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# Integrative analysis of genetic data sets reveals a shared innate immune component in autism spectrum disorder and its co-morbidities

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## Abstract

**Background:** Autism spectrum disorder (ASD) is a common neurodevelopmental disorder that tends to co-occur with other diseases, including asthma, inflammatory bowel disease, infections, cerebral palsy, dilated cardiomyopathy, muscular dystrophy, and schizophrenia. However, the molecular basis of this co-occurrence, and whether it is due to a shared component that influences both pathophysiology and environmental triggering of illness, has not been elucidated. To address this, we deploy a three-tiered transcriptomic meta-analysis that functions at the gene, pathway, and disease levels across ASD and its co-morbidities.

**Results:** Our analysis reveals a novel shared innate immune component between ASD and all but three of its co-morbidities that were examined. In particular, we find that the Toll-like receptor signaling and the chemokine signaling pathways, which are key pathways in the innate immune response, have the highest shared statistical significance. Moreover, the disease genes that overlap these two innate immunity pathways can be used to classify the cases of ASD and its co-morbidities vs. controls with at least 70 % accuracy.

**Conclusions:** This finding suggests that a neuropsychiatric condition and the majority of its non-brain-related co-morbidities share a dysregulated signal that serves as not only a common genetic basis for the diseases but also as a link to environmental triggers. It also raises the possibility that treatment and/or prophylaxis used for disorders of innate immunity may be successfully used for ASD patients with immune-related phenotypes.

**Keywords:** Autism spectrum disorder, Co-morbidities of ASD, Innate immunity pathways, Three-tiered meta-analysis, Gene expression

## Background

While at an organismal level, two or more diseases may appear unrelated, at the molecular level, it is unlikely that they arise entirely independently of one another. Studies of the human interactome—the molecular network of physical interactions (e.g., protein–protein, gene, metabolic, regulatory etc.) between biological entities in cells—demonstrate that gene function and regulation are

integrated at the level of an organism. Extensive patterns of shared co-occurrences also evidence molecular commonalities between seemingly disparate conditions [1].

Indeed, different disorders may share molecular components so that perturbations causing disease in one organ system can affect another [2]. Yet, since the phenotypes appear so different, medical sub-disciplines address the conditions with sometimes wildly differing treatment protocols. If investigators can uncover the molecular links between seemingly dissimilar conditions, the connections may help explain why certain groups of diseases arise together and assist clinicians in their decision-making about best treatments. Knowledge of shared molecular

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pathology may also provide therapeutic insights for repositioning of existing drugs [3].

Such thinking has emerged most recently in neuropsychiatry, where many such illnesses do not have clear boundaries in terms of their pathophysiology or diagnosis [4, 5]. Indeed, there is now growing evidence that rare variants ranging from chromosomal abnormalities and copy number variation (CNV) to single nucleotide variation have implications for autism spectrum disorder (ASD) and other neuropsychiatric conditions [6–13]. For example, single nucleotide polymorphisms (SNPs), which overlap genes in common molecular pathways, such as calcium channel signaling, are shared in ASD, attention deficit-hyperactivity disorder, bipolar disorder, major depressive disorder, and schizophrenia [14]. CNVs, especially the rare ones, can explain a portion of the risk for multiple psychiatric disorders [10, 13]. For example, the 16p11.2 CNV encompassing around 600 kb (chr 16:29.5, 30.2 Mb) has been implicated in multiple psychiatric disorders with the deletions being associated with ASD, developmental delay, and intellectual disability, and duplications being associated with ASD, schizophrenia, bipolar disorder, and intellectual disability [10, 13, 15–19]. However, pathogenic variations are observed in only about 30 % of the ASD-affected individuals [12, 20–23] and these variations often fail to explain the idiopathic (non-syndromic) ASD cases as well as why ASD-affected individuals suffer from many other non-neuropsychiatric conditions.

To complement the evidence of genome-wide pleiotropy across neuropsychiatric diseases, rather than looking at one neurodevelopmental disease (ASD) and comparing it to other seemingly, brain-related diseases, we expand our exploration outside of the brain to conditions related to other organ systems that co-occur with ASD. Recent studies based on electronic health records [24, 25] have identified various co-morbidities in ASD, including seizures [26, 27], gastrointestinal disorders [28, 29], ear infections and auditory disorders, developmental disorders, sleep disorders [30], muscular dystrophy [31–33], cardiac disorders, and psychiatric illness [34, 35].

In this paper, we introduce an integrative gene expression analysis to identify a shared pathophysiological component between ASD and 11 other diseases, namely, asthma, bacterial and viral infection, chronic kidney disease, cerebral palsy, dilated cardiomyopathy, ear infection, epilepsy, inflammatory bowel disease (IBD), muscular dystrophy, schizophrenia, and upper respiratory infection, that have at least 5 % prevalence in ASD patients [24, 25]. We asked the question, “Do these disease states—which are not included in the definition of ASD but co-occur at a significantly high frequency—illuminate dysregulated pathways that are important in ASD?” We reasoned that

such pathways may offer previously hidden clues to shared molecular pathology.

Other investigators have integrated genomic data from genome-wide association studies and non-synonymous SNP studies for multiple immune-related diseases, revealing that combining genetic results better identified shared molecular commonalities [36]. We believe that adopting an integrative approach not only at the gene level but also at the biochemical pathway and disease levels will power the results still further.

Here we describe results from a novel three-tiered meta-analysis approach to determine molecular similarities between ASD and 11 of its co-morbid conditions. For every disease condition, we (i) looked for statistically significant differentially expressed genes, (ii) identified their enrichment in canonical pathways, and (iii) determined the statistical significance of the shared pathways across multiple conditions. We are unaware of any analyses that go from population-based co-morbidity clusters of ASD to a multi-level molecular analysis at anywhere near this breadth.

Our results unearth several innate-immunity-related pathways—specifically, the Toll-like receptor and chemokine signaling pathways—as significant players in ASD and all but three of its examined co-morbidities. Candidate genes in these two pathways significantly overlap in conditions of ASD, asthma, bacterial and viral infection, chronic kidney disease, dilated cardiomyopathy, ear infection, IBD, muscular dystrophy, and upper respiratory infection. Candidate genes did not appear to be significantly shared in cerebral palsy, epilepsy, or schizophrenia. Notably, although bacterial and viral infection, respiratory infection, ear infection, IBD, and asthma have well-known connections with the immune system, we demonstrate that innate immunity pathways are shared by ASD and its co-morbidities, *irrespective* of whether they are immunity-related diseases or not.

Since both Toll-like receptor signaling and chemokine signaling pathways play crucial roles in innate immunity, the results suggest that this first-line defense system (which protects the host from infection by pathogens and environmental triggers) may be involved in ASD and specific co-morbidities. If the profiles of genetic susceptibility pathways in relation to environmental triggers can be ascertained, they may help in defining new treatments, such as vaccination [37] or other tolerization therapies [38]. Those may help individuals and families who are at high risk for ASD to prevent and/or treat immune-related phenotypes of the illness.

## Results

### Three-tiered meta-analysis pipeline

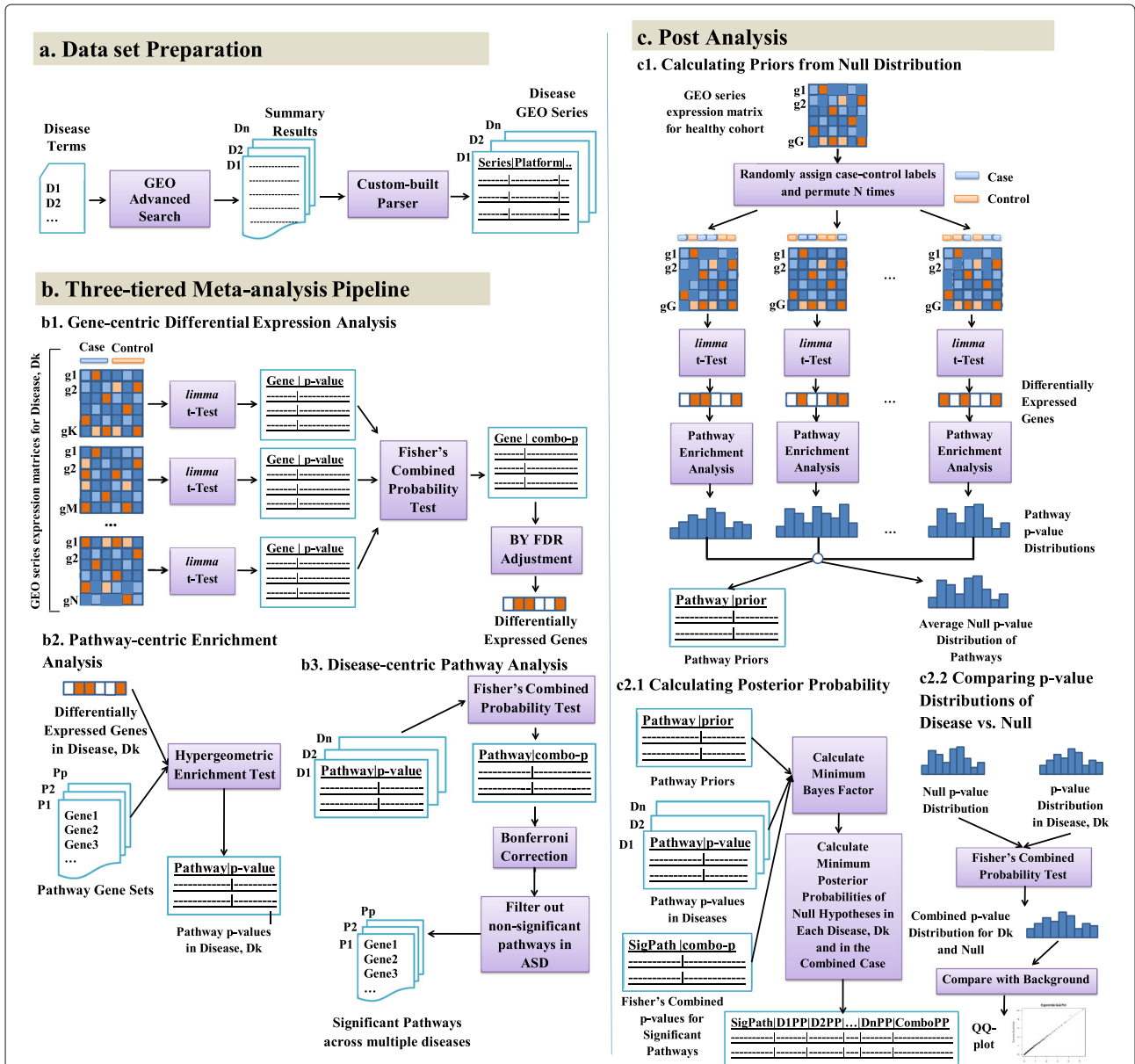
We examined ASD and 11 of its most common co-morbidities (Table 1) through a three-tiered lens of gene,

**Table 1** Co-morbidities of autism spectrum disorders

Disease group	Clinical manifestations	References
Multisystem disorders (congenital anomalies, auditory disorders, infections, gastro-intestinal disorders, cardiac disorders etc.)	Asthma	Becker, 2007 [100]; Doshi-Velez, Ge, and Kohane, 2014 [25]
	Bacterial and viral infections	Atladóttir et al., 2010 [78]; Atladóttir et al., 2012 [79]; Garbett et al., 2012 [63]; Hagberg, Gressens, and Mallard, 2012 [101]
	Chronic kidney disease	Curatolo et al., 2004 [102]; Loirat et al., 2010 [103]
	Cerebral palsy	Surén et al., 2012 [104]; Doshi-Velez, Ge, and Kohane, 2014 [25]
	Dilated cardiomyopathy	Witchel, Hancox, and Nutt, 2003 [105]; Bilder et al., 2013 [106]
	Ear infection/otitis media	Konstantareas and Homatidis, 1987 [107]; Rosenhall et al., 1999 [108]; Porges et al., 2013 [109]
	Inflammatory bowel disease (Crohn's disease, ulcerative colitis)	Horvath et al., 1999 [28]; Horvath and Perman, 2002 [29]; Walker et al., 2013 [110]
	Muscular dystrophy	Wu et al., 2005 [31]; Hendriksen and Vles, 2008 [32]; Hinton et al., 2009 [33]; Kohane et al., 2012 [24]
	Upper respiratory infection	Shavelle, Strauss, and Pickett, 2001 [111]; Porges et al., 2013 [109]; Bilder et al., 2013 [106]
	Seizures	Epilepsy
Psychiatric disorders	Schizophrenia	Morgan, Roy, and Chance, 2003 [34]; Tabarés-Seisdedos and Rubenstein, 2009 [112]; Ingason et al., 2011 [113]; Smoller et al., 2013 [14]; Murdoch and State, 2013 [114]

pathway, and disease. Figure 1 shows our three-tiered meta-analysis pipeline. Differential analysis of expression data from 53 microarray studies (see Additional file 1: Table S1) related to the 12 disease conditions revealed

different numbers of significant genes per disease depending on different false discovery rate (FDR) corrections (shown in Table 2). The complete lists of  $p$  values per gene per disease under different FDR corrections are given in



**Fig. 1** Three-tiered meta-analysis pipeline. **a** Data preparation: Select the GEO series relevant to ASD and co-morbid diseases. **b** Three tiers: (1) For each disease, select significant genes from differential expression analysis of GEO series with a Fisher's combined test with  $p < 0.05$  after Benjamini–Yekutieli (BY) FDR adjustment. (2) For each disease, select significant pathways from hypergeometric enrichment analysis with  $p < 0.05$ . (3) Identify significant shared pathways across diseases using Fisher's combined test with  $p < 0.05$  after Bonferroni FDR correction. Exclude the non-significant pathways in ASD. **c** Post analysis. (1) Using the gene expression data from a healthy cohort, generate a null distribution of pathway  $p$  values and calculate prior probabilities of pathways being significant by chance. (2.1) Using the prior probabilities, pathway  $p$  values in each individual disease, and the Fisher's combined  $p$  values of significant pathways across diseases, calculate minimum Bayes factors and minimum posterior probabilities of null hypotheses for each significant pathway in each disease and in the combined case. (2.2) Combine the pathway  $p$  value distribution of each disease with the average null distribution of  $p$  values using Fisher's combined probability test and compare the combined  $p$  value distribution with the background chi-squared distribution using a QQ plot for significance. Identify the significant pathways using the combined  $p$  values, minimum posterior probabilities, and QQ plots. ASD autism spectrum disorder, BY Benjamini–Yekutieli correction, FDR false discovery rate, GEO Gene Expression Omnibus, QQ plot, quantile–quantile plot

**Table 2** Number of differentially expressed genes selected under different FDR corrections for different diseases

	Bonferroni	BY	BH	None <sup>a</sup>
ASD	157	1258	5104	9176
Asthma	238	852	2501	5555
Bacterial and viral infection	1613	3630	6016	8183
Chronic kidney disease	66	416	3771	12577
Cerebral palsy	93	220	646	2352
Dilated cardiomyopathy	146	349	908	3455
Ear infection/otitis media	1629	3867	6708	6708
Epilepsy	5	4	12	2242
Inflammatory bowel disease	831	2547	4771	6897
Muscular dystrophy	207	517	1303	3885
Schizophrenia	54	149	508	2881
Upper respiratory infection	32	59	172	2664

Significance cutoff of  $p < 0.05$ 

ASD autism spectrum disorder, BY Benjamini–Yekutieli, BH Benjamini–Hochberg, FDR false discovery rate

<sup>a</sup>No FDR correction

**Additional file 2.** To select the most informative FDR correction test, we looked at the accuracy of classification of cases vs. controls for each disease using the disease gene sets selected under different FDR corrections. We found the Benjamini–Yekutieli (BY) adjustment to be the most informative and accurate—classification accuracy being at least 63 % using the genes selected under BY adjustment as features for a support vector machine (SVM) classifier. This was true for all the diseases examined (see “Methods” section as well as Additional file 3: Figure S1 for details).

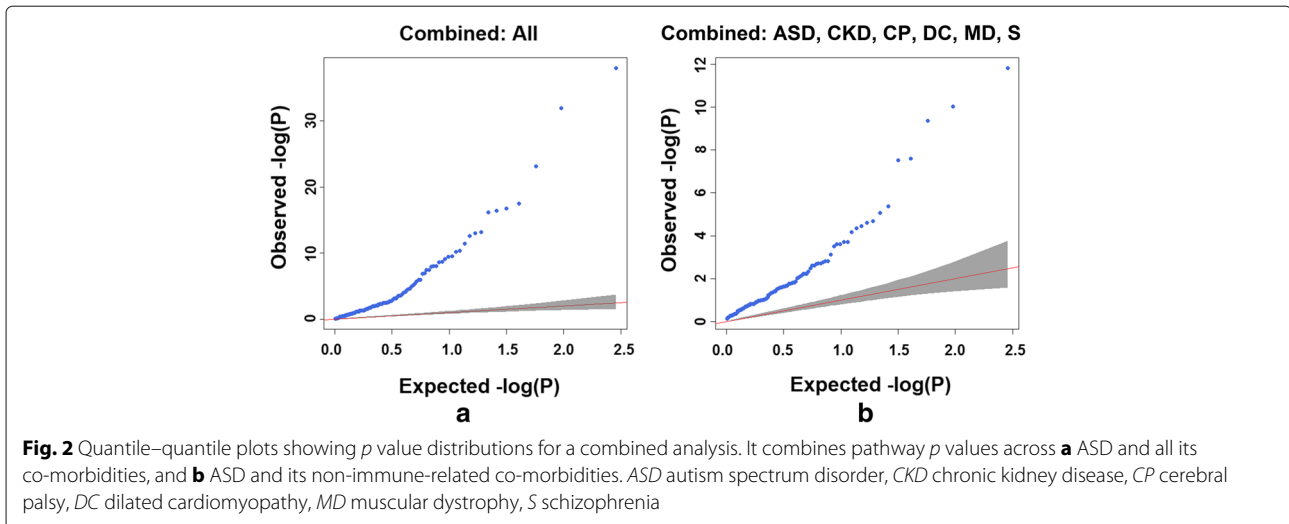
Hypergeometric enrichment analysis on individual pathway gene sets from the Kyoto Encyclopedia of Genes and Genomes (KEGG), BioCarta, Reactome, and the Pathway Interaction Database (PID) collections, as well as on the combined gene set of all canonical pathways, helped us to obtain a  $p$  value per pathway per disease. For different pathway gene set collections, the complete lists of  $p$  values per pathway in each disease are provided in Additional file 4. Combining the  $p$  values per pathway across all the diseases using Fisher’s combined probability test [39] and correcting for multiple comparisons using Bonferroni correction, we measured the shared significance of pathways across ASD and its co-morbidities (see “Methods” section for details). After selecting any pathway that had an adjusted  $p$  value  $< 0.05$  as significant and filtering out the pathways that are not significant in ASD, we found a list of pathways that are dysregulated in ASD and at least one of its co-morbidities (see Additional file 4).

To confirm that the presence of multiple significant pathways among ASD and its co-morbidities was due

to shared biology, we estimated minimum Bayes factors (BFs) and minimum posterior probabilities of the null hypothesis for each of the significant KEGG pathways in ASD and its co-morbidities (Fig. 1 and Additional file 5). The priors for the pathways were estimated from 100 null distributions of  $p$  values generated by differential expression analysis and pathway analysis performed on the gene expression data of a healthy cohort (GEO accession GSE16028) (see Fig. 1 and “Methods” section for details). Looking at the significant pathway  $p$  values in each disease and their corresponding posterior probabilities of the null hypothesis, we found that, for the significant  $p$  values ( $p < 0.05$ ), the posterior probabilities of the  $p$  values being significant by chance were always less than 5 %. The quantile–quantile (QQ) plot of combined  $p$  values of pathways across ASD and its co-morbidities shows marked enrichment of significant  $p$  values indicative of shared disease biology captured by the pathways tested (Fig. 2a). The QQ plots of hypergeometric  $p$  values of pathways in ASD and its co-morbid diseases against theoretical quantiles also show significant enrichment (see Additional file 3: Figure S2). For contrast, we combined pathway  $p$  values from each disease separately with the null  $p$  value distribution. When the pathway  $p$  value distribution in a disease is combined with the null  $p$  value distribution, the QQ plots do not show much deviation from the background distribution (see Additional file 3: Figure S3), indicating both that there is a lack of shared biology (as expected) and that our analysis does not cause systematic inflation.

#### Involvement of innate immunity pathways in ASD and its co-morbidities

The results demonstrate that pathways that are dysregulated across ASD and its co-morbidities with the highest statistical significance (i.e., the lowest Bonferroni-corrected combined  $p$  value) are all related to innate immunity. For the KEGG, BioCarta, and PID gene sets, the Toll-like receptor signaling pathway was found to be the most significant (Additional file 4). For the KEGG database, the top two significant pathways were Toll-like receptor signaling and chemokine signaling (Table 3 and Additional file 4). The top three significant pathways, revealed from the analysis of the Reactome data set, include chemokine receptor signaling, innate immunity, and Toll-like receptor signaling (Additional file 4). When we expanded our aperture of analysis to the gene sets from all canonical pathways, the Toll-like receptor signaling and chemokine signaling pathways were still found to be the most significantly dysregulated in the disease conditions (Additional file 4). Thus, we primarily focused our attention on these two pathways in ASD and its co-morbidities and then, for completeness, extended to other innate immunity KEGG pathways that were found significantly dysregulated (Table 3).



Both Toll-like receptor signaling and chemokine signaling pathways are key pathways in the innate immune response mechanism. Toll-like receptors are the most common pattern recognition receptors that recognize distinct pathogen-associated molecular patterns and participate in the first line of defense against invading pathogens. They also play a significant role in inflammation, immune cell regulation, survival, and proliferation. Toll-like receptors activate various signal transduction pathways, which in turn activate expression and synthesis of chemokines, which together with cytokines, cell adhesion molecules, and immunoreceptors, orchestrate the early host response to infection. At the same time they represent an important link in the adaptive immune response [40]. Our study revealed that the KEGG Toll-like receptor signaling pathway, by itself, was significantly dysregulated (with a combined  $p$  value of  $1.7 \times 10^{-30}$  after Bonferroni correction) in ASD, asthma, chronic kidney disease, dilated cardiomyopathy, ear infection, IBD, muscular dystrophy, and upper respiratory infection with the minimum posterior probability of appearing significant by chance being at most 1 %. In addition, the KEGG chemokine signaling pathway was found significantly dysregulated (with a combined  $p$  value of  $1.02 \times 10^{-21}$  after Bonferroni correction) in ASD, asthma, bacterial and viral infection, dilated cardiomyopathy, ear infection, IBD, and upper respiratory infection with the minimum posterior probability of appearing significant by chance being at most 2.4 % in each case. These findings indicate the role of immune dysfunction in this wide range of seemingly unconnected disease conditions. Although there is some experimental evidence linking an abnormal chemokine response to Toll-like receptor ligands associated with autism [41, 42], no study so far has linked them to the co-morbidities suffered by ASD-affected individuals.

When we looked at the other significant KEGG pathways, we found two others involved in innate immunity, namely, the NOD-like receptor signaling and leukocyte transendothelial migration pathways. The NOD-like receptor signaling pathway, by itself, was significantly dysregulated (with a combined  $p$  value of  $2.6 \times 10^{-15}$  after Bonferroni correction and a minimum posterior probability of the null hypothesis at most 4 %) in ASD, asthma, bacterial and viral infection, chronic kidney disease, dilated cardiomyopathy, ear infection, IBD, and upper respiratory infection. The leukocyte transendothelial migration pathway was significantly dysregulated (with a combined  $p$  value of  $1.4 \times 10^{-6}$  after Bonferroni correction and a minimum posterior probability of the null hypothesis at most 1.7 %) in ASD, asthma, cerebral palsy, and muscular dystrophy. Some NOD-like receptors recognize certain types of bacterial fragments; others induce caspase-1 activation through the assembly of multi-protein complexes called inflammasomes, which are critical for generating mature pro-inflammatory cytokines in concert with the Toll-like receptor signaling pathway. While the Toll-like receptor, chemokine, and NOD-like receptor signaling pathways have more to do with the recognition of infectious pathogens and initiating response, the leukocyte transendothelial migration pathway orchestrates the migration of leukocytes from blood into tissues via a process called diapedesis, which is vital for immune surveillance and inflammation. During this diapedesis of leukocytes, the leukocytes bind to endothelial cell adhesion molecules and then migrate across the vascular endothelium to the site of infection. Notably, increased permeability of the blood–brain barrier favoring leukocyte migration into the brain tissue has been implicated in ASD before [43], but not as a shared transcriptomic commonality among its co-morbidities.

**Table 3** KEGG pathways significantly shared among ASD and its co-morbidities<sup>a</sup>

Pathway	ASD	Asthma	Bacterial and viral infection	Chronic kidney disease	Cerebral palsy	Dilated cardiomyopathy	Ear infection	Epilepsy	Inflammatory bowel disease	Muscular dystrophy	Schizophrenia	Upper respiratory infection	Chi-square value	p value from chi-square distribution	Bonferroni corrected p value
Toll-like receptor signaling pathway	0.0048*	5.52E-06*	0.0762	0.0114*	0.6550	0.0034*	4.28E-16*	1	5.93E-05*	0.0210*	1	1.14E-10*	189.1151	1.1745E-32*	1.703E-30*
Chemokine signaling pathway	0.0145*	0.0003*	0.000051*	0.2197	0.8628	0.0194*	3.21E-10*	1	1.37E-06*	0.5703	1	8.89E-09*	170.8496	7.0449E-26*	1.022E-21*
NOD-like receptor signaling pathway	0.0342*	9.02E-05*	0.0136*	0.0019*	0.4760	0.0019*	1.99E-08*	1	0.0036*	0.7335	1	9.04E-05*	116.9434	1.7813E-17*	2.583E-15*
Ribosome	6.49E-13*	0.9647	4.84E-10*	0.1720	0.6006	1	0.984089	1	0.9460	0.0026*	1	1	119.0004	3.68E-17*	5.336E-15*
Spliceosome	6.70E-05*	0.9541	6.39E-06*	0.2965	0.3831	0.2746	0.920135	1	1.36E-05*	0.5081	0.1721	1	82.5337	9.9149E-09*	1.438E-06*
Leukocyte trans-endothelial migration	0.0023*	0.8201	0.0110*	0.0797	0.0002*	0.8164	0.097372	1	0.1238	7.63E-06*	0.5000	1	75.6280	9.962E-09*	1.445E-06*
Regulation of actin cyto-skeleton	0.0234*	0.9080	0.2734	0.1131	0.0745	0.0355*	0.227981	1	0.2032	5.90E-05*	0.1330	1	73.9701	2.7324E-05*	0.003962*
Tight junction	0.0359*	0.5613	0.4111	0.1064	0.0005*	0.8542	0.303934	1	0.1900	0.0006*	1	1	56.2763	6.9114E-05*	0.010022*

ASD autism spectrum disorder, KEGG Kyoto Encyclopedia of Genes and Genomes

<sup>a</sup>The entries with value '1' indicate where there was no overlap between the pathway and the disease gene set

\*Entries with significant p values



To confirm that the presence of multiple significant innate-immunity-related pathways among ASD and its co-morbidities was due to shared biology, we repeated the combined *p* value analysis excluding the immune-related diseases (bacterial and viral infection, asthma, IBD, upper respiratory infection, and ear infection). Innate immunity pathways (leukocyte transendothelial migration, Toll-like receptor signaling, and NOD-like receptor signaling pathways) still appeared among the most significant dysregulated pathways shared by ASD, cerebral palsy, chronic kidney disease, and muscular dystrophy. The QQ plot of combined *p* values of pathways across ASD and its non-immune-related co-morbidities shows marked enrichment of significant *p* values indicative of the shared disease biology of these conditions (Fig. 2b). Additional file 1: Table S2 shows the most significant KEGG pathways that are shared by ASD and its non-immune-related co-morbidities. For other pathway gene set collections, the complete lists of Fisher's combined *p* values per pathway per disease are provided in Additional file 6.

#### Disease–innate immunity pathway overlap at gene level

To examine the shared innate immunity KEGG pathways through a finer lens, we examined the genes that overlapped with them (Table 4 and Additional file 3: Figure S4). Although these pathways have a broad involvement in a variety of diseases, a small number of genes in these pathways appear dysregulated most often in ASD and its co-morbidities. Thus, we took a closer look at the genes that are shared by ASD and at least one of its co-morbid conditions.

In the Toll-like receptor signaling pathway, as shown in Fig. 3a, commonly shared, differentially expressed genes include CD14 and LY96 (also known as MD-2), responsible for mediating the lipopolysaccharide response, which itself has been shown to create an autism-like phenotype in murine model systems [44], but has never been linked to the shared biology of ASD, cerebral palsy, dilated cardiomyopathy, muscular dystrophy, and IBD. The widely expressed Toll-like receptors, especially, TLR1, TLR2, and TLR9, mediate the recognition of foreign substances, including infectious pathogens, and the regulation of the subsequent cytokine production required for the immune response. Although these genes have been known to be involved in immunity-related conditions, they have not been implicated in the co-occurrence of such conditions in ASD patients. Other genes involved were CCL4, also known as Macrophage inflammatory protein 1 $\beta$  (MIP-1 $\beta$ ), which is the most upregulated chemokine in natural killer cells of children with autism [45]; MAPK21, a gene upstream of the MAP-kinases that mediates multiple intra- and extra-cellular signals; JUN (a subunit of transcription factor AP-1), which regulates gene expression in response to a variety of stimuli, including cytokines,

growth factors, stress, and bacterial and viral infections; SPP1 (also known as OPN), a cytokine that upregulates expression of interferon- $\gamma$  (IFN- $\gamma$ ), which itself has been implicated in ASD and other diseases characterized by social dysfunction [46]; and TBK1, a gene that can mediate NF $\kappa$ B activation in response to certain growth factors and is often considered as a therapeutic target for inflammatory diseases.

In the chemokine pathway, as shown in Fig. 3b, the commonly shared genes include the chemokines (e.g., CCL4, which had altered expression levels in asthma and ear infection) and MAP-kinases (e.g., MAP2K1, which had altered expression levels in ASD, dilated cardiomyopathy, ear infection, and muscular dystrophy). The HCK gene, which belongs to the Src family of tyrosine kinases, showed altered expression levels in ASD, asthma, IBD, ear infection, bacterial and viral infection, and muscular dystrophy. Considering HCK's role in microglia and macrophages in controlling proliferation and cell survival [47], this finding is not surprising. JAK2, which is dysregulated in ASD and its multiple immune-related co-morbidities, regulates STAT3 activity, which in turn transduces interleukin-6 (IL-6) signals. Increased IL-6 in the maternal serum has been known to alter fetal brain development, impairing social behaviors in the offspring [48, 49]. The alpha and beta subunits of G-proteins, dysregulated in ASD, asthma, IBD, and bacterial and viral infections, are important signaling molecules, which are often considered to have weak links to a number of brain conditions. The RAP1B gene, a member of the RAS family, regulates multiple cellular processes including cell adhesion, growth and differentiation, and integrin-mediated cell signaling. This protein also plays a role in regulating outside-in signaling in platelets, and G-protein coupled receptor signaling. Thus, it may be of importance.

In the NOD-like receptor signaling pathway, the genes NOD1 and NOD2 drive the activation of NF $\kappa$ B and MAPK, the production of cytokines, and apoptosis. The BIRC2 and BIRC3 genes (which had altered expressions in ASD, asthma, ear infection, and bacterial and viral infections) are members of the inhibitor-of-apoptosis protein family and are key regulators of NOD1 and NOD2 innate immunity signaling. In the leukocyte transendothelial migration pathway, the TXK gene, which is a non-receptor tyrosine kinase (with altered expression in ASD, ear infection, IBD, and bacterial and viral infections), specifically regulates IFN- $\gamma$  gene transcription and the development, function, and differentiation of conventional T cells and nonconventional NKT cells. Mutation of the TXK gene has been identified to be a segregating factor for a number of neurodevelopmental disorders, including ASD, bipolar disorder, and intellectual disabilities [50].

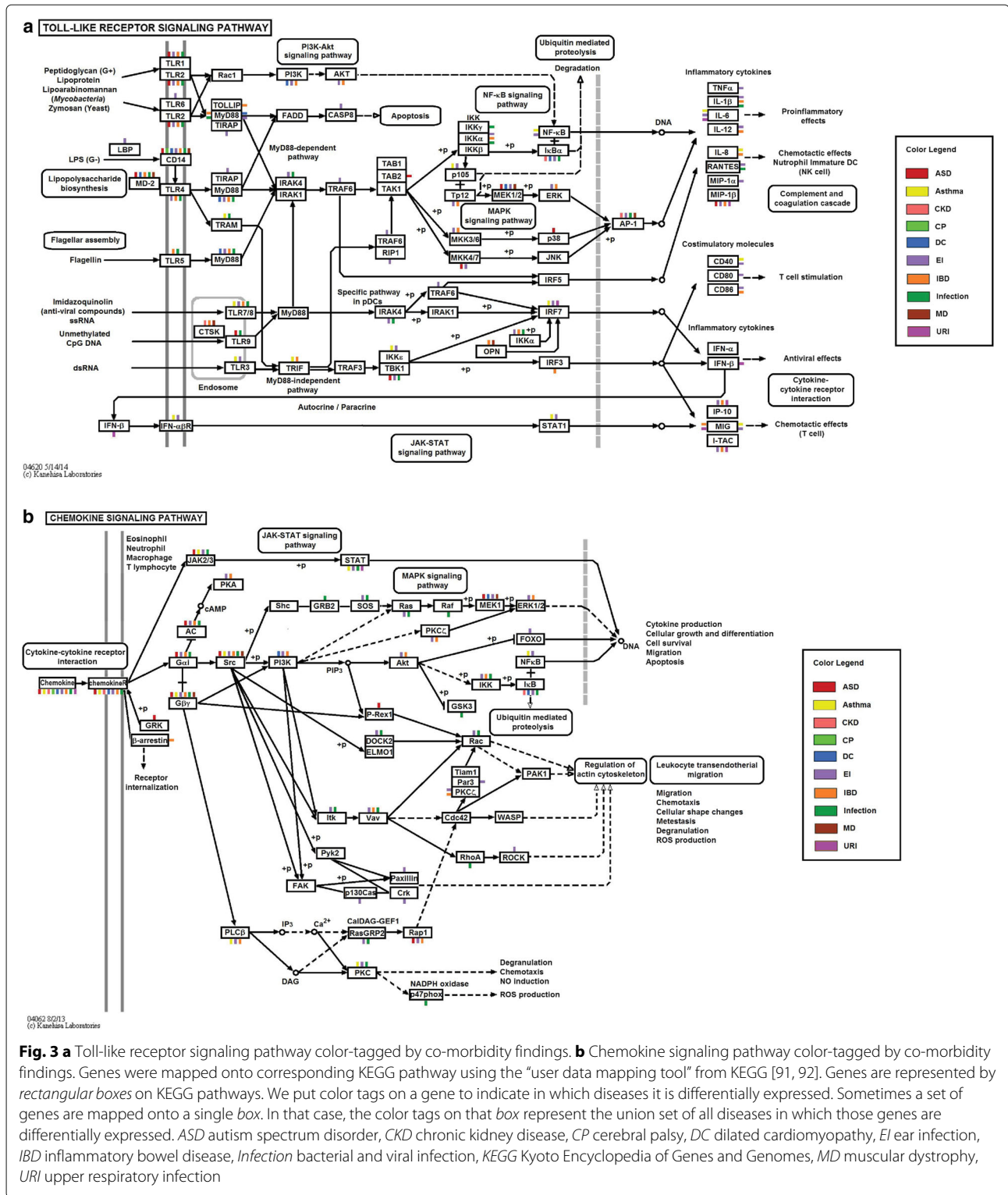
**Table 4** Differentially expressed genes in ASD and co-morbidities that overlap with innate immunity pathways

	Toll-like receptor signaling pathway	Chemokine signaling pathway	NOD-like receptor signaling pathway	Leukocyte transendothelial migration pathway
Autism spectrum disorder	TLR9, MAP2K4, CCL4, LY96, CD14, TAB2, MAP2K2, MAPK13, MAP2K1, TBK1, TLR1, TLR2	CCL4, JAK2, GRK7, CCL17, CCL21, CCL22, GNB3, GNAI2, CCR2, CXCR3, GNAI3, CCR10, ADCY6, PREX1, HCK, MAP2K1, RAP1B	BIRC3, MAPK13, SUGT1, PSTPIP1, PYCARD, TAB2, BIRC2	TXK, NCF2, JAM2, GNAI2, GNAI3, CLDN23, ACTN3, ICAM1, ACTN1, MAPK13, CD99, RAP1B, CLDN14, MSN
Asthma	STAT1, IKBKE, NFKB1, RELA, TLR7, TICAM1, IL8, IFNAR1, IFNAR2, TICAM2, CD40, CXCL9, TLR3, IL6, IRF7	STAT1, CCL2, GNB4, JAK2, CCL20, NFKB1, RELA, CXCL5, XCR1, PLCB1, CXCL1, PRKCD, HCK, IL8, CCL1, CXCL2, CXCL9, LYN	CXCL1, RIPK2, BIRC3, CCL2, IL8, CASP5, NFKB1, RELA, CXCL2, IL6	TXK, ACTN2, ICAM1
Chronic kidney disease	JUN, CTSK, NFKBIA, FOS	CCL17, NFKBIA, CXCR6	NFKBIA, HSP90AA1, NLRC4, HSP90AB1	CLDN16, ACTN4, CLDN9
Cerebral palsy	CD14	CCL2	CCL2	JAM3, MMP2, VCAM1, ACTN4, ACTG1, MSN, CTNNA3
Dilated cardiomyopathy	MYD88, LY96, CD14, NFKBIA, MAP2K1, PIK3R1	CCL2, CCL11, CCL8, NFKBIA, MAP2K1, CCR1, PIK3R1	RIPK2, CCL2, CCL11, CCL8, NFKBIA	PIK3R1
Ear infection	JUN, CD86, STAT1, CCL3, MYD88, CCL5, CCL4, LBP, TLR6, MAP3K8, CD14, IKBKE, NFKB1, NFKBIA, PIK3R5, TLR5, ITIRAP, RELA, TOLLIP, CXCL11, TLR7, TLR8, CXCL10, CASP8, TNF, IL12B, MAP2K3, MAP2K1, MAP2K6, MAPK3, IL1B, CD40, TBK1, CXCL9, TLR3, TLR4, TLR1, FOS, TLR2, IL6, IRF7, PIK3R2	STAT3, STAT1, STAT2, CCL2, CCL3, CCL5, CCL4, CX3CR1, CCL11, CCL7, CXCL14, JAK3, JAK2, CCL17, CCL20, CCL19, CCL22, NFKB1, NFKBIB, NFKBIA, PIK3R5, RELA, GNG7, FGR, GNG11, GNGT2, XCL1, CXCL5, ADCY3, CXCL11, ADCY2, CXCL10, CXCL1, PLCB3, CXCR5, CXCR2, GNG8, HCK, MAP2K1, CCR5, CCR7, MAPK3, CXCL16, CCR1, CXCL13, CXCL2, CXCL3, CXCL9, LYN, PIK3R2	CASP8, CXCL1, RIPK2, TNF, BIRC3, CCL2, CCL5, CCL11, CCL7, MEFV, CASP1, TNFAIP3, MAPK3, NFKB1, NFKBIB, NFKBIA, IL18, RELA, IL1B, NLRP3, BIRC2, CXCL2, IL6	TXK, NCF4, VCAM1, PIK3R5, CLDN23, CLDN10, CLDN8, MYL9, CLDN5, ICAM1, ACTN4, CLDN19, CLDN22, RASSF5, CLDN7, CLDN4, PIK3R2
Epilepsy	-	-	-	-

**Table 4** Differentially expressed genes in ASD and co-morbidities that overlap with innate immunity pathways (*Continued*)

	Toll-like receptor signaling pathway	Chemokine signaling pathway	NOD-like receptor signaling pathway	Leukocyte transendothelial migration pathway
Inflammatory bowel disease	CD86, MYD88, CCL4, LY96, MAP3K8, AKT1, CD14, CTSK, SPP1, TOLLIP, CXCL11, TLR8, CXCL10, TICAM1, CHUK, IL8, MAP2K3, IL12A, MAPK3, IRF3, IL1B, PIK3CA, CXCL9, TLR4, TLR1, TLR2	CCL2, CCL4, CCL11, CCL7, AKT1, CCL18, ARRB2, CCL20, GNG5, GNB3, GNB2, CCL24, PRKX, GNG10, GNAI2, GNG11, XCL1, CXCL11, CXCL6, CCR10, CXCL10, ADCY6, CHUK, CXCL1, PLCB3, CXCR2, CXCR1, HCK, IL8, ADCY4, PRKCZ, MAPK3, CCR1, XCL2, CXCL13, RAP1A, PF4, CXCL2, CXCL3, PF4V1, PIK3CA, CXCL9, PPBP, VAV3, LYN	CHUK, CXCL1, BIRC3, CCL2, CARD6, CCL11, CCL7, IL8, CASP5, CASP1, MAPK3, IL1B, NLRP1, CXCL2, HSP90AB1	TXK, ITGA4, NCF4, MMP9, NCF2, MYL12A, THY1, GNAI2, MYL5, RHOH, CLDN8, MYL9, CLDN15, MYL12B, RAP1A, PIK3CA, MSN, VAV3
Bacterial and viral infection	TLR9, MYD88, CCL5, LY96, CD14, NFKBIA, TLR5, TLR8, CHUK, IRAK4, MAP2K7, IRF3, IKBK, TICAM2, IL1B, TBK1, TLR4, TLR1, FOS, TLR2, IRF7	STAT3, STAT2, CCL5, DOCK2, JAK2, VAV2, NRAS, NFKBIA, GNG7, GNG5, GNB2, GNG10, GNG11, ADCY3, CXCR3, CCL4L1, GNAI3, GNB1, SOS2, CHUK, RAF1, RHOA, CXCR2, CXCR1, PRKCD, HCK, RAC2, RASGRP2, ADCY4, CCR3, CCR4, CXCR6, CCR7, GRB2, IKBK, HRAS, GSK3B, CCR1, ITK, NCF1, PPBP, LYN	CHUK, RIPK2, CCL5, NOD1, CARD6, CCL8, CARD8, CASP5, NOD2, NFKBIA, IKBK, PYCARD, NLR4, IL1B, BIRC2	TXK, ITGAM, NCF4, MMP9, NCF2, VAV1, VASP, MYL12A, VAV2, ITGB2, CTNNA1, GNAI3, EZR, PLCG1, RHOH, PRKCA, ESAM, RAC2, CD99, ITK, NCF1, CYBA, CYBB, MYL5, RHOA
Muscular dystrophy	LY96, CD14, CTSK, SPP1, MAP2K1, FOS	GNG10, HCK, MAP2K1, GNG12	PYCARD	JAM3, MMP2, NCF2, VCAM1, ITGB2, JAM2, MYL5, CD99, ACTG1, MYL12B, MSN, CYBA
Schizophrenia	–	–	–	CD99
Upper respiratory infection	CCL4, CXCL11, CXCL10, IFNB1, CXCL9, IL6, IRF7	STAT2, CCL2, CCL4, CCL7, CXCL11, CXCL10, CXCL9	CCL2, CCL7, IL6	–

ASD autism spectrum disorder



Besides the immune-related ones, Table 3 documents several other pathways and gene sets including the ribosome and spliceosome gene sets, which have roles in genetic information processing and translation and the

actin cytoskeleton regulation pathway, which controls various cellular processes like cell motility. Neuronal signal processing and neuron motility have often been associated with ASD, thus these findings are not surprising. The

genes in the tight junction pathway mediate cell adhesion and are thought to constitute the intra-membrane and para-cellular diffusion barriers. These findings implicate the involvement of these cellular processes in the shared pathology of ASD and its co-morbidities.

#### Discriminatory power of innate immunity pathway genes

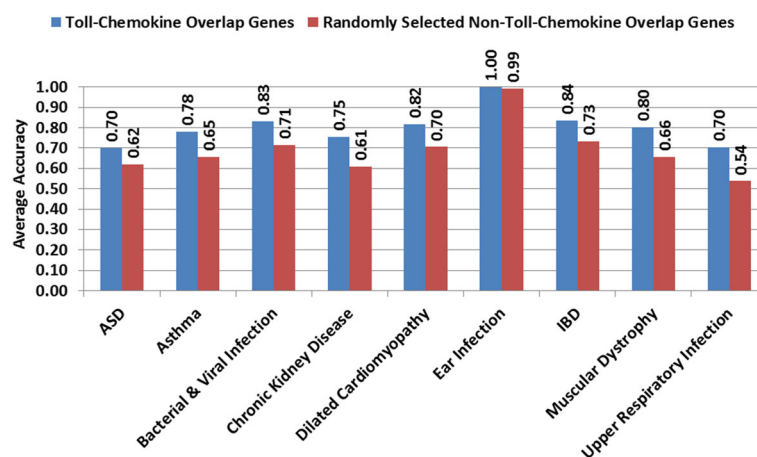
We assessed the discriminatory power of the innate immunity pathway genes, by taking the union of the genes from the chemokine signaling and Toll-like receptor signaling pathways and performing threefold SVM classification of cases vs. controls for each of the 12 disease conditions. We could achieve an average accuracy of at least 70 % (Fig. 4). We also performed the same classification using the same number of randomly selected genes that do not overlap with these pathways. With randomly selected genes, the classification accuracy was much lower. This result suggests that the genes that have altered expressions in the diseases examined and are present in these innate immunity pathways were sufficient to partially distinguish the disease states from the controls. When we included the overlapping genes in the NOD-like receptor signaling and transendothelial migration pathways in this analysis, the classification accuracy was at least 65 % (see Additional file 3: Figure S5), which was still better than for the randomly selected non-immune genes. In fact, a recent functional genomic study showed that immune/inflammation-related genes can provide reasonable accuracy in the diagnostic classification of male infants and toddlers with ASD [51].

#### Discussion

This study bridges previous analyses based on the electronic health records of the co-morbidities of large

populations of individuals with ASD and the gene expression profiles of each of these co-morbid diseases as well as ASD against their respective control cases. We have identified that the most significantly and consistently dysregulated pathways shared by these diseases are the innate immunity signaling pathways. For most of these disorders, the genes in these pathways can classify the disorders with respect to their controls with moderate accuracy, further evidence of the extent of the dysregulation in these pathways.

In contrast to traditional approaches that look at a group of disorders of the same organ system, we have focused on ASD and its co-morbidities, which often occur in different organ systems, with a view to finding their shared genetics. It would have been ideal to perform the study on a sufficiently large cohort of ASD patients having enough representatives of all the co-morbid diseases, but in practice, such a study is currently infeasible due to cost constraints and/or patient availability. Thus, to perform this study with existing data sets for ASD and its co-morbidities, we make use of the power of statistics and computation. First, we look at the functional genomic makeup of patients with ASD and its co-morbid diseases separately, and then find the commonalities between them. Some of the microarray studies we looked at have small sample sizes, which gives rise to the possibility of poor random error estimates and inaccurate statistical tests for differential expression. For this reason, we selected limma *t*-statistics, an empirical Bayes method [52], which is reportedly one of the most effective methods for differential expression analysis even for very small data sets [53]. To find the combined significance of the pathways across multiple diseases, we used Fisher's combined probability test [39], because, it gives a single test



**Fig. 4** Accuracy of classification for case-control groups in different diseases using differentially expressed genes that overlap in the KEGG Toll-like receptor signaling and chemokine signaling pathways versus randomly selected disease genes that do not overlap in the innate immunity pathways. Diseases for which the differentially expressed genes are not over-represented in the Toll-like receptor signaling and chemokine signaling pathways, are omitted here. *ASD* autism spectrum disorder, *IBD* inflammatory bowel disease, *KEGG* Kyoto Encyclopedia of Genes and Genomes

of significance for a number of not-so-correlated tests of significance performed on very heterogeneous data sets. When the individual tests do not appear as significant, yet have a combined effect, Fisher's combined  $p$  value can indicate whether the probability of the combined effect is on the whole lower than would often have been obtained by chance. Notably, a significant statistic from Fisher's test implies that the pathway is involved in the biology of at least one of the diseases. Thus, to ensure that the combined significant statistic is due to the shared biology of multiple diseases, we calculate minimum BFs and minimum posterior probabilities of significance by chance for each significant pathway, and also compare the combined  $p$  value distributions of diseases and the null data set using QQ plots. We draw our conclusions using a combination of the  $p$  values and the posteriors to avoid any systematic bias inherent to the methods used.

As expected for a neurological disease, the pathways that are most significantly dysregulated in ASD are often the pathways involved in neuronal signaling and development, synapse function, and chromatin regulation [12]. Similarly, for immune-related diseases, like, asthma, IBD, and various infections, the role of innate immunity pathways is well documented in individual studies [54–60]. Despite some controversy, in the last 15 years, experimental evidence has also pointed in the direction of dysregulated immunological signaling in at least some subsets of individuals with autism. This evidence includes findings of an abnormal chemokine response to Toll-like receptor ligands associated with autism in experimental studies [41, 42], and differential gene and protein expression in the central nervous system and peripheral blood of patients with ASD [35, 41, 61–68]. Many reports suggest the alteration of the activation, amount, and distribution of microglia, a representative immune cell in the brain, and its autophagy to be involved in ASD [69–72]. A recent study implicates adaptive immune dysfunction, in particular, disruption of the IFN- $\gamma$  signaling driven anti-pathogen response, to be related to ASD and other diseases characterized by social dysfunction [46]. However, that dysregulation of innate immunity pathways connects ASD with some of its non-immune-related co-morbidities (e.g., chronic kidney disease, cerebral palsy, and muscular dystrophy) is rather intriguing.

That the innate immunity pathways are shared between ASD and the other co-morbid states does not mean that all cases of ASD are characterized by a disorder in these pathways. For example, in our previous work we have shown that although, on average, the gene expression profile of children with ASD shows dysregulated innate immunity signaling, this is a reflection of the smaller number of individuals with ASD who are outliers in this pathway [73]. With our growing understanding of the heterogeneity of ASD and the characterization of ASD populations

with distinct co-morbidity associations [25], the integrative analysis we describe here may, therefore, implicate a subset of individuals with ASD with innate immune dysregulation that is either the result of genetic vulnerabilities [74] or particular exogenous stimuli such as infections or disordered microbiome ecologies [75].

Although it is tempting to consider that innate immunity signaling is primarily driven by external environmental stimuli such as infection, we have to recognize that the same signaling mechanisms may be repurposed by different organs for different purposes. For example, 21 % of the genes described in the KEGG long-term potentiation pathway (one of the mechanisms underlying synaptic plasticity) overlap with the genes in the Gene Ontology's collection of immune genes. It may be, as suggested by large epidemiological studies, that sometimes the disorder is in the signaling system and at other times it is because of an external stimulus. Specifically, nationally scaled studies have demonstrated increased autoimmune disease frequency in the parents of children with ASD [76], increased gestational C-reactive protein in mothers of children with ASD [77], and increased frequency of ASD after pregnancies complicated by infection [78, 79]. Some early studies also suggest the infectious exposure may be directly from the gastrointestinal microbiome [80–84], which also can engage the innate immune system. The success of treatment and/or prophylaxis for disorders of innate immunity in some of the diseases that are co-morbid with ASD raises the possibility that similar treatments may also be successful for subsets of those with ASD.

## Conclusions

Over the years, ASD has baffled researchers not only with its heterogeneity, but also its co-occurrence with a number of seemingly unrelated diseases of different organ systems. In this study, we introduced a three-tier meta-analysis approach to capture the shared genetic signals that form the basis of ASD's co-occurrence with other diseases. For ASD and 11 of its most frequently occurring co-morbidities, we extracted significant differentially expressed genes, measured their enrichment in canonical pathways, and determined the pathways that are shared by the diseases in question in a statistically rigorous fashion. An analysis of this scale for studying ASD and its co-morbidities is unheard of as per our knowledge. Our results reveal the involvement of two disrupted innate immunity pathways – Toll-like receptor signaling and chemokine signaling – in ASD and several of its co-morbidities irrespective of whether they are immune-related diseases or not. We also showed that the disease genes that overlapped with these pathways could discriminate between patients and controls in each disease with at least 70 % accuracy, further proving their

importance. As innate immunity pathways are imperative in orchestrating the first line-of-defense mechanism against infection-causing pathogens and environmental triggers, their involvement in ASD and its co-morbidities can be thought of as the missing genetic link for environmental factors in the pathophysiology of ASD. This mindset also raises the possibility that successful treatments for innate immunity disorders may help ASD patients.

## Methods

### Overview of the three-tiered meta-analysis

To analyze genome-wide expression studies across ASD and 11 of its co-morbidities (Table 1), we introduced a step-wise three-tiered meta-analysis pipeline (Fig. 1). Our meta-analysis started at the gene level, in which we first identified the genes that are differentially expressed among cases and controls for a given disease. We then extended this analysis to the pathway level, where we investigated the pathways that were significantly enriched in candidate genes for a given disease. Finally, we identified the pathways that were significant across multiple diseases by newly combining pathway-level results across diseases and performing a Bayesian posterior probability analysis of null hypotheses for pathways in each disease as well as in the combined case. Details are described below.

### Gene-centric expression analysis per disease

Using the GEOquery package [85] from Bioconductor in R, we downloaded the gene expression data for each disease in gene matrix transposed (GMT) format from the Gene Expression Omnibus (GEO). The accession identifiers for the disease studies are listed in Additional file 1: Table S1. We removed 'NA' values from the data and log-normalized the expression values for subsequent analysis. Then, we performed differential expression analysis on each data set using the limma package [52] from Bioconductor in R, and obtained  $p$  values for each gene in each experiment.

To determine the degree of correlation between the differential expression analyses of the  $p$  values of data sets selected under each disease, we calculated the pairwise Pearson correlation coefficient of  $p$  values (Additional file 1: Table S3). Considering a Pearson correlation coefficient of at least 0.30 with  $p < 0.05$  as significant, we found that the  $p$  values are not significantly correlated. This lack of correlation allowed us to use Fisher's combined probability test to calculate combined  $p$  values for the genes in each disease condition. We used Fisher's combined probability test as follows:

$$P \sim \chi^2 = -2 \sum_{i=1}^k \ln(p_i).$$

Here,  $p_i$  is the  $p$  value of test  $i$ ,  $\chi^2$  is the chi-squared distribution,  $k$  is the number of tests, and  $P$  is the adjusted  $p$  value ( $p < 0.05$  was considered significant).

### Selecting the most informative FDR correction test for multiple comparisons

To adjust the combined  $p$  values, we considered different FDR corrections [i.e., Bonferroni, Benjamini–Yekutieli (BY), and Benjamini–Hochberg (BH)]. We also considered the 'no correction' case for completeness. We selected the most informative one, based on the level of accuracy we could achieve in classifying cases of a particular disease, vs. controls, using the genes selected under a specific test with a significance cutoff of  $p < 0.05$ . We tested the accuracy of the case–control classification for each of the 53 disease data sets using four different classification methods, namely, naive Bayes method, Fisher's linear discriminant analysis,  $k$  nearest neighbor, and SVM. The set of significant genes selected under different FDR corrections was considered as a feature of the classification methods. We performed threefold cross validation and calculated the average accuracy. We selected the FDR correction test that produced the best average accuracy in each disease. See Additional file 3: Figure S1 and the supplementary text on different classification techniques for microarray gene expression data provided in Additional file 7 for more details.

### Pathway-centric enrichment analysis per disease

From the disease-level gene-centric expression analysis, we obtained a list of significant genes per disease. For each disease, we then performed a hypergeometric enrichment test for each pathway. This test uses the hypergeometric distribution to calculate the statistical significance of  $k$  or more significant disease genes, out of  $n$  total genes, appearing in a specific pathway gene set. It helps identify whether or not the specific disease gene set is over-represented in a certain pathway, by providing a  $p$  value per pathway per disease.

### Disease-centric analysis of pathways

Once we obtained the  $p$  values for the pathways per disease, first we calculated the pairwise Pearson correlation of pathway  $p$  values across diseases (Additional file 1: Table S4). Since the distributions were not significantly correlated (Pearson correlation coefficient  $< 0.30$  with  $p$  value  $< 0.05$ ), we safely assumed the distributions to be independent. Next, we calculated combined  $p$  values for each pathway across all the diseases using Fisher's combined probability test. We corrected for multiple comparisons using Bonferroni correction. We defined a significance threshold of adjusted  $p$  value  $< 0.05$  and called any pathway that passed this threshold, significant. We restricted our results to the pathways that appeared significant in ASD.

### Calculation of priors, minimum BFs, and minimum posterior probabilities of null hypotheses

To estimate the prior probability of pathways, we selected a publicly available GEO study of 109 gene expression profiles of blood drawn from healthy individuals enrolled at a single site (GEO accession: GSE16028). We assigned case-control labels randomly to the samples and performed differential expression analysis using R package limma. We selected differentially expressed genes using uncorrected  $p$  values ( $<0.05$ ), because after BY correction none of the genes remained significant. On the significant gene list, we performed hypergeometric enrichment analysis to obtain a pathway  $p$  value distribution. We repeated this process 100 times to obtain 100 null  $p$  value distributions. We calculated the prior for each pathway by looking at how many times the pathway appeared significant ( $p$  value  $<0.05$ ) during these 100 runs. We took an average of the 100 distributions to obtain the null  $p$  value distribution.

The null hypothesis for pathway  $p$  values is that  $p$  values are uniformly distributed and the alternative hypothesis is that smaller  $p$  values are more likely than larger  $p$  values. Following the approach of Sellke, Bayarri, and Berger [86], we estimated the minimum BF using the following formula:

$$\text{BF} = \begin{cases} -ep \log(p), & \text{if } p < \frac{1}{e}, \\ 1, & \text{otherwise,} \end{cases}$$

where  $e$  is Euler's constant.

For calculating minimum BF for  $\chi^2$ -distributed test statistics, we used Johnson's formula [87]:

$$\text{BF} = \begin{cases} \left(\frac{\nu}{x}\right)^{-\frac{\nu}{2}} \exp\left(-\frac{x-\nu}{2}\right), & \text{for } x > \nu, \\ 1, & \text{otherwise,} \end{cases}$$

where  $x$  is the chi-square statistic that gave rise to the observed  $p$  value and  $\nu$  is the degrees of freedom.

Following Goodman's approach [88], we used the prior probability distribution drawn from the null data set and the minimum BF to estimate a lower bound on the posterior probability of the null hypothesis based on Bayes' theorem as follows:

$$\text{Minimum Posterior Probability} = \left(1 + \left(\frac{\text{BF} \times q}{1 - q}\right)^{-1}\right)^{-1}$$

where  $q$  is the prior probability.

The null distributions and priors for all KEGG pathways and the minimum BFs, and minimum posterior probabilities of null hypotheses for KEGG pathways are given in Additional file 5.

### Measuring the discriminatory power of overlapping innate immunity genes

We performed threefold classification and measured the average accuracy of the case-control classification for

each disease with the SVM classifier using the union set of the genes from KEGG Toll-like receptor signaling and chemokine signaling pathways shared across ASD and its co-morbidities to see how well the overlapping genes could distinguish the disease state from controls and compared it with the classification accuracy using randomly selected genes that do not overlap with these two pathways (Fig. 4). We repeated the same test for the overlapping genes in the four innate immunity KEGG pathways and compared the classification accuracy with the discriminatory power of randomly selected non-immunity genes (Additional file 3: Figure S5).

### Data set selection

#### Gene expression data sets

We selected 11 disease conditions that co-occur most commonly in ASD patients. Each of these diseases has at least 5 % prevalence in ASD patients [25]. The prevalence of a co-morbid condition can be defined in two ways: (i) the percentage of ASD patients having a co-morbid disease and (ii) the percentage of patients with a co-morbid disease having ASD [24]. The diseases that satisfy either of these criteria include asthma, bacterial and viral infection, cerebral palsy, chronic kidney disease, dilated cardiomyopathy, ear infection/otitis media, epilepsy, IBD, muscular dystrophy, schizophrenia, and upper respiratory infection. Table 1 shows the disease groups along with the literature references.

To identify publicly available studies relevant to these co-morbidities, we performed an extensive literature search of the GEO of the National Center for Biotechnology Information (NCBI) [89, 90]. Using the advanced search tool provided by GEO, we searched series data sets from studies that performed expression profiling by array on either human or mouse. The search results were parsed using a custom-built parser. It identified 1329 GEO studies for ASD and 11 of its co-morbidities that have been publicly available since 2002. We verified the search results by hand to remove false positives. From the hand-curated results, we retained only those series that corresponded to case-control studies and had complete gene annotations supplied by either NCBI or the submitter. We investigated whether case-control studies had matched controls for the disease cases as well as to reduce noise. We made sure that we had at least 30 samples under each disease. For each selected GEO series, the accession identifier as well as abridged study details including the organism, tissue type, platform, and number of samples is provided in Additional file 1: Table S1. To remove the potential for biases that could arise from using gene expression data sets from different array platforms, tissues, and species, we avoided combining the actual measurements of expression values across platforms, tissues, and diseases.



Instead, we performed differential expression analysis on each study separately and then combined the  $p$  values only.

### Pathway gene sets

We collected 1320 curated pathway gene sets, including those from the KEGG pathways [91, 92], Reactome pathways [93, 94], BioCarta pathways [95], PID pathways [96], SigmaAldrich gene sets, Signaling Gateway gene sets, Signal Transduction KE gene sets, and SuperArray gene sets from the Molecular Signatures Database (MSigDb) version 4.0 [97]. The gene sets were downloaded in GMT format. Of the available gene sets, we used those that were expert-curated: C2:CP (canonical pathways), C2:CP-BioCarta (BioCarta gene sets), C2:CP-KEGG (KEGG gene sets), C2:CP-Reactome (Reactome gene sets), and PID (Pathway Interaction Database gene sets extracted from C2). From the KEGG collection, we excluded the disease- and drug-related gene sets. After excluding too large (>300 genes) and too small (<10 genes) gene sets, 1261, 146, 211, 629, and 196 gene sets remained in these categories, respectively.

### Additional files

**Additional file 1:** Supplementary tables. This PDF file contains supplementary Tables S1 through S4. (PDF 218 kb)

**Additional file 2:** Differential expression analysis of genes. This Excel file contains differential expression analysis  $p$  values per gene per disease under different FDR corrections [i.e., Bonferroni, Benjamini–Yekutieli (BY), and Benjamini–Hochberg (BH)] as well as ‘no correction’. For presentation purposes, the genes that are not significant even under the ‘no correction’ case, have been omitted. (XLS 54 kb)

**Additional file 3:** Supplementary figures. This PDF file contains supplementary Figures S1 through S5 and their captions. (PDF 8110 kb)

**Additional file 4:** Pathway enrichment analysis. This Excel file contains hypergeometric test  $p$  values per pathway per disease for KEGG, BioCarta, Reactome, and PID pathway collections as well as all canonical pathway gene sets collected from MSigDB version 4.0, and Fisher’s combined  $p$  values for ASD and its co-morbidities. (XLS 1444 kb)

**Additional file 5:** Posterior probability analysis. This Excel file contains the minimum Bayes factors and minimum posterior probabilities for null hypotheses for the significant KEGG pathways. It also contains the 100 pathway  $p$  value distributions generated by permuting the null data set. Finally, it has Fisher’s combined  $p$  values for ASD and its co-morbidities with the null  $p$  value distribution (one sheet per disease). (XLSX 380 kb)

**Additional file 6:** Pathway enrichment analysis for non-immune diseases. This Excel file contains hypergeometric test  $p$  values per pathway per disease for KEGG, BioCarta, Reactome, and PID pathway collections as well as all canonical pathway gene sets collected from MSigDB version 4.0, and Fisher’s combined  $p$  values for ASD and its co-morbidities excluding the immune-related diseases: bacterial and viral infection, asthma, inflammatory bowel disease, upper respiratory infection, and ear infection. (XLS 673 kb)

**Additional file 7:** Supplementary text. This PDF file contains the supplementary text describing the four different classification techniques we used for classifying cases vs. controls in microarray gene expression data from ASD and its co-morbidities. (PDF 196 kb)

### Abbreviations

ASD: Autism spectrum disorder; BF: Bayes factor; BH: Benjamini–Hochberg correction; BY: Benjamini–Yekutieli correction; CNV: Copy number variation; FDR: False discovery rate; GEO: Gene Expression Omnibus; GMT: Gene matrix transposed; IBD: Inflammatory bowel disease; IFN: Interferon; IL: interleukin; KEGG: Kyoto Encyclopedia of Genes and Genomes; MSigDB: Molecular Signatures Database; NCBI: National Center for Biotechnology Information; PID: Pathway Interaction Database; QQ plot: Quantile–quantile plot; SNP: Single nucleotide polymorphism; SVM: Support vector machine

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### Availability of data and materials

All microarray expression studies included in this analysis are publicly available via the GEO website [98]. The accession ID for each study is provided in Additional file 1: Table S1. All the pathway gene sets used for the analysis are publicly available from the MSigDB website upon registration [99]. All calculations were performed in R version 2.15.1 and Microsoft Excel 2010. Some pre- and post-processing was performed in Python version 2.7.6. The source code and instructions for performing the analysis are licensed under the terms of the MIT License (<https://opensource.org/licenses/MIT>), and are available from <https://github.com/snz20/3TierMA> (DOI:10.5281/zenodo.159288).

### Authors’ contributions

SN contributed to the study design and data set curation, performed statistical analyses, and wrote the manuscript. NP contributed to the study design and data curation, and advised on statistical analyses. BB and IK led the study design, advised on statistical analyses, and wrote the manuscript with SN. All authors have read and approved the manuscript for publication.

### Competing interests

The authors declare that they have no competing interests.

### Ethics approval and consent to participate

No ethical approval was required for this study. This study analyses only existing publicly available data generated by others who obtained ethical approval.

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### References

- Loscalzo J, Kohane I, Barabási AL. Human disease classification in the postgenomic era: a complex systems approach to human pathobiology. *Mol Syst Biol.* 2007;3(1):124.
- Barabási A-L, Gulbahce N, Loscalzo J. Network medicine: a network-based approach to human disease. *Nat Rev Genet.* 2011;12(1):56–68.
- Sirota M, Dudley JT, Kim J, Chiang AP, Morgan AA, Sweet-Cordero A, et al. Discovery and preclinical validation of drug indications using compendia of public gene expression data. *Sci Transl Med.* 2011;3(96):77.
- Cuthbert BN, Insel TR. Toward the future of psychiatric diagnosis: the seven pillars of RDoC. *BMC Med.* 2013;11(1):126.

5. Insel TR. Mental disorders in childhood: shifting the focus from behavioral symptoms to neurodevelopmental trajectories. *JAMA*. 2014;311(17):1727–8.
6. Iossifov I, Ronemus M, Levy D, Wang Z, Hakker I, Rosenbaum J, et al. De novo gene disruptions in children on the autistic spectrum. *Neuron*. 2012;74(2):285–99.
7. Neale BM, Kou Y, Liu L, Ma'Ayan A, Samocha KE, Sabo A, et al. Patterns and rates of exonic de novo mutations in autism spectrum disorders. *Nature*. 2012;485(7397):242–5.
8. Sanders SJ, Murtha MT, Gupta AR, Murdoch JD, Raubeson MJ, Willsey AJ, et al. De novo mutations revealed by whole-exome sequencing are strongly associated with autism. *Nature*. 2012;485(7397):237–41.
9. O'Roak BJ, Vives L, Girirajan S, Karakoc E, Krumm N, Coe BP, et al. Sporadic autism exomes reveal a highly interconnected protein network of de novo mutations. *Nature*. 2012;485(7397):246–50.
10. Malhotra D, Sebat J. CNVs: harbingers of a rare variant revolution in psychiatric genetics. *Cell*. 2012;148(6):1223–41.
11. Yu TW, Chahrour MH, Coulter ME, Jiralerspong S, Okamura-Ikeda K, Ataman B, et al. Using whole-exome sequencing to identify inherited causes of autism. *Neuron*. 2013;77(2):259–73.
12. Pinto D, Delaby E, Merico D, Barbosa M, Merikangas A, Klei L, et al. Convergence of genes and cellular pathways dysregulated in autism spectrum disorders. *Am J Hum Genet*. 2014;94(5):677–94.
13. Lin GN, Corominas R, Lemmens I, Yang X, Tavernier J, Hill DE, et al. Spatiotemporal 16p11.2 protein network implicates cortical late mid-fetal brain development and KCTD13-Cul3-RhoA pathway in psychiatric diseases. *Neuron*. 2015;85(4):742–54.
14. Smoller J, Craddock N, Kendler K, Lee P, Neale B, Nurnberger J, et al. Identification of risk loci with shared effects on five major psychiatric disorders: a genome-wide analysis. *Lancet*. 2013;381(9875):1371–9.
15. Levy D, Ronemus M, Yamrom B, Lee Y-H, Leotta A, Kendall J, et al. Rare de novo and transmitted copy-number variation in autistic spectrum disorders. *Neuron*. 2011;70(5):886–97.
16. Marshall CR, Noor A, Vincent JB, Lionel AC, Feuk L, Skaug J, et al. Structural variation of chromosomes in autism spectrum disorder. *Am J Hum Genet*. 2008;82(2):477–88.
17. Bijlsma E, Gijbbers A, Schuurs-Hoeijmakers J, Van Haeringen A, Van De Putte DF, Anderlid BM, et al. Extending the phenotype of recurrent rearrangements of 16p11.2: deletions in mentally retarded patients without autism and in normal individuals. *Eur J Med Genet*. 2009;52(2):77–87.
18. McCarthy SE, Makarov V, Kirov G, Addington AM, McClellan J, Yoon S, et al. Microduplications of 16p11.2 are associated with schizophrenia. *Nat Genet*. 2009;41(11):1223–7.
19. Weiss LA, Shen Y, Korn JM, Arking DE, Miller DT, Fossdal R, et al. Association between microdeletion and microduplication at 16p11.2 and autism. *N Engl J Med*. 2008;358(7):667–75.
20. Pinto D, Pagnamenta AT, Klei L, Anney R, Merico D, Regan R, et al. Functional impact of global rare copy number variation in autism spectrum disorders. *Nature*. 2010;466(7304):368–72.
21. Béna F, Bruno DL, Eriksson M, van Ravenswaaij-Arts C, Stark Z, Dijkhuizen T, et al. Molecular and clinical characterization of 25 individuals with exonic deletions of NRXN1 and comprehensive review of the literature. *Am J Med Genet B Neuropsychiatr Genet*. 2013;162(4):388–403.
22. Moreno-De-Luca D, Sanders S, Willsey A, Mulle J, Lowe J, Geschwind D, et al. Using large clinical data sets to infer pathogenicity for rare copy number variants in autism cohorts. *Mol Psychiatr*. 2013;18(10):1090–5.
23. Carter M, Scherer S. Autism spectrum disorder in the genetics clinic: a review. *Clin Genet*. 2013;83(5):399–407.
24. Kohane IS, McMurry A, Weber G, MacFadden D, Rappaport L, Kunkel L, et al. The co-morbidity burden of children and young adults with autism spectrum disorders. *PLoS ONE*. 2012;7(4):33224.
25. Doshi-Velez F, Ge Y, Kohane IS. Comorbidity clusters in autism spectrum disorders: an electronic health record time-series analysis. *Pediatrics*. 2014;133(1):54–63.
26. Mouridsen SE, Rich B, Isager T. Epilepsy in disintegrative psychosis and infantile autism: a long-term validation study. *Dev Med Child Neurol*. 1999;41(02):110–14.
27. Tuchman R, Rapin I. Epilepsy in autism. *Lancet Neurol*. 2002;1(6):352–8.
28. Horvath K, Papadimitriou JC, Rabsztyan A, Drachenberg C, Tildon JT. Gastrointestinal abnormalities in children with autistic disorder. *J Pediatr*. 1999;135(5):559–63.
29. Horvath K, Perman JA. Autistic disorder and gastrointestinal disease. *Curr Opin Pediatr*. 2002;14(5):583–7.
30. Richdale AL, Schreck KA. Sleep problems in autism spectrum disorders: prevalence, nature, & possible biopsychosocial aetiologies. *Sleep Med Rev*. 2009;13(6):403–11.
31. Wu JY, Kuban KC, Allred E, Shapiro F, Darras BT. Association of Duchenne muscular dystrophy with autism spectrum disorder. *J Child Neurol*. 2005;20(10):790–5.
32. Hendriksen J, Vles J. Neuropsychiatric disorders in males with duchenne muscular dystrophy: frequency rate of attention-deficit hyperactivity disorder (ADHD), autism spectrum disorder, and obsessive-compulsive disorder. *J Child Neurol*. 2008;23(5):477–81.
33. Hinton VJ, Cyrulnik SE, Fee RJ, Batchelder A, Kiefel JM, Goldstein EM, et al. Association of autistic spectrum disorders with dystrophinopathies. *Pediatr Neurol*. 2009;41(5):339–46.
34. Morgan CN, Roy M, Chance P. Psychiatric comorbidity and medication use in autism: a community survey. *Psychiatrist*. 2003;27(10):378–81.
35. Meyer U, Feldon J, Dammann O. Schizophrenia and autism: both shared and disorder-specific pathogenesis via perinatal inflammation?. *Pediatr Res*. 2011;69:26–33.
36. Zhenakova A, van Diemen CC, Wijmenga C. Detecting shared pathogenesis from the shared genetics of immune-related diseases. *Nat Rev Genet*. 2009;10(1):43–55.
37. Robinson WH, Fontoura P, Lee BJ, de Vegvar HEN, Tom J, Pedotti R, et al. Protein microarrays guide tolerizing DNA vaccine treatment of autoimmune encephalomyelitis. *Nat Biotechnol*. 2003;21(9):1033–9.
38. Lutterotti A, Yousef S, Spettek A, Stürmer KH, Stellmann JP, Breiden P, et al. Antigen-specific tolerance by autologous myelin peptide-coupled cells: a phase 1 trial in multiple sclerosis. *Sci Transl Med*. 2013;5(188):75.
39. Fisher RA. Statistical methods for research workers, 5th edn. Biological monographs and manuals. Edinburgh: Oliver and Boyd Ltd; 1934.
40. Mogensen TH. Pathogen recognition and inflammatory signaling in innate immune defenses. *Clin Microbiol Rev*. 2009;22(2):240–73.
41. Ashwood P, Krakowiak P, Hertz-Picciotto I, Hansen R, Pessah I, Van de Water J. Elevated plasma cytokines in autism spectrum disorders provide evidence of immune dysfunction and are associated with impaired behavioral outcome. *Brain Behav Immun*. 2011;25(1):40–5.
42. Enstrom AM, Onore CE, Van de Water JA, Ashwood P. Differential monocyte responses to TLR ligands in children with autism spectrum disorders. *Brain Behav Immun*. 2010;24(1):64–71.
43. Verkhatsky A, Rodríguez JJ, Parpura V. Neuroglia in ageing and disease. *Cell Tissue Res*. 2014;357(2):493–503.
44. Malkova NV, Collin ZY, Hsiao EY, Moore MJ, Patterson PH. Maternal immune activation yields offspring displaying mouse versions of the three core symptoms of autism. *Brain Behav Immun*. 2012;26(4):607–16.
45. Enstrom AM, Lit L, Onore CE, Gregg JP, Hansen RL, Pessah IN, et al. Altered gene expression and function of peripheral blood natural killer cells in children with autism. *Brain Behav Immun*. 2009;23(1):124–33.
46. Filiano AJ, Xu Y, Tustison NJ, Marsh RL, Baker W, Smirnov I, et al. Unexpected role of interferon- $\gamma$  in regulating neuronal connectivity and social behaviour. *Nature*. 2016. doi:10.1038/nature18626.
47. Suh HS, Kim MO, Lee SC. Inhibition of granulocyte-macrophage colony-stimulating factor signaling and microglial proliferation by anti-CD45RO: role of Hck tyrosine kinase and phosphatidylinositol 3-kinase/Akt. *J Immunol*. 2005;174(5):2712–19.
48. Fatemi SH. Multiple pathways in prevention of immune-mediated brain disorders: implications for the prevention of autism. *J Neuroimmunol*. 2009;217(1–2):8.
49. Parker-Athill E, Luo D, Bailey A, Giunta B, Tian J, Shytle RD, et al. Flavonoids, a prenatal prophylaxis via targeting JAK2/STAT3 signaling to oppose IL-6/MIA associated autism. *J Neuroimmunol*. 2009;217(1):20–7.
50. Polan MB, Pastore MT, Steingass K, Hashimoto S, Thrush DL, Pyatt R, et al. Neurodevelopmental disorders among individuals with duplication of 4p13 to 4p12 containing a GABAA receptor subunit gene cluster. *Eur J Hum Genet*. 2014;22(1):105–9.
51. Pramparo T, Pierce K, Lombardo MV, Barnes CC, Marinero S, Ahrens-Barbeau C, et al. Prediction of autism by translation and immune/inflammation coexpressed genes in toddlers from pediatric community practices. *JAMA Psychiatr*. 2015;72(4):386–94.

52. Smyth GK. Linear models and empirical Bayes methods for assessing differential expression in microarray experiments. *Stat Appl Genet Mol Biol*. 2004;3(1):1–25.
53. Murie C, Woody O, Lee AY, Nadon R. Comparison of small  $n$  statistical tests of differential expression applied to microarrays. *BMC Bioinform*. 2009;10(1):1.
54. Tesse R, Pandey R, Kabesch M. Genetic variations in Toll-like receptor pathway genes influence asthma and atopy. *Allergy*. 2011;66(3):307–16.
55. Zuany-Amorim C, Hastewell J, Walker C. Toll-like receptors as potential therapeutic targets for multiple diseases. *Nat Rev Drug Discov*. 2002;1(10):797–807.
56. Lin J, Caye-Thomasen P, Tono T, Zhang QA, Nakamura Y, Feng L, et al. Mucin production and mucous cell metaplasia in otitis media. *Int J Otolaryngol*. 2012;2012:745325. doi:10.1155/2012/745325.
57. Kimura H, Yoshizumi M, Ishii H, Oishi K, Ryo A. Cytokine production and signaling pathways in respiratory virus infection. *Front Microbiol*. 2013;4(276):2.
58. Hennessy EJ, Parker AE, O'Neill LA. Targeting Toll-like receptors: emerging therapeutics?. *Nat Rev Drug Discov*. 2010;9(4):293–307.
59. Neurath MF. Cytokines in inflammatory bowel disease. *Nat Rev Immunol*. 2014;14(5):329–42.
60. Gijbsbers K, Van Assche G, Joossens S, Struyf S, Proost P, Rutgeerts P, et al. CXCR1-binding chemokines in inflammatory bowel diseases: down-regulated IL-8/CXCL8 production by leukocytes in Crohn's disease and selective GCP-2/CXCL6 expression in inflamed intestinal tissue. *Eur J Immunol*. 2004;34(7):1992–2000.
61. Ramos PS, Sajuthi S, Langefeld CD, Walker SJ. Immune function genes CD99L2, JARID2 and TPO show association with autism spectrum disorder. *Mol Autism*. 2012;3(1):1–5.
62. Saxena V, Ramdas S, Ochoa CR, Wallace D, Bhide P, Kohane I. Structural, genetic, and functional signatures of disordered neuro-immunological development in autism spectrum disorder. *PLoS ONE*. 2012;7(12):48835.
63. Garbett KA, Hsiao EY, Kálmán S, Patterson PH, Mirnics K. Effects of maternal immune activation on gene expression patterns in the fetal brain. *Transl Psychiatr*. 2012;2(4):98.
64. Moscovitch SD, Szyper-Kravitz Z, Shoenfeld Y. Autoimmune pathology accounts for common manifestations in a wide range of neuro-psychiatric disorders: the olfactory and immune system interrelationship. *Clin Immunol*. 2009;130(3):235–43.
65. Li X, Chauhan A, Sheikh AM, Patil S, Chauhan V, Li XM, et al. Elevated immune response in the brain of autistic patients. *J Neuroimmunol*. 2009;207(1):111–16.
66. Smith SE, Li J, Garbett K, Mirnics K, Patterson PH. Maternal immune activation alters fetal brain development through interleukin-6. *J Neurosci*. 2007;27(40):10695–702.
67. Kong S, Shimizu-Motohashi Y, Campbell M, Lee I, Collins C, Brewster S, et al. Peripheral blood gene expression signature differentiates children with autism from unaffected siblings. *Neurogenetics*. 2013;14(2):143–52.
68. Voineagu I, Eapen V. Converging pathways in autism spectrum disorders: interplay between synaptic dysfunction and immune responses. *Front Hum Neurosci*. 2013;7:738.
69. Estes ML, McAllister AK. Immune mediators in the brain and peripheral tissues in autism spectrum disorder. *Nat Rev Neurosci*. 2015;16(8):469–86.
70. Suzuki K, Sugihara G, Ouchi Y, Nakamura K, Futatsubashi M, Takebayashi K, et al. Microglial activation in young adults with autism spectrum disorder. *JAMA Psychiatr*. 2013;70(1):49–58.
71. Gupta S, Ellis SE, Ashar FN, Moes A, Bader JS, Zhan J, et al. Transcriptome analysis reveals dysregulation of innate immune response genes and neuronal activity-dependent genes in autism. *Nat Commun*. 2014;5:5748.
72. Kim H, Cho M, Shim W, Kim J, Jeon E, Kim D, et al. Deficient autophagy in microglia impairs synaptic pruning and causes social behavioral defects. *Mol Psychiatr*. 2016. doi:10.1038/mp.2016.103.
73. Campbell MG, Kohane IS, Kong SW. Pathway-based outlier method reveals heterogeneous genomic structure of autism in blood transcriptome. *BMC Med Genet*. 2013;6(1):34.
74. Jyonouchi H, Geng L, Davidow AL. Cytokine profiles by peripheral blood monocytes are associated with changes in behavioral symptoms following immune insults in a subset of ASD subjects: an inflammatory subtype?. *J Neuroinflamm*. 2014;11(1):187.
75. West PR, Amaral DG, Bais P, Smith AM, Egnash LA, Ross ME, et al. Metabolomics as a tool for discovery of biomarkers of autism spectrum disorder in the blood plasma of children. *PLoS ONE*. 2014;9(11):112445.
76. Atladóttir HÓ, Pedersen MG, Thorsen P, Mortensen PB, Deleuran B, Eaton WW, et al. Association of family history of autoimmune diseases and autism spectrum disorders. *Pediatrics*. 2009;124(2):687–94.
77. Brown AS, Sourander A, Hinkka-Yli-Salomäki S, McKeague I, Sundvall J, Surcel H. Elevated maternal C-reactive protein and autism in a national birth cohort. *Mol Psychiatr*. 2014;19(2):259–64.
78. Atladóttir HO, Thorsen P, Østergaard L, Schendel DE, Lemcke S, Abdallah M, et al. Maternal infection requiring hospitalization during pregnancy and autism spectrum disorders. *J Autism Dev Disord*. 2010;40(12):1423–30.
79. Atladóttir HÓ, Henriksen TB, Schendel DE, Parner ET. Autism after infection, febrile episodes, and antibiotic use during pregnancy: an exploratory study. *Pediatrics*. 2012;130(6):1447–54.
80. Yap IK, Angley M, Veselkov KA, Holmes E, Lindon JC, Nicholson JK. Urinary metabolic phenotyping differentiates children with autism from their unaffected siblings and age-matched controls. *J Proteome Res*. 2010;9(6):2996–3004.
81. Kang DW, Park JG, Ilhan ZE, Wallstrom G, LaBaer J, Adams JB, et al. Reduced incidence of *Prevotella* and other fermenters in intestinal microflora of autistic children. *PLoS ONE*. 2013;8(7):68322.
82. Wang L, Christophersen CT, Sorich MJ, Gerber JP, Angley MT, Conlon MA, et al. Increased abundance of *Sutterella spp.* and *Ruminococcus torques* in feces of children with autism spectrum disorder. *Mol Autism*. 2013;4(1):42.
83. De Angelis M, Piccolo M, Vannini L, Siragusa S, De Giacomo A, Serrazanetti DI, et al. Fecal microbiota and metabolome of children with autism and pervasive developmental disorder not otherwise specified. *PLoS ONE*. 2013;8(10):e76993.
84. Mayer EA, Padua D, Tillisch K. Altered brain-gut axis in autism: comorbidity or causative mechanisms?. *Bioessays*. 2014;36(10):933–9.
85. Davis S, Meltzer PS. GEOquery: a bridge between the Gene Expression Omnibus (GEO) and BioConductor. *Bioinformatics*. 2007;23(14):1846–7.
86. Sellke T, Bayarri M, Berger JO. Calibration of  $p$  values for testing precise null hypotheses. *Am Stat*. 2001;55(1):62–71.
87. Johnson VE. Bayes factors based on test statistics. *J R Stat Soc Ser B Stat Methodol*. 2005;67(5):689–701.
88. Goodman SN. Toward evidence-based medical statistics. 2: The Bayes factor. *Ann Intern Med*. 1999;130(12):1005–13.
89. Edgar R, Domrachev M, Lash AE. Gene Expression Omnibus: NCBI gene expression and hybridization array data repository. *Nucleic Acids Res*. 2002;30(1):207–10.
90. Barrett T, Wilhite SE, Ledoux P, Evangelista C, Kim IF, Tomashevsky M, et al. NCBI GEO: archive for functional genomics data sets—update. *Nucleic Acids Res*. 2013;41(D1):991–5.
91. Kanehisa M, Goto S. KEGG: Kyoto Encyclopedia of Genes and Genomes. *Nucleic Acids Res*. 2000;28(1):27–30.
92. Kanehisa M, Goto S, Sato Y, Furumichi M, Tanabe M. KEGG for integration and interpretation of large-scale molecular data sets. *Nucleic Acids Res*. 2012;40(D1):D109–D114.
93. Croft D, Mundo AF, Haw R, Milacic M, Weiser J, Wu G, et al. The Reactome pathway knowledgebase. *Nucleic Acids Res*. 2014;42(D1):472–9.
94. Monaco MK, Stein J, Naithani S, Wei S, Dharmawardhana P, Kumari S, et al. Gramene 2013: comparative plant genomics resources. *Nucleic Acids Res*. 2014;42(D1):1193–9.
95. Nishimura D. BioCarta. *Biotech Softw Internet Rep Comput Softw J Scient*. 2001;2(3):117–20.
96. Schaefer CF, Anthony K, Krupa S, Buchoff J, Day M, Hannay T, et al. PID: the pathway interaction database. *Nucleic Acids Res*. 2009;37(suppl 1):674–9.
97. Subramanian A, Tamayo P, Mootha VK, Mukherjee S, Ebert BL, Gillette MA, et al. Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles. *Proc Natl Acad Sci USA*. 2005;102(43):15545–50.
98. Gene Expression Omnibus. <https://www.ncbi.nlm.nih.gov/geo/>, Accessed 29 Sept 2016.
99. Molecular Signatures Database. <http://www.broadinstitute.org/gsea/msigdb/collections.jsp>, Accessed 29 Sep 2016.
100. Becker KG. Autism, asthma, inflammation, and the hygiene hypothesis. *Med Hypotheses*. 2007;69(4):731–40.

101. Hagberg H, Gressens P, Mallard C. Inflammation during fetal and neonatal life: implications for neurologic and neuropsychiatric disease in children and adults. *Ann Neurol*. 2012;71(4):444–57.
102. Curatolo P, Porfiro MC, Manzi B, Seri S. Autism in tuberous sclerosis. *Eur J Paediatr Neurol*. 2004;8(6):327–32.
103. Loirat C, Bellanné-Chantelot C, Husson I, Deschênes G, Guignon V, Chabane N. Autism in three patients with cystic or hyperechogenic kidneys and chromosome 17q12 deletion. *Nephrol Dial Transpl*. 2010;25(10):3430–3.
104. Surén P, Bakken IJ, Aase H, Chin R, Gunnes N, Lie KK, et al. Autism spectrum disorder, ADHD, epilepsy, and cerebral palsy in Norwegian children. *Pediatrics*. 2012;130(1):152–8.
105. Witchel HJ, Hancox JC, Nutt DJ. Psychotropic drugs, cardiac arrhythmia, and sudden death. *J Clin Psychopharmacol*. 2003;23(1):58–77.
106. Bilder D, Botts EL, Smith KR, Pimentel R, Farley M, Viskochil J, et al. Excess mortality and causes of death in autism spectrum disorders: a follow up of the 1980s Utah/UCLA autism epidemiologic study. *J Autism Dev Disord*. 2013;43(5):1196–204.
107. Konstantareas MM, Homatidis S. Brief report: ear infections in autistic and normal children. *J Autism Dev Disord*. 1987;17(4):585–94.
108. Rosenhall U, Nordin V, Sandström M, Ahlsen G, Gillberg C. Autism and hearing loss. *J Autism Dev Disord*. 1999;29(5):349–57.
109. Porges SW, Macellario M, Stanfill SD, McCue K, Lewis GF, Harden ER, et al. Respiratory sinus arrhythmia and auditory processing in autism: modifiable deficits of an integrated social engagement system?. *Int J Psychophysiol*. 2013;88(3):261–70.
110. Walker SJ, Fortunato J, Gonzalez LG, Krigsman A. Identification of unique gene expression profile in children with regressive autism spectrum disorder (ASD) and ileocolitis. *PLoS ONE*. 2013;8(3):58058.
111. Shavelle RM, Strauss DJ, Pickett J. Causes of death in autism. *J Autism Dev Disord*. 2001;31(6):569–76.
112. Tabares-Seisdedos R, Rubenstein J. Chromosome 8p as a potential hub for developmental neuropsychiatric disorders: implications for schizophrenia, autism and cancer. *Mol Psychiatr*. 2009;14(6):563–89.
113. Ingason A, Rujescu D, Cichon S, Sigurdsson E, Sigmundsson T, Pietiläinen O, et al. Copy number variations of chromosome 16p13.1 region associated with schizophrenia. *Mol Psychiatr*. 2011;16(1):17–25.
114. Murdoch JD, State MW. Recent developments in the genetics of autism spectrum disorders. *Curr Opin Genet Dev*. 2013;23(3):310–5.

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