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Cytosolic Monothiol Glutaredoxins Function in Intracellular Iron Sensing and Trafficking via Their Bound Iron-Sulfur Cluster

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SUMMARY

Iron is an essential nutrient for cells. It is unknown how iron, after its import into the cytosol, is specifically delivered to iron-dependent processes in various cellular compartments. Here, we identify an essential function of the conserved cytosolic monothiol glutaredoxins Grx3 and Grx4 in intracellular iron trafficking and sensing. Depletion of Grx3/4 specifically impaired all iron-requiring reactions in the cytosol, mitochondria, and nucleus, including the synthesis of Fe/S clusters, heme, and di-iron centers. These defects were caused by impairment of iron insertion into proteins and iron transfer to mitochondria, indicating that intracellular iron is not bioavailable, despite highly elevated cytosolic levels. The crucial task of Grx3/4 is mediated by a bridging, glutathione-containing Fe/S center that functions both as an iron sensor and in intracellular iron delivery. Collectively, our study uncovers an important role of monothiol glutaredoxins in cellular iron metabolism, with a surprising connection to cellular redox and sulfur metabolisms.

INTRODUCTION

Iron is essential for virtually all organisms, because it functions as a cofactor in central cellular processes such as respiration, DNA synthesis and repair, ribosome biogenesis, and metabolism. Research over the past decade has uncovered sophisticated systems facilitating the specific transport of iron across the plasma and various intracellular membranes (Hentze et al., 2004; Kaplan and Kaplan, 2009; Philpott and Protchenko, 2008; Vergara and Thiele, 2008). Despite its central metabolic function, little is known about the passage of iron through the eukaryotic cytosol to become incorporated into proteins and transported into various subcellular compartments. A soluble, low-molecular-mass form of iron was described in the 1970s, but ever since the discovery of this “labile iron pool,” its physiological importance and composition have been under debate (Crichton and Charloteaux-Wauters, 1987; Richardson and Ponka, 1997). Presumably, iron may also be bound to dedicated proteins ensuring specific delivery and insertion into iron-requiring sites. A metallo-chaperone function has been worked out for insertion of copper and nickel into respective metal-containing enzymes (Finney and O’Halloran, 2003; Lyons and Eide, 2007), but proteins performing a general role in iron trafficking or insertion are unknown. An iron donor function has been suggested for frataxin in mitochondrial Fe/S cluster biosynthesis (Bencze et al., 2006; Lill, 2009). In humans, the poly (rC) binding protein 1 (PCBP1) was shown to specifically deliver bound iron to ferritin, the major iron storage protein in higher eukaryotes (Shi et al., 2008). The apparently specific role of the PCBP1 iron chaperone and the fact that both ferritin and PCBP1 are not universally conserved leave open the possibility that other proteins with a general importance for iron trafficking exist. Clearly, the mode of specific iron delivery within the eukaryotic cytosol remains one of the fundamental unresolved problems of iron homeostasis.

Because iron is not only essential but also toxic at higher levels, cells have developed sophisticated systems for ensuring a tightly regulated iron homeostasis (Hentze et al., 2004; Kaplan and Kaplan, 2009). In mammals this process is executed by iron-regulatory proteins in a posttranscriptional fashion, and the yeast Saccharomyces cerevisiae uses the iron-sensing transcription factors Aft1 and Aft2. Under iron deprivation, Aft1-Aft2 activate transcription of genes of the iron regulon encoding cell-surface iron transporters and proteins involved in intracellular iron utilization (Kaplan and Kaplan, 2009; Philpott and Protchenko, 2008). Sensing of intracellular iron by Aft1 also requires the regulatory proteins Fra1-Fra2 and the cytosolic-nuclear monothiol glutaredoxins Grx3 and Grx4, which are essential for the nuclear export of Aft1 in response to iron sufficiency (Kumanovics et al., 2008; Ojeda et al., 2006; Pujol-Carrion et al., 2006). The regulatory role of Grx3/4 is functionally conserved in fungi that utilize iron-regulated transcription systems unrelated to those from S. cerevisiae (Haas et al., 2008; Kaplan and Kaplan, 2009; Mercier and Labbe, 2009).

Grx3/4 belong to the large thioredoxin (Trx) fold family and are composed of an N-terminal Trx and a C-terminal monothiol
glutaredoxin (Grx) domain (Herrero and de la Torre-Ruiz, 2007; Lillig et al., 2008). Although the Grx3/4 subfamily of multidomain monothiol glutaredoxins is conserved in eukaryotes, no universal function has been assigned to this family so far. In contrast to most members of the Grx protein family that catalyze diithio-di-sulfide redox reactions, monothiol Grx proteins rarely possess oxidoreductase activity. Instead, after in vitro reconstitution and upon overexpression in *Escherichia coli*, they are capable of binding a bridging [2Fe-2S] cluster utilizing the active-site cysteine residue of the Grx domain and glutathione (GSH) as ligands (Li et al., 2009; Picciocchi et al., 2007). The existence of this unusual Fe/S center under physiological conditions, however, has not been demonstrated, and its functional role has remained unclear.

Here, we have used yeast as a model to define an essential role of Grx3/4 in intracellular iron trafficking. Depletion of Grx3/4 led to functional impairment of virtually all iron-dependent processes, including heme biosynthesis, mitochondrial and cytosolic Fe/S protein biogenesis, and the formation of di-iron centers in mitochondria and the cytosol, eventually leading to the loss of cell viability. We provide evidence for the in vivo binding of a bridging Fe/S cluster to Grx3/4 and we assign a crucial physiological function to this cofactor both in cytosolic iron trafficking and as an iron sensor. Thus, the conserved cytosolic monothiol glutaredoxins use their bound Fe/S cofactor for a general role in intracellular iron trafficking.

RESULTS

Deficiency in Grx3/4 Is Associated with Defects in Iron-Dependent Enzymes

To facilitate the functional analysis of Grx3/4, we constructed a regulatable yeast strain (Gal-GRX4; strain background W303-1A; see Table S1 available online) in which GRX3 was deleted and GRX4 was expressed under the control of the glucose-repressible GAL-L promoter. Upon Grx4 depletion, Gal-GRX4 cells failed to grow on both fermentable and nonfermentable carbon sources (Figure 1A; see below). Likewise, double deletion of GRX3-GRX4 was lethal in the W303 strain background, distinguishing these cells from strain BY4742 grx3/4Δ, which shows only severely retarded growth (Figure 1A) (Ojeda et al., 2006). The strong effect of Grx3/4 deficiency on cell viability is not explained by their role in iron regulation, since Aft1 is not essential under iron-replete conditions (Kaplan and Kaplan, 2009). These data and the general conservation of Grx3/4 in eukaryotes suggest that these proteins perform a so far unknown, important function.

Gal-GRX4 cells were used to investigate the immediate consequences of Grx3/4 deficiency. Gal-GRX4 cells were cultivated in minimal medium supplemented with glucose and iron chloride to gradually deplete Grx4 (Figures 1B and 1C). A strong activation of the Aft1-dependent FET3 gene was observed using a FET3 promoter-GFP fusion as a reporter (Figure 1B) (Ojeda et al., 2006). Surprisingly, the activities of the mitochondrial Fe/S protein aconitase and cytosolic catalase, a heme-containing protein, drastically decreased, despite the presumed sufficient cellular iron supply. These effects resemble those upon depletion of Ssq1, a component of the iron-sulfur cluster (ISC) assembly machinery, even though the changes occurred later as a result of slower depletion of Ssq1 (strain Gal-SSQ1). Grx3/4 deficiency was associated with a severe activity loss of respiratory complexes II (succinate dehydrogenase) and IV (cytochrome oxidase) but was fully complemented by expression of GRX4 from a plasmid (Figure 1D). Likewise, low activities of both aconitase and respiratory complexes III and IV were observed in BY4742 grx3/4Δ cells (Figure S1A), consistent with our earlier observation of an impaired 55Fe/S cluster incorporation into aconitase (Ojeda et al., 2006). Immunostaining of cell extracts from Grx4-depleted Gal-GRX4 cells and BY4742 grx3/4Δ cells further revealed changes in the levels of several iron-containing proteins, including the aconitase-type Fe/S proteins Aco1 and Leu1, ferrochelatase Hem15, and the core mitochondrial ISC assembly protein Iso1 (Figure 1C). These changes of protein levels correlate with those of the transcriptome of both iron-deprived and ISC machinery–compromised cells (Hausmann et al., 2008; Shakoury-Elizeh et al., 2004). In contrast, other iron-dependent proteins, such as succinate dehydrogenase subunit 2 (Sdh2) and the ubiquinone biosynthesis enzyme Coq7, were hardly changed and behaved similarly to the noniron proteins mitochondrial cytochrome oxidase subunit 4 (Cox4) and porin (Por1), cytosolic Hsp70, and ribosomal subunit Rps3. Together, these findings indicate that Grx3/4-deficient cells develop severe defects in several mitochondrial and cytosolic iron-dependent proteins, despite the induction of the Aft1-dependent iron uptake system. Notably, these global iron-related defects are not detected upon the constitutive activation of Aft1 (Hausmann et al., 2008; Ihrig et al., 2010), suggesting that these consequences of Grx3/4 deficiency occur independently of Aft1 and a deregulated iron homeostasis.

Deficiency in Grx3/4 Impairs the De Novo Synthesis of Cellular Fe/S Clusters and Heme

We asked whether the decreased Fe/S protein activities in Grx3/4-depleted cells might be explained by an impaired de novo synthesis of their Fe/S clusters and addressed this problem by using an established 55Fe radiolabeling and immunoprecipitation assay (Molik et al., 2007). First, the essential cytosolic Fe/S proteins Rli1, Dre2, and Nar1 were analyzed by expressing these proteins from a high-copy vector in wild-type and Gal-GRX4 cells. Fe/S cluster insertion into all three Fe/S protein targets was decreased 4–10-fold upon Grx4 depletion (Figure 2A). The amount of Dre2 in Gal-GRX4 cells was comparable to that in wild-type cells, indicating a specific Fe/S cluster assembly defect (Figure 2A, inset). In the case of Rli1 and Nar1, protein levels were diminished, likely suggesting that the apoforms of these Fe/S proteins were degraded. Similar apoprotein instability is frequently observed upon strong defects in Fe/S protein biogenesis (Balk et al., 2004). Analysis of 55Fe incorporation into the mitochondrial Fe/S proteins Bio2 (biontin synthase) and Iv3 (dihydroxyacid dehydratase) and the essential mitochondrial ISC scaffold protein Iso1 revealed an up to 4-fold lower 55Fe incorporation upon Grx4 depletion (Figures 2B and 2C). Protein levels of Bio2, Iv3, and Iso1 did not change upon depletion of Grx4 (Figures 2B and 2C, insets). These findings indicate a general impairment in the de novo assembly of Fe/S proteins upon depletion of Grx3/4.

In principle, the defect of Fe/S protein maturation upon Grx3/4 depletion could be explained by a primary mitochondrial Fe/S
protein assembly defect, because mitochondria are involved in generation of all cellular Fe/S proteins (Lill and Mühlenhoff, 2008). However, we note that the observed effects were less severe in mitochondria compared to the cytosol. To directly test the functionality of the mitochondrial ISC assembly machinery, we made use of an anaerobic in vitro system analyzing the capacity of mitochondrial detergent extracts to support Fe/S cluster insertion into the apoform of purified Yah1, a [2Fe-2S] ferredoxin (Molik et al., 2007). Apo-Yah1 was incubated with mitochondrial lysates and $^{55}$Fe. The radiolabeled Yah1 holoprotein was bound to Q-sepharose, and the amount of radioactivity associated with holo-Yah1 was quantified by scintillation counting. Remarkably, extracts derived from Grx3/4-depleted mitochondria were even more competent in synthesizing the Fe/S cluster on Yah1 than Grx4-complemented Gal-GRX4 cells (Figure 2D). This finding strongly suggests that the mitochondrial
ISC assembly system is functional in Grx3/4-depleted cells, rendering it likely that the decreased Fe/S cluster incorporation into apoproteins is explained by impaired iron supply.

Consistent with this idea, 55Fe insertion into heme was 5.5-fold lower in Grx4-depleted Gal-GRX4 cells, compared with wild-type cells or Gal-GRX4 cells complemented by GRX4 (Figure 2E). This diminished heme synthesis activity may explain the loss of function of heme-dependent enzymes, such as catalase and cytochrome oxidase, upon depletion of Grx3/4 (see above). In summary, Grx3/4-depleted cells are strongly impaired in both cellular Fe/S protein maturation and heme biosynthesis. Such defects are not observed in Aft1-activated cells (Hausmann et al., 2008; Ihrig et al., 2010).

Deficiency in Grx3/4 Leads to Impairment of Di-Iron Enzymes Despite Cytosolic Iron Overload

The strong decrease of cellular Fe/S clusters and heme in Grx3/4-deficient cells is somewhat paradoxical, because these cells are expected to accumulate iron as the result of a constitutively activated cellular iron uptake system (Ojeda et al., 2006). To verify this idea, we measured the cellular iron content by ICP-MS analysis of wild-type or Grx4-depleted Gal-GRX4 cells grown in minimal medium supplemented with 10 μM FeCl3. Total cellular iron increased 6-fold upon depletion of Grx4 (Figure 3A). The level of chelatable iron increased similarly and was mainly present as Fe2+ (Figure S2). Cellular levels of other metals, with the exception of Zn (3-fold higher), were hardly changed. In contrast, mitochondrial iron levels were up to 2.3-fold lower in Grx4-depleted Gal-GRX4 cells compared with wild-type cells (Figure 3B). Mitochondria from BY4742 grx3/4D cells contained even 7.5-fold less iron. Mitochondrial Mn, Co, and Zn levels were hardly altered, but Cu changed in parallel to iron. The decrease in mitochondrial iron levels in Grx3/4-depleted cells is the more remarkable, because cells with mitochondrial Fe/S protein assembly defects usually display strongly elevated mitochondrial iron levels (Lill and Mühlenhoff, 2008). The fact that this was not observed, despite high levels of total cellular iron, indicates a defective delivery of iron to mitochondria in Grx3/4-deficient cells.

A reasonable explanation for these general defects in iron handling in the absence of Grx3/4 may be a sequestration of iron into the vacuole, the major iron storage compartment in fungi (Kaplan and Kaplan, 2009; Philpott and Prochenko, 2008). To test this idea, we varied the amount of Ccc1, the major importer of divalent metals into the vacuole. Deletion of CCC1 did not restore growth of Grx4-depleted Gal-GRX4 cells and did not increase the low enzyme activities of aconitase and catalase (Figure 3C, top panel, and Figure 3D). Similarly,
overproduction of Ccc1 failed to restore growth (Figure 3C, middle panel). These data suggest that the accumulated iron is not stored in the vacuole.

Grx4-depleted Gal-GRX4 cells failed to grow under anaerobic conditions (Figure 3C, bottom panel). Thus, reactive oxygen species (due to increased iron levels) are not responsible for the lethal phenotype of Grx3/4-depleted cells. Moreover, oxidized glutathione levels (GSSG; measured under anaerobic conditions) were below the detection limit (not shown). Rather, reduced glutathione (GSH) was strongly elevated in Grx4-depleted Gal-GRX4 cells, but not in BY4742 grx3/4Δ (Figure 3E). Together, these results and the predominant presence of iron in its ferrous form (Figure S2) indicate that reducing conditions prevail in Grx3/4-deficient cells.

Figure 3. Deficiency in Grx3/4 Results in Cytosolic Iron and GSH Accumulation
(A and B) The metal content of (A) wild-type (WT) and Gal-GRX4 cells (depleted for 64 hr) and (B) mitochondria isolated from the indicated strains was determined by ICP-MS.

(C) The indicated strains lacking (ccc1Δ) or overproducing Ccc1 (Ccc1↑) were cultivated in SD medium for 40 hr. Ten-fold serial dilutions were spotted onto SC agar plates containing glycerol (Glyc) or glucose (Glc), and were cultivated at 30°C under aerobic (+O2) or anaerobic (−O2) conditions.

(D) WT, Gal-GRX4 and Gal-GRX4/ccc1Δ cells were grown in SD medium for 64 hr, and aconitase and catalase enzyme activities were determined.

(E) GSH levels were determined in cell extracts from WT, BY4742 grx3/4Δ, Gal-GRX4 (depleted for 40 hr or 64 hr) and Gal-GRX4 cells expressing GRX4 from a plasmid (+Grx4).

The experiments presented above showed a maturation defect in cellular Fe/S and heme proteins in Grx3/4-deficient cells despite a cytosolic iron accumulation. Because this indicated a defective delivery of iron, we asked whether other iron-dependent enzymes were affected by Grx3/4 deficiency. First, the iron status of ribonucleotide reductase (Rnr), a cytosolic diferric-tyrosyl radical enzyme essential for deoxyribonucleotide synthesis, was analyzed (Perlstein et al., 2005). Upon depletion of Grx4 in Gal-GRX4 cells, the protein levels of Rnr2 decreased slightly (Figure S3). Nevertheless, the specific activity of Rnr was 6-fold lower compared with wild-type cells (Figure 4A). This was likely due to inefficient metallation, because 55Fe insertion into Rnr in vivo (followed by immunoprecipitation of subunit Rnr2) was 5–7-fold decreased in Grx4-depleted cells (Figure 4B).
As a second di-iron protein, we analyzed the mitochondrial mono-oxygenase Coq7, which catalyzes the hydroxylation of demethoxyubiquinol (DMQ₆), the penultimate reaction of ubiquinone (CoQ₆) biosynthesis (Tran et al., 2006). HPLC analysis of mitochondrial lipid extracts revealed diminished CoQ₆ and increased DMQ₆ levels upon Grx4 depletion (Figure 4C). This effect was reversed to wild-type ratios by expression of GRX4 from a plasmid (Figure 4D). The simultaneous accumulation of the substrate (DMQ₆) and decrease of the product (CoQ₆) of the enzyme Coq7 indicates a diminished activity of this di-iron mono-oxygenase upon depletion of Grx3/4. Collectively, these data demonstrate that a Grx3/4 deficiency causes a severe defect in cellular di-iron enzymes.

Are the observed defects in Grx3/4-depleted cells specific for iron-dependent proteins? This question was addressed by analyzing the activities and protein levels of several metal-dependent enzymes. The in-gel activities of the endogenous Cu/Zn- and Mn-dependent superoxide dismutases (Sod1 and Sod2, respectively) remained unchanged in Grx4-depleted Gal-GRX4 cells (Figure 4E). In marked contrast, both the level and activity of ectopically expressed iron-only superoxide dismutase (FeSod) from E. coli strongly declined. Furthermore,
the Zn-dependent alcohol dehydrogenase (ADH) was 3-fold more active upon Grx4 depletion (Figure S1B). This increased ADH activity is characteristic for a switch toward fermentative metabolism and is typically observed upon iron deprivation. The normal function of several metal-reliant enzymes indicates that the described defects in Grx3/4-depleted cells are specific for iron-related processes. We conclude that Grx3/4 perform an essential role in cellular iron trafficking, in addition to and independently of their nonessential function in iron uptake regulation.

**Grx3/4 Assemble a Bridging Fe/S Cluster Independently of the CIA Machinery**

For a function of monothiol glutaredoxins in cellular iron trafficking, iron binding may be a necessary prerequisite. Recent in vitro studies have shown that various glutaredoxins, including yeast Grx3/4, can bind a bridging, GSH-ligated [2Fe-2S] cluster (Bandyopadhyay et al., 2008; Berndt et al., 2007; Johansson et al., 2007; Li et al., 2009). We asked whether this unusual Fe/S cofactor is of physiological relevance and can be observed in a native environment using the 55Fe radiolabelling assay. Significant amounts of 55Fe were coimmunoprecipitated with anti-Grx4-antibodies from cell extracts derived from wild-type cells (strain BY4742), whereas only background levels were observed in grx3/4A cells (Figure 5A). Similar amounts of 55Fe were coimmunoprecipitated from both grx3A and grx4A cells, indicating that the homologous Grx3 and Grx4 bind iron with similar efficiency and independently of each other. The amount of Grx4-bound 55Fe was unchanged in both wild-type cells of our standard strain W303-1A, and, remarkably, under anaerobic or oxidative stress conditions prevailing after addition of H2O2 or upon deletion of SOD1 (sod1Δ cells), indicating that iron binding to Grx3/4 was insensitive to oxidative stress (Figure 5B).

To analyze whether the iron bound to Grx3/4 is part of an Fe/S cluster, we investigated the requirement of 55Fe binding for components of Fe/S protein biogenesis (Lill and Mühlenhoff, 2008). We first analyzed the role of the cytosine desulphurase complex Nfs1-Isd11, which serves as the sulfur donor for Fe/S protein biogenesis. For regulated depletion of Nfs1 or Isd11, we used Gal-NFS1 or Gal-ISD11 cells, respectively. 55Fe binding to Grx4 (Figure 5C) and to the cytosolic Fe/S protein Leu1 as a control (Figure S4) declined to background levels upon depletion of either Nfs1 or Isd11. The same result was seen in Gal-ISU1Δisu2 cells depleted for the core ISC scaffold proteins Isu1/2. No recovery of iron binding to Grx3/4 was observed when a cytosolic-nuclear version of Nfs1 (ΔN-Nfs1) was produced in Gal-NFS1 cells lacking mitochondrial Nfs1 (Figure 5C). We found during these experiments that the Grx4 protein levels decreased upon Nfs1 depletion, whereas the Grx3 levels remained unchanged. Although the reason for this specific decrease is unknown, we note that the remaining Grx3 did not bind 55Fe above background levels, unlike in grx4Δ cells (Figure 5A). The dependence of iron binding to Grx3/4 on core components of the mitochondrial ISC assembly system demonstrates that Grx3/4 bind an Fe/S cluster in vivo.

Does incorporation of the Fe/S cluster into Grx3/4 also require the cytosolic iron-sulfur protein assembly (CIA) system (Lill, 2009)? Depletion of the essential CIA components Nbp35 and Dre2 in the respective GAL promoter-exchange mutants significantly increased rather than impaired 55Fe binding to Grx3/4 (Figure 5D), whereas hardly any 55Fe binding was observed to Leu1 as a control (Figure S4). This surprising finding suggests that incorporation of the Fe/S cluster into Grx3/4, although requiring mitochondrial Nfs1-Isd11-Isu1, does not depend on the cytosolic CIA machinery.

The noncanonical pathway used for Fe/S cluster assembly on Grx3/4 may be due to the special nature of this cofactor, with a conserved Cys and GSH serving as ligands of a [2Fe-2S] cluster bridged between two Grx monomers, as observed after in vitro reconstitution or after overexpression in *E. coli* (Li et al., 2009; Picciochi et al., 2007). We therefore sought to obtain in vivo evidence for this configuration. First, we determined which ligands might coordinate the Fe/S cluster. The importance of the conserved active-site cysteine residues C171 in the C-terminal Grx domain and C37 in the N-terminal Trx domain (Figure 5E, top) for 55Fe binding was tested in wild-type cells expressing a plasmid-encoded Myc-tagged Grx4 (Grx4-Myc) in which these residues were mutated to serine (mutant proteins C171S and C37S) or alanine (C171A). Myc-tagged wild-type Grx4 was fully functional (see below) and was produced at similar levels as the mutant proteins (Figure 5E, bottom). Mutant proteins C171S and C171A did not bind 55Fe above background levels (Figure 5E, middle). In contrast, mutant protein C37S bound wild-type amounts of 55Fe, suggesting that iron is bound via C171. Second, the importance of GSH for 55Fe binding to Grx3/4 in vivo was tested using the GSH synthesis-deficient mutant gshΔ, that can be depleted for GSH upon growth in media lacking GSH. Although significant amounts of 55Fe were bound to Grx4-Myc in the presence of exogenously added GSH, only background levels of 55Fe were found in GSH-deprived cells (Figure 5F). Together, these findings suggest that the active-site C171 of the Grx domain and GSH serve as ligands of the Grx3/4-bound Fe/S cluster.

Finally, we tried to find in vivo evidence for the presence of a bridging Fe/S cluster on Grx3/4. A yeast strain was constructed that simultaneously expressed C-terminally HA- and Myc-tagged Grx4. Cell extracts were subjected to immunoprecipitation with anti-HA or anti-Myc immunobeads followed by immunoblotting. Immunoprecipitation with anti-HA antibodies led to coisolation of Grx4-HA and a smaller amount of Grx4-Myc (Figure 5G). The same result, yet with inversed intensities, was observed using anti-Myc beads, whereas no cross-reacting bands were visible in wild-type cells. In cells expressing a C171S mutant Grx4-Myc and wild-type Grx4-HA, immunoprecipitation was far less efficient but still detectable. These observations document the importance of residue C171 for efficient Grx4 dimer formation and thus are consistent with the idea of a bridging Fe/S cluster between two Grx monomers. Collectively, our findings suggest that, under physiological conditions, Grx3/4 bind a bridging Fe/S cluster that is coordinated by the active-site cysteine and GSH. These results are consistent with the structure of glutaredoxins reconstituted in vitro.

**The Grx3/4-Bound Fe/S Cluster Is Important for Iron Metabolism**

Is the Grx3/4-bound Fe/S cluster important for the function of Grx3/4 in iron trafficking and is this cofactor also required for
DISCUSSION

Our analysis of the cytosolic-nuclear monothiol glutaredoxins Grx3 and Grx4 provides strong evidence for an essential and general role of these proteins in intracellular iron trafficking, a so-far poorly understood process (Figure S5). This function of Grx3/4 is additional to their involvement in Aft1-dependent iron sensing, because the phenotypes of Grx3/4-depleted and Aft1-activated cells differ markedly in that only the former cells display global and severe biochemical defects in multiple iron-dependent enzymes (Hausmann et al., 2008; Ihrig et al., 2010). For instance, the function of cellular Fe/S-containing, heme-containing, and other iron-containing proteins is strongly impaired upon depletion of Grx3/4 but is not affected when Aft1 is activated. Many of these proteins are involved in essential processes, such as DNA synthesis and ribosome function, explaining why Grx3/4-depleted cells eventually lose their viability. The two functions of Grx3/4 in iron trafficking and sensing are mediated by their crucial cofactor, a GSH-ligated and bridging [2Fe-2S] center. Destruction of this Fe/S center elicits a highly similar phenotype as the depletion of the Grx proteins. The observed biochemical consequences of Grx3/4 depletion are specific for iron, because the functions of other metal-dependent enzymes, such as Cu,Zn- and Mn-dependent superoxide dismutases, were not affected. Together, our findings demonstrate a crucial role of Grx3/4 in intracellular iron delivery from the cytosolic labile iron pool to virtually all iron-binding proteins or pathways in the cell (Figure S5). Hence, Grx3/4 represent long-sought general factors that facilitate the proper assembly of various iron centers in proteins and cofactors. Grx3/4 are unlikely to make specific and direct contacts with the potential iron-dependent targets. Therefore, they may not function as classical metallo-chaperones that deliver their metal specifically to individual targets (Finney and O’Halloran, 2003; Lyons and Eide, 2007). Rather, the iron activated by Grx3/4 may be utilized by dedicated assembly factors, such as components of the mitochondrial ISC or cytosolic CIA systems (Lill and Mühlenhoff, 2008) or the cytosolic ferritin-specific iron chaperone PCBP1 (Shi et al., 2008), for proper insertion into recipient apo-proteins.
Several observations make it unlikely that the severe iron trafficking defect in the absence of Grx3/4 is related to oxidative stress damage, even though, in principle, Fenton chemistry is possible in Grx3/4-depleted cells as a result of their massive accumulation of intracellular ferrous iron. First, the lethal phenotype of Grx3/4-depleted cells was not cured under anaerobic conditions, and the activities of iron-dependent enzymes were not recovered. Second, BY4742 grx3/4Δ cells display only weak signs of oxidative stress, possibly because the cytosolic redox balance in yeast is maintained by the dithiol glutaredoxins Grx1 and Grx2 (Rodriguez-Manzaneque et al., 1999). Third, binding of the essential Fe/S center to Grx3/4 was rather insensitive to oxidative stress.
Finally, Grx3/4-depleted cells show hyper-accumulation of reduced GSH with no signs for an increase in oxidized GSSG. The observed cytosolic iron trafficking defect in Grx3/4-deficient cells cannot be explained by low levels of iron in the cytosol. Rather, these cells accumulate high amounts of chelatable iron, which, in all likelihood, is predominantly present in the cytosolic compartment. An iron efflux into the vacuole, the main storage organelle for excess iron in plants and fungi (Kaplan and Kaplan, 2009; Philpott and Protchenko, 2008), is unlikely because the deletion of the vacuolar divalent metal transporter gene CCC1 did not rescue any of the defects of Grx3/4-depleted cells. Mitochondria, which accumulate iron under conditions of hampered Fe/S protein biogenesis (Lill and Mühlenhoff, 2008), displayed diminished iron levels in Grx3/4-depleted cells. These data suggest that the surplus iron in Grx3/4-deficient cells is not bioavailable for efficient funnelling into iron-dependent processes.

Our $^{55}$Fe radiolabeling experiments demonstrate that the Fe/S cluster of monothiol glutaredoxins, as initially characterized in vitro or after overexpression in E. coli (Li et al., 2009; Picciocchi et al., 2007), is also assembled in its native environment and is of utmost physiological relevance. The essential involvement of the core components of the mitochondrial ISC assembly system provides strong evidence that the bound iron is part of an Fe/S cluster and that the sulfide ions of the [2Fe-2S] cluster are derived from the mitochondrial cysteine desulfurase complex Nfs1-Isd11. Our in vivo results support the unusual bridging character of the Fe/S cluster, which is coordinated by the Grx domain active-site Cys171 and GSH. Hence, GSH performs a dual role in cellular iron metabolism. As part of the Grx3/4 complex, it is central for intracellular iron delivery and sensing, and as a component of the ISC export machinery, it cooperates with the mitochondrial ABC transporter Atm1 both in the assembly of cytosolic-nuclear Fe/S proteins and in mitochondrial-cytosolic iron homeostasis (Lill, 2009; Sipos et al., 2002). Surprisingly, the Grx3/4 Fe/S cofactor is assembled independently of the CIA machinery, an observation that differs from all other known, nonscaffold cytosolic Fe/S proteins analyzed so far (Lill and Mühlenhoff, 2008). This unique assembly mode is likely related to the special type of Fe/S cluster on Grx3/4. The CIA machinery–independent maturation of Grx3/4 is consistent with earlier observations that this machinery is not important for cellular iron regulation in yeast (Hausmann et al., 2008; Kaplan and Kaplan, 2009).

Although it is clear that the bridging Fe/S cluster of Grx3/4 is crucial for iron trafficking, the question remains how it may function. Currently, it cannot be discerned whether the iron moiety of the Fe/S cluster is mobilized and used for insertion into iron-dependent proteins, or whether the Fe/S cluster plays a more indirect role by activating cytosolic iron for specific trafficking. In any case, it is well known for Fe/S clusters that they can reversibly loose one of their iron ions, the most prominent example being aconitase (Beinert et al., 1997). Thus, the Fe/S cluster of Grx3/4 may not be fully disassembled during its action in iron delivery. The role of the Grx3/4 Fe/S center in iron trafficking is clearly distinct from the proposed function of other monothiol glutaredoxins, as Fe/S scaffold (or Fe/S carrier) proteins facilitating Fe/S cluster transfer to target apoproteins (Bandyopadhyay et al., 2008; Picciocchi et al., 2007). Thus, monothiol glutaredoxins may use diverse mechanisms for maturation of iron-containing proteins.

In principle, the essential requirement of the Grx active-site C171 for iron homeostasis could be explained by thiol-dependent redox chemistry. However, with few exceptions, monothiol glutaredoxins, including yeast Grx3/4, are inactive in GSH-dependent redox reactions in vitro, and there is little evidence that monothiol glutaredoxins perform specific catabolic functions (Herrero and de la Torre-Ruiz, 2007). In addition, BY4742 grx3/4Δ cells display wild-type glutathione reductase activity (Rodríguez-Manzaneque et al., 1999). Most convincingly, the high-copy suppressor phenotype of the C171S mutation in the Grx domain excludes impaired thiol-related (redox) chemistry as an explanation for the functional defects prevailing in Grx3/4-depleted cells. High levels of the C171S mutant protein, but not the corresponding Ala variant, supported wild-type growth and almost fully restored the activities of iron-dependent proteins. This finding suggests that the C171S protein is still capable of coordinating the Fe/S cofactor, as this is known for other proteins with Cys to Ser mutations (da Silva and Williams, 2001; Johnson et al., 2005). However, iron binding to the C171S mutant protein may be more labile, thus precluding its detection by our in vivo $^{55}$Fe labeling and immunoprecipitation assay. As a consequence, the amount of bound Fe/S cluster may be too low to support Grx4 function in Aft1 sensing. Consistent with this interpretation, in reconstituted human Grx2 and poplar GrxC1, the [2Fe-2S] cluster is coordinated in a binding pocket at the interface of two Grx monomers by the two active-site Cys residues and two noncovalently bound GSH molecules (Berndt et al., 2007; Johansson et al., 2007; Rouhier et al., 2007). The replacement of cysteine with serine may still provide a low-affinity iron binding site.

Grx3/4 play a role in iron-uptake regulation by cooperating with the iron-responsive transcription factor Aft1, the major regulator of cellular iron uptake in yeast (Philpott and Protchenko, 2008). In fact, the physical interaction between Grx3/4 and Aft1 was the initial evidence linking Grx3/4 and cellular iron homeostasis (Ojeda et al., 2006; Pujol-Carrion et al., 2006). Our study shows that the Grx3/4-bound Fe/S cluster functions as the long-sought iron sensor that communicates the cytosolic iron status to Aft1 (Figure S5). Removal of the bound Fe/S cluster (e.g., by mutation of the active-site Cys of the Grx domain or by depletion of GSH) abolished the ability of Grx3/4 to regulate Aft1. The iron-sensing role of Grx3/4 appears to be conserved in other fungi that utilize iron-responsive transcription factors, even if they are structurally unrelated to S. cerevisiae Aft1 (Kaplan and Kaplan, 2009; Mercier and Labbe, 2009).

The crucial function of Grx3/4 and its unique Fe/S center in intracellular iron trafficking and sensing opens an additional chapter of the intimate connection between the iron metabolism and Fe/S protein biogenesis. Not only is the iron metabolism regulated by (mitochondrial) Fe/S protein biogenesis (Kaplan and Kaplan, 2009), but, conversely, iron insertion into Fe/S proteins requires its activation by the Fe/S cluster-containing Grx3/4. The mutual dependence of these two important pathways is further linked to the cellular redox balancing systems via GSH, which is crucial for both iron insertion into iron-dependent factors and regulation of cellular iron homeostasis (Lill and Mühlenhoff, 2008). These considerations reserve a central role.
for monothiol glutaredoxins Grx3/4 and their bound Fe/S center in cellular metabolism. The central function of Grx3/4 in intracellular iron trafficking may provide the explanation for the conservation of this protein family in eukaryotes. The discovery of this glutaredoxin function now paves the way toward mechanistic investigations of how iron is specifically activated for incorporation into iron-requiring processes.

**EXPERIMENTAL PROCEDURES**

### Yeast Strains, Cell Growth, and Plasmids

Yeast strains used in this study are listed in Table S1. If not stated otherwise, the strain background is W303-1A. Cells were cultivated in minimal medium containing all recommended supplements (SC), 2% w/v glucose (SD), or 3% w/v glycerol (Sherman, 2002). Iron-replete media were supplemented with 50 μM FeCl3. Media for anaerobic growth were supplemented with Tween80, ergosterol, and methionine. Gal-GRX4 cells were depleted for Grx4 to critical levels by cultivation in SD medium for 40–64 hr prior to analysis. Repression of other conditional Gal-strains was performed as described in Table S2, gsh1Δ cells were cultivated in the presence or absence of 1 mM GSH for 64 hr prior to analysis (Sipos et al., 2002). Wild-type and site-directed mutant Grx4 proteins were expressed in yeast with a C-terminal Myc-tag from plasmid pCM189 (Ojeda et al., 2006) under the control of the tetO2 or the endogenous GRX4 promoter (Table S2).

### Biochemical Analyses In Vivo

Radio labeling of yeast cells with 55FeCl3 (Perkin-Elmer) and measurement of 55Fe incorporation into proteins by immunoprecipitation and into heme was performed as described elsewhere (Pierik et al., 2009). Antibodies were raised in rabbits against recombinant proteins expressed in E. coli. 55Fe-labeling of added apo-ferredoxin in detergent extracts of isolated mitochondria was performed as described elsewhere (Molik et al., 2007). DMQ and CoQ were quantified according to Tran et al. (2006), and GSH and GSSG were quantified according to Edlidge and Davis (1987).

### Miscellaneous Methods

The following published methods were used: manipulation of DNA and PCR (Sambrook and Russel, 2001), preparation of yeast mitochondria (Molik et al., 2007), immunological techniques (Harlow and Lane, 1988), FET3 promoter assays (Molik et al., 2007), determination of cellular and mitochondrial metal contents by ICP-MS (Mühlenhoff et al., 2003), and enzyme activities of iron proteins, ADH, and MDH (Molik et al., 2007), ribonucleotide reductase (Wang et al., 2009), and superoxide dismutase (Flohe and Otting, 1984).

### SUPPLEMENTAL INFORMATION

Supplemental information includes two tables and five figures and may be found with this article online at doi:10.1016/j.cmet.2010.08.001.

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**REFERENCES**


