**Helicobacter pylori Infection Promotes Methylation and Silencing of Trefoil Factor 2, Leading to Gastric Tumor Development in Mice and Humans**

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**Helicobacter pylori** Infection Promotes Methylation and Silencing of *Trefoil Factor 2*, Leading to Gastric Tumor Development in Mice and Humans

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**Abstract**

**BACKGROUND & AIMS**—Trefoil factors (TFFs) regulate mucosal repair and suppress tumor formation in the stomach. *Tff1* deficiency results in gastric cancer, whereas *Tff2* deficiency increases gastric inflammation. *TFF2* expression is frequently lost in gastric neoplasms, but the nature of the silencing mechanism and associated impact on tumorigenesis have not been determined.

**METHODS**—We investigated the epigenetic silencing of *TFF2* in gastric biopsy specimens from individuals with *Helicobacter pylori*-positive gastritis, intestinal metaplasia, gastric cancer, and disease-free controls. *TFF2* function and methylation were manipulated in gastric cancer cell
lines. The effects of Tff2 deficiency on tumor growth were investigated in the gp130F/F mouse model of gastric cancer.

RESULTS—In human tissue samples, DNA methylation at the TFF2 promoter began at the time of H pylori infection and increased throughout gastric tumor progression. TFF2 methylation levels were inversely correlated with TFF2 messenger RNA levels and could be used to discriminate between disease-free controls, H pylori-infected, and tumor tissues. Genome demethylation restored TFF2 expression in gastric cancer cell lines, so TFF2 silencing requires methylation. In Tff2-deficient gp130F/F/Tff2−/− mice, proliferation of mucosal cells and release of T helper cell type-1 (Th-1) cytokines increased, whereas expression of gastric tumor suppressor genes and Th-2 cytokines were reduced, compared with gp130F/F controls. The fundus of gp130F/F/Tff2−/− mice displayed glandular atrophy and metaplasia, indicating accelerated preneoplasia. Experimental H pylori infection in wild-type mice reduced antral expression of Tff2 by increased promoter methylation.

CONCLUSIONS—TFF2 negatively regulates preneoplastic progression and subsequent tumor development in the stomach, a role that is subverted by promoter methylation during H pylori infection.

Keywords
Tumor Suppressor; Epigenetics; Stomach Cancer; Trefoil Factor 2

The trefoil factors (TFF) 1, TFF2, and TFF3 form a family of secreted proteins characterized by a triple loop structure, the trefoil domain. Gastric TFF1 expression is restricted to surface mucus and pit epithelium of the fundus and antrum, whereas TFF2 is expressed in fundic mucus neck cells (MNCs), antrum, and duodenal Brunner’s glands. TFF3 expression is restricted to intestinal goblet cells. TFF functions include mucus stabilization and barrier protection, regulation of cell migration during wound healing, gastric cell differentiation, proliferation, and apoptosis.

A body of published evidence shows that TFF1 is a stomach-specific tumor suppressor gene (TSG). Tff1−/− mice spontaneously develop antral/pyloric tumors, and antral tumors in the gp130Y757F/Y757F (gp130F/F) mouse model of gastric cancer show reduced Tff1 expression. Approximately half of all human gastric cancers show loss of TFF1 expression through loss of heterozygosity and promoter methylation as well as transcriptional inhibition by regulatory molecules. TFF1 inhibits gastric cancer cell line growth, blocks G1/S phase cell cycle progression, increases Retinoblastoma gene expression, and promotes gastric differentiation, all consistent with a tumor suppressor role in the stomach.

By contrast, the role of TFF2 in gastric cancer progression is less well understood. TFF2 expression is rapidly induced in gastrointestinal ulcerative pathologies particularly in regenerating epithelium or after nonsteroidal anti-inflammatory (NSAID) drug treatment. Moreover, in vivo delivery of recombinant TFF2 ameliorates mucosal damage.

TFF2 gives its former name to a mucus metaplasia, of mainly fundic origin known as spasmolytic polypeptide expressing metaplasia (SPEM). Whereas SPEM is associated with Helicobacter pylori-dependent fundic gland atrophy and may be a precursor to gastric cancer, TFF2 is dispensable for the development of SPEM-like metaplasia. In addition, Tff2-deficient mice display only subtle alterations in mucosal proliferation, parietal cell activation, and reduced reparative function.
An anti-inflammatory role for Tff2 has recently been described. Tff2-deficient mice display immune deregulation and are more susceptible to dextran sodium sulphate-induced colitis and Helicobacter felis–induced gastritis, developing more severe inflammatory pathology than wild-type controls. Furthermore, H pylori-infected Tff2-deficient mice develop more advanced premalignant lesions of atrophy, metaplasia, and dysplasia than wild-type littermates. Therefore, in addition to immune modulation, TFF2 may impose growth restraint and limit neoplastic progression in the context of chronic inflammation.

Here, we demonstrate that loss of TFF2 expression occurs during the progression of human intestinal-type gastric cancer via the acquisition of aberrant promoter methylation. To determine the likely functional impact of TFF2 loss (via promoter methylation) on gastric tumor growth, we have employed a genetic approach to disrupt Tff2 function in murine gastric cancer. The gp130Y757F/Y757F (gp130F/F) mouse is a model of gastric tumorigenesis that develops antral tumors with histopathologic similarities to human intestinal-type gastric cancer. We show that genetic Tff2 deficiency substantially accelerates tumor growth in gp130F/F mice, primarily via further loss of gastric TSGs and misexpression of hormones and cytokines that promote preneoplastic fundic atrophy and chronic gastritis. These data support the view that TFF2 may act locally to suppress preneoplastic change and subsequent gastric tumor development, a role that is ultimately subverted by epigenetic silencing after H pylori infection.

Materials and Methods

Mice

Tff2+/− and gp130F/+ heterozygous mice11,18 were crossed to obtain wild-type, Tff2−/−, gp130F/F, and compound mutant gp130F/F/Tff2−/− mice and maintained on a C57BL6/129-Sv mixed background. Age-matched littermate controls were used in all experiments. Wild-type (C57BL6) mice were infected with H pylori Sydney strain 1 (SS1) as described.26 Experiments were performed in accordance with either the Royal Melbourne Hospital (approval No. 010/2005) or Murdoch Children’s Research Institute Animal Ethics Committee (approval No. A583).

Human Tissue Sources

Gastric mucosal tissue was obtained from H pylori-positive individuals and disease-free controls as described.28 Gastric cancers and matched preneoplastic adjacent to cancer tissues were obtained with informed consent (approval No. 174.2008; Kanazawa University Ethics Committee for Human Genome Research).

Morphometric and Histologic Analysis

Mouse stomachs were removed, pinned, and photographed for macroscopic morphometry. Paraffin sections (4 µm) stained with H&E or alcian blue periodic acid-Schiff (AB/PAS) were used for microscopic morphometry and histologic assessment. Morphometric analysis was performed using ImageJ software (http://rsb.info.nih.gov/ij/index.html; National Institutes of Health, Bethesda, MD). The gastric mucosal area and length were outlined with the calibrated software drawing tool at the appropriate magnification. Inflammation was assessed by microscopy in a blinded fashion on AB/PAS and H&E-stained sections. Antral tumor tissues were analyzed with a minimum of 3 separate samples per animal cut and scored according to the density of inflammatory cells from minimum = 0 to maximum = 3 including infiltration around the muscularis mucosa, submucosal inflammation, and intramucosal infiltration of the tumors.
Mammalian Cell Culture

AGS and MKN28 gastric cancer cell lines were maintained in RPMI 1640 Glutamax I growth medium, 10% fetal bovine serum, 2 mmol/L nonessential amino acids, 50 IU/mL penicillin, 50 µg/mL streptomycin (all Invitrogen, Carlsbad, CA), at 37°C in a humidified incubator with 5% CO₂/air.

For proliferation analysis, AGS cells were serum starved overnight then seeded in 6-well plates at 2 × 10⁵ cells/well in growth medium supplemented with 0.2% fetal bovine serum. Recombinant human (rh) TFF2 was used at 50 µg/mL. Viable cell counts were performed by 0.4% trypan blue dye exclusion on a hemocytometer. For apoptosis, cells were stained with annexin V-Alexa Fluor 488 conjugate (Invitrogen Molecular Probes, Carlsbad, CA) and propidium iodide according to the manufacturer’s protocols. Apoptotic (annexin V positive, propidium iodide negative) cells (10,000 events) were resolved on an LSRII flow cytometer (BD Biosciences, San Diego, CA) and analyzed using FACS Diva software (BD Biosciences). For genome demethylation studies, AGS and MKN28 cells were plated in 25-cm² flasks at 5 × 10⁵ cells and allowed to attach overnight. Cells were treated with 10 µmol/L 5′-aza-2′-deoxycytidine (5′-Aza-dC) dissolved in dimethyl sulfoxide (Sigma) or dimethyl sulfoxide alone for 72 hours, with fresh 5′-Aza-dC added at 24-hour intervals.

Quantitative DNA Methylation Analysis

Genomic DNA was isolated from tissue using the DNeasy tissue kit (Qiagen, Hilden, Germany) or using Trizol (Invitrogen) and 200 – 500 ng of each sample was bisulphite converted using Zymo EZ DNA methylation reagents (Zymo Research, Orange, CA) according to the manufacturer’s protocols. TFF2 promoter fragments were recovered from bisulphite-converted DNA by polymerase chain reaction (PCR) (full details in Supplementary Materials and Methods). Quantitative methylation analysis of bisulphite PCR-amplified products was performed with the EPITYPER system (Sequenom Inc, San Diego, CA) using MassCLEAVE reagents (Sequenom) followed by matrix-assisted laser desorption ionization time-of-flight mass spectrometry. Data cleaning and 2-way hierarchical cluster analysis were performed using the R script gplots software (available: http://www.r-project.org/).

Results

**TFF2 Epigenetic Silencing by Progressive Promoter Methylation During Human Intestinal-Type Gastric Cancer Development**

To examine potential epigenetic mechanisms underlying loss of TFF2 expression, we performed correlative TFF2 messenger RNA (mRNA) expression and promoter methylation analysis on gastric mucosal specimens collected from 4 pathologic subgroups. These included normal stomach, *H pylori*-positive gastritis (Supplementary Table 1), intestinal-type gastric cancer, and patient matched preneoplastic adjacent tissue showing intestinal metaplasia (IM) (Supplementary Table 2). Quantitative reverse transcription and polymerase chain reaction (QRT-PCR) (Supplementary Materials and Methods) revealed progressive loss of TFF2 expression in *H pylori*-positive gastritis (fold change: −3.33 ± 0.77; *P* = .035), IM (fold change: −4.22 ± 0.679; *P* = .009), and gastric cancer (fold change: −5021.53 ± 2513.59; *P* < .001) compared with normal stomach (Figure 1A). Next, we quantified CpG methylation in the TFF2 promoter region and first exon (Figure 1B) using the Sequenom EPI-TYPER assay. EPITYPER is equivalent to the bisulphite mutagenesis and sequencing technique in resolving methylation on individual CpG dinucleotides (Supplementary Figure 1) but superior in terms of quantitation, intrasample precision, and sample throughput. TFF2 promoter methylation was significantly increased to a similar degree in *H pylori*-positive gastritis and IM tissues, respectively, but substantially increased in gastric cancer compared
Accelerated Tumor Growth in Tff2-Deficient Murine Gastric Cancer

To determine the role of Tff2 in gastric tumor progression, in particular to recapitulate the likely functional impact of TFF2 silencing by promoter methylation in human gastric cancer, we compared antral tumor growth between gp130<sup>F/F</sup> mice carrying wild-type or Tff2-null alleles (Figure 3A). Macroscopic morphometric analysis revealed that antral tumors in 6-week-old gp130<sup>F/F</sup>/Tff2<sup>−/−</sup> compound mutant mice were more than double the size (105 ± 2.2 mm<sup>2</sup>) of gp130<sup>F/F</sup> single mutant controls (48 ± 17 mm<sup>2</sup>) (Figure 3B), and, by 12 weeks, gp130<sup>F/F</sup>/Tff2<sup>−/−</sup> tumors were 35% larger than tumors of gp130<sup>F/F</sup> single mutant and gp130<sup>F/F</sup>/Tff2<sup>−/−</sup> heterozygous null controls (154 ± 14 mm<sup>2</sup>, 103 ± 68 mm<sup>2</sup>, and 95 ± 92 mm<sup>2</sup>, respectively) (Figure 3B). Microscopic analysis of antral tumor area at 12 weeks confirmed the increase in tumor size in gp130<sup>F/F</sup>/Tff2<sup>−/−</sup> mice (0.53 ± 0.03 mm<sup>2</sup>) compared with gp130<sup>F/F</sup> (0.41 ± 0.01 mm<sup>2</sup>) and gp130<sup>F/F</sup>/Tff2<sup>−/−</sup> littermate controls (0.41 ± 0.04 mm<sup>2</sup>) (Figure 3C). Consistent with these observations, mean antral gland height, cell number per antral gland, and Ki-67-positive cells (Figure 3D) were all increased in gp130<sup>F/F</sup>/Tff2<sup>−/−</sup> antral tumors compared with gp130<sup>F/F</sup> controls. No difference in apoptotic cell number was observed between gp130<sup>F/F</sup>/Tff2<sup>−/−</sup> and gp130<sup>F/F</sup> antral tumors (Figure 3E), thus excluding decreased apoptosis as a mechanism for the accelerated tumor growth. In addition, expression of the gastric endocrine regulators gastrin and somatostatin were substantially reduced in gp130<sup>F/F</sup>/Tff2<sup>−/−</sup> mice compared with gp130<sup>F/F</sup> controls (Figure 3F) indicating that increased tumor growth occurs under hypogastrinemic conditions. We conclude that antral tumor growth is accelerated in gp130<sup>F/F</sup>/Tff2<sup>−/−</sup> mice, independently of apoptosis or gastrin, suggesting that Tff2 negatively regulates gastric tumor progression.

Accelerated Tumorigenesis in gp130<sup>F/F</sup>/Tff2<sup>−/−</sup> Mice Is Independent of Signal Transducer and Activator of Transcription 3 but Correlated With Decreased Tumor Suppressor Gene Expression

Tumorigenesis in gp130<sup>F/F</sup> mice is due to hyperactivation of the signal transducer and activator of transcription (Stat)3 and reduced Tff1 TSG expression downstream of disrupted Shp2-ras-Erk/Ap1 signalling. We found no difference in phosphorylated Stat3 or extracellular signal-regulated kinase (Erk) protein levels between gp130<sup>F/F</sup>/Tff2<sup>−/−</sup> and gp130<sup>F/F</sup> antrum (Supplementary Figure 2). Therefore, increased tumor growth in the
absence of functional Tff2 is independent of gastric gp130 signalling. By contrast, Tff1 protein (Figure 4A) and mRNA expression (Figure 4B) were significantly reduced in gp130+/−/Tff2−/− antral tumors compared with gp130+/+. We also examined the recently described gastric TSGs Gastrokine (Gkn)33 and Gkn234 and, similar to Tff1, found that the mRNA expression of both was more substantially reduced in the antrum of gp130+/+/Tff2−/− mice compared with gp130+/+ controls (Figure 4B). Increased tumor growth in gp130+/+ Tff2−/− mice is therefore independent of gp130 signalling via Stat3, but correlates with reduced expression of gastric TSGs: Tff1, Gkn1, and Gkn2.

To substantiate further the impact of TFF2 on tumor growth and regulation of gastric TSGs, we examined proliferation and apoptosis in AGS cells treated with rhTFF2. Treatment with rhTFF2 slowed the growth of AGS cells by 25% ± 4.3% (P < .05) after 48 hours compared with mock-treated cells (Figure 4C) and was dose dependent (Supplementary Figure 3). Apoptosis was unchanged between rhTFF2-treated cells and controls (Figure 4D), confirming our in vivo observation that Tff2 inhibits proliferation of gastric epithelial cells without affecting programmed cell death. We hypothesized that altered TSG expression accounts for the observed rhTFF2-mediated growth inhibition of AGS cells. Consistent with this, TFF1 expression was increased (fold change: 1.22 ± 0.06, P < .05) in rhTFF2 treated AGS cells (Figure 4E). Intriguingly, TFF2 expression (fold change: 1.3 ± 0.1, P < .05) was also increased with rhTFF2 treatment, suggesting the existence of a positive feedback mechanism on TFF2 transcription (Figure 4E).

**Th1 Polarized Immune Response in gp130+/+ Tff2−/− Antral Tumors**

Tumors in gp130+/+ mice are associated with extensive inflammation. Therefore, we next examined inflammatory mediators associated with gp130+/+ Tff2−/− compound mutant tumors. We observed increased leukocyte infiltration in gp130+/+ Tff2−/− compared with gp130+/+ mice (Figure 5A and B), although numbers of activated macrophages (F4/80 positive) and neutrophils (myeloperoxidase [MPO] positive) were not significantly different (Figure 5C). Inflammatory leukocytes were mainly confined to desmin-positive lamina propria at the luminal margin of the epithelium (data not shown). In agreement with increased immunocyte infiltration, we observed a Th type 1 polarized immune response with significantly increased expression of the proinflammatory cytokines interleukin (IL)-1α, IL-1β, and interferon (IFN)-γ in gp130+/+ Tff2−/− antral tumors compared with gp130+/+ controls (Figure 5D). We observed a concomitant decrease in the expression of the Th2 cytokine IL-13, with IL-4 also trending to decrease (Figure 5E). Thus, ablation of Tff2 in the context of gastric tumorigenesis results in a Th1 polarized response that may contribute directly to tumor progression by permitting increased leukocyte infiltration.

**Increased Fundic Atrophy in gp130+/+ Tff2−/− Mice**

Histologic analysis revealed significant glandular atrophy in the fundus of 12-week gp130+/+ Tff2−/− compared with wild-type and gp130+/+ mice (Figure 6A and B). To characterize the cellular composition of the fundus in gp130+/+ Tff2−/− mice, sections were stained for neutral and acidic mucins (AB/PAS; surface mucus and MNCs), Tff2, Griffonia simplicifolia lectin II (GSI) (MNCs), and intrinsic factor (chief cells). H+K+ATPase mRNA (parietal cells) was quantified by QRT-PCR. In gp130+/+ Tff2−/− and to a lesser extent in gp130+/+ Tff2−/− mice, the fundic mucosa consisted predominantly of a GSII and AB/PAS-positive MNC-like population and was almost entirely devoid of parietal and chief cells (Figure 6A–C). Therefore, genetic ablation of Tff2 in the context of pre-existing gastric pathology accelerates the loss of differentiated epithelial cell types leading to profound glandular atrophy.
H pylori Induces Tff2 Promoter Methylation After Chronic Infection in Mice

To show definitively that H pylori infection can induce Tff2 promoter methylation, we next analysed stomachs from wild-type (C57BL/6) mice experimentally infected with the mouse-adapted H pylori SS1 strain for 12 and 15 months. Tff2 expression was reduced in H pylori 12-month postinfection (MPI) stomach (Figure 7A), whereas Tff2 promoter methylation was markedly increased at 12 MPI (and 15 MPI) compared with age-matched noninfected controls (Figure 7B). These observations were supported by hierarchical cluster analysis which effectively discriminated H pylori-infected mice from noninfected mice on the basis of Tff2 methylation profiles (Figure 7C). Collectively, these results show that chronic H pylori infection directly induces Tff2 promoter methylation in vivo. Because the progression of gastric pathology is clearly accelerated in gp130F/F/Tff2−/− mice, and H pylori infection leads to gastric Tff2-deficiency via epigenetic silencing (Figure 7A–C), we also investigated whether short-term (3 months) H pylori infection in gp130F/F mice can phenocopy the effects of genetic Tff2 deficiency by augmenting Tff2 promoter methylation. We observed no effect of the gp130F/F mutation, H pylori, or their combined effect on Tff2 expression or methylation (Supplementary Figure 4) in antral stomach after 3 months of infection.

Discussion

We have established TFF2 epigenetic silencing, by promoter methylation, as a novel paradigm for loss of TFF2 expression in human intestinal-type gastric cancer and following experimental H pylori infection in the mouse stomach. We also assessed Tff2 function in the context of mouse gastric tumor growth. Disruption of Tff2 in gp130F/F/Tff2−/− mice produced a polarized immune response, with subduced Th2 and exacerbated Th1-type inflammation. Tff2 deficiency in 6- and 12-week gp130F/F/Tff2−/− mice caused larger tumors compared with gp130F/F mice because of increased proliferation and accumulation of mucus cells without affecting apoptosis. This demonstrates a novel function for Tff2 in restraining tumorigenesis and acting as a TSG in the stomach (Supplementary Figure 5). Concordantly, development of gastric neoplasia is accelerated in Tff2-deficient mice compared with wild-type littermates after H pylori infection.

Whereas tumorigenesis in gp130F/F mice is attributed to dysregulated gp130 signalling and repression of Tff1, genetic ablation of Tff2 conferred no additional dysregulation upon gp130 signalling via Stat3 or Shp2-ras-Erk/Ap1 pathways. However, Tff2 deficiency led to reduced expression of Tff1 and putative TSGs, Gkn1, and Gkn2. These 2 gastrokines are emerging as important endogenous regulators of gastric homeostasis and show profound loss of expression in H pylori infection, aspirin ingestion, and neoplastic progression. Thus, TFF2 is a likely upstream regulator of gastrokine gene expression, and its inhibition may promote tumor growth because of loss of anti-proliferative gastrokine and TFF1 genes. Additionally, human GKN2 protein exists predominantly as a heterodimer with TFF1, thus attenuated expression of either protein may potentiate their combined loss of function.

Tff2 deficiency in gp130F/F/Tff2−/− mice led to reduced Tff1 expression. This finding provides a mechanism for increased proliferation because TFF1 has previously been shown to inhibit the growth of the human gastric cancer AGS cell line. Here we have shown that exogenous TFF2 peptide inhibits the growth of AGS cells without affecting apoptosis. Treatment with rhTFF2 increased TFF1 and TFF2 mRNA expression in cell lines, consistent with reported cross- and autoregulatory functions for TFF2. Moreover, down-regulation of both TFF1 and TFF2 expression frequently occurs in gastric cancer, consistent with a tumor suppressor function.

TFF1 and TFF2 play critical roles in maintaining mucosal integrity, and combined functional loss likely compromises the gastric mucosa, permitting increased inflammation...
and damage while subduing repair processes. In this context, compound gp130\(^{F/F}\)/Tff2\(^{−/−}\) mice displayed pervasive fundic atrophy by 12 weeks of age with significant loss of parietal and chief cell lineages and accumulation of undifferentiated GSII-positive MNC-like cells. Similarly, fundic atrophy is accelerated in Tff2-deficient mice chronically infected with *Helicobacter* species.\(^{20,26}\)

Tumorigenesis in gp130\(^{F/F}\)/Tff2\(^{−/−}\) mice was associated with augmented Th1 cytokine release and attenuated Th2 cytokine expression, particularly IL-13. Th1 cytokines, in particular IL-1\(\beta\), promote mucosal inflammation, inhibit gastric acid secretion\(^{43}\) and induce fundic atrophy leading to gastric adenocarcinoma.\(^{44}\) Our data suggest that Tff2 may modulate and prevent imbalanced levels of Th1 and Th2 cytokines leading to mucosal destruction and disrepair. It is unknown whether TFF2 directly regulates inflammatory cytokines; however, these results highlight the important homeostatic role for TFF2 in the normal gastric mucosa. TFF2 loss after *H pylori* infection provides a mechanism by which the bacterium may modulate the local immune response.

Expression of the hormones, gastrin and somatostatin, required for normal gastric mucosal development, and the former for parietal cell activation,\(^{45}\) was almost completely ablated in gp130\(^{F/F}\)/Tff2\(^{−/−}\) tumors. Gastrin deficiency results in chronic atrophic gastritis and predisposition to tumors via IFN\(\gamma\)/STAT3 activity,\(^{46}\) thus decreased gastrin and somatostatin may contribute to the observed fundic atrophy. Additionally, because gastrin is a direct transcriptional regulator of Tff1,\(^{47}\) its profound suppression in gp130\(^{F/F}\)/Tff2\(^{−/−}\) mice may also contribute to loss of Tff1 expression, independent but in concert with the effects of Tff2.

Quantitative analysis of normal human antral stomach, *H pylori*-dependent gastritis, intestinal metaplasia, and tumor cohorts revealed progressive loss of TFF2 expression during gastric cancer development and concomitant gain of promoter methylation, particularly at CpG dinucleotides overlapping the transcription start site. This is the first description of TFF2 promoter methylation in precancerous or cancerous stomach, supporting the notion that TFF2 is a TSG targeted by epigenetic silencing. Whereas we believe that promoter methylation represents a principle means of TFF2 silencing, it is also possible that metaplastic induction leads to antral mucus cell loss and depletion of the source of TFF2 synthesis. Increased tumor growth in gp130\(^{F/F}\)/Tff2\(^{−/−}\) mice clearly illustrates how perturbed TFF2 expression might influence the rate of tumor progression in human gastric cancer. This is supported by our in vitro studies in which genome demethylation in AGS and MKN28 gastric cancer cell lines restores TFF2 transcription, and recombinant TFF2 treatment in AGS cells activates TFF1 transcription and inhibits cell proliferation.

Our human and mouse data show that TFF2 methylation is likely initiated during *H pylori* infection, although the mechanism is unclear. This may not be a direct function of the bacterium itself but more likely arises from infection-dependent inflammation, in particular the activity of proinflammatory cytokines such as IL-1\(\beta\), the expression of which is inversely correlated with that of TFF2. IL-1\(\beta\) expression is up-regulated during *H pylori* infection\(^ {48}\) and reportedly induces nitric oxide production, which stimulates DNA methyltransferases responsible for inappropriate hypermethylation and transcriptional gene silencing.\(^ {49}\) Thus, increased IL-1\(\beta\) release may indirectly mediate TFF2 epigenetic silencing in human gastric cancer.

Inappropriate epigenetic silencing is one of the earliest detectable molecular defects in gastric tumorigenesis.\(^ {50}\) We demonstrate that TFF2 promoter methylation effectively discriminates *H pylori*-positive gastritis/intestinal metaplasia, gastric cancer, and disease-free control tissue thereby warranting further evaluation of its diagnostic potential. This also raises the possibility of future clinical application of nontoxic DNA methylation inhibitors.
such as 5′-Aza-C to reverse the inappropriate gene silencing of key gastric regulator genes. DNA methylation profiles are far more stable than mRNA markers and, as exemplified by our analysis of TFF2, provide a robust surrogate for transcriptional silencing during the formative stages of tumorigenesis. Determination of gastric TFF2 methylation profiles may provide a novel approach for early intervention in gastric adenocarcinoma, a disease in which late diagnosis continues to present a major therapeutic challenge. In summary, we demonstrate that TFF2 expression is epigenetically silenced during the progression to human intestinal-type gastric cancer by promoter hypermethylation. Taken together with our mouse genetic analysis, we conclude that TFF2 functions as a novel stomach-specific and epigenetically regulated tumor suppressor gene.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

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Abbreviations used in this paper

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<td>5′-Aza-C</td>
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<td>Trefoil factor</td>
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<td>TSG</td>
<td>tumor suppressor gene</td>
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Figure 1. Increased TFF2 promoter methylation correlates with loss of expression in gastric cancer. (A) QRT-PCR analysis of TFF2 messenger RNA (mRNA) levels in human gastric mucosal tissues (n = 91); graph shows mRNA fold-change relative to normal stomach; black bars show mean fold-change. Asterisks show statistical significance (P < .05). (B) TFF2 promoter region scanned by EPITYPER methylation analysis. Wide black box shows coding regions; narrow black box shows 5' UTR; arrowheads show transcriptional orientation; TFF2 transcription start site (TSS); base pair (bp) positions are indicated; solid-line circles show CpGs covered by the analysis; broken-line circles show nonanalyzed CpGs. (C) EPITYPER
DNA methylation analysis in human gastric mucosal tissues described in A. *Bar graph* shows mean CpG methylation ratios (levels). CpG units in pairs 8–9 and 11–12, respectively, are shown as combined averages (the constituent CpG units were not resolved by the EPITYPER assay). *Asterisks* show statistical significance (*P < .05*). (D) Two-way hierarchical cluster analysis of *TFF2* promoter methylation levels (*columns*) against human tissue samples (*rows*). Classification of the human gastric mucosal tissues is shown on the right: *open boxes* (normal stomach), *shaded boxes* (*H pylori*-positive gastritis/preneoplastic adjacent to cancer with IM), *solid boxes* (gastric cancer). CpG methylation ratios 0 (*green*) to 1.0 (*red*; see color key). *Histogram* shows CpG/methylation ratio frequency.
Figure 2.

*TFF2* is re-expressed following genome demethylation. (A) Semiquantitative RT-PCR analysis of *TFF2* messenger RNA (mRNA) expression in normal gastric antrum, MKN28 cells, and AGS cells. *GAPDH* demonstrates complementary DNA (cDNA) integrity. (B) EPITYPER analysis of *TFF2* promoter methylation of samples shown in A. Epigrams show CpG methylation ratios: graded from blue (0; hypomethylated) to yellow (1; hypermethylated). Base pair (bp) positions are indicated; solid circles show CpGs covered by the analysis; faded circles show nonanalyzed CpGs. (C) QRT-PCR analysis of *TFF2* expression in MKN28 cells and AGS cells treated with 10 µmol/L 5′Aza-dC for 72 hours compared with mock-treated controls (n = 6). Histograms show mean mRNA fold-change relative to untreated control. Asterisks show statistical significance (P < .05).
Figure 3.
Morphometric analysis of antral tumors in gp130^{F/F}/Tff2^{−/−} mice. (A) Images of antral tumors in 12-week-old mice; tumor areas are defined with dotted outlines; scale bar shows 5 mm. (B) Macroscopic morphometric analysis of antral tumor area in 6- and 12-week-old mice. (C) Microscopic morphometric analysis of tumor area (standardized to length of muscularis mucosa) in 12-week-old mice. (D) Antral gland height; cell number per antral gland; proliferating cells by quantitative Ki-67 immunohistochemistry. (E) Localization of apoptotic cells by quantitative activated caspase (casp) 3 immunohistochemistry. (F) QRT-PCR analysis of Gastrin and Somatostatin messenger RNA (mRNA) expression.
Histograms show mean mRNA fold change relative to wild-type. Asterisks show statistical significance ($P < .05$).
Figure 4.
Loss of gastric tumor suppressor gene expression in $gp130^{W/Tff2^{-/-}}$ antral tumors. (A) Immunohistochemical localization of Tff1 in antral tumors. (B) QRT-PCR analysis of Tff1, Gkn1, and Gkn2. Histograms show mean messenger RNA (mRNA) fold change relative to wild type. (C) Proliferation determined by viable cell counts in AGS cells following treatment with 50 µg/mL recombinant human (rh) TFF2 for 48 hours ($n = 20$). (D) Apoptosis measured by Annexin V flow cytometry in AGS cells treated with 50 µg/mL rhTFF2 for 48 hours ($n = 5$). (E) QRT-PCR analysis of TFF1 and TFF2 mRNA expression in AGS cells treated with 50 µg/mL rhTFF2 for 48 hours ($n = 6$). Asterisks show statistical significance ($P < .05$).
Figure 5.
Analysis of inflammatory mediators in gp130F/F/Tff2−/− antral tumors. (A) Representative H&E sections showing intramucosal inflammation in the lamina propria within tumors; scale bar shows 50 µm. (B) Semiquantitative gradation of tumor-associated inflammatory leukocyte infiltrate. (C) Quantitative immunohistochemistry of tumor-associated, activated macrophages (F4/80) and neutrophils (MPO). QRT-PCR expression analysis in antral tumors of (D) Th1 cytokines: IL1α, IL1β, IFNγ. (E) Th2 cytokines: IL-13, IL-4. Histograms show mean messenger RNA fold change relative to wild type. Asterisks show statistical significance (P < .05).
Figure 6.
Analysis of fundic atrophy in gp130	extsuperscript{F/F}/Tff2	extsuperscript{-/-} mice. (A) Immunohistochemical localization of Tff2 (MNCs), intrinsic factor (chief cells), GSII (MNCs); neutral/acidic mucins (AB/PAS). Scale bar shows 50 µm. (B) Quantitative morphometry of normal vs atrophic fundic mucosa. (C) QRT-PCR analysis of H⁺K⁺ATPase expression; histogram shows mean messenger RNA fold change relative to wild type. Asterisks show statistical significance (P < .05).
Figure 7.
Tff2 promoter methylation is induced by experimental H pylori infection in mice. (A) Immunoblot analysis of Tff2 protein expression in stomachs of wild-type (C57BL6) mice at 12-month postinfection (12-MPI) with H pylori SS1 (Hp SS1) compared with noninfected controls. Horizontal black bars show mean optical density (OD) of Tff2 protein bands normalized to GAPDH. Representative protein bands from each group are shown below the scatterplots. (B) Quantitative Tff2 promoter methylation analysis (EPITYPER) in 12 and 15 MPI Hp SS1 and control stomach. Histograms show mean CpG methylation ratios. Asterisks show statistical significance (P < .05). Wide black box shows exon1 coding region; narrow
black box shows exon1 5' UTR; transcription start site (TSS); base pair (bp) positions are indicated; solid-line circles show CpGs covered by the analysis; broken-line circles show nonanalyzed CpGs. (C) Two-way hierarchical cluster analysis of Tff2 promoter methylation data (columns) against mouse samples (rows). Hp SS1 infection status classification is shown on the right: white boxes (12/15 MPI noninfected controls), black boxes (12/15 MPI Hp SS1). CpG methylation ratios: 0.2 (green; low) to 0.6 (red; high; see color key).