediate concomitant with Y• generation (0.08-0.15 s\textsuperscript{-1}).

3.1.1. Information required for experimental design and spectral deconvolution—To design the SF and RFQ-EPR experiments that led to Scheme 3, a number of parameters were determined for the NrdI/NrdF system. First, the UV-vis absorption and EPR spectra (where appropriate) of starting materials and products were obtained. Second, a knowledge of the affinity between NrdI\textsubscript{hq} and Mn\textsuperscript{II}\textsubscript{2}-NrdF is required to maximize complex formation. Third, because minimal studies of the reaction of flavodoxins, and no studies of the reaction of NrdIs, with O\textsubscript{2} have been reported, the reaction of NrdI\textsubscript{hq} with O\textsubscript{2} was assessed. The results of these experiments are described first.
3.2. UV-visible absorption spectra of NrdI, Mn\(^{III}\)\(^2\)-NrdF, and Y•

Analysis of the SF data requires knowledge of the UV-vis absorption spectra of the stable redox states of NrdI and NrdF. The UV-vis absorption spectra of NrdI in hq, sq, and ox states are shown in Figure 1A. Because NrdI accumulates only 30% sq during anaerobic titration with sodium dithionite,\(^{10}\) the spectrum of NrdI\(_{sq}\) was estimated by correlation of UV-vis and EPR spectra of solutions of NrdI partially reduced with known amounts of dithionite. The spectrum is similar to those of flavodoxins and other NrdIs.\(^{26,27,38}\) NrdI\(_{sq}\) is entirely in the neutral form under the conditions used in all experiments (pH 7.6), based on UV-vis, EPR, and SF absorption experiments (section 2.1, Table S1, and Figure S1).

The UV-vis absorption spectrum of NrdF reconstituted with Mn\(^{III}\)\(^2\)-Y• cofactor is shown in the inset of Figure 1B. Figure 1B also shows the spectrum resolved into its two components, the Mn\(^{III}\)\(^2\) cluster and the Y•. To obtain these spectra, Mn\(^{III}\)\(^2\)-Y• NrdF (200 μM) was incubated with hydroxylamine (0.5 mM) (sections 2.8 and S1.2). The deconvolution is complicated as Y• reduction is accompanied by slower reduction of the Mn\(^{III}\)\(^2\) cluster.\(^{7}\) By 10 min, when Y• is fully reduced, ~60% of the Mn is also reduced, based on AA analysis of NrdF subsequent to chelation with EDTA and Sephadex G25 chromatography to remove Mn\(^{II}\)-EDTA. The spectrum of Y• was then determined by spectral subtractions and the quantification of Y• by EPR spectroscopy. The spectrum of the Mn\(^{III}\)\(^2\) cluster exhibits a weak, broad feature at 460 nm with a shoulder at 485 nm, similar to the Mn\(^{III}\)\(^2\) form of Mn catalase, whose active site is structurally related to that of the class I RNRs.\(^{39}\) The spectrum of Y• exhibits a diagnostic, sharp peak at 410 nm, a shoulder at 392 nm, and a broad feature from 470 to 670 nm. The analysis indicated that the UV-vis absorption spectrum intensity of the Mn\(^{III}\)\(^2\) cluster increased by 190% upon reduction of Mn\(^{III}\)\(^2\)-Y• cofactor to Mn\(^{III}\)\(^2\) cluster (Figure S2). Similar 2-3 fold changes in absorption intensities of biological Mn complexes have also been reported for anion binding to Mn superoxide dismutase\(^{40}\) (SOD) and for mutation of a tyrosine in the second coordination sphere of Mn catalase.\(^{41}\) These changes have been assigned to structural reorganizations, which suggests that, in NrdF, Y• reduction may alter the coordination sphere of the accompanying Mn\(^{III}\)\(^2\) cluster. The absorption spectrum of Mn\(^{II}\)-loaded NrdF (not shown) is identical to that of apo-NrdF, featureless in the visible region.

3.3. EPR spectra of Mn\(^{II}\)-loaded NrdF, Mn\(^{III}\)\(^2\)-Y• NrdF, and NrdI\(_{sq}\)

Characterization of the EPR spectra of Mn\(^{II}\)-loaded and Mn\(^{III}\)\(^2\)-Y• NrdF is important for the RFQ-EPR analysis of the cluster assembly reaction described below. The EPR spectrum of NrdI\(_{sq}\) (77 K, Figure S1) is similar to previously described flavodoxin neutral sqs.\(^{42}\)

3.3.1. Mn\(^{II}\)-loaded NrdF—Initially, NrdF was loaded with Mn\(^{II}\) following the protocol reported for E. coli NrdF,\(^{7}\) by incubation of the apoprotein with 4 Mn\(^{II}/\beta_2\) followed by passage through a Sephadex G25 column to remove unbound Mn\(^{II}\). For E. coli NrdF, this procedure yielded protein with a complex, multoline (~40 lines) EPR signal with negligible amounts of mononuclear Mn\(^{II}\.\)\(^{7}\) However, when this same protocol was followed for B. subtilis NrdF, its EPR signal revealed a substantial contribution from mononuclear Mn\(^{II}\) in addition to a signal associated with a coupled Mn\(^{II}\)\(^2\) cluster. Titration of 75 μM apoNrdF with Mn\(^{II}\), monitored by EPR spectroscopy at 10 K, show that the mononuclear Mn\(^{II}\) is present even at 1 Mn\(^{II}/\beta_2\) (Figure S3A). Measurements at 293 K, where only unbound Mn\(^{II}\) is detectable, suggest that <10% of the total Mn\(^{II}\) is unbound at 3.5 Mn\(^{II}/\beta_2\), whereas 20% is unbound at 4.0 Mn\(^{II}/\beta_2\). Therefore, to minimize the unbound Mn\(^{II}\) present, EPR samples were generally prepared using 3.5 Mn\(^{II}/\beta_2\) (Figure 2A). Similar titrations of E. coli apoNrdF exhibit negligible unbound Mn\(^{II}\) unless the Mn\(^{II}/\beta_2\) was greater than 3.5, indicating much stronger Mn\(^{II}\) binding than to B. subtilis NrdF (Figure S3B).
As for the Mn$^{II}_2$ cluster of *B. subtilis* NrdF itself, the low-temperature X-band CW EPR spectrum possesses several resonances (Figures 2A, S4) across the explored field range (0—900 mT). The features at 270-300 and 380-440 mT show evidence of hyperfine structure with splittings of approximately 4.2 mT. Such a splitting pattern is consistent with nearly equal hyperfine interactions from two exchange-coupled $^{55}$Mn$^{II}$ ($I = 5/2$) ions [$A(55\text{Mn}) = 285$ MHz, typically] 

As the temperature is raised, the total integrated intensity of the EPR spectrum increases (Figure S4, inset). This behavior indicates that the two Mn$^{II}$ ions are antiferromagnetically coupled (i.e., that $J < 0$; using the Heisenberg-Dirac-van Vleck Hamiltonian for exchange $\hat{H} = -2JS_1S_2$) and, at higher temperatures, higher energy spin manifolds on the spin-ladder become thermally populated at the expense of the EPR-silent $S_T = 0$ ground spin level.

### 3.3.2. Mn$^{III}_2$-Y$^•$ NrdF—The EPR spectrum of *B. subtilis* Mn$^{III}_2$-Y$^•$ NrdF (10 K, Figure 2B) is similar to that of *E. coli* NrdF. The five-line pattern present in the low-temperature EPR spectra of the Mn$^{III}_2$-Y$^•$ cofactors of class Ib RNRs has been attributed to the interaction of Y$^•$ with the Mn$^{III}_2$ cluster. 

As the temperature is increased, the signal appears to collapse, yielding, at 100 K, what appears to be a spectrum of solely a Y$^•$ (Figure S5). Similar temperature-dependent behavior has been observed for a nitroxide coupled to a ferric-heme complex and for the acetate-inhibited $S_2$-form of photosystem II (in this case, a Y$^•$ is coupled to a paramagnetic tetranuclear Mn cluster). With increasing temperature, the rate of electronic relaxation of the metal-based paramagnet increases and decouples this spin center from the nearby organic radical. This decoupling removes any contribution to the spectrum from dipolar and/or exchange interactions between the two spin centers. Thus, the high-temperature spectrum of NrdF described here is essentially that of Y$^•$ isolated from the Mn$^{III}_2$ cluster.

As a result of this decoupling, the structure evident in the 100 K spectrum can be assigned as hyperfine interaction (HFI) of the unpaired electron on tyrosine with four protons: those at the 3 and 5 positions of the phenoxyl ring, and the two bound to the β-carbon of the sidechain. To simulate this spectrum, the HFI for H3/5 were held fixed at values typically found for Y$^•$s in class Ib RNRs (section S2.2). The contribution to the spectrum by the two β-protons was determined using the McConnell relations by varying the value of the dihedral angle $\Theta$ until the simulated spectrum best-agrees with the experimental one. This was achieved with an angle of 44 (±2)°, slightly larger than the dihedral angles of 25 and 32° observed in the two chains of the crystal structure of *B. subtilis* Mn$^{III}_2$-NrdF (see Figure S6 for a comparison of simulations achieved using these other values of $\Theta$). This increase in dihedral angle observed for Mn$^{III}_2$-Y$^•$ compared to that found in the structure of the Mn$^{III}_2$ form suggests that the phenoxyl plane has rotated with respect to the β-carbon upon oxidation of NrdF in the presence of NrdI and O$_2$.

Interestingly, in the crystal structure of the Mn$^{III}_2$ form of *C. ammoniagenes* NrdF, the tyrosine adjacent to the Mn cluster (Y115) has a dihedral angle of 40° and 42°, in the first and second chains, respectively.

### 3.4. $K_d$ for NrdI$_{hq}$ and Mn$^{II}_2$-NrdF

Knowledge of the affinity of NrdI$_{hq}$ for Mn$^{II}_2$-NrdF is also important to maximize complex formation in the rapid kinetics studies described subsequently. To make this measurement, we took advantage of the previous observation that the hq forms of flavodoxins display weak fluorescence with a broad excitation maximum centered at ~370 nm and emission maxima in the 500-530 nm region. Our initial experiments demonstrated that the NrdI$_{hq}$ FMNH$^-$ cofactor displays similar fluorescence properties and that the intensity of its fluorescence emission spectrum is sensitive to the presence of NrdF (Figure S7A); this property was exploited to determine the $K_d$ for NrdI$_{hq}$ binding to Mn$^{II}_2$-NrdF. A control
titration of *B. subtilis* NrdI<sub>hq</sub> with *E. coli* Mn<sup>II</sup>-NrdF exhibited no change in fluorescence, demonstrating that this method reports on specific NrdI-NrdF interaction. Titrations of Mn<sup>II</sup>-loaded *B. subtilis* NrdF (1 μM, 4 Mn<sup>II</sup>/β<sub>2</sub>) with NrdI<sub>hq</sub> were analyzed using a non-cooperative binding model, as described in the SI (section S1.1, Figure S7B). The analysis gives 1.6 ± 0.1 NrdIs per NrdF dimer with a K<sub>d</sub> of 0.6 ± 0.2 μM. Figures 2A and S3A suggest that NrdF is not fully loaded with Mn<sup>II</sup> under these conditions, but a similar K<sub>d</sub> was obtained with apoNrdF. The unusual binding stoichiometry may therefore indicate error in the computationally derived extinction coefficient of NrdF, or that a fraction of NrdF is incompetent to bind NrdI. This K<sub>d</sub> value is significantly lower than that previously reported for *B. anthracis* NrdI<sub>ox</sub>-NrdF, 23 μM. The sequence similarity between the *B. subtilis* and *B. anthracis* systems (75% for NrdF, 63% for NrdI) suggests that the difference in K<sub>d</sub>s reflects tighter binding of NrdF to NrdI<sub>hq</sub> than to NrdI<sub>ox</sub>. The K<sub>d</sub> of 0.6 μM indicates that at the concentrations of NrdF and NrdI<sub>hq</sub> used in subsequent rapid kinetics experiments, >95% of NrdI<sub>hq</sub> is complexed.

3.5. Reaction of NrdI<sub>hq</sub> with O<sub>2</sub> (± NrdF) monitored by SF absorption

Typical flavodoxins react with O<sub>2</sub> to produce O<sub>2</sub>•<sup>-</sup>; however, based on the unusual redox properties of *E. coli* and *B. subtilis* NrdIs, we previously proposed that NrdI<sub>hq</sub> would react with O<sub>2</sub> to produce H<sub>2</sub>O<sub>2</sub>. To resolve this issue, and as a prelude to our investigation of the kinetics of Mn<sup>III</sup><sub>2</sub>-Y• cofactor assembly from NrdI<sub>hq</sub>, Mn<sup>II</sup>-NrdF, and O<sub>2</sub>, we initially investigated the kinetics of the reaction of NrdI<sub>hq</sub> alone with O<sub>2</sub>. In these experiments, 20 μM NrdI<sub>hq</sub> and O<sub>2</sub>-saturated buffer (1.3 mM O<sub>2</sub>) were mixed in a 1:1 ratio at 25 °C, in the presence of 250 U/mL SOD, and the reaction was monitored by SF absorption spectroscopy from 310 to 700 nm at 10 nm increments. SOD was included to minimize kinetic complexity associated with production of O<sub>2</sub>•<sup>-</sup>, which can also react with NrdI<sub>hq</sub> and NrdI<sub>sq</sub> (section S2.3). Singular value decomposition (SVD) analysis of the data indicated significant contribution from three absorbing species: NrdI<sub>hq</sub> (E-FMNH<sup>-</sup>), NrdI<sub>sq</sub> (E-FMNH•), and NrdI<sub>ox</sub> (E-FMN). The data were fit using the global analysis software KinTek Explorer with SpectraFit in order to extract the spectra of these species. The ability to reproduce the known NrdI<sub>sq</sub> spectrum (Figure 1A) was one of the criteria used to judge the appropriateness of the kinetic model. The full analysis is described in section S2.3 (Figures S8A and S9). Three reactions (eq. 1-3) were required in the kinetic model to reproduce the known NrdI<sub>sq</sub> spectrum.

\[
\begin{align*}
E - \text{FMNH}^- + O_2 \rightarrow E - \text{FMNH}^+ + O_2^- & \quad (1) \\
E - \text{FMNH}^+ + O_2 \rightarrow E - \text{FMN} + O_2^- & \quad (2) \\
E - \text{FMNH}^- + O_2 + H^+ \rightarrow E - \text{FMN} + H_2O_2 & \quad (3)
\end{align*}
\]

This model gives rate constants of \( k_1 = 1.6 \pm 0.1 \text{ mM}^{-1} \text{s}^{-1} \), \( k_2 = 2.0 \pm 0.1 \text{ mM}^{-1} \text{s}^{-1} \), and \( k_3 = 0.7 \pm 0.1 \text{ mM}^{-1} \text{s}^{-1} \), for eq. 1, 2, and 3, respectively. Based on this model and the relative values of \( k_1 \) and \( k_3 \), 70% of NrdI<sub>hq</sub> reacts with O<sub>2</sub> to initially form NrdI<sub>sq</sub> and O<sub>2</sub>•<sup>-</sup>. The NrdI<sub>sq</sub> then goes on to react with O<sub>2</sub> to produce NrdI<sub>ox</sub> and O<sub>2</sub>•<sup>-</sup>. Thirty percent of NrdI<sub>hq</sub> reacts by a second pathway to form NrdI<sub>ox</sub> and H<sub>2</sub>O<sub>2</sub>.

To investigate whether complexation with NrdF affects the kinetics of NrdI<sub>hq</sub> oxidation, SF experiments were also carried out with NrdI<sub>hq</sub> (20 μM) mixed in a 1:1 ratio with O<sub>2</sub>-saturated buffer containing NrdF (50 μM, 98% complex), apo or Mn<sup>II</sup>-loaded, and SOD.
Unlike the reaction of \( \text{NrdI}_{\text{hq}} \) with \( \text{O}_2 \), these SF data could not be satisfactorily modeled by the reactions in eq. 1-3, although SVD again indicated the presence of only three spectrally distinct species. Therefore, we instead focused on the pseudo-first-order rate constants for \( \text{NrdI}_{\text{sq}} \) formation and decay, which can be extracted directly from SF traces acquired at 610 nm, where only \( \text{NrdI}_{\text{sq}} \) contributes significantly (Figures 1A and 3). \( \text{NrdI}_{\text{sq}} \) formation is accelerated in the presence of apo\( \text{NrdF} \) and even further accelerated in the presence of Mn\( ^{\text{II}} \)-loaded \( \text{NrdF} \) (Figure 3). Whereas the \( A_{610} \) SF traces for reaction of \( \text{NrdI}_{\text{hq}} \) alone with \( \text{O}_2 \) can be fit to a sum of two exponentials, those in the presence of \( \text{NrdF} \) are best fit to three exponentials, with two phases required for \( \text{sq} \) formation and one phase for \( \text{sq} \) decay (Table 1, Figure S10). We attribute the presence of two phases for \( \text{sq} \) formation to interaction of \( \text{NrdI} \) with conformationally heterogeneous populations of \( \text{NrdF} \) (note that even in the presence of Mn\( ^{\text{II}} \)-loaded \( \text{NrdF} \), only 80% of \( \text{NrdF} \) contains Mn\( ^{\text{II}2} \) cluster). The rate constants for the major phases (75-80% of total amplitude) of \( \text{sq} \) formation in the presence of apo- and Mn\( ^{\text{II}} \)-loaded \( \text{NrdF} \) were 5 and 25 times faster, respectively, than that for \( \text{NrdI}_{\text{sq}} \) formation in the absence of \( \text{NrdF} \). The sensitivity of the rate constants for \( \text{NrdI}_{\text{sq}} \) formation to the presence of \( \text{NrdF} \) and Mn\( ^{\text{II}} \) suggests the relevance of eq. 1 to Mn\( ^{\text{III}2} \)-\( \text{Y}^* \) cofactor assembly, discussed subsequently. By contrast, the rate constant for \( \text{NrdI}_{\text{sq}} \) decay (eq. 2) was very similar in the presence and absence of \( \text{NrdF} \) (Table 1).

Control experiments indicated that comproportionation and disproportionation reactions (eq. 4) of \( \text{NrdI}'s \) FMN cofactor were too slow to contribute significantly to the overall reactions of \( \text{NrdI} \) with \( \text{O}_2 \) in the presence and absence of \( \text{NrdF} \) (section S2.4, Figure S11, Table S2). This observation indicates that the acceleration in \( \text{NrdI}_{\text{sq}} \) formation in the presence of \( \text{NrdF} \) is due to an increase in \( k_1 \), rather than a large increase in \( k_3 \) followed by comproportionation. Therefore, the data suggest that the reaction of \( \text{NrdI}_{\text{hq}} \) with \( \text{O}_2 \) to produce \( \text{NrdI}_{\text{sq}} \) and \( \text{O}_2^*^- \) also predominates in the presence of apo- or Mn\( ^{\text{II}} \)-loaded \( \text{NrdF} \).

\[
E - \text{FMNH}^- + E - \text{FMN}^{2+} \xrightleftharpoons[k_4]{k_{-4}} 2E - \text{FMNH}^* \tag{4}
\]

### 3.6. Mn\( ^{\text{III}2} \)-\( \text{Y}^* \) cofactor assembly monitored by SF absorption and RFQ-EPR spectroscopies

The experiments described above provided the information required to design the central experiment: the reaction of \( \text{NrdI}_{\text{hq}} \) with \( \text{O}_2 \) in the presence of Mn\( ^{\text{II}} \)-loaded \( \text{NrdF} \), monitored by SF absorption and RFQ-EPR spectroscopies.\(^57\) The experimental design is challenging as it is necessary to minimize excess Mn\( ^{\text{II}} \), which interferes with the EPR analysis, and to maximize \( \text{NrdI}_{\text{hq}} \)-\( \text{NrdF} \) complex formation, to avoid free \( \text{NrdI}_{\text{hq}} \) reacting with \( \text{O}_2 \). As described in section 3.3 (Figure S3A), adding 3.5 Mn\( ^{\text{II}/}\beta2 \) to 75 \( \mu \text{M} \) apo\( \text{NrdF} \) strikes a balance between high metal loading of \( \text{NrdF} \) (3.2 Mn\( ^{\text{II}/}\beta2 \) bound) and minimizing unbound Mn\( ^{\text{II}} \). With 75 \( \mu \text{M} \) Mn\( ^{\text{II}} \)-loaded \( \text{NrdF} \) and 50 \( \mu \text{M} \) \( \text{NrdI}_{\text{hq}} \) immediately after 1:1 mixing with \( \text{O}_2 \)-saturated buffer, >97% of \( \text{NrdI}_{\text{hq}} \) is complexed to \( \text{NrdF} \). As noted above, 0.36 \( \text{Y}^* /\beta2 \) and 1.0 Mn\( ^{\text{III}/}\beta2 \) are produced (27 \( \mu \text{M} \) Mn\( ^{\text{III}2} \)-\( \text{Y}^* \), 11 \( \mu \text{M} \) Mn\( ^{\text{III}2} \) not associated with \( \text{Y}^* \)), whereas 2.2 Mn\( ^{\text{II}/}\beta2 \) is not oxidized. These yields are important in interpreting the kinetics results. The same conditions were used for both SF and RFQ-EPR analyses.

Given the number of absorbing species and the complexity of their spectra, in the initial SF experiments, the reaction was monitored between 310 and 700 nm in 10 nm increments, as well as at 405 and 415 nm (to better resolve the \( \text{Y}^* \)). The spectra reconstructed from the individual SF traces are shown in Figure 4 in three time regimes for clarity. Based on these results, we carried out a more detailed analysis of the kinetic traces at 610, 340, and 410 nm (Figure S12). At 610 nm, \( \text{NrdI}_{\text{sq}} \) formation and disappearance predominated, along with a small contribution (~5% of total amplitude) from Mn\( ^{\text{III}2} \) cluster and \( \text{Y}^* \). Analysis at 340 nm allowed detection of a Mn\( ^{\text{III}2}\text{Mn}^4 \) intermediate, and 410 nm was chosen as it is the \( \lambda_{\text{max}} \)
the Y•; all three oxidation states of NrdI also contribute at these two wavelengths. In the following sections these results are used to formulate the mechanistic proposal for Mn$^{III-2}$-Y• cofactor assembly in Scheme 3 (all rate constants and amplitudes are in Table S3).

### 3.6.1. NrdI$_{sq}$ production and decay

The full spectrum analysis of the first time regime (83 ms) revealed NrdI$_{sq}$ formation at 610 nm and three isosbestic points, at 330, 390, and 415 nm, indicating direct conversion of NrdI$_{hq}$ to NrdI$_{sq}$ (Figure 4A). In the second and third regimes, NrdI$_{sq}$ decays by ~4 s to NrdI$_{ox}$, with an isosbestic point at ~500 nm. The plot of A$_{610}$ (Figure 5, red trace, fit in black line), is fit best to four exponentials (Table 2, Figure S12A) – two increasing phases at 610 nm (NrdI$_{sq}$ formation), with rate constants of 40 s$^{-1}$ (~75% of total NrdI) and 8 s$^{-1}$ (~20%), followed by a decreasing phase of 0.65 s$^{-1}$ (NrdI$_{sq}$ oxidation to NrdI$_{ox}$), and finally a small increase of 0.15 s$^{-1}$ (Mn$^{III-2}$ and Y• formation, section 3.6.3).

For comparison with the SF results, the reaction of NrdI$_{hq}$ with Mn$^{II}$-loaded NrdF and O$_2$ under identical conditions was monitored by the RFQ method (12 ms – 60 s) and analyzed by EPR spectroscopy. NrdI$_{sq}$ can be quantified at 77 K, even in the presence of multiple Mn-derived signals and Y•, because its signal saturates at microwave powers orders of magnitude below the other paramagnetic species. The samples from 12 ms to 7 s were analyzed at 5 $\mu$W at 77 K and the results overlaid on the A$_{610}$ trace of the SF reactions (Figure 5). The data show accumulation of 31 ± 5 $\mu$M sq at 83 ms – 20% lower than the maximal accumulation of NrdI$_{sq}$ as determined from the SF trace at 610 nm (38 $\mu$M).

The RFQ data can be fit to a two-exponential model with $k_1 = 88 \pm 38$ s$^{-1}$ and $k_2 = 0.8 \pm 0.2$ s$^{-1}$. The larger than expected value of $k_1$ based on the SF data, and the large uncertainty in $k_1$ and its amplitude, reflect the observation that the NrdI$_{sq}$ formation reaction is 50% complete by the first timepoint (Figure 5).$^{60}$ $k_2$ agrees well with the rate constant for sq decay determined by SF absorption (Scheme 3). Thus, the SF and RFQ-EPR data are in agreement that ~60-70% NrdI$_{hq}$ reacts with O$_2$ at ~40-60 s$^{-1}$ to form NrdI$_{sq}$ and O$_2^\cdot$-.

This rate constant is ~5 times faster than that of the major phase of NrdI$_{sq}$ formation in the presence of apoNrdF (7.4 s$^{-1}$) and ~40 times faster than the rate constant for NrdI$_{sq}$ formation in the absence of NrdF (1.6 mM$^{-1}$ s$^{-1}$; 1 s$^{-1}$ at 0.6 mM O$_2$).

### 3.6.2. Formation and decay of a Mn$^{III}$Mn$^{IV}$ intermediate

The reconstructed spectra in Figure 4B in the 83 ms – 3 s regime are dominated in the visible region by the conversion of NrdI$_{sq}$ to NrdI$_{ox}$, with an isosbestic point at ~500 nm. Absence of the NrdI$_{sq}$/NrdI$_{ox}$ isosbestic point at ~350 nm (Figure 1A) indicates formation of an additional, UV-absorbing species; the 300-350 nm region displays significantly higher absorbance than can be attributed to NrdI$_{ox}$ (Figure 1A). This absorbance is more clearly observed in the spectra from the 3-60 s regime, in which features in the 300-350 nm region decay as the sharp feature of the Y• grows in at 410 nm (arrows, Figure 4C). The A$_{340}$ SF traces (Figure S12B, Table 3) suggest formation and decay of this UV absorbing feature at 2.5 and 0.08 s$^{-1}$, respectively.

The UV-visible spectrum of this species was estimated from the spectrum of the 2 s timepoint reconstructed from the SF data. At this timepoint, substantial intermediate is present, little Y• is apparent (Figure 4B) and NrdI is entirely in the sq and ox forms. After subtraction of the contributions of NrdI$_{sq}$ (13 $\mu$M, Figure 5), NrdI$_{ox}$ (37 $\mu$M), and protein scattering, the relatively featureless spectrum with little visible absorption shown in Figure S14 was obtained. This spectrum is similar to that of the Mn$^{III}$Mn$^{IV}$ form of Mn catalase$^{62}$ and synthetic models.$^{63}$
The identity of the intermediate was determined by analysis of the RFQ samples at 10 K, where possible EPR-active intermediates in cofactor formation (Mn^{II}Mn^{III} and Mn^{III}Mn^{IV}) may be observable. A spectral component that has 16 resonance lines is present in the X-band EPR data of RFQ reaction intermediates trapped between 12 ms and 30 s (Figure S15). These features are very similar to those of strongly exchange-coupled Mn^{III}Mn^{IV} dimers.

Indeed, the best fit of this contribution to the 2 s spectrum (obtained after subtraction of a scaled spectrum of the starting material to remove the contribution from unreacted Mn^{II}-loaded NrdF) requires two Mn ions with effective HFI of [—465 —435 —310] MHz and [230 230 240] MHz for the Mn^{III} and Mn^{IV} centers, respectively (Figure 6, section S2.5). These values are nearly identical to those determined for the superoxidized form of Mn catalase. Importantly, there is no spectroscopic evidence for the presence of a Mn^{II}Mn^{III} form of the cluster at any point in the reaction. Owing to the large intrinsic HFI for Mn^{II} centers as well as the larger projection factors, the EPR spectrum of an antiferromagnetically-coupled Mn^{II}Mn^{III} dimer is expected to be spread over a much larger field range than that typically observed for a Mn^{III}Mn^{IV} dimer.

For the lowest field feature of the Mn^{III}Mn^{IV} multiline signal, there is a noticeable shift upfield from 270.1 mT to 271.3 mT as the reaction progresses (see Figure S15). Additional changes in spectral lineshape are also observed for the highest field features. These small perturbations of the effective Mn HFI likely result from changes in either the exchange coupling parameter $J$, or in the Mn^{III} zero-field splitting constant, possibly due to a change in protonation state of a bridging oxo group or a solvent-derived ligand to the Mn^{III} ion, respectively.

In order to quantify the Mn^{III}Mn^{IV} intermediate and determine its rates of formation and decay, we first used the intensity of peak 14 at ~373 mT (Figure 6) to estimate the relative concentrations of the Mn^{III}Mn^{IV} species at each timepoint. The results of this analysis are plotted in Figure 7, fit to rate constants of $2.2 \pm 0.4$ s$^{-1}$ and $0.12 \pm 0.02$ s$^{-1}$. The similarity of these rate constants to those from the SF analysis strongly suggests that the UV-absorbing intermediate and the EPR-active Mn^{III}Mn^{IV} are the same species. Interestingly, the rate constant for formation of Mn^{III}Mn^{IV} is 20 times slower than the rate of NrdI$_{sq}$ generation (section 4.3).

The relative concentrations of the intermediate were then converted to absolute concentrations (Figure S16) by determination of a scaling factor between the peak intensities in Figure 7 and the concentrations of Mn^{III}Mn^{IV} obtained by quantitative simulations (30% uncertainty) of three of the RFQ samples (section 2.9). The presence of multiple paramagnetic species in the reaction mixture – Mn^{II}_2-NrdF, Mn^{II}, NrdI$_{sq}$, and Y$^\bullet$ – requires that, before simulation of Mn^{III}Mn^{IV}, the contribution from unreacted Mn^{II}-loaded NrdF be subtracted from the spectra using the broad features centered at ~290 and 410 mT, outside the envelope of Mn^{II}, Y$^\bullet$, and NrdI$_{sq}$ (Figure 2A). As shown in Table 3 and Figure S16, the maximal concentration of Mn^{III}Mn^{IV} estimated by this method was $19 \pm 8$ μM, at ~1.5 s. While Mn^{III}Mn^{IV} could, in principle, be on pathway to form Mn^{II}_2-Y$^\bullet$, Mn^{III}_2 cluster, or both, the $19 \pm 8$ μM value most likely underestimates the amount of Mn^{III}Mn^{IV} formed, as the RFQ-EPR result underestimated the concentration of NrdI$_{sq}$ by 20% (Figure 5). This is consistent with the intermediate being a precursor to the 27 μM Mn^{III}_2-Y$^\bullet$ cofactor formed under these conditions. This conclusion is supported by the kinetics of Y$^\bullet$ formation, as described below.

In an effort to facilitate characterization and quantification of the Mn^{III}Mn^{IV} species, we also generated the Y105F mutant of B. subtilis NrdF (sections S1.3 and S2.6). This mutant bound Mn^{II} more tightly than wt NrdF did (Figure S17), but little fluorescence change was observed in titrations to determine NrdI$_{hq}$/Y105F-NrdF affinity, indicating that NrdI$_{hq}$
bound Mn$^{II}_2$-Y105F NrdF either weakly or incorrectly (Figure S18). SF studies monitoring reaction of NrdI$^{hq}$ and Mn$^{II}_2$-Y105F NrdF with O$_2$ revealed no evidence of a Mn$^{III}$Mn$^{IV}$ intermediate (Figure S19).

### 3.6.3. Y• generation

To determine the kinetics of Y• generation and whether the Mn$^{III}$Mn$^{IV}$ intermediate is kinetically competent for its formation, the single wavelength trace at 410 nm, the $\lambda_{\text{max}}$ of the Y•, was fit to three exponentials (Figure S12C, Table 3, Table S3). The fastest phase, a small decrease at 48 s$^{-1}$, is attributed to the oxidation of NrdI$^{hq}$ to NrdI$^{sq}$ ($\varepsilon_{410}$ of these species differ by only 0.35 mM$^{-1}$ cm$^{-1}$, Figure 1A). The second and major phase is an increase with $k = 0.74$ s$^{-1}$ and an amplitude consistent with oxidation of NrdI$^{sq}$ to NrdI$^{ox}$. The slowest phase ($k = 0.09$ s$^{-1}$) is associated with the appearance of the sharp feature of Y• (Figure 4C). The observed amplitude of this phase is consistent with decay of 25 ± 8 $\mu$M Mn$^{III}$Mn$^{IV}$ (Table 3). Because NrdI$^{sq}$ and/or Mn$^{III}$Mn$^{IV}$ contribute significantly to the 77 K EPR spectra of all but the 30, 40, and 60 s RFQ samples, independent determination of the rate constant for Y• formation by EPR spectroscopy is problematic. However, the rate constants for Mn$^{III}$Mn$^{IV}$ decay by RFQ-EPR (0.12 ± 0.02 s$^{-1}$) and SF (0.09-0.15 s$^{-1}$ at 340 and 610 nm) and Y• formation by SF (0.08 s$^{-1}$ at 410 nm) are in agreement (Table 3). Therefore, despite the somewhat lower than expected maximal concentration of Mn$^{III}$Mn$^{IV}$ determined by simulation of the RFQ-EPR data, the kinetic results together strongly suggest that the Mn$^{III}$Mn$^{IV}$ intermediate is responsible for tyrosine oxidation to Y•.

### 3.6.4. Interpretation of the rate constant for NrdI$^{sq}$ decay

In all rapid kinetics experiments, regardless of the presence/absence of NrdF and as long as SOD is present, the rate constant for reaction of NrdI$^{sq}$ with O$_2$ (eq. 2) is ~1 s$^{-1}$ (Table 2). This rate constant is lower than that for Mn$^{III}$Mn$^{IV}$ formation (2.2 ± 0.4 s$^{-1}$), which suggests that the O$_2^•$− produced by NrdI$^{sq}$ oxidation is not involved in Mn$^{III}$Mn$^{IV}$ generation and that O$_2^•$− generated by NrdI$^{hq}$ oxidation is predominantly responsible for Mn$^{II}_2$ oxidation. Furthermore, the observation that the rate of NrdI$^{sq}$ oxidation is not accelerated in the presence of Mn$^{II}$-loaded NrdF also suggests that no further reducing equivalents, such as those required to reduce a tryptophan radical,$^{20,21}$ are needed for cluster assembly. Thus O$_2^•$− provides the three oxidizing equivalents required for Mn$^{III}_2$-Y• cofactor generation.

### 4. DISCUSSION

#### 4.1. Kinetic complexity in Mn$^{III}_2$-Y• cofactor assembly

Together, the SF and EPR results described in this manuscript strongly support the mechanistic model of Mn$^{III}_2$-Y• cofactor assembly in Scheme 3. Following a treatment of several of the complexities that have been encountered in the course of the analysis, we discuss each reaction in Scheme 3 in turn.

Interestingly, 31-37 $\mu$M NrdI$^{sq}$ is generated in the fastest phase of the cluster assembly reaction, in conditions that give 27 $\mu$M Y• and 75 $\mu$M Mn oxidized. Thus ~2 Mn$^{II}$ are oxidized for every O$_2^•$− produced in the fast phase, which may suggest that all of the Mn$^{II}$ oxidized is present in dinuclear Mn$^{III}_2$ clusters. It is not clear why Y• is only generated in ~70% of these clusters – but the analogous question in class Ia diferric-Y• assembly, in which only 1.2 Y•/β2 are generated despite oxidation of 3.6 Fe/β2, is also unanswered despite twenty years of mechanistic study.

The formation of substoichiometric levels of NrdI$^{sq}$ could be related to heterogeneity in NrdF due to incomplete/incorrect Mn$^{II}$ binding or to NrdI’s ability to react with O$_2$ by two pathways (eq. 1-3). Approximately 70% of NrdI$^{hq}$ reacts with O$_2$ to form NrdI$^{sq}$ in the absence or presence of Mn$^{II}$-loaded NrdF, suggesting that the complexity may be the result
of using NrdI stoichiometrically, whereas it likely acts catalytically in vivo based on the reported amounts of NrdI and NrdF in *B. subtilis* and *E. coli*.

In vivo, NrdI<sub>hq</sub> can be rapidly regenerated by a NrdI reductase. This reductase has not been identified, but NrdI’s role as a one electron donor suggests that the reductant is likely flavodoxin (ferredoxin) reductase. A flavodoxin reductase has yet to be identified in *B. subtilis*. Interaction with the reductase may also influence the reaction of NrdI<sub>hq</sub> with O<sub>2</sub> to favor O<sub>2</sub>•− production.

**4.2. Reaction of NrdI<sub>hq</sub> with O<sub>2</sub>**

The rate acceleration of NrdI<sub>hq</sub> oxidation in the presence of NrdF and O<sub>2</sub> may be caused by positively charged residues we have previously noted on NrdF at the NrdI binding site (R196 and K27 in *B. subtilis* NrdF). Positively charged groups in the vicinity of the reactive C4a position of the flavin, thought to facilitate formation of the sq-O<sub>2</sub>•− pair, are key features of the active sites of many flavoproteins that react with O<sub>2</sub> as part of their catalytic cycles. Our observed rate constants for reaction of NrdI<sub>hq</sub> with O<sub>2</sub> in the presence of NrdF to form O<sub>2</sub>•− are comparable to the rate constants for reaction of many flavoprotein oxidases with O<sub>2</sub>.

The reaction of NrdI<sub>sq</sub> with O<sub>2</sub> in *B. subtilis* NrdI (0.65-0.8 s<sup>−1</sup>, 0.6 mM O<sub>2</sub>) is nearly two orders of magnitude slower than the reaction of the hq and insensitive to the presence of NrdF. The hq and neutral sq forms of NrdI (and flavodoxins in general) are protonated at the N5 position of the flavin, whereas the ox form is deprotonated at this position. Crystal structures of flavodoxins and NrdIs have revealed that a peptide loop region (the “40s loop” in *B. subtilis*) in the vicinity of C4a undergoes a conformational change upon flavin reduction to allow a hydrogen bond between N5H and a peptide backbone carbonyl. The proton transfer and accompanying conformational change are reasonable sources of a kinetic barrier to sq oxidation (e.g. ref. 68). This hypothesis is supported by the decrease in the rate constant for sq decay with decreasing pH (Table S1). The observation that the rate constant for NrdI<sub>sq</sub> oxidation is not significantly changed in the presence of NrdF indicates either that the conformational change and proton transfer are rate limiting even if NrdI<sub>sq</sub> is bound to NrdF, or that NrdI<sub>sq</sub> dissociates from NrdF before reacting with O<sub>2</sub>. Further studies are required to determine when in the course of the cluster assembly reaction NrdI dissociates from NrdF, as sq or ox.

**4.3. Formation of the Mn<sup>III</sup>Mn<sup>IV</sup> intermediate**

Having identified O<sub>2</sub>•− as the oxidant in our current studies, we suggest that O<sub>2</sub>•− oxidizes the Mn<sup>II</sup><sub>2</sub> cluster by inner- or outer-sphere electron transfer, possibly coupled with proton transfer. It is also possible that superoxide is transported to the metal site in the protonated form, HO<sub>2</sub>•<sup>+</sup> (pK<sub>a</sub> 4.9), analogous to the case of azide binding to hemerythrin, in which HN<sub>3</sub> (pK<sub>a</sub> 4.75) is proposed to be the species that binds to the metal site. From our biochemical and crystallographic data on the *E. coli* class Ib system, we proposed that the oxidant channels from NrdI to the metal site via a hydrophilic tunnel within the NrdI•NrdF complex. The orientation of the channel suggests that (H)O<sub>2</sub>• would first encounter Mn2, the Mn<sup>II</sup> site farthest from Y105. Therefore, we propose initial formation of a Mn<sup>II</sup>Mn<sup>III</sup> intermediate or Mn<sup>II</sup>Mn<sup>III</sup>OO(OH) adduct, with Mn<sup>III</sup> being at site 2. To cleave the O-O bond and generate a Mn<sup>III</sup>Mn<sup>IV</sup> intermediate similar to superoxidized Mn catalase (μ-oxo, μ-hydroxo bridged), a (hydro)peroxo-bridged Mn<sup>II</sup>Mn<sup>III</sup> species, possibly analogous to the μ-1,2-peroxodiferric intermediate proposed in class Ia RNR Fe<sup>III</sup>-Y• cofactor assembly (Scheme 1), may be formed at least transiently.

A Mn<sup>II</sup>Mn<sup>III</sup> species that forms and accumulates should be detectable by UV-vis absorption and EPR spectroscopy. The UV-vis spectrum of the Mn<sup>II</sup>Mn<sup>III</sup> would be expected to be similar to that of the Mn<sup>III</sup><sub>2</sub> cluster (Figure 1B) but with half the extinction coefficient; the
4.4. Tyrosine oxidation

The kinetic data indicate that Mn$^{II}$-Mn$^{IV}$ decays concomitantly with Y• generation, suggesting that it is the oxidant directly responsible for tyrosine oxidation. This reaction is notable for its slow rate constant: 0.1 s$^{-1}$ (25 °C), compared to 1 s$^{-1}$ and 5 s$^{-1}$ (both at 5 °C) for oxidation of tyrosine by X in E. coli and mouse class Ia RNRs, respectively. Although it is not known whether electron, proton, or coupled electron/proton transfer is rate-limiting for tyrosine oxidation in these systems, the very slow oxidation of tyrosine by Y• by a Mn$^{II}$Mn$^{IV}$ intermediate may reflect its lower reduction potential compared to X, as has been suggested on the basis of calculations on other RNR systems.

4.5. Concluding remarks

Comparison of the general mechanisms of Mn$^{II}$-Y• and Fe$^{II}$-Y• cofactor assembly (Schemes 1 and 3) demonstrates Nature’s elegant and efficient strategy to balance two inherent problems in enzymatic Y• generation: how to activate O$_2$ and the need for an odd number of electrons for Y• generation. In Fe$^{II}$-Y• cofactor assembly, reaction of O$_2$ with a Fe$^{II}$ center is facile, but an extra electron is delivered after O$_2$ activation to form the stable Fe$^{III}$-Y• cofactor. To our knowledge, only one biological Mn$^{II}$ center has been suggested to be transiently oxidized by O$_2$ at a physiologically relevant rate, and it does so with the help of an electron-donating co-substrate. In NrdF, the most efficient strategy for
circumventing this problem is to deliver the extra electron as the first step in the process in the form of O$_2$$^{•−}$, with which Mn$^{\text{II}}$ reacts readily. Reduction of O$_2$ requires NrdI$hq$; efforts to assemble Mn$^{\text{III}}$Y• using exogenous O$_2$$^{•−}$ have been unsuccessful (section S2.7). Once O$_2$$^{•−}$ is generated, the three oxidizing equivalents necessary for formation of the Mn$^{\text{III}}$Mn$^{\text{IV}}$ intermediate and, subsequently, a Y• are present. This is a more economical mechanism than use of H$_2$O$_2$ as oxidant, which would require 2 O$_2$ and 5 added reducing equivalents to form one Y• (Scheme 2).

An important question stemming from these results is how class I b RN Rs assemble both Mn$^{\text{III}}$Y• and Fe$^{\text{III}}$Y• cofactors, whereas class I a RN Rs assemble only Fe$^{\text{III}}$Y• cofactors. The structural$^{51}$ and mechanistic similarities between these systems suggest that the most incisive answer to this question may not be found at the metal site, but instead in the ways in which the unique difficulties of O$_2$$^{•−}$ production and transport to the metal site have been solved for cluster formation in class I b RN Rs. Production of O$_2$$^{•−}$ requires a specific accessory protein, NrdI. The oxidant channels$^{23}$ in class I b RN Rs appear configured for transport of a hydrophilic molecule like O$_2$$^{•−}$ or HO$_2$$^{•−}$ – distinct from the proposed O$_2$ access route to the diferrous site in the class I a RN Rs, which is largely hydrophobic.$^{80,81}$ Thus Mn$^{\text{III}}$Y• cofactor assembly in the class I b RN Rs represents a remarkable example of how Nature has expanded the range of chemistry that can be performed by the dimetal-carboxylate structural motif, by creating and harnessing a normally deleterious oxidant for an essential cellular purpose.

**Supplementary Material**

Refer to Web version on PubMed Central for supplementary material.

**Acknowledgments**

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**REFERENCES**

(32). These concentrations were chosen to simplify the kinetics of NrdI oxidation, rather than to maximize Y• yield (0.6 Y•/β2), see section 3.6.


(49). \[\Theta\] corresponds to the dihedral angle between a vector that is normal to the phenoxyl plane and the \[\beta\]1-proton. The \[\beta\]2-proton is assumed to have a dihedral angle equal to 120°–\[\Theta\].


(53). The extinction coefficient was calculated using ExPASy (see ref. 10). However, stoichiometries of metal binding are generally ~3.5 per dimer in \[\beta\]2 subunits purified to date.


(57). SOD was not included in these experiments as initial studies demonstrated that the reaction rates were affected only at the highest concentrations of SOD, 250 U/mL, and then only slightly (Figure S8B). We attribute the negligible effect of SOD in the presence of Mn\[III\]II-loaded NrdF to \[O_2•−\] channeling to the metal site rather than its reacting further with NrdI.

(58). The monotonic increase in NrdI\[sq\] evident from the first ~160 ms of the multiwavelength SF data (Figure 4A) suggests that the 8 s\(^{-1}\) phase corresponds to a second, slower phase of NrdI\[sq\] formation. The amplitude of the 8 s\(^{-1}\) phase is too large to be attributable to formation of Mn\[III\]II\[IV\] intermediate, which does not absorb significantly at 610 nm (section 3.6.2), and 5 times too large to be attributable to formation of Mn\[III\]II\[II\]Y• (\[\epsilon_{610} = 0.3 \text{ mM}^{-1} \text{ cm}^{-1}\]) or Mn\[III\]II\[II\]Y• (\[\epsilon_{610} = 0.3 \text{ mM}^{-1} \text{ cm}^{-1}\]).

(59). Studies of the dependence of these rate constants on \[O_2\] (Figure S13) indicated that the 40 s\(^{-1}\) NrdI\[sq\] formation and 0.65 s\(^{-1}\) NrdI\[sq\] decay phases exhibited O\(^2−\) dependence, whereas the 8 s\(^{-1}\) NrdI\[sq\] rise phase did not. The latter observation is not understood at present but is likely related to reversibility of the reactions of NrdI with O\(^2−\). As expected if a metal-centered intermediate precedes tyrosine oxidation, the rate constant for formation of Mn\[III\]II\[II\] cluster and Y• (~0.15 s\(^{-1}\) at 610 nm) was zero order in O\(^2−\).

(60). In support of the data fitting overestimating the value of \(k_1\), a half-time of 11 to 16 ms for NrdI\[hq\] formation corresponds to a rate constant of 43-63 s\(^{-1}\), in good agreement with the rate constant derived from the SF studies.

(61). Note that, even if the other 30% of NrdI\[hq\] reacts with O\(^2−\) to form H\(_2\)O\(_2\) (the amplitudes of the SF fits suggest that all NrdI\[hq\] reacts to form O\(^2−\)), only 15 \(\mu\)M H\(_2\)O\(_2\) would be produced under these reaction conditions. This amount of H\(_2\)O\(_2\) could only generate 7.5 \(\mu\)M Y•, whereas 27 \(\mu\)M is produced. This indicates that O\(^2−\) is the primary, and probably only, oxidant involved in cluster assembly.


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Figure 1.
UV-vis absorption spectra of NrdI and NrdF. A) NrdI_{ox} (black), NrdI_{sq} (neutral form, blue), and NrdI_{hq} (red). The sq spectrum was estimated as described in section 2.3. B) The Y• (red) and Mn_{III2} cluster (black, es given for cluster following Y• reduction) in NrdF, after removal of the contribution of protein end absorption. Inset: 200 μM Mn_{III2}-Y• NrdF, containing 1.0 Mn/β2 and 0.36 Y•/β2.
Figure 2.
X-band EPR spectra of (A) Mn$^{II}$-loaded NrdF (150 μM NrdF, 3.4 Mn$^{II}$/β2, acquired at 0.1 mW, 10 K), (B) Mn$^{III2}$-Y• NrdF (0.1 mW, 10 K). Other acquisition parameters are described in section 2.9.
Figure 3. Comparison of representative A_{610} traces for reaction of 20 μM NrdI_{eq} mixed 1:1 with O_{2}-saturated buffer containing no NrdF (black), 50 μM apo-NrdF (red), and 50 μM Mn^{II}-loaded NrdF (blue, 3.5 Mn^{II}/β2). The first and second reaction also contained SOD (100 U/mL). The reactions were carried out at 25 °C, pH 7.6. The small increase at >5 s in the blue trace is due to Mn^{III}_{2}-Y• cofactor assembly.
Figure 4.
Reaction of 50 μM NrdI_hq and 75 μM Mn^{II}-loaded NrdF with 0.6 mM O_2, as monitored by SF absorption spectroscopy, divided into three time regimes: 0 – 0.083 s (A), 0.083 – 3 s (B), and 3 – 60 s (C). The spectra are point-by-point reconstructions from kinetic traces acquired every 10 nm between 310 and 700 nm, as well as 405 and 415 nm. One shot per wavelength is shown, but the data is representative of further experiments conducted at this and lower concentrations (10 μM NrdI_hq, 25 μM Mn^{II}_2-NrdF).
Figure 5.
Formation and decay of NrdI$_{sq}$ in the reaction of NrdI$_{hq}$ and Mn$^{II}$-loaded NrdF with O$_2$, monitored by SF absorption and RFQ-EPR spectroscopies. NrdI$_{sq}$ was monitored by SF absorption spectroscopy at 610 nm (right axis, red line) and the trace fit to four exponentials (black line, Table 2). NrdI$_{sq}$ was quantified in RFQ timepoints (left axis, black squares) quenched at the indicated times by EPR spectroscopy (77 K, 5 $\mu$W). Note that NrdI$_{sq}$ does not reach zero concentration due to a contribution from Y• at >2 s timepoints, resulting in the smaller than expected amplitude for sq decay as determined by RFQ (Table 2). The two y-axes are scaled for direct comparison of the SF and RFQ-EPR data according to the extinction coefficient of NrdI$_{sq}$ at 610 nm, 4.9 ± 0.7 mM$^{-1}$ cm$^{-1}$.10
Figure 6. Simulation of the EPR spectrum of the Mn$^{III}$Mn$^{IV}$ intermediate. The spectrum was obtained by subtraction of a scaled spectrum of the Mn$^{II}$-loaded NrdF starting material from the spectrum of the RFQ mixture of 75 μM NrdF, 50 μM NrdI, and oxygenated buffer after 2 s aging time. Simulation of the spectrum was performed as described (section 2.9). Peaks 11-14 are labeled. Spectrometer settings: temperature: 10 K; microwave frequency = 9.38 GHz; power = 20 μW; modulation amplitude = 0.5 mT; modulation frequency = 100 kHz; sweep rate = 5.3 mT/s. The off-scale feature at ~330 mT is due to NrdI$_{sq}$. 
Figure 7.
Concentration of the Mn$^{III}$Mn$^{IV}$ intermediate, followed by the peak to trough intensity of peak 14 (Figure 6). The data (black, mean ± SD for 2-4 independent experiments) are fitted to a two phase model (red) with rate constants given in Table 3.
Scheme 1.
Mechanism of diferric-Y• cofactor assembly in class Ia RNRs.
Scheme 2.
(A) Stoichiometry of diferric-Y• cofactor assembly in the *E. coli* class Ia RNR. (B) and (C) Proposed stoichiometries of dimanganese-Y• cofactor assembly in *E. coli* and *B. subtilis* class Ib RNRs with H₂O₂ and O₂•− as oxidants.
Scheme 3. Proposed mechanism of Mn$^{\text{III}}_2$-Y• cofactor assembly in *B. subtilis* NrdF$^{a}$

$^a$Rate constants were measured in this study. The detailed structures of the proposed Mn$^{\text{II}}$Mn$^{\text{III}}$OO(H) and Mn$^{\text{III}}$Mn$^{\text{IV}}$ intermediates, as well as the oxidation state of NrdI when it dissociates from NrdF, are unknown. Mn at site 1 (Mn1) is closest to the Y that is oxidized, Mn2 is indicated in red.
Table 1

Apparent rate constants (amplitudes in parentheses) for the fits to the SF traces shown in Figure 3.

<table>
<thead>
<tr>
<th>Phase</th>
<th>k (s⁻¹)</th>
</tr>
</thead>
<tbody>
<tr>
<td>NrdI</td>
<td>1.6 (0.052)</td>
</tr>
<tr>
<td>NrdI + apoNrdF²</td>
<td></td>
</tr>
<tr>
<td>NrdI + Mn²-loaded NrdF³</td>
<td></td>
</tr>
<tr>
<td>Sq formation (phase 1)</td>
<td>18 (0.009)</td>
</tr>
<tr>
<td>Sq formation (phase 2)</td>
<td>NA⁵</td>
</tr>
<tr>
<td>Sq decay (eq. 2)</td>
<td>1.1 (−0.053)</td>
</tr>
</tbody>
</table>

²100 U/mL SOD.

³No SOD.

⁵NA: not applicable (phase not observed)
Table 2

Apparent rate constants and amplitudes for the reaction of NrdI\textsubscript{hq} and Mn\textsuperscript{II}-loaded NrdF with O\textsubscript{2}, determined by fits to single-wavelength SF (610 nm) or RFQ-EPR (77 K) data in Figure 5.\textsuperscript{a}

<table>
<thead>
<tr>
<th>Phase</th>
<th>SF, 610 nm</th>
<th>RFQ-EPR</th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>k (s\textsuperscript{-1})</td>
<td>ΔA</td>
<td>Amplitude (μM)</td>
<td>k (s\textsuperscript{-1})</td>
</tr>
<tr>
<td>sq formation (phase 1)</td>
<td>40</td>
<td>0.18</td>
<td>37</td>
<td>88 ± 38\textsuperscript{c}</td>
</tr>
<tr>
<td>sq formation (phase 2)</td>
<td>8</td>
<td>0.05</td>
<td>10</td>
<td></td>
</tr>
<tr>
<td>sq decay</td>
<td>0.65</td>
<td>−0.24</td>
<td>49</td>
<td>0.8 ± 0.2</td>
</tr>
<tr>
<td>Mn\textsuperscript{III}\textsubscript{2}-Y\textsuperscript{*} formation</td>
<td>0.15</td>
<td>0.012</td>
<td>27</td>
<td></td>
</tr>
</tbody>
</table>

\textsuperscript{a}Values represent the data from 3 traces (SF) or 2-4 experiments (RFQ-EPR). Similar rate constants were obtained in SF studies at lower NrdI\textsubscript{hq} and NrdF concentrations (Figure S13 and Table 1).

\textsuperscript{b}Adjusted for packing factor

\textsuperscript{c}The reaction is ~50% complete by the first timepoint, explaining the large uncertainty in these values (see text).

\textsuperscript{d}Mn\textsuperscript{III} not associated with Y\textsuperscript{*} is also produced in the assembly reaction. Assuming 27 μM Mn\textsuperscript{III}\textsubscript{2}-Y\textsuperscript{*} and 11 μM Mn\textsuperscript{III}\textsubscript{2} (section 2.8.1) gives ΔA \textsuperscript{ΔA} = 0.011, in good agreement with the observed ΔA.
Table 3

Apparent rate constants and amplitudes for Mn\textsubscript{III}Mn\textsubscript{IV} formation and decay in the reaction of NrdI\textsubscript{hq}, Mn\textsuperscript{II}, loaded NrdF, and O\textsubscript{2}, determined by fits to single-wavelength SF (340 and 410 nm) or RFQ-EPR (10 or 77 K) data.\textsuperscript{a}

<table>
<thead>
<tr>
<th>Phase</th>
<th>340 nm</th>
<th>410 nm</th>
<th>RFQ-EPR</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>$k$ (s\textsuperscript{-1})</td>
<td>$\Delta A$</td>
<td>Amplitude (μM)</td>
</tr>
<tr>
<td>Mn\textsuperscript{III}Mn\textsuperscript{IV} formation</td>
<td>2.5</td>
<td>0.17</td>
<td>13 ± 4 \textsuperscript{b}</td>
</tr>
<tr>
<td>Mn\textsuperscript{III}Mn\textsuperscript{IV} decay</td>
<td>0.08</td>
<td>−0.16</td>
<td>19 ± 8 \textsuperscript{e}</td>
</tr>
</tbody>
</table>

\textsuperscript{a}Values represent the data from 3 traces (SF) or 2-4 experiments (RFQ-EPR). Similar rate constants were obtained in SF studies at lower NrdI\textsubscript{hq} and NrdF concentrations (Figure S13).

\textsuperscript{b}For phases involving Mn\textsuperscript{III}Mn\textsuperscript{IV} formation or decay, we have assumed $\varepsilon_{340} = 13 \pm 4$ mM\textsuperscript{-1} cm\textsuperscript{-1} and $\varepsilon_{410} = 3 \pm 1$ mM\textsuperscript{-1} cm\textsuperscript{-1}. These values were determined from Figure S14 on the basis of the estimated UV-vis spectrum of the intermediate and the quantification of the EPR spectrum of Mn\textsuperscript{III}Mn\textsuperscript{IV} by simulation at 3 timepoints (section 2.9).

\textsuperscript{c}NA: not applicable (not observable at this wavelength). Given that the Mn\textsuperscript{III}Mn\textsuperscript{IV} intermediate should contribute $\sim 0.07$ to $\Delta A_{410}$ (Figure S14), it is probable that this phase was not resolved from that of sq decay in the fits because its rate constant for formation is somewhat similar to that for sq decay, and of the same sign (see Table S3).

\textsuperscript{d}Uncertainty includes uncertainties in fit and in concentration of Mn\textsuperscript{III}Mn\textsuperscript{IV} (30%)

\textsuperscript{e}Concentrations of Mn\textsubscript{II}2-Y• (27 μM) and Mn\textsuperscript{II}2 (11 μM) are taken as given (section 2.8.1)