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Aza-Tryptamine Substrates in Monoterpene Indole Alkaloid Biosynthesis

Hyang-Yeol Lee,1 Nancy Yerkes,1 and Sarah E. O’Connor1,*

1Massachusetts Institute of Technology, Department of Chemistry, 77 Massachusetts Avenue, Cambridge, MA 02139, USA
*Correspondence: soc@mit.edu
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SUMMARY

Biosynthetic pathways can be hijacked to yield novel compounds by introduction of novel starting materials. Here we have altered tryptamine, which serves as the starting substrate for a variety of alkaloid biosynthetic pathways, by replacing the indole with one of four aza-indole isomers. We show that two aza-tryptamine substrates can be successfully incorporated into the products of the monoterpene indole alkaloid pathway in Catharanthus roseus. Use of unnatural heterocycles in precursor-directed biosynthesis, in both microbial and plant natural product pathways, has not been widely demonstrated, and successful incorporation of starting substrate analogs containing the aza-indole functionality has not been previously reported. This work serves as a starting point to explore fermentation of aza-alkaloids from other tryptophan- and tryptamine-derived natural product pathways.

INTRODUCTION

Aza-indoles are important pharmacophores that mimic the indole scaffold. These unique heterocycles are isosteric to the indole moiety, but display improved water solubility and unique hydrogen bonding properties (Merour and Joseph, 2001). Several natural products contain aza-indole moieties (Perry et al., 1994; Schumacher and Davidson, 1995; Trimurtulu et al., 1994), and the antitumor properties of a synthetic aza analog of the indole natural product rebeccamycin have also been examined (Marminon et al., 2003) (Figure 1). Moreover, a number of synthetic drug candidates contain the aza-indole scaffold (Leblanc et al., 2009; Tsai et al., 2008; Wang et al., 2003) (Figure 1). We synthesized aza-tryptamines 1-4 and demonstrated that two of these compounds, 1 and 4, could serve as surrogates for tryptamine in the monoterpene indole alkaloid biosynthetic pathway to generate aza-alkaloids (Figure 2). Notably, these two deactivated aza-indole substrates were successfully utilized in an enzymatic Pictet-Spengler reaction, a key step in this biosynthetic pathway.

A general synthetic procedure for the production of 4-, 5-, 6-, and 7-aza-tryptamine 1-4 was developed. Ortho-iodoanilines, which were either commercially available or were prepared by copper-catalyzed halogen exchange reaction using the diamine ligand trans-N,N'-dimethylcyclohexane (Klapars and Buchwald, 2002) (Figure 2), could be subjected to palladium-catalyzed heteroannulation with a trimethylsilylated alkyne, followed by the acid hydrolysis to give all four 3-substituted aza-indole products in good yields (Pullagurla et al., 2005; Ujjainwalla and Warner, 1998) (Figure 2).

We investigated the use of aza-tryptamines in a Pictet-Spengler reaction, which is widely used in synthetic organic chemistry (Royer et al., 2004), and also serves as the first committed enzymatic step in a number of alkaloid biosynthetic pathways (O’Connor and Maresh, 2006) (Figure 3). In the Pictet-Spengler reaction, tryptamine 5 reacts with an aldehyde to form an iminium ion. The nucleophilic indole then attacks the iminium ion, resulting in the formation of a tetrahydro-beta-carboline product (Figure 3) (Maresh et al., 2008). Because aza-tryptamines are more electron deficient than tryptamine 5, the efficiency of the Pictet-Spengler reaction is expected to be greatly decreased with substrates 1-4. However, when incubated with both the aldehyde secologanin 6 and the “Pictet-Spenglerase” strictosidine synthase from the monoterpene indole alkaloid biosynthetic pathway in Catharanthus roseus, 4- and 7-aza-tryptamines 1 and 4 were converted into the corresponding aza-tetrahydro-beta-carboline products 7 and 10. No product formation was observed with 5- and 6-aza-tryptamines 2 and 3. Overall, the catalytic efficiency of 7-aza-tryptamine 4 (kcat: 11.8 ± 0.4 min⁻¹ and KM: 65 μM ± 9.1) was approximately 100-fold lower compared with tryptamine 5 (kcat: 96 ± 4.7 min⁻¹ and KM: 5 μM ± 1.4), and 4-aza-tryptamine 1 (kcat: 10.2 ± 0.3 min⁻¹ and a KM: 151 μM ± 9.3) was 280-fold reduced relative to tryptamine 5.

Strictosidine synthase, in general, has been shown to display a greater tolerance for substituents at the 4 and 7 positions of the indole ring compared with substituents at the 5 and 6 positions (Figure 3) (Bernhardt et al., 2007; Loris et al., 2007; McCoy et al., 2006), a pattern consistent with the reactivity observed with aza-tryptamines 1-4. Although steric factors did not seem likely to play a major role in the recognition of the aza-tryptamines—the size of nitrogen (van der Waals radius 1.55 Å) is similar to carbon (1.70 Å)—hydrogen bonding properties of the aza-indoles could affect whether the substrate interacts productively with the enzyme. Given that 7-aza-tryptamine 4 is the best substrate of the series, it is notable that the 7-position of the indole ring is pointed toward the solvent, away from any protein contact, while 4, 5, and 6 positions are located in the interior of the protein (Ma et al., 2006).

We asked whether the differing electronic properties of 1-4 also played a role in the relative rates of enzymatic reaction, since the Pictet-Spengler reaction depends upon the electron density of the aromatic amine substrate. After formation of the iminium ion, the Pictet-Spengler reaction can proceed by one
and 7-aza-indoles (Zhang et al., 2002), the electronic density of reports that 4- and 6-aza-indoles are more deactivated than 5-
(137.5 ppm). Although these data are consistent with previous the C2 position of 7-aza-tryptamine
4-aza-tryptamine
1
electron rich, followed by 5-aza-tryptamine
2
and
3
of two mechanisms. In one mechanism, the C2 carbon can attack the iminium moiety to form the cyclized intermediate shown in Figure 3. Alternatively, the C3 carbon can attack the iminium moiety to form a spiro intermediate, which then undergoes a 1,2 shift to rearrange to the cyclized intermediate shown in Figure 3. Computational evidence suggests that C2, rather than C3, specifically acts as the nucleophile that attacks the iminium moiety, and that the spiro intermediate is not part of the productive reaction mechanism (Figure 3) (Kowalski et al., 1995; Maresh et al., 2008). Therefore, it is the electron density of the C2 position that should have the most impact upon the relative reactivity of 1-4. Because electron density can be inferred from
13C
chemical shifts (Lauterbur, 1961),
1H-
13C-HSQC spectroscopy was used to measure the chemical shifts of the C2 atom in 1-4. These measurements indicated that the C2 position of 7-aza-tryptamine
4
(115.9 ppm) is the most electron rich, followed by 5-aza-tryptamine
2
(131.0 ppm), then 4-aza-tryptamine
1
(135.2 ppm) and lastly 6-aza-tryptamine
3
(137.5 ppm). Although these data are consistent with previous reports that 4- and 6-aza-indoles are more deactivated than 5- and 7-aza-indoles (Zhang et al., 2002), the electronic density of these aza-indole substrates does not correspond to the order of enzyme reactivity, where 7-aza-tryptamine
4
> 4-aza-tryptamine
1
> > > 5-aza-tryptamine
2
~ 6-aza-tryptamine
3. Therefore, we conclude that the electron density of the aza-tryptamines is not the dominating factor in enzymatic reactivity.

It is also known that the pyridine moieties of 5- and 6-aza-indoles are more basic than 4- and 7-aza-indoles (7-aza-indole pKα: 4.6, 4-aza-indole pKα: 6.9, 6-aza-indole pKα: 7.9, 5-aza-indole pKα: 8.3 [Adler and Albert, 1963]). Therefore, 5- and 6-aza-tryptamines
2
and
3
are more likely to be protonated at neutral pH. Protonation of the pyridine would reduce the electron density of the aza-indole, and the rate of the Pictet-Spengler reaction would be further diminished. However, although strictosidine synthase is active at pH 8 (Maresh et al., 2008), both
2
and
3
still failed to form product when incubated with strictosidine synthase and 6 at this more basic pH value. Additionally, the tryptamine-binding pocket of strictosidine synthase consists primarily of hydrophobic residues (Ma et al., 2006), suggesting that an amino acid of the enzyme does not protonate the pyridine ring of any of the aza-tryptamines. Therefore, we conclude that neither the higher pKa values of 5- and 6-aza-tryptamines, nor the corresponding protonation state of the pyridine moiety, are responsible for the lack of enzymatic reactivity with 2 and 3. Instead, the reactivity is consistent with the previously reported substrate specificity of strictosidine synthase, in which tryptamine analogs with a variety of substituents at the 4 and 7 positions are accepted, while most substrates substituted at the 5 and 6 positions are not (Bernhardt et al., 2007; Loris et al., 2007; McCoy et al., 2006). Importantly, the aza group has different hydrogen bonding properties and is much more hydrophilic than the corresponding carbocycle. In addition to discriminating against 5- and 6 substituted tryptamines for steric reasons, strictosidine synthase also discriminates against these isosteric changes.

It is notable that strictosidine synthase allows turnover of chemically deactivated substrates. We speculate that for successful turnover of 1 and 4, the enzyme uses the active site glutamic acid residue to keep the iminium ion nitrogen protonated (Maresh et al., 2008) (Figure 3), and therefore electrophilic, while at the same time ensuring that the pyridine moiety of the aza-indole remains neutral. This highlights the superb capacity of enzymes to catalyze reactions by selectively protonating spatially separated functional groups.

The enzymatically active substrates, 4- and 7-aza-tryptamine
1
and
4
, were chosen for further analysis in this biosynthetic pathway, in the steps beyond the Pictet-Spengler reaction catalyzed by strictosidine synthase (Figure 3). To examine whether the aza-strictosidine analogs 7 and 10 formed by strictosidine synthase could be subjected to further enzymatic

Figure 1. The Aza-indole Moiety (Highlighted in Red) in Natural and Synthetic Compounds
(A) Aza-indoles in natural products.
(B) Aza-indole moiety in an indole natural product analog.
(C) Synthetic pharmaceutically important compounds containing an aza-indole group.

Figure 2. Synthesis of Aza-tryptamine Compounds 1-4

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transformations, these compounds were incubated with *Catharanthus roseus* cell culture that is known to produce a variety of monoterpene indole alkaloids derived from strictosidine (O’Connor and Maresh, 2006). Coculture of *C. roseus* with varying concentrations of 1 and 4, or the corresponding strictosidine synthase products, aza-strictosidines 7 and 10, did not result in the formation of any new alkaloid compounds as evidenced by analysis of the cell culture extracts by liquid chromatography mass spectrometry (LC-MS). However, when 4- and 7-aza-strictosidines 7 and 10 were deglycosylated in vitro by the strictosidine glucosidase enzyme that acts directly downstream from strictosidine synthase, and then cocultured with *C. roseus*, we observed a variety of new compounds that were not present in control cultures lacking aza-indole starting substrates (Figures 3 and 4). Monoterpene indole alkaloid biosynthesis is a highly branched pathway in *C. roseus*, with over 100 compounds arising from the deglycosylated strictosidine intermediate. The molecular weight of the new compounds formed appeared to correlate with aza analogs of some of these known alkaloids.

Several compounds with *m/z* 356 and *m/z* 354 derived from 4-aza-tryptamine 1 and 7-aza-tryptamine 4 were observed (Figure 4). Natural tryptamine derived compounds with *m/z* 353 (corresponding to *m/z* 354 for the aza-tryptamine analog) in these cell lines are heteroyohimbine alkaloids, with ajmalicine 12 as the major compound. Additionally, there are several known sitsirikine type alkaloids 13 with *m/z* 355 (corresponding to *m/z* 356 for the aza-indole analog) that have been previously isolated from *C. roseus*. One of the most abundant unnatural products (*m/z* 356, HPLC retention time at 3.6 min), which was derived from 7-aza-tryptamine 4, was isolated and characterized by 1H-NMR and g-correlated spectroscopy. The 1H-NMR spectrum of the isolated compound most closely matched that of

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**Figure 3. Reaction of Aza-tryptamines in Monoterpene Indole Alkaloid Biosynthesis**

(A) Reaction of aza-tryptamine compounds 1-4 in a Pictet-Spengler reaction. (B) Formation of downstream alkaloid and aza-alkaloid products.

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**Figure 4. LC-MS Spectra Showing Production of New Compounds by Catharanthus roseus When Supplemented with Deglycosylated Strictosidine Derived from 1 and 4**

Starred peaks indicate compounds found only in cultures supplemented with deglycosylated strictosidine derived from 1 or 4. LC-MS spectra showing the results of feeding deglycosylated strictosidine 11 derived from natural tryptamine 5 to cell cultures are shown in the Supplemental Data.
isosistsirikine. Notably, isosistsirikine isomers have been shown to exhibit antineoplastic activity (Mukhopadhyay et al., 1983). NMR analysis of all isosistsirikine stereoisomers have been previously reported (Lounasmaa et al., 1994). The most abundant of the aza-alkaloids from catharanthine 15 or tabersonine 16 (m/z 337 for parent, m/z 338 for aza analog) or akuammicine 17 (m/z 323 for parent, m/z 324 for aza analog), the other major alkaloids normally observed in C. roseus culture, were observed (Figure 3). Although heteroyohimbine type alkaloids such as ajmalicine 12 and isosistsirikine 13 result from enzymatic reduction of deglycosylated strictosidine (Bernhardt et al., 2009; Hemscheidt and Zenk, 1985; Stockigt et al., 1983), formation of both catharanthine 15, tabersonine 16, and akuammicine 17 require a complex rearrangement of the deglycosylated strictosidine intermediate (Qureshi and Scott, 1968; Scott et al., 1971). Although the mechanism of this rearrangement remains poorly understood, it clearly involves the C2 and C3 positions of the indole moiety. The diminished electron density of the aza-tryptamine substrates 1 and 4 do not appear to be compatible with this rearrangement. In contrast, formation of alkaloid products such as 12 and 13 entail a reduction reaction that does not involve participation of the indole group, enabling the biosynthesis of milligram quantities of aza-heteroyohimbine products.

**SIGNIFICANCE**

By introducing unnatural starting materials to a producing organism, biosynthetic pathways can be hijacked to yield novel compounds. Here we have altered tryptamine 5, which serves as the starting substrate for a variety of alkaloid biosynthetic pathways, by replacing the indole with one of four aza-indole isomers. We show that two aza-tryptamine substrates can be successfully incorporated into the products of the monoterpene indole alkaloid pathway in *Catharanthus roseus*. A key step of this alkaloid biosynthetic pathway is an enzymatic Pictet-Spengler reaction that converts tryptamine 5 and secologanin 6 into strictosidine 11, a tetrahydro-β-carboline product. Two of the electron deficient aza-tryptamines, 1 and 4, were able to undergo this enzymatic reaction, resulting in the formation of pyrrolopyridazines 7 and 10, the aza analogs of tetrahydro-β-carbolines. These pyrrolopyridazines could then be transformed by *C. roseus* into heteroyohimbine type alkaloids. Use of unnatural heterocycles in precursor-directed biosynthesis, in both microbial and plant natural product pathways, has not been widely demonstrated, and successful incorporation of starting substrate analogs containing the aza-indole functionality has not, to the best of our knowledge, been previously reported. The most abundant of the aza-alkaloids produced in *C. roseus* culture was isolated, characterized and shown to be an analog of isosistsirikine, a natural product that has demonstrated anti-neoplastic activity. Normally, the level of isosistsirikine in *C. roseus* culture is low; however, aza-tryptamine substrate 6 appeared to form this particular product preferentially compared with other classes of monoterpene alkaloids. Future efforts will focus on evaluation of the bioactivity of isolated aza-analogs. Additionally, this work serves as a starting point to explore fermentation of aza-alkaloids from other tryptophan- and tryptamine-derived natural product pathways.

**EXPERIMENTAL PROCEDURES**

**Characterization of Compounds 1-4**

**Compound 1:**

\[
\text{H-NMR (500 MHz, } \text{D}_2\text{O)} \delta 8.40 \text{ (d, } J = 9.0 \text{ Hz, 1H), 8.37 (d, } J = 6.0 \text{ Hz, 1H); 7.93 (s, 1H), 7.56} \text{ (d, } J = 7.5 \text{ Hz, 1H), 7.63 (d, } J = 8.0 \text{ Hz, 1H); 3.33 (t, } J = 7.0 \text{ Hz, 2H), 3.19 (d, } J = 7.2 \text{ Hz, 2H).}
\]

13C-NMR (125 MHz, D2O) \δ 135.2, 134.5, 133.6, 132.7, 128.4, 116.7, 105.0. Exact mass (ESI-TOF): m/z calc (M+H) = 162.1024, found: 162.1022, 162.1026.

**Compound 2:**

\[
\text{H-NMR (500 MHz, } \text{D}_2\text{O)} \delta 8.94 \text{ (s, 1H), 8.16 (d, } J = 6.5 \text{ Hz, 1H), 7.74 (dd, } J = 6.5 \text{ Hz, 1H), 7.60 (s, 1H), 3.30 (t, } J = 7.2 \text{ Hz, 2H), 3.16 (t, } J = 7.2 \text{ Hz, 2H).}
\]

13C-NMR (125 MHz, D2O) \δ 143.1, 134.4, 131.0, 130.9, 123.9, 113.6, 109.8, 40.2, 22.7. Exact mass (ESI-TOF): m/z calc (M+H) = 162.1026, found: 162.1028, 162.1030.

**Compound 3:**

\[
\text{H-NMR (500 MHz, } \text{D}_2\text{O)} \delta 8.79 \text{ (s, 1H), 8.05 (d, } J = 6.5 \text{ Hz, 1H), 7.96 (s, 1H), 7.92 (d, } J = 6.5 \text{ Hz, 1H), 3.30 (t, } J = 7.2 \text{ Hz, 2H), 3.14 (t, } J = 7.2 \text{ Hz, 2H).}
\]

13C-NMR (125 MHz, D2O) \δ 137.5, 136.2, 131.1, 127.5, 126.7, 115.3, 112.0, 39.7, 21.9. Exact mass (ESI-TOF): m/z calc (M+H) = 162.1026, found: 162.1030.

**Compound 4:**

\[
\text{H-NMR (500 MHz, } \text{D}_2\text{O)} \delta 8.53 (d, } J = 8.0 \text{ Hz, 1H), 8.23 (d, } J = 5.5 \text{ Hz, 1H), 7.49 (s, 1H), 7.44 (t, } J = 6.8 \text{ Hz, 1H), 3.26 (t, } J = 6.8 \text{ Hz, 2H), 3.09 (t, } J = 7.0 \text{ Hz, 2H).}
\]

13C-NMR (125 MHz, D2O) \δ 138.8, 137.1, 133.1, 128.2, 125.9, 115.9, 111.9. Exact mass (ESI-TOF): m/z calc (M+H) = 162.1026, found: 162.1024.

**Enzyme Assays**

Strictosidine synthase (*C. roseus*) was expressed in *E. coli* and purified as previously described (Bernhardt et al., 2009; Maresh et al., 2008; McCoy et al., 2006). Activity was measured using a high-performance liquid chromatography (HPLC) assay following previously reported procedures (Maresh et al., 2008). Secologanin (2.5 mM), 1-naphthalene acetic acid as an internal standard (60 µM), and tryptamine analogs (1–500 µM) in phosphate-buffered saline (PBS) buffer (50 mM, pH 7.0) were preincubated at 30°C. Reactions were initiated by addition of strictosidine synthase (1–200 nM). The concentration of 1 and 4 was calculated from the extinction coefficients (ε = 6068 M⁻¹cm⁻¹, 4: ε = 7890 M⁻¹cm⁻¹, 290 nm, PBS buffer [50 mM, pH 7.0]). Enzymatic reactions were quenched by addition of 2.0 M aqueous sodium hydroxide. Quenched aliquots of the reaction were directly injected onto an analytical HPLC using a solvent gradient of 10% to 70% acetonitrile in 0.1% aqueous trifluoroacetic acid. The absorbance of tryptamine analogs, strictosidine analogs and NAA were measured at 280 and 290 nm. Peak areas were integrated and normalized to the internal standard. Initial rates were determined from the slope by linear least-squares fit of four normalized data points for each concentration of varied substrate. Kinetic parameters Vmax for each reaction were estimated from the initial rates directly fit to appropriate forms of the Michaelis-Menten equation by nonlinear least-squares fit using Origin 6.0 software (OriginLab Corp.).

**Feeding Studies with 1 and 4**

To a solution of strictosidine synthase and secologanin (10 mg, 25.8 mmol) in 50 mM PBS buffer (3 ml, pH 7.0) was added 4- or 7-aza-tryptamine 1 or 4 (4.6 mg, 20 mmol) in 50 mM PBS buffer (1 ml, pH 7.0). The reaction mixture was incubated at 30°C for 48 hr. Strictosidine glucosidase was then added and the mixture was allowed to incubate for an additional 48 hr. The solution was evaporated, dissolved in 150 µl ethanol and added to *C. roseus* cell
superculture culture (150 ml) that had been subcultured for 10 days. The cultures were allowed to incubate for an additional 7 days, at which point the cells were filtered and ground in methanol. The methanol extract was dried, redissolved in ethyl acetate, and washed with water. The organic layer was concentrated, dissolved in methanol and subjected to LC-MS analysis. Ultra-performance LC analysis was performed on an Acquity Ultra Performance BEH C18 column with a 1.7 µm particle size, 2.1 x 100 mm dimension, and a flow rate of 0.6 ml min⁻¹ in tandem with a Micromass LCT Premier TOF Mass Spectrometer with an ESI source (Waters Corporation). The capillary and sample cone voltages were 3000 V and 30 V, respectively. The desolvation and source temperatures were 300 and 100°C, respectively. The cone and desolvation gas flow rates were 60 and 800 l hr⁻¹. Analysis was performed with MassLynx 4.1. Compound 14 was purified from reverse phase preparative HPLC using a gradient of 0%-30% acetone in water with 0.1% trifluoroacetic acid over 40 min.

7-Aza-isositsirikine (3z, 16R, 19E) 14

¹H-NMR (800 MHz, CD₃OD) δ 8.25 (d, J = 3.7 Hz, 1H), 8.00 (dd, J = 7.7 Hz, 1H), 7.17 (dd, J = 7.7 Hz, 4.9 Hz, 1H), 6.04 (q, J = 6.7 Hz, 1H), 3.88 (m, 3H), 3.78 (s, 3H), 3.70 (m, 3H), 3.56 (m, 2H), 3.19 (m, 1H), 3.06 (m, 1H), 2.62 (m, 2H), 2.46 (m, 2H). Exact mass (ESI-TOF): m/z calc (M+H): 356.1969, found: 356.1974.

SUPPLEMENTAL DATA

Supplemental Data include thirteen figures, six tables, and Supplemental Experimental Procedures and can be found with this article online at http://www.cell.com/chemistry-biology/supplemental/51074-5521(09)00408-6.

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