

MIT Open Access Articles

*Directed Palladium-Catalyzed Acetoxylation of Indolines.
Total Synthesis of *N*-Benzoylcylindrocarine*

The MIT Faculty has made this article openly available. **Please share** how this access benefits you. Your story matters.

Citation: Flynn, Kristen M, White, Kolby L and Movassaghi, Mohammad. 2022. "Directed Palladium-Catalyzed Acetoxylation of Indolines. Total Synthesis of *N*-Benzoylcylindrocarine." *The Journal of Organic Chemistry*, 87 (5).

As Published: 10.1021/acs.joc.1c02811

Publisher: American Chemical Society (ACS)

Persistent URL: <https://hdl.handle.net/1721.1/141146>

Version: Original manuscript: author's manuscript prior to formal peer review

Terms of use: Creative Commons Attribution-Noncommercial-Share Alike



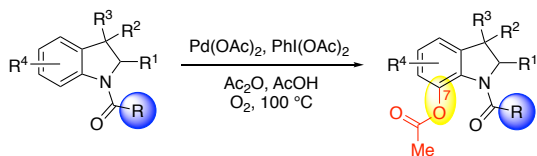
Directed Palladium Catalyzed Acetoxylation of Indolines. Total Synthesis of *N*-Benzoylcylindrocarine

Kristen M. Flynn, Kolby L. White, and Mohammad Movassaghi*

Department of Chemistry, Massachusetts Institute of Technology, Cambridge, Massachusetts 02139, United States

E-mail: movassag@mit.edu

Abstract Graphic

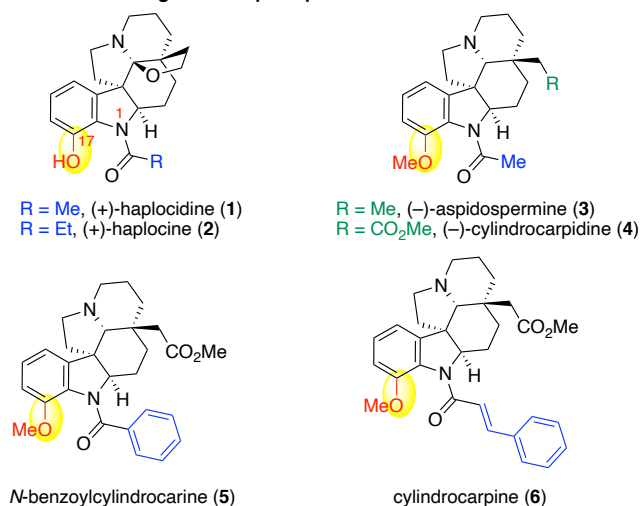


ABSTRACT: We describe a palladium catalyzed C7-acetoxylation of indolines with a range of amide directing groups. While a variety of substituents are tolerated on the indoline-core and the N1-acyl group, the acetoxylation is most sensitive to the C2- and C6-indoline substituents. The practicality of this indoline C7-acetoxylation is demonstrated using a cinnamamide substrate on mmol-scale. Several N1-acyl groups, including those present in natural alkaloids, guide C7-acetoxylation of indoline substrates over a competitive C5-oxidation. The application of this chemistry allowed for the first synthesis of *N*-benzoylcylindrocarine by late-stage C17-acetoxylation of *N*-benzoylfendleridine.

Introduction

The indoline substructure is ubiquitous in many bioactive indole alkaloids.¹ The aspidosperma family of indole alkaloids includes members of current interest for chemical synthesis given their structural complexity, boasting contiguous stereocenters and varying degrees of oxidation and substitution on a polycyclic core.^{1,2} Many members of this alkaloid family have a C17–O bond on the indoline substructure (Figure 1A).^{1b,3,4} Strategies for synthesis of C17-oxygenated aspidosperma alkaloids largely rely on the use of oxygenated starting materials that are converted into the indoline substructure.⁵ Notably, recent advances in transition metal catalyzed introduction of C–O bonds via arene functionalization⁶ have not been applied to the synthesis of C17-oxygenated aspidosperma alkaloids. Inspired by the biosynthesis of monoterpene indoline alkaloids wherein the polycyclic core undergoes enzymatic modifications including methylation, acylation, and C–H oxygenation,⁷ we sought a chemoselective C17-oxygenation of aspidosperma alkaloids, serving as a platform for late-stage diversification of complex synthetic intermediates.

A. Representative C17-oxygenated aspidosperma alkaloids:



B. This report:

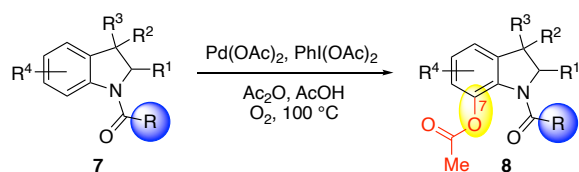


Figure 1. A. Representative aspidosperma alkaloids with C17-oxygenation of their N1-acylindoline substructures. B. Evaluation of C7-acetoxylation of indoline substrates with various N1-amides and substitution.

During our total synthesis efforts directed at (+)-haplocidine (**1**) and (+)-haplocine (**2**, Figure 1A), we found an opportunity to develop a method for selective metal-catalyzed late-stage C17–O bond formation.⁸ Our interest in palladium catalysis for this transformation stemmed from its versatility in regioselective C7–H indoline functionalization⁹ and precedent for palladium-catalyzed regioselective acetoxylation of aryl and bi-aryl systems.¹⁰ Alternative contemporary strategies for this transformation required the use of toxic lead or thallium reagents.¹¹ Subsequent to our report⁸ additional advances in C7-oxygenation of indolines have been reported involving the use of an *N*-pyrimidyl auxiliary with ruthenium, copper, and rhodium catalysts.¹² The prospects of using simple N1-acyl groups already present in targets to guide C7-oxygenation of indolines, without the need for removal of an auxiliary,⁸ prompted further investigation of the generality of this approach to facilitate its application in complex settings. Herein we describe our findings concerning the palladium catalyzed C7-acetoxylation of indolines guided by a range of N1-acyl groups, including naturally occurring benzoyl, cinnamoyl, and acetyl groups (Figure 1A). We discuss the impact of C2- and C6-indoline substituents and the electron density of the arene, highlight this indoline oxidation on mmol-scale, and report its application in the first synthesis of *N*-benzoylcylindrocarine (**5**).

Results and Discussion

The impact of each reaction component on the overall efficiency of the directed acetoxylation was examined using acetamide **7a** (Table 1). Starting with our optimized conditions (Pd(OAc)₂, PhI(OAc)₂, Ac₂O–AcOH, 100 °C, O₂, 7 h, 66% for **8a**),^{8,13} we note that exclusion of palladium (II) acetate or stoichiometric oxidant led to no desired product and resulted in recovery of starting material. The stoichiometric oxidant alone cannot oxidize the substrate to give the desired C7-acetoxyacetamide **8a** but is necessary for the oxidation of palladium during the catalytic cycle.¹⁴ We observed no advantage and more decomposition when conducting the reaction under an argon atmosphere rather than dioxygen.¹⁵ When acetic anhydride was excluded from the reaction mixture, a lower yield of the C7-acetoxyacetamide **8a** (56%) was obtained, consistent with related studies that support its role in facilitating catalytic turnover.^{10b}

We next evaluated various *N*-substituted indolines under standard conditions for directed acetoxylation. A variety of acyl directing groups are competent such as those with an aliphatic substituent providing the desired C7-acetylated product (**8a-8f**, Table 1). Conjugated amides are also effective directing groups (**8g-8h**, Table 1) and the C7-acetylated cinnamamide product **8g** was obtained from the corresponding C7-H cinnamamide substrate on 1 mmol-scale (64%). We then investigated indoline substrates with varying substitution (**8i-8m**, Table 1). While oxidation of acetamide **7m** afforded the desired C7-acetylated amide **8m** (67%), the oxidation of the corresponding C2 desmethyl substrate was significantly less effective (ca. 40%) and returned starting material. The observed negative impact on the efficiency of C7-H oxidation in the absence of a C2-substituent is consistent with prior report concerning palladium-catalyzed C7-arylation of indolines.¹⁶

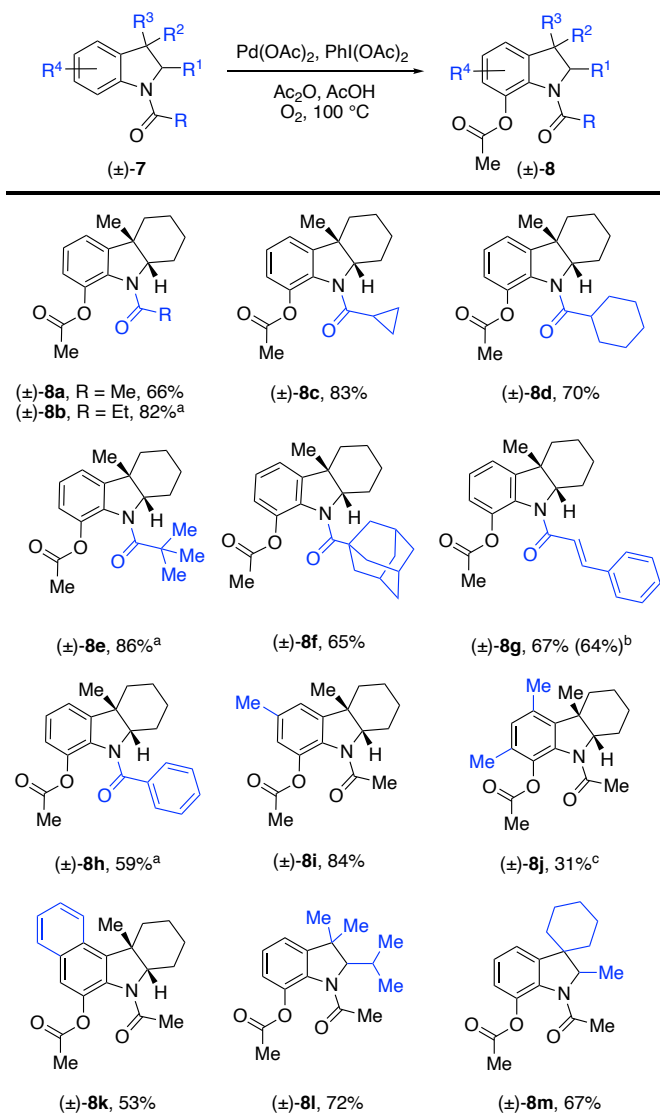
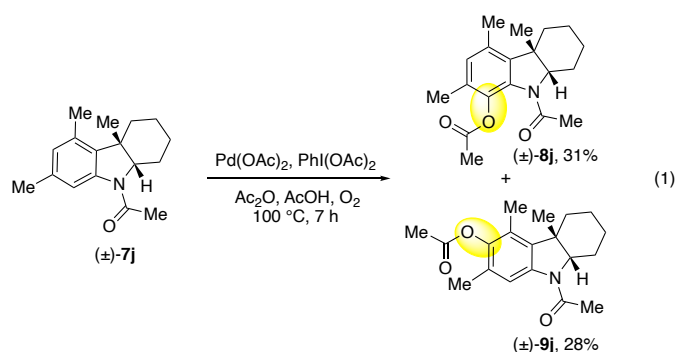


Table 1. Substrate scope for palladium catalyzed acetoxylation of indolines. Conditions: $\text{Pd}(\text{OAc})_2$ (15 mol %), $\text{PhI}(\text{OAc})_2$ (2.5 equiv), Ac_2O - AcOH (1:10, v/v), $100\text{ }^\circ\text{C}$, O_2 , 7–8 h. Yields are average of two independent runs. ^a See reference 8. ^b Isolated yield on 1 mmol scale. ^c C5-acetylated regioisomer isolated in 28% yield, see equation 1.

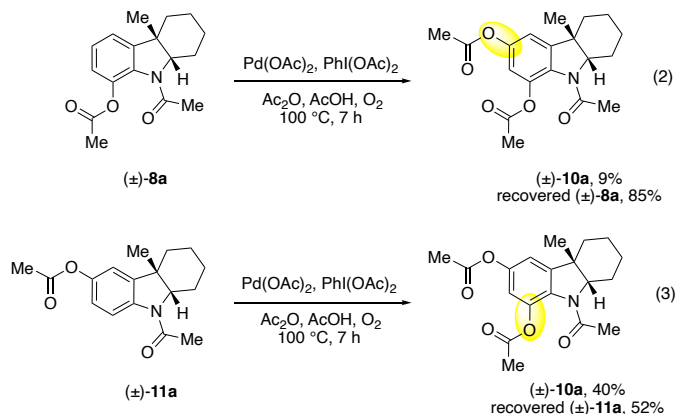
Indolines with stronger electron-withdrawing substituents at N1 (e.g., trifluoroacetyl, 2,2,2-trichloroethoxycarbonyl, methanesulfonyl, and methyl 2-oxoacetyl) led to minimal acetylated product (ca. 5–10%) with diminished C7 vs. C5 regioselectivity. This observation may be attributed to decreased Lewis basicity of the directing group necessary for C7-H metalation.^{10,13}

While the C5-methyl substituted amide **7i** efficiently afforded the desired C7-acetylated product **8i** in 84% yield, the corresponding C5-chloro derivative mostly returned starting material (76%) and provided only 19% yield of the desired C7-acetylated product. This observation is consistent with the corresponding calculated highest occupied molecular orbital (HOMO) energy levels for these substrates where an increase in the HOMO energy level affords superior acetoxylation outcome.¹³ Similarly, the use of a C5-methoxylated variant of amide **7a** gave the corresponding C6- and C7-acetylated products in 18% and 29% yield, respectively, along with recovered starting material (21%). This is consistent with the greater calculated electron density at C6 compared to C7¹³ allowing competitive regioisomeric acetoxylation despite the directed C7-oxidation rate enhancement.^{10d}

A similar observation for a decrease in selectivity was seen with acetamide **7j**. When amide **7j** was exposed to standard reaction conditions the desired acetoxylation product **8j** was isolated in 31% yield along with the regioisomeric **9j** in comparable yield (28%, equation 1). The non-directed C5-acetoxylation leading to product **9j** is likely due to both an increased electron density, evidenced by comparable calculated atomic charge densities at C5 and C7,¹³ and the adverse steric impact of the C6-substituent on the directed formation of the palladacycle intermediate.



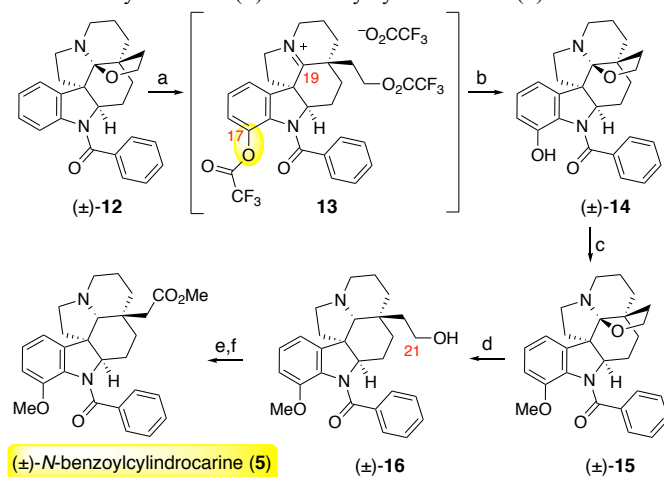
Given the presence of minor double oxidation side products in several cases (*ca.* 5–15%) we considered if an undesired C5-acetoxylation product could lead to these undesired over oxidation side products. The regioisomeric C5- and C7-acetylated acetamides **11a** and **8a**, respectively (equations 2 and 3), were used to evaluate their propensity for further oxidation. While exposure of amide **8a** to our standard acetoxylation conditions led to slow second C5-oxidation to give product **10a** in 9% yield along with significant recovery of the starting amide (85%, equation 2), amide **11a** progressed faster to the bisacetylated product **10a** (40%) under identical conditions (equation 3). These results highlight that while the directed C7-acetylated products may slowly oxidize further, any undirected C5-acetylated side-product can undergo a second oxidation rapidly leading to the minor double oxidation side-products observed in the acetoxylation of many substrates (Table 1).



Consistent with the observations described above, computational analysis of the C5- and C7-acetoxyacetamides **11a** and **8a**, respectively, revealed that they both have decreased atomic charge at the respective acetoxylation sites

compared to amide **7a**,¹³ leading to an overall attenuated rate of second oxidation (equations 2 and 3). Interestingly, analysis of the N1-amide bond conformers revealed that while acetamide **7a** does not have a preference for the *s*-cis conformation (49.4% *s*-cis vs. 50.6% *s*-trans), the optimal conformation for C–H functionalization,⁸ the C5-acetoxyacetamide **11a** preferred the *s*-cis conformation (74.5% *s*-cis vs. 25.5% *s*-trans), likely contributing to its susceptibility toward a second oxidation.^{8,13}

Scheme 1. Synthesis of (±)-*N*-benzoylcylindrocarine (**5**).^a



^a Reagents and conditions: (a) TFAA, TFA, 60 °C; Pd(OAc)₂ (20 mol%), PhI(OAc)₂ (4 equiv), 75 °C; (b) Et₃N, MeOH, 63% over 2 steps; (c) MeI, Cs₂CO₃, acetone, 80%; (d) TFAA, 1,2-DCE; NaBH(OMe)₃, 0 → 23 °C; Et₃N, MeOH, 75% over two steps; (e) TPAP, NMO, CH₂Cl₂, 50%; (f) NaClO₂, NaH₂PO₄·H₂O, resorcinol, H₂O, DMSO, 0 °C; TMS-diazomethane, MeOH, PhH, 44% over two steps .

To demonstrate the versatility of this chemistry, we utilized an N1-benzamide directed acetoxylation of a complex substrate for the first total synthesis of *N*-benzoylcylindrocarine (**5**, Scheme 1). The synthesis begins by benzoylation of synthetic fendleridine, prepared via our amide electrophilic activation chemistry,⁸ to give *N*-benzoylfendleridine (**12**) in 90% yield. Electrophilic opening of the C19-hemiaminal led to deactivation of the tertiary nitrogen⁸ and allowed benzamide directed C17-trifluoroacetoxylation which upon methanolysis provided the C17-hydroxy benzamide **14** in 63% yield over two steps. Synthesis of the methyl ether (80%) followed by C19-reduction gave the pentacyclic C21-alcohol **16** in 75% over two steps. A Ley Griffith oxidation¹⁷ of the C21-primary alcohol **16** afforded the corresponding aldehyde in 50% yield which upon further C21-oxidation under Pinnick conditions¹⁸ and methyl ester formation gave *N*-benzoylcylindrocarine (**5**) in 44% yield over two steps. All spectroscopic data for our synthetic (±)-*N*-benzoylcylindrocarine (**5**) was consistent with the previously reported values.^{4b}

Conclusions

In summary, we describe a regioselective palladium catalyzed C7-acetoxylation of indolines. A variety of N1-amides, including those found in natural alkaloids, serve as an effective directing group for this C–H oxidation reaction. The overall efficiency of this transformation was demonstrated by mmol-scale synthesis of cinnamamide **8g**. We also employed experimental and computational methods to correlate the observed oxidation outcome with various factors including substitution, energy and atomic charge calculations, and conformational preferences in the substrate to inform future applications. The versatility of this transformation was demonstrated through application of an N1-benzamide directed C–H oxidation en route to the first total synthesis of *N*-benzoylcylindrocarine (**5**).

EXPERIMENTAL SECTION

General Methods. All palladium catalyzed oxidation reactions were performed in oven-dried or flame-dried modified Schlenk (Kjeldahl shape) flasks unless noted otherwise. The flasks were fitted with Teflon-wrapped glass stoppers, conducted under oxygen atmosphere, and heated in an oil bath. All other reactions were performed in oven-dried or flame-dried round-bottom flasks fitted with rubber septa and were conducted under positive argon pressure using standard Schlenk techniques. Flash column chromatography was performed as described by Still et al.¹⁹ using granular silica gel (60 Å pore size, 40–63 μm, 4–6% H₂O content). Analytical thin-layer chromatography (TLC) was performed using glass plates precoated with 0.25 mm 230–400 mesh silica gel impregnated with a fluorescent indicator (254 nm). Thin-layer chromatography plates were visualized by exposure to short wave ultraviolet light (254 nm) and irreversibly stained by treatment with an aqueous solution of ceric ammonium molybdate (CAM) followed by heating (1 min) on a hot plate (250 °C). Organic solutions were concentrated at 29–30 °C on rotary evaporators capable of achieving a minimum pressure of 2 Torr and then at 0.5 Torr (vacuum pump) unless otherwise indicated. Proton nuclear magnetic resonance (¹H NMR) spectra were recorded with a 600, 500, and 400 MHz spectrometers. Chemical shifts are recorded in parts per million on the δ scale and are referenced from the residual protium in the NMR solvent (CHCl₃: δ 7.26, CD₂HClN: 1.94). Data are reported as follows: chemical shift [multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, br = broad), coupling constant(s) in Hertz, integration, assignment]. Carbon-13 nuclear magnetic resonance (¹³C) spectra were recorded with 150.9, 125.8, and 100.6 MHz spectrometers. Chemical shifts are recorded in parts per million on the δ scale and are referenced from the carbon resonances of the solvent (CDCl₃: δ 77.16, CD₃CN: 118.26). Data are reported as follows: chemical shift [assignment]. Infrared data was obtained with an FTIR and are reported as follows: [frequency of absorption (cm⁻¹), intensity of absorption (s = strong, m = medium, w = weak, br = broad), assignment]. High resolution mass spectra (HRMS) was recorded on a Fourier transform ion cyclotron resonance mass spectrometer (FTICR-MS) using electrospray ionization (ESI) or direct analysis in real time (DART) source.

Information for Compounds. For complete experimental procedures and full characterization data for C7-acetoxylated acetamide (±)-**8a**, C7-acetoxylated propionamide (±)-**8b**, C7-acetoxylated pivalamide (±)-**8c**, and C7-acetoxylated benzamide (±)-**8h**, please see the Supporting Information of our previous report.⁸ For complete experimental procedures and full characterization data for all other products reported in Table 1, please see below.

C7-Acetoxylated cyclopropylamide (±)-8c. Diacetoxyiodobenzene (62.1 mg, 195 μmol, 2.50 equiv), palladium acetate (2.6 mg, 12 μmol, 0.15 equiv), and cyclopropylamide (±)-**7c** (19.9 mg, 77.9 μmol, 1 equiv) were dissolved in acetic acid (1.70 mL) followed by addition of acetic anhydride (170 μL) at 23 °C. The reaction was sparged with dioxygen for 10 minutes, sealed with a Teflon wrapped glass stopper, and heated to 100 °C. After 7 h, the reaction mixture was cooled to 23 °C and diluted with ethyl acetate (5 mL). Saturated aqueous sodium carbonate solution (10 mL) was carefully added. The resulting mixture was extracted with ethyl acetate (3 × 10 mL). The combined organic extracts were further washed with saturated aqueous sodium chloride solution (10 mL), dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure. The resulting residue was purified by flash column chromatography on silica gel (eluent: 10 → 35% ethyl acetate in hexanes) to give C7-acetoxylated cyclopropylamide (±)-**8c** (20.5 mg, 84%) as a colorless film. Structural assignments were made using additional information from gCOSY, gHSQC, and gHMBC experiments. ¹H NMR (400 MHz, CDCl₃, 25 °C): δ 7.13 (app-t, *J* = 7.6 Hz, 1H, C5H), 6.99 (app-d, *J* = 7.4, 1.2 Hz, 1H, C4H), 6.94 (app-d, *J* = 8.1, 1.2 Hz, 1H, C6H), 4.24 (app-q, *J* = 10.6, 6.4 Hz, 1H, C2H), 2.33–2.26 (m, 1H, C3CH_a), 2.25 (s, 3H, C7OC(O)CH₃), 2.16–2.05 (m, 1H, C2CH_a), 1.76 (app-sept, *J* = 7.8 Hz, 4.6 Hz, 1H, N1C(O)CH), 1.66–1.52 (m, 3H, C3CH_b, C3CH₂CH_a, C2CH₂CH_a), 1.42–1.28 (m, 3H, C2CH_b, C2CH₂CH_b), 1.16 (s, 3H, C3CH₃), 1.13–1.07 (m, 1H, C3CH₂CH_b), 1.01–0.74 (m, 3H). ¹³C NMR (125.8 MHz, CDCl₃, 25 °C): δ 171.5 (N1C(O)), 168.2 (C7OC(O)), 143.8 (C4a), 140.9 (C7), 132.9 (C7a), 125.6 (C5), 122.3 (C6), 119.2 (C4), 69.9 (C2), 44.8 (C3), 33.0 (C3CH₂), 30.2 (C3CH₃), 28.9 (C2CH₂), 23.2 (C2CH₂CH₂), 22.1 (C3CH₂CH₂), 21.1

(C7OC(O)CH₃), 13.2 (N1C(O)CH), 8.8 (N1C(O)CHCH₂), 8.4 (N1C(O)CHCH₂). FTIR (thin film) cm⁻¹: 2926 (m), 2856 (w), 1764 (s), 1652 (s), 1608 (w), 1457 (s), 1409 (s), 1364 (m), 1206 (m), 1173 (s). HRMS (ESI) (*m/z*): calc'd for C₁₉H₂₄NO₃ [M+H]⁺: 314.1751, found: 314.1745. TLC (25% ethyl acetate in hexanes), R_f: 0.26 (UV, CAM).

C7-Acetoxyated cyclohexylamide (±)-8d. Diacetoxyiodobenzene (52.8 mg, 164 μmol, 2.50 equiv), palladium acetate (2.2 mg, 9.8 μmol, 0.15 equiv), and cyclohexylamide (±)-7d (19.5 mg, 65.6 μmol, 1 equiv) were dissolved in acetic acid (1.43 ml) followed by addition of acetic anhydride (143 μL) at 23 °C. The reaction was sparged with dioxygen for 10 minutes, sealed with a Teflon wrapped glass stopper, and heated to 100 °C. After 7 h, the reaction mixture was cooled to 23 °C and diluted with ethyl acetate (5 mL). Saturated aqueous sodium carbonate solution (10 mL) was carefully added. The resulting mixture was extracted with ethyl acetate (3 × 10 mL). The combined organic extracts were further washed with saturated aqueous sodium chloride solution (10 mL), dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure. The resulting residue was purified by flash column chromatography on silica gel (eluent: 10 → 35% ethyl acetate in hexanes) to give C7-acetoxyated cyclohexylamide (±)-8d (16.4 mg, 70%) as a colorless film. Structural assignments were made using additional information from gCOSY, gHSQC, and gHMBC experiments. ¹H NMR (400 MHz, CDCl₃, 25 °C): δ 7.11 (app-t, *J* = 7.7 Hz, 1H, C5H), 6.95 (m, 2H, C4H, C6H), 3.89 (br-s, 1H, C2H), 2.45 (s, 1H, N1C(O)CH), 2.34–2.26 (m, 1H, C3CH_a), 2.24 (s, 3H, C7OC(O)CH₃), 2.09–2.00 (m, 1H, C2CH_a), 1.88–1.75 (m, 4H, N1C(O)CHCH_a, N1C(O)CH-CH₂CH₂CH₂), 1.75–1.51 (m, 7H, C3CH_b, C3CH₂CH_a, C2CH₂CH_a, N1C(O)CHCH_b, N1C(O)CHCH₂CH_a), 1.42–1.17 (m, 4H, C2CH_b, C2CH₂CH_b, N1C(O)CHCH₂CH_b), 1.14 (s, 3H, C3CH₃), 1.12–1.05 (m, 1H, C3CH₂CH_b). ¹³C NMR (125.8 MHz, CDCl₃, 25 °C): δ 173.3 (N1C(O)), 167.6 (C7OC(O)CH₃), 143.5 (C4a), 141.1 (C7), 132.5 (C7a), 125.7 (C5), 122.2 (C6), 118.8 (C4), 69.3 (C2), 44.8 (C4), 43.4 (N1C(O)CH), 33.0 (C2CH₂), 30.3 (C3CH₃), 30.2 (N1C(O)CHCH₂), 29.5 (N1C(O)CHCH₂CH₂), 29.1 (C2CH₂), 25.8 (N1C(O)CHCH₂CH₂CH₂), 23.3 (C2CH₂CH₂), 22.0 (C3CH₂CH₂), 21.0 (C7OC(O)CH₃). FTIR (thin film) cm⁻¹: 2922 (s), 2853 (s), 1761 (s), 1659 (s), 1447 (s), 1404 (m), 1363 (m), 1261 (m), 1206 (s), 1192 (s), 1174 (s). HRMS (ESI) (*m/z*): calc'd for C₂₂H₃₀NO₃ [M+H]⁺: 356.2220, found: 356.2230. TLC (25% ethyl acetate in hexanes), R_f: 0.46 (UV, CAM).

C7-Acetoxyated adamantylamide (±)-8f. Diacetoxyiodobenzene (33.9 mg, 105 μmol, 2.50 equiv), palladium acetate (1.4 mg, 6.3 μmol, 0.15 equiv), and adamantylamide (±)-7f (14.7 mg, 42.1 μmol, 1 equiv) were dissolved in acetic acid (0.92 mL) followed by addition of acetic anhydride (92 μL) at 23 °C. The reaction was sparged with dioxygen for 10 minutes, sealed with a Teflon wrapped glass stopper, and heated to 100 °C. After 7 h, the reaction mixture was cooled to 23 °C and diluted with ethyl acetate (5 mL). Saturated aqueous sodium carbonate solution (10 mL) was carefully added. The resulting mixture was extracted with ethyl acetate (3 × 10 mL). The combined organic extracts were further washed with saturated aqueous sodium chloride solution (10 mL), dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure. The resulting residue was purified by flash column chromatography on silica gel (eluent: 10 → 50% ethyl acetate in hexanes) to give C7-acetoxyated adamantylamide (±)-8f (11.0 mg, 63%) as a white solid. Structural assignments were made using additional information from gCOSY, gHSQC, and gHMBC experiments. ¹H NMR (400 MHz, CDCl₃, 25 °C): δ 7.12 (app-t, *J* = 7.7 Hz, 1H, C5H), 6.95 (m, 2H, C4H, C6H), 4.20 (app-q, *J* = 10.3, 5.7 Hz, 1H, C2H), 2.33–2.25 (app-d, *J* = 13.5 Hz, 1H, C3CH_a), 2.22 (s, 3H, C7OC(O)CH₃), 2.21–2.11 (m, 3H, C2CH_a, N1C(O)CCH₂), 2.11–2.05 (m, 3H, N1C(O)CCH₂CH), 2.00–1.92 (m, 4H, N1C(O)CCH₂), 1.80–1.73 (m, 6H, N1C(O)CCH₂CHCH₂), 1.65–1.51 (m, 3H, C2CH₂CH_a, C3CH₂CH_a, C3CH_b), 1.31–1.12 (m, 3H, C2CH_b, C2CH₂CH_b, C3CH₂CH_b), 1.11 (s, 3H, C3CH₃). ¹³C NMR (125.8 MHz, CDCl₃, 25 °C): δ 176.8 (N1C(O)), 168.0 (C7OC(O)CH₃), 143.2 (C4a), 142.6 (C7), 135.6 (C7a), 125.7 (C5), 121.9 (C6), 118.6 (C4), 69.3 (C2), 46.9 (C3), 43.1 (N1C(O)C), 40.0 (N1C(O)CCH₂), 36.7 (N1C(O)CCHCH₂CH₂), 33.1 (C3CH₂), 29.5 (C3CH₃), 29.0 (C2CH₂), 28.6 (N1C(O)CCH₂CH), 23.8 (C2CH₂CH₂), 22.1 (C3CH₂CH₂), 21.2 (C7OC(O)CH₃). FTIR (thin film) cm⁻¹: 2899 (s), 2846 (m), 1766 (s), 1645 (s), 1614 (m), 1300 (m), 1446 (m), 1211 (s), 1175 (s). HRMS (ESI) (*m/z*): calc'd for C₂₆H₃₄NO₃ [M+H]⁺: 408.2533, found: 408.2522. TLC (25% ethyl acetate in hexanes), R_f: 0.52 (UV, CAM).

C7-Acetoxylation of cinnamamide (±)-7g to cinnamamide (±)-8g. Diacetoxyiodobenzene (805 mg, 2.50 mmol, 2.50 equiv), palladium acetate (34 mg, 150 μmol, 0.15 equiv), and cinnamamide (±)-7g (317 mg, 1.00 mmol, 1 equiv) were dissolved in acetic acid (21.9 mL) followed by addition of acetic anhydride (2.19 mL) at 23 °C. The reaction was sparged with dioxygen for 10 minutes, sealed with a Teflon wrapped glass stopper, and heated to 100 °C. After 7 h, the reaction mixture was cooled to 23 °C and diluted with ethyl acetate (15 mL). Saturated aqueous sodium carbonate solution (30 mL) was carefully added. The resulting mixture was extracted with ethyl acetate (3 × 30 mL). The combined organic extracts were further washed with saturated aqueous sodium chloride solution (30 mL), dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure. The resulting residue was purified by flash column chromatography on silica gel (eluent: 10 → 35% ethyl acetate in hexanes) to give C7-acetoxylation cinnamamide (±)-8g (241 mg, 64%) as a yellow solid. Cinnamamide (±)-7g (21.0 mg, 66.1 μmol, 1 equiv) provided C7-acetoxylation cinnamamide (±)-8g (16.7 mg, 67%) as a yellow solid. Structural assignments were made using additional information from gCOSY, gHSQC, and gHMBC experiments. ¹H NMR (400 MHz, CDCl₃, 25 °C): δ 7.78 (d, *J* = 15.5 Hz, 1H, N1C(O)CHCH), 7.56 (app-d, *J* = 7.5, 2.0 Hz, 2H, CH=CHCH_{ar}), 7.38 (m, 3H, CH=CHCH_{ar}, CH=CHCH_{ar}), 7.16 (app-t, *J* = 7.7 Hz, 1H, C5H), 7.03 (app-d, *J* = 7.4, 1.2 Hz, 1H, C4H), 7.00 (app-d, *J* = 8.1, 1.2 Hz, 1H, C6H), 6.76 (d, *J* = 15.5 Hz, 1H, N1C(O)CH), 4.29 (br-s, 1H, C2H), 2.33–2.24 (m, 1H, C3CH_a), 2.24–2.16 (m, 1H, C2CH_a), 2.10 (s, 3H, C7OC(O)CH₃), 1.68–1.51 (m, 3H, C3CH_b, C3CH₂CH_a, C2CH₂CH_a), 1.37–1.19 (m, 2H, C2CH_b, C2CH₂CH_b), 1.16 (s, 3H, C3CH₃), 1.14–1.06 (m, 1H, C3CH₂CH_b). ¹³C NMR (125.8 MHz, CDCl₃, 25 °C): δ 168.4 (C7OC(O)CH₃), 164.2 (N1C(O)), 144.4 (C4a), 142.7 (N1C(O)CHCH), 140.6 (C7), 135.1 (CH=CHC_{ar}), 132.5 (C7a), 130.0 (CH=CHC_{ar}), 129.0 (CH=CHC_{ar}), 129.0 (CH=CHC_{ar}), 125.9 (C5), 122.4 (C6), 119.6 (C4), 119.4 (N1C(O)CH), 70.4 (C2), 44.8 (C3), 33.0 (C3CH₂), 30.0 (C3CH₃), 28.7 (C2CH₂), 23.1 (C2CH₂CH₂), 22.1 (C3CH₂CH₂), 21.0 (C7OC(O)CH₃). FTIR (thin film) cm⁻¹: 2923 (m), 2854 (m), 1759 (m), 1652 (s), 1616 (m), 1448 (s), 1397 (s), 1253 (w), 1206 (s), 1177 (s). HRMS (ESI) (*m/z*): calc'd for C₂₄H₂₆NO₃ [M+H]⁺: 376.1907, found: 376.1905. TLC (25% ethyl acetate in hexanes), *R*_f: 0.33 (UV, CAM).

C7-Acetoxylation of acetamide (±)-7i to acetamide (±)-8i. Diacetoxyiodobenzene (67.5 mg, 0.210 mmol, 2.50 equiv), palladium acetate (2.8 mg, 13 μmol, 0.15 equiv), and acetamide (±)-7i (20.4 mg, 83.0 μmol, 1 equiv) were dissolved in acetic acid (1.83 mL) followed by addition of acetic anhydride (183 μL) at 23 °C. The reaction was sparged with dioxygen for 10 minutes, sealed with a Teflon wrapped glass stopper, and heated to 100 °C. After 8 h, the reaction mixture was cooled to 23 °C and diluted with ethyl acetate (5 mL). Saturated aqueous sodium carbonate solution (10 mL) was carefully added. The resulting mixture was extracted with ethyl acetate (3 × 10 mL). The combined organic extracts were further washed with saturated aqueous sodium chloride solution (10 mL), dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure. The resulting residue was purified by flash column chromatography on silica gel (eluent: 10 → 40% ethyl acetate in hexanes) to give C7-acetoxylation acetamide (±)-8i (20.6 mg, 81%) as an off-white solid. Structural assignments were made using additional information from gCOSY, gHSQC, and gHMBC experiments. ¹H NMR (400 MHz, CDCl₃, 45 °C): δ 6.78 (app-s, 1H, C4H), 6.75 (app-s, 1H, C6H), 3.96 (br-s, 1H, C2H), 2.33 (s, 3H, C5CH₃), 2.26 (s, 3H, C7OC(O)CH₃), 2.25–2.21 (m, 1H, C3CH_a), 2.20 (s, 3H, N1C(O)CH₃), 2.08–1.99 (m, 1H, C2CH_a), 1.65–1.49 (m, 3H, C3CH_b, C3CH₂CH_a, C2CH₂CH_a), 1.36–1.15 (m, 2H, C2CH_b, C2CH₂CH_b), 1.14 (s, 3H, C3CH₃), 1.13–1.04 (m, 1H, C3CH₂CH_b). ¹³C NMR (125.8 MHz, CDCl₃, 45 °C): δ 168.0 (2C, N1C(O), C7OC(O)CH₃), 143.6 (C4a), 140.6 (C7), 136.1 (C5), 130.3 (C7a), 122.7 (C6), 120.0 (C4), 70.6 (C2), 44.7 (C3), 33.0 (C3CH₂), 30.2 (C3CH₃), 28.5 (C2CH₂), 23.2 (C2CH₂CH₂), 22.7 (N1C(O)CH₃), 22.1 (C3CH₂CH₂), 21.2 (C5CH₃ or C7OC(O)CH₃), 21.1 (C5CH₃ or C7OC(O)CH₃). FTIR (thin film) cm⁻¹: 2927 (s), 2858 (m), 1770 (s), 1662 (s), 1476 (m), 1393 (m), 1364 (m), 1202 (s), 1185 (w), 1137 (m). HRMS (ESI) (*m/z*): calc'd for C₁₈H₂₄NO₃ [M+H]⁺: 302.1751, found: 302.1744. TLC (25% ethyl acetate in hexanes), *R*_f: 0.11 (UV, CAM).

C7-Acetoxylation of acetamide (±)-7j and C5-acetoxylation of acetamide (±)-9j. Diacetoxyiodobenzene (48.2 mg, 0.150 mmol, 2.50 equiv), palladium acetate (2.0 mg, 9.0 μmol, 0.15 equiv), and acetamide (±)-7j (15.4 mg, 59.8 μmol, 1 equiv) were dissolved in acetic acid (1.31 mL) followed by addition of acetic anhydride (131 μL) at 23 °C. The reaction was sparged with dioxygen for 10 minutes, sealed with a Teflon wrapped glass stopper, and heated to 100 °C.

After 7 h, the reaction mixture was cooled to 23 °C and diluted with ethyl acetate (5 mL). Saturated aqueous sodium carbonate solution (10 mL) was carefully added. The resulting mixture was extracted with ethyl acetate (3 × 10 mL). The combined organic extracts were further washed with saturated aqueous sodium chloride solution (10 mL), dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure. The resulting residue was purified by flash column chromatography on silica gel (eluent: 1 → 15% acetone in dichloromethane) to give C7-acetoxylated acetamide (±)-**8j** (6.0 mg, 31%) as a colorless film and C5-acetoxylated acetamide (±)-**9j** (5.3 mg, 28%) as a colorless film. Structural assignments were made using additional information from gCOSY, gHSQC, gHMBC and NOESY experiments. ¹H NMR (400 MHz, CDCl₃, 50 °C): δ 6.73 (s, 1H, C5H), 3.93 (m, 1H, C2H), 2.55 (app-d, *J* = 14.6, 4.3, 2.6 Hz, 1H, C3CH_a), 2.29 (s, 3H, C4CH₃), 2.26 (s, 3H, C7OC(O)CH₃), 2.19 (s, 3H, N1C(O)CH₃), 2.16 (s, 3H, C6CH₃), 2.02–1.92 (m, 1H, C2CH_a), 1.67–1.53 (m, 2H, C2CH₂CH_a, C3CH₂CH_a), 1.52–1.42 (m, 1H, C3CH_b), 1.42–1.20 (m, 2H, C2CH_b, C2CH₂CH_b), 1.18 (s, 3H, C3CH₃), 1.17–1.08 (m, 1H, C3CH₂CH_b). ¹³C NMR (100.6 MHz, CDCl₃, 50 °C): δ 168.2 (N1C(O)), 167.7 (C7OC(O)CH₃), 138.5 (C7), 137.5 (C4a), 134.1 (C4 or C6), 131.0 (C4 or C6), 130.4 (C2, C5, C7a), 71.0 (C2), 47.0 (C3), 33.5 (C3CH₂), 28.2 (C2CH₃), 27.3 (C3CH₃), 23.4 (C2CH₂CH₂), 22.8 (N1C(O)CH₃), 22.0 (C3CH₂CH₂), 20.8 (C7OC(O)CH₃), 18.7 (C4CH₃), 16.5 (C6CH₃). FTIR (thin film) cm⁻¹: 2957 (m), 2924 (s), 2856 (m), 1760 (s), 1669 (s), 1415 (s), 1369 (s), 1304 (m), 1266 (w), 1184 (s). HRMS (ESI) (*m/z*): calc'd for C₁₉H₂₆NO₃ [M+H]⁺: 316.1907, found: 316.1911. TLC (10% acetone in dichloromethane), *R*_f: 0.44 (UV, CAM).

C5-Acetoxylated acetamide (±)-9j. Structural assignments were made using additional information from gCOSY, gHSQC, gHMBC and NOESY experiments. ¹H NMR (400 MHz, CDCl₃, 25 °C): δ 7.94 (br-s, 1H, C7H), 3.68 (br-s, 1H, C2H), 2.53–2.42 (m, 1H, C2CH_a), 2.33 (s, 3H, C5OC(O)CH₃), 2.25 (br-s, 3H, N1C(O)CH₃), 2.12 (s, 3H, C6CH₃), 2.11 (s, 3H, C4CH₃), 1.95 (s, 1H, C2CH_a), 1.65–1.48 (m, 3H, C3CH_b, C2CH₂CH_a, C3CH₂CH_a), 1.38–1.21 (m, 2 H, C2CH₂CH_b, C3CH₂CH_b), 1.19 (s, 3H, C3CH₃). ¹³C NMR (125.8 MHz, CDCl₃, 25 °C): δ 169.2 (C5OC(O)CH₃), 168.1 (N1C(O)), 144.6 (C5), 139.0 (C7a), 134.5 (C4a), 129.0 (C6), 126.2 (C4), 118.3 (C7), 69.1 (C2), 46.2 (C3), 33.1 (C3CH₂), 28.5 (C2CH₂), 28.5 (C3CH₃), 23.5 (N1C(O)CH₃), 22.5 (C2CH₂CH₂ or C3CH₂CH₂), 21.4 (C2CH₂CH₂ or C3CH₂CH₂), 20.6 (C5OC(O)CH₃), 17.1 (C6CH₃), 11.9 (C4CH₃). FTIR (thin film) cm⁻¹: 2927 (m), 2857 (m), 1759 (s), 1657 (s), 1470 (m), 1448 (m), 1415 (m), 1371 (m), 1309 (m), 1215 (m). HRMS (ESI) (*m/z*): calc'd for C₁₉H₂₆NO₃ [M+H]⁺: 316.1907, found: 316.1893. TLC (10% acetone in dichloromethane), *R*_f: 0.50 (UV, CAM).

C7-Acetoxylated acetamide (±)-8k. Diacetoxiodobenzene (44.9 mg, 0.140 mmol, 2.50 equiv), palladium acetate (1.9 mg, 8.4 μmol, 0.15 equiv), and acetamide (±)-**7k** (15.6 mg, 55.8 μmol, 1 equiv) were dissolved in acetic acid (1.22 mL) followed by addition of acetic anhydride (122 μL) at 23 °C. The reaction was sparged with dioxygen for 10 minutes, sealed with a Teflon wrapped glass stopper, and heated to 100 °C. After 7 h, the reaction mixture was cooled to 23 °C and diluted with ethyl acetate (5 mL). Saturated aqueous sodium carbonate solution (10 mL) was carefully added. The resulting mixture was extracted with ethyl acetate (3 × 10 mL). The combined organic extracts were further washed with saturated aqueous sodium chloride solution (10 mL), dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure. The resulting residue was purified by flash column chromatography on silica gel (eluent: 1 → 12% acetone in dichloromethane) to give C7-acetoxylated acetamide (±)-**8k** (9.6 mg, 51%) as tan solid. Structural assignments were made using additional information from gCOSY, gHSQC, and gHMBC experiments. ¹H NMR (400 MHz, CDCl₃, 25 °C): δ 8.05 (app-d, *J* = 8.4, 1.3 Hz, 1H, C4CH), 7.81 (app-d, *J* = 7.8, 1.7 Hz, 1H, C5CH), 7.50 (s, 1H, C6H), 7.41 (m, 2H, C4CHCH, C5CHCH), 4.04 (br-s, 1H, C2H), 2.84 (d, *J* = 14.5 Hz, 1H, C3CH_a), 2.32 (s, 3H, C7OC(O)CH₃), 2.26 (s, 3H, N1C(O)CH₃), 2.07–1.99 (m, 1H, C2CH_a), 1.80 (app-t, *J* = 14.6, 11.9, 3.0 Hz, 1H, C3CH_b), 1.67–1.44 (m, 3H, C2CH_b, C2CH₂CH_a, C3CH₂CH_a), 1.40 (s, 3H, C3CH₃), 1.39–1.32 (m, 1H, C2CH₂CH_b), 1.23–1.09 (m, 1H, C3CH₂CH_b). ¹³C NMR (125.8 MHz, CDCl₃, 25 °C): δ 168.7 (C2, N1C(O), C7OC(O)CH₃), 140.2 (C7), 136.5 (C4a), 133.2 (C7a), 132.9 (C4), 129.1 (C5CH), 128.3 (C5), 126.2 (C5CHCH or C4CHCH), 125.1 (C5CHCH or C4CHCH), 122.4 (C4CH), 121.0 (C6), 71.1 (C2), 48.1 (C3), 34.2 (C3CH₂), 27.7 (C2CH₂), 27.6 (C3CH₃), 22.9 (N1C(O)CH₃), 22.7 (C2CH₂CH₂), 21.4 (C7OC(O)CH₃), 21.3 (C3CH₂CH₂). FTIR (thin

film) cm^{-1} : 2933 (m), 2863 (w), 1765 (s), 1664 (s), 1446 (m), 1395 (s), 1368 (m), 1338 (w), 1305 (m), 1194 (s). HRMS (ESI) (m/z): calc'd for $\text{C}_{21}\text{H}_{24}\text{NO}_3$ [$\text{M}+\text{H}$] $^+$: 338.1751, found: 338.1748. TLC (10% acetone in dichloromethane), R_f: 0.56 (UV, CAM).

C7-Acetoxyalted acetamide (±)-8l. Diacetoxyiodobenzene (59.5 mg, 185 μmol , 2.50 equiv), palladium acetate (2.5 mg, 11 μmol , 0.15 equiv), and acetamide (±)-**7l** (17.1 mg, 73.9 μmol , 1 equiv) were dissolved in acetic acid (1.61 mL) followed by addition of acetic anhydride (161 μL) at 23 °C. The reaction was sparged with dioxygen for 10 minutes, sealed with a Teflon wrapped glass stopper, and heated to 100 °C. After 7 h, the reaction mixture was cooled to 23 °C and diluted with ethyl acetate (5 mL). Saturated aqueous sodium carbonate solution (10 mL) was carefully added. The resulting mixture was extracted with ethyl acetate (3 × 10 mL). The combined organic extracts were further washed with saturated aqueous sodium chloride solution (10 mL), dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure. The resulting residue was purified by flash column chromatography on silica gel (eluent: 10 → 35% ethyl acetate in hexanes) to give C7-acetoxyalted acetamide (±)-**8l** (15.5 mg, 72%) as a yellow oil. Structural assignments were made using additional information from gCOSY, gHSQC, and gHMBC experiments. ^1H NMR (400 MHz, CDCl_3 , 50 °C): δ 7.11 (app-t, $J = 8.2, 7.3$ Hz, 1H, C5H), 6.96 (app-d, $J = 9.8, 7.8, 1.2$ Hz, 2H, C4H, C6H), 4.12 (br-s, 1H, C2H), 2.29 (s, 3H, C7OC(O)CH₃), 2.24 (s, 3H, N1C(O)CH₃), 1.97 (app-t, $J = 6.8, 3.8$ Hz, 1H, C2CH), 1.40 (s, 3H, C3CH₃), 1.19 (s, 3H, C3CH₃), 1.06 (d, $J = 6.8$ Hz, 3H, C2CHCH₃), 0.57 (d, $J = 6.8$ Hz, 3H, C2CHCH₃). ^{13}C NMR (100.6 MHz, CDCl_3 , 50 °C): δ 171.0 (N1C(O)), 168.2 (C7OC(O)CH₃), 146.8 (C4a), 140.4 (C7), 134.3 (C7a), 126.2 (C5), 122.6 (C6), 118.6 (C4), 79.2 (C2), 44.6 (C3), 31.1 (C3CH₃), 29.9 (C2CH), 22.7 (N1C(O)CH₃), 21.9 (C2CHCH₃), 21.2 (C7OC(O)CH₃), 20.3 (C3CH₃), 18.0 (C2CHCH₃). FTIR (thin film) cm^{-1} : 2962 (m), 1769 (s), 1662 (s), 1460 (m), 1369 (s), 1297 (m), 1247 (m), 1207 (m), 1179 (s), 1012 (w). HRMS (ESI) (m/z): calc'd for $\text{C}_{17}\text{H}_{24}\text{NO}_3$ [$\text{M}+\text{H}$] $^+$: 290.1751, found: 290.1749.

C7-Acetoxyalted acetamide (±)-8m. Diacetoxyiodobenzene (64.9 mg, 202 μmol , 2.50 equiv), palladium acetate (2.7 mg, 12 μmol , 0.15 equiv), and acetamide (±)-**7m** (19.7 mg, 80.6 μmol , 1 equiv) were dissolved in acetic acid (1.76 mL) followed by addition of acetic anhydride (176 μL) at 23 °C. The reaction was sparged with dioxygen for 10 minutes, sealed with a Teflon wrapped glass stopper, and heated to 100 °C. After 7 h, the reaction mixture was cooled to 23 °C and diluted with ethyl acetate (5 mL). Saturated aqueous sodium carbonate solution (10 mL) was carefully added. The resulting mixture was extracted with ethyl acetate (3 × 10 mL). The combined organic extracts were further washed with saturated aqueous sodium chloride solution (10 mL), dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure. The resulting residue was purified by flash column chromatography on silica gel (eluent: 10 → 35% ethyl acetate in hexanes) to give C7-acetoxyalted acetamide (±)-**8m** (15.5 mg, 64%) as a white solid. Structural assignments were made using additional information from gCOSY, gHSQC, and gHMBC experiments. ^1H NMR (400 MHz, CDCl_3 , 25 °C): δ 7.11 (app-t, $J = 7.8$ Hz, 1H, C5H), 7.01 (app-d, $J = 7.4$ Hz, 1H, C4H), 6.94 (app-d, $J = 8.1$ Hz, 1H, C6H), 4.39 (br-s, 1H, C2H), 2.27 (s, 3H, C7OC(O)CH₃), 2.23 (s, 3H, N1C(O)CH₃), 1.87–1.56 (m, 4H), 1.56–1.41 (m, 2H), 1.41–1.22 (m, 4H), 1.18 (d, $J = 6.5$ Hz, 3H, C2CH₃). ^{13}C NMR (125.8 MHz, CDCl_3 , 25 °C): δ 168.1 (2C, N1C(O), C7OC(O)CH₃), 144.8 (C4a), 140.3 (C7), 131.8 (C7a), 125.9 (C5), 122.5 (C6), 120.0 (C4), 64.9 (C2), 48.0 (C3), 36.9 (C3CH₂), 29.2 (C3CH₂), 25.7 (C3CH₂CH₂CH₂), 23.5 (C3CH₂CH₂), 23.4 (C3CH₂CH₂), 22.8 (N1C(O)CH₃), 21.1 (C7OC(O)CH₃), 14.9 (C2CH₃). FTIR (thin film) cm^{-1} : 2921 (m), 2842 (w), 1754 (s), 1663 (s), 1456 (m), 1400 (m), 1310 (m), 1265 (w), 1212 (s), 1192 (s). HRMS (ESI) (m/z): calc'd for $\text{C}_{18}\text{H}_{24}\text{NO}_3$ [$\text{M}+\text{H}$] $^+$: 302.1751, found: 302.1738. TLC (25% ethyl acetate in hexanes), R_f: 0.43 (UV, CAM).

C5,C7-Diacetoxyalted acetamide (±)-10a: Structural assignments were made using additional information from gCOSY, gHSQC, and gHMBC experiments. ^1H NMR (600 MHz, CDCl_3 , 25 °C): δ 6.77 (d, $J = 2.2$ Hz, 1H, C4H), 6.75 (d, $J = 2.2$ Hz, 1H, C6H), 3.95 (br-s, 1H, C2H), 2.27 (s, 3H, C5OC(O)CH₃ or C7OC(O)CH₃), 2.26 (s, 3H, C5OC(O)CH₃ or C7OC(O)CH₃), 2.21 (s, 3H, N1C(O)CH₃), 2.19–2.17 (m, 1H, C3CH_a), 2.09–2.02 (m, 1H, C2CH_a), 1.66–1.60 (app-d, 1H, C2CH₂CH_a), 1.59–1.51 (m, 2H, C3CH_b, C3CH₂CH_a), 1.36–1.17 (m, 2H, C2CH_b, C2CH₂CH_b),

1.16 (s, 3H, C3CH₃), 1.15–1.07 (m, 1H, C3CH₂CH_b). ¹³C NMR (150.9 MHz, CDCl₃, 25 °C): δ 169.2 (C5OC(O)CH₃ or C7OC(O)CH₃), 167.6 (2C, N1C(O), C5OC(O)CH₃ or C7OC(O)CH₃), 148.3 (C5 or C7), 144.4 (C4a), 140.8 (C5 or C7), 130.2 (C7a), 115.6 (C6), 112.9 (C4), 70.6 (C2), 44.9 (C3), 32.9 (C3CH₂), 30.0 (C3CH₃), 28.5 (C2CH₂), 23.1 (C2CH₂CH₂), 22.8 (N1C(O)CH₃), 22.0 (C3CH₂CH₂), 21.3 (C5OC(O)CH₃ or C7OC(O)CH₃), 21.0 (C5OC(O)CH₃ or C7OC(O)CH₃). FTIR (thin film) cm⁻¹: 2926 (m), 2857 (w), 1769 (m), 1666 (m), 1614 (w), 1468 (m), 1397 (m), 1367 (m), 1198 (s). HRMS (ESI) (*m/z*): calc'd for C₁₉H₂₄NO₅ [M+H]⁺: 346.1649, found: 346.1652. TLC (50% ethyl acetate in hexane), R_f: 0.30 (UV, CAM).

C17-Hydroxy benzamide (±)-14. (±)-*N*-Benzoylfendleridine⁸ (**12**, 4.7 mg, 12 μmol, 1 equiv) was dissolved in trifluoroacetic anhydride (33 μL) and trifluoroacetic acid (330 μL) in a pressure tube. The reaction was sealed and heated to 60 °C. After 4 h, the reaction mixture was cooled to 23 °C. Palladium acetate (0.5 mg, 2 μmol, 0.2 equiv) and diacetoxyiodobenzene (15 mg, 47 μmol, 4.0 equiv) were added. The reaction vessel was sealed and heated to 75 °C. After 2 h, the reaction was concentrated under reduced pressure and subsequently dissolved in methanol (0.65 mL) and triethylamine (130 μL). After 12 h, the reaction was concentrated under reduced pressure. The resulting residue was purified by flash column chromatography on silica gel (eluent: 3% triethylamine and 0 → 30% ethyl acetate in hexanes) to give C17-hydroxy benzamide (±)-**14** (3.1 mg, 63%) as a white solid. Structural assignments were made using additional information from gCOSY, gHSQC, and gHMBC experiments. ¹H NMR (500 MHz, CDCl₃, 25 °C): δ 10.18 (s, 1H, OH), 7.54–7.48 (m, 5H, N1C(O)C₆H₅), 7.20 (app-d, *J* = 7.6, 1.3 Hz, 1H, C14H), 7.15 (t, *J* = 7.8 Hz, 1H, C15H), 6.91 (app-d, *J* = 8.1, 1.2 Hz, 1H, C16H), 4.16 (t, *J* = 8.5 Hz, 1H, C21H_a), 4.06 (app-q, *J* = 10.9, 8.2, 6.2 Hz, 1H, C21H_b), 3.73 (app-d, *J* = 11.2, 5.2 Hz, 1H, C2H), 2.87 (t, *J* = 7.5 Hz, 2H, C10H₂), 2.71 (app-t, *J* = 11.6, 2.8 Hz, 1H, C8H_a), 2.52 (app-d, *J* = 11.3, 2.7, 2.1 Hz, 1H, C8H_b), 2.10–2.01 (m, 1H, C11H_a), 1.94–1.82 (m, 3H, C3H_a, C11H_b, C20H_a), 1.82–1.74 (m, *J* = 13.5, 4.1 Hz, 1H, C3H_b), 1.70–1.56 (m, 3H, C4H_a, C6H_a, C7H_a), 1.48–1.42 (m, 1H, C7H_b), 1.35–1.24 (m, 2H, C4H_b, C6H_b), 1.21 (app-q, *J* = 12.0, 6.1 Hz, 1H, C20H_b). ¹³C NMR (125.8 MHz, CDCl₃, 25 °C): δ 170.2 (N1C(O)), 147.3 (C17), 141.4 (C13), 136.3 (N1C(O)C₆H₅), 130.4 (N1C(O)C₆H₅), 129.1 (N1C(O)C₆H₅), 128.9 (C15), 127.9 (C18), 126.3 (N1C(O)C₆H₅), 118.2 (C16), 116.1 (C14), 101.9 (C19), 70.8 (C2), 65.3 (C21), 58.7 (C12), 48.8 (C10), 43.8 (C8), 39.8 (C5), 35.6 (C11), 34.6 (C20), 32.9 (C6), 26.4 (C4), 25.7 (C3), 20.9 (C7). FTIR (thin film) cm⁻¹: 2937 (br-s), 2861 (m), 2832 (m), 2760 (m), 1620 (s), 1598 (s), 1566 (s), 1495 (m), 1465 (s), 1445 (s), 1428 (s). HRMS (ESI) (*m/z*): calc'd for C₂₆H₂₉N₂O₃ [M+H]⁺: 417.2173, found: 417.2176. TLC (3% triethylamine and 30% ethyl acetate in hexanes), R_f: 0.38 (UV, CAM).

C17-Methyl ether (±)-15. Cesium carbonate (58 mg, 0.17 mmol, 10 equiv) and iodomethane (10 μL, 0.2 mmol, 10 equiv) were added to a solution of C17-hydroxy benzamide (±)-**14** (6.9 mg, 17 μmol, 1 equiv) in acetone (530 μL) at 23 °C. After 3 h, a solution of saturated aqueous ammonium chloride (5 mL) and dichloromethane (5 mL) were added and the layers were separated. The aqueous layer was extracted with dichloromethane (2 × 5 mL) and the combined organic layers were dried with anhydrous sodium sulfate, were filtered, and were concentrated under reduced pressure. The resulting residue was purified by flash column chromatography on silica gel (eluent: 3% triethylamine and 5 → 25% ethyl acetate in hexanes) to give C17-methyl ether (±)-**15** (5.7 mg, 80%) as a white solid. Structural assignments were made using additional information from gCOSY, gHSQC, and gHMBC experiments. ¹H NMR (500 MHz, CDCl₃, 25 °C): δ 7.52–7.42 (m, 2H, N1C(O)C₆H₅), 7.39 (t, *J* = 7.4 Hz, 1H, N1C(O)C₆H₅), 7.35–7.26 (m, 3H, C14H, N1C(O)C₆H₅), 7.05 (t, *J* = 7.9 Hz, 1H, C15H), 6.62 (d, *J* = 8.1 Hz, 1H, C16H), 4.22 (br-s, 1H, C2H), 4.17–4.03 (m, 2H, C21H₂), 3.25 (br-s, 3H, OCH₃), 3.04–2.88 (m, 1H, C10H₂), 2.77 (t, *J* = 11.1 Hz, 1H, C8H_a), 2.61 (d, *J* = 11.2 Hz, 1H, C8H_b), 2.23 (br-s, 1H, C11H_a), 2.16–2.07 (m, 1H, C3H_a), 1.99–1.83 (m, 3H, C4H_a, C11H_b, C20H_a), 1.79–1.66 (m, 3H, C3H_b, C6H_a, C7H_a), 1.49 (m, 1H, C7H_b), 1.44–1.29 (m, 2H, C4H_b, C6H_b), 1.21 (app-d, *J* = 12.3, 6.1 Hz, 1H, C20H_b). ¹³C NMR (125.8 MHz, CDCl₃, 25 °C): δ 169.7 (N1C(O)), 149.1 (C17), 141.8 (C13), 137.7 (N1C(O)C₆H₅), 130.5 (N1C(O)C₆H₅), 130.3 (C18), 128.1 (N1C(O)C₆H₅), 127.2 (N1C(O)C₆H₅), 126.4 (C15), 117.3 (C14), 111.0 (C16), 102.3 (C19), 70.9 (C2), 65.2 (C21), 59.8 (C12), 54.8 (OCH₃), 49.0 (C10), 43.9 (C8), 39.8 (C5), 35.1 (C11), 34.7 (C20), 33.1 (C6), 26.5 (C4), 24.5 (C3), 21.1 (C7). FTIR (thin film) cm⁻¹: 2936 (s), 2834 (m), 1634 (s), 1579 (m),

1487 (s), 1454 (s), 1384 (s), 1338 (m), 1295 (m). HRMS (DART) (m/z): calc'd for C₂₇H₃₁N₂O₃ [M+H]⁺: 431.2329, found: 431.2340. TLC (3% triethylamine and 40% ethyl acetate in hexanes), Rf: 0.47 (UV, CAM).

C21-Alcohol (±)-16. C17-methyl ether (±)-**15** (36.9 mg, 0.100 mmol, 1 equiv) was azeotropically dried by concentration from anhydrous benzene (3 × 200 µL) and the residue was dissolved in 1,2-dichloroethane (0.75 mL). Trifluoroacetic anhydride (0.7 M in 1,2-dichloroethane, 50 µL, 40 µmol, 5 equiv) was added dropwise at 23 °C. After 1 h, the reaction mixture was cooled to 0 °C and sodium trimethoxyborohydride (9.8 mg, 77 µmol, 10 equiv) was added as a solid. After 5 min, the reaction was warmed to 23 °C. After 30 min, a solution of saturated aqueous sodium bicarbonate (5 mL) and dichloromethane (5 mL) were added and the layers were separated. The aqueous layer was extracted with dichloromethane (3 × 5 mL) and the combined organic layers were dried over anhydrous sodium sulfate, were filtered, and were concentrated under reduced pressure. The combined organic layers were dried over anhydrous sodium sulfate, were filtered, and were concentrated under reduced pressure. The residue was used directly in the next step. Triethylamine (150 µL) was added to a solution of the residue in methanol (0.75 mL) at 23 °C. After 1 h, the reaction was concentrated under reduced pressure. The resulting residue was purified by flash column chromatography on silica gel (eluent: 3% triethylamine and 10 → 50% ethyl acetate in hexanes) to give C21-alcohol (±)-**16** (2.5 mg, 75%) as a white solid. Structural assignments were made using additional information from gCOSY, gHSQC, and gHMBC experiments. ¹H NMR (600 MHz, CD₃CN, 60 °C): δ 7.46 (d, *J* = 7.4 Hz, 3H, N1C(O)CCH, N1C(O)CCHCHCH), 7.39 (app-t, *J* = 8.3, 7.0 Hz, 2H, N1C(O)CCHCH), 7.09 (t, *J* = 7.9 Hz, 1H, C15H), 6.94 (app-d, *J* = 7.5, 1.0 Hz, 1H, C14H), 6.75 (d, *J* = 8.2 Hz, 1H, C16H), 4.13 (app-q, *J* = 11.4, 5.8 Hz, 1H, C2H), 3.51–3.44 (m, 1H, C21H_a), 3.44–3.37 (m, 1H, C21H_a), 3.35 (s, 3H, C17OCH₃), 3.02 (app-t, *J* = 9.0, 3.5 Hz, 1H, C10H_a), 2.96 (app-d, *J* = 10.9, 4.0, 1.8 Hz, 1H, C8H_a), 2.35 (s, 1H, C19H), 2.26 (app-q, *J* = 10.6, 8.9, 7.3 Hz, 1H, C10H_b), 2.15–2.06 (m, 2H, C3H_a, C11H_a), 2.03–1.95 (m, 1H, C4H_a, C8H_b), 1.75–1.54 (m, 4H, C3H_b, C6H_a, C7H_a, C11H_b), 1.53–1.42 (m, 2H, C7H_b, C20H_b), 1.24 (app-t, *J* = 13.6, 4.7 Hz, 1H, C6H_b), 1.14–1.05 (m, 2H, C4H_b, C20H_b). ¹³C NMR (150.9 MHz, CD₃CN, 60 °C): δ 171.1 (N1C(O)), 151.1 (C17), 143.7 (C13), 139.1 (N1C(O)C), 131.6 (1C or 2C, N1C(O)CCH or N1C(O)CCHCHCH), 131.4 (C18), 129.3 (N1C(O)CCHCH), 128.1 (1C or 2C, N1C(O)CCH or N1C(O)CCHCHCH), 127.4 (C15), 116.8 (C14), 112.7 (C16), 72.1 (C2), 71.7 (C19), 58.3 (C21), 55.9 (C17OCH₃), 55.6 (C12), 54.4 (C8), 53.1 (C10), 41.6 (C20), 38.7 (C11), 36.4 (C5), 36.2 (C6), 26.6 (C3), 25.3 (C4), 22.6 (C7). FTIR (thin film) cm⁻¹: 3402 (br-s), 2932 (s), 2868 (m), 2783 (m), 2725 (w), 1620 (s), 1578 (m), 1489 (s), 1458 (s), 1385 (s), 1323 (m), 1274 (m). HRMS (DART) (m/z): calc'd for C₂₇H₃₃N₂O₃ [M+H]⁺: 433.2486, found:433.2492. TLC (3% triethylamine and 60% ethyl acetate in hexanes), Rf: 0.34 (UV, CAM).

(±)-*N*-Benzoylcylindrocarine (5). C21-alcohol (±)-**16** (6.8 mg, 16 µmol, 1 equiv) was azeotropically dried by concentration from anhydrous benzene (3 × 200 µL) and the residue was dissolved in dichloromethane (1.70 mL). *N*-Methylmorpholine *N*-oxide (NMO, 5.4 mg, 47 µmol, 3.0 equiv) was added followed by tetrapropylammonium perruthenate (TPAP, 0.1 M in dichloromethane, 40 µL, 2 µmol, 0.1 equiv) at 23 °C. After 20 min, the reaction mixture was diluted with dichloromethane (10 mL) and filtered through a pad of celite. The filter cake was rinsed with dichloromethane (2 × 10 mL) and the combined organic liquid was concentrated under reduced pressure. The resulting residue was purified by flash column chromatography on silica gel (eluent: 3% triethylamine and 10 → 40% ethyl acetate in hexanes) to give C21- aldehyde **S1** (3.4 mg, 50%) as a thin film.

C21-aldehyde **S1** (3.4 mg, 7.9 µmol, 1 equiv) was dissolved in dimethylsulfoxide (DMSO) (1.30 mL) and water (0.10 mL) then cooled to 0 °C. Resorcinol (4.3 mg, 39 µmol, 5.0 equiv) was added followed by a solution of sodium phosphate monobasic (NaH₂PO₄·H₂O, 5.4 mg, 39 µmol, 5.0 equiv) in water (50 µL). Sodium chlorite (NaClO₂, 3.6 mg, 39 µmol, 5.0 equiv) in water (50 µL) was added dropwise at 0 °C. After 10 min, a solution of saturated aqueous ammonium chloride (5 mL) and diethyl ether (5 mL) were added and warmed to 23 °C. The layers were separated and the aqueous layer was extracted with diethyl ether (3 × 10 mL). The combined organic extracts were further washed with saturated aqueous sodium chloride solution (10 mL), dried with anhydrous sodium sulfate, were filtered, and were concentrated under reduced pressure. The yellow residue was used directly in the next step.

The yellow residue was dissolved in methanol (260 μ L) and benzene (1.0 mL). Trimethylsilyldiazomethane²⁰ (TMS-diazomethane, 2 M in hexanes, 40 μ L, 80 μ mol, 10 equiv) was added at 23 °C. After 30 min, the reaction mixture was concentrated under reduced pressure. The resulting residue was purified by flash column chromatography on silica gel (eluent: 3% triethylamine and 0 \rightarrow 30% ethyl acetate in hexanes) to give (\pm)-*N*-benzoylcylindrocaine (**5**) (1.6 mg, 44%) as a thin film. Structural assignments were made using additional information from gCOSY, gHSQC, and gHMBC experiments. ¹H NMR (600 MHz, CD₃CN, 60 °C): δ 7.50–7.44 (m, 3H, N1C(O)CCHCHCH, N1C(O)CCH), 7.39 (app-t, *J* = 8.3, 6.9 Hz, 2H, N1C(O)CCHCH), 7.09 (t, *J* = 7.9 Hz, 1H, C15H), 6.94 (d, *J* = 8.5 Hz, 1H, C14H), 6.75 (d, *J* = 8.2 Hz, 1H, C16H), 4.15 (app-q, *J* = 11.4, 6.0 Hz, 1H, C2H), 3.53 (s, 3H, C21OCH₃), 3.34 (s, 3H, C17OCH₃), 3.05 (app-t, *J* = 9.1, 3.6 Hz, 1H, C10H_a), 2.97 (app-d, *J* = 12.8, 4.0, 1.8 Hz, 1H, C8H_a), 2.52 (s, 1H, C19H), 2.30 (app-q, *J* = 10.8, 8.9, 7.0 Hz, 1H, C10H_b), 2.20 (d, *J* = 14.3 Hz, 1H, C20H_a), 2.18–2.08 (m, 3H, C3H_a, C4H_a, C11H_a), 2.02–1.98 (m, 1H, C8H_b), 1.96–1.92 (m, 1H, C20H_b),²¹ 1.77–1.55 (m, 4H, C3H_b, C6H_a, C7H_a, C11H_b), 1.49 (app-d, *J* = 13.2, 4.9, 2.4 Hz, 1H, C7H_b), 1.42–1.28 (m, 1H, C6H_b), 1.28–1.22 (m, 1H, C4H_b). ¹³C NMR (150.9 MHz, CD₃CN, 60 °C): δ 172.8 (C21), 171.0 (N1C(O)), 151.2 (C17), 142.9 (C13), 139.0 (N1C(O)C), 131.6 (1C or 2C, N1C(O)CCH or N1C(O)CCHCHCH), 131.5 (C18), 129.4 (2C, N1C(O)CCHCH), 128.1 (1C or 2C, N1C(O)CCH or N1C(O)CCHCHCH), 127.5 (C15), 116.6 (C14), 113.0 (C16), 71.6 (C2), 70.4 (C19), 56.0 (C17OCH₃), 55.5 (C12), 54.2 (C8), 53.0 (C10), 51.7 (C21OCH₃), 43.2 (C20), 38.3 (C11), 37.1 (C5), 35.9 (C6), 26.5 (C3), 25.4 (C4), 22.6 (C7). FTIR (thin film) cm⁻¹: 2943 (s), 2871 (w), 1733 (s), 1636 (s), 1579 (w), 1489 (s), 1459 (s), 1386 (s), 1334 (m), 1277 (m), 1204 (m), 1157 (m), 1042 (w), 922 (w), 889 (w), 793 (m), 782 (m), 720 (m), 703 (m).²² HRMS (DART) (*m/z*): calc'd for C₂₈H₃₃N₂O₄ [M+H]⁺: 461.2435, found: 461.2431. TLC (3% triethylamine and 40% ethyl acetate in hexanes), R_f: 0.37 (UV, CAM).

Energy and Atomic Charge Calculations. The geometries (gas phase) in the ground state of indoline amides (6-10 depending on substrate) were optimized with Merck Molecular Force Field (MMFF)²³ followed by density functional theory at B3LYP level with 6-311+g(d,p) as basis set (Gaussian09, by Gaussian, Inc.).²⁴ The electrostatic potential (ESP) derived charge were calculated using the lowest energy *s*-cis conformation. The total percent *s*-cis and *s*-trans were calculated using the Boltzmann distribution to determine the average contribution of each conformation (6-8 depending on substrate) at 23 °C and the percent *s*-cis was added up independently to *s*-trans.

ASSOCIATED CONTENT

Supporting Information

Copies of ¹H and ¹³C NMR spectra of all compounds described in the experimental section. Coordinates and computational data of compounds discussed are described in the supporting information.

ACKNOWLEDGMENTS

We are grateful for financial support by NIH-NIGMS (GM-074825 and GM-141963). We thank the NSF CCI Center for Selective C–H Functionalization (CHE-1700982). K.M.F. acknowledges an MIT Dean of Science graduate fellowship.

REFERENCES

(1) (a) Siegler, D. S. Indole Alkaloids. In *Plant Secondary Metabolism*; Springer Science+Business: New York, 1998; pp 628–654. (b) Saxton, J. E. Alkaloids of the Aspidospermine Group. In *the Alkaloids, Chemistry and Biology*; Cordell, G. A., Ed.; Academic Press: San Diego, 1998; Vol. 51, pp 1–197. (c) O'Connor, S. E. Alkaloids. In *Comprehensive Natural Products II*; Mander, L., Liu, H.-W., Eds.; Elsevier: Amsterdam, 2010; Vol. 1, pp 977–1007.

(2) (a) Lopchuk, J. M. Recent Advances in the Synthesis of *Aspidosperma*-Type Alkaloids. In *Progress in Heterocyclic Chemistry*; Gribbe, G. W., Joule, J. A., Eds.; Elsevier: Amsterdam, 2011; Vol. 23, pp 1–25. (b) Saya, J. M.; Ruijter, E.; Orru, R. V. A. Total Synthesis of *Aspidosperma* and *Strychnos* Alkaloids Through Indole Dearomatization. *Chem. Eur. J.* **2019**, *25*, 8916–8935.

(3) (a) Cava, M. P.; Talapatra, S. K.; Nomura, K.; Weisbach, J. A.; Douglas, B.; Shoop, E. C. Haplocine and Haplocidine: New *Aspidosperma*-Type Alkaloids from *Haplophyton cimidum*. *Chem. and Ind.* **1963**, 1242–1243. (b) Cava, M. P.; Nomura, K.; Talapatra, S. K. The Stereochemistry of Limaspermine, Haplocine and Haplocidine. *Tetrahedron* **1964**, *20*, 581–583. (c) Walser, A.; Djerassi, C. Alkaloid-Studien LII. Die Alkaloide aus *Vallesia dichotoma* RUIZet PAV. *Helv. Chim. Acta* **1965**, *48*, 391–404. (d) Dastoor, N. J.; Gorman, A. A.; Schmid, H. Über die Alkaloide von *Aspidosperma discolor* A. DC. *Helv. Chim. Acta* **1967**, *50*, 213–231.

(4) (a) Djerassi, C.; Archer, A. A. P. G.; George, T.; Gilbert, B.; Shoolery, J. N.; Johnson, L. F. Alkaloid Studies XXV. The Structures of the *Aspidosperma* Alkaloids Cyndrocarpine and Cyndrocarpidine. *Experientia* **1960**, *16*, 532–534. (b) Milborrow, B. V.; Djerassi, C. Alkaloid Studies. Part LXI. The Structure of Twelve New Alkaloids from *Aspidosperma cyndrocarpon*. *J. Chem. Soc. C* **1969**, 417–424.

(5) (a) Lajiness, J.P.; Jiang, W.; Boger, D. L. Divergent Total Synthesis of (–)-*Aspidospermine* and (+)-*Spegazzine*. *Org. Lett.* **2012**, *14*, 2078–2081. (b) Satoh, H.; Ueda, H.; Tokuyama, H. Divergent Total Synthesis of (–)-*Aspidophytine* and its Congeners via Fischer Indole Synthesis. *Tetrahedron* **2013**, *69*, 89–95. (c) Shen, X.-L.; Zhao, R.-R.; Mo, M.-J.; Peng, F.-Z.; Zhang, H.-B.; Shao, Z.-H. Catalytic Enantioselective and Divergent Total Synthesis of (+)-10-Oxocylindrocarpinol, (+)-Cylindrocarpidine, (–)-*N*-Acetylcylindrocarpinol, and (+)-*Aspidospermine*. *J. Org. Chem.* **2014**, *79*, 2473–2480. (d) Wang, N.; Du, S.; Li, D.; Jiang, X. Divergent Asymmetric Total Synthesis of (+)-*Vincadifformine*, (–)-*Quebrachamine*, (+)-*Aspidospermidine*, (–)-*Aspidospermine*, (–)-*Pyrifolidine*, and Related Natural Products. *Org. Lett.* **2017**, *19*, 3167–3170.

(6) Evano, G.; Wang, J.; Nitelet, A. Metal-Mediated C–O Bond Forming Reactions in Natural Product Synthesis. *Org. Chem. Front.* **2017**, *4*, 2480–2499.

(7) (a) O'Connor, S. E.; Maresh, J. J. Chemistry and Biology of Monoterpene Indole Alkaloid Biosynthesis. *Nat. Prod. Rep.* **2006**, *23*, 532–547. (b) El-Sayed, M.; Verpoorte, R. Catharanthus Terpenoid Indole Alkaloids: Biosynthesis and Regulation. *Phytochem. Rev.* **2007**, *6*, 277–305.

(8) White, K. L.; Movassaghi, M. Concise Total Syntheses of (+)-*Haplocidine* and (+)-*Haplocine* via Late-Stage Oxidation of (+)-*Fendleridine* Derivatives. *J. Am. Chem. Soc.* **2016**, *138*, 11383–11389.

(9) Jiao, L.-Y.; Oestreich, M. C–H Bond Functionalization at the Benzene Core of Indoles and Indolines (Excluding C-2 and C-3). In *Strategies for Palladium-Catalyzed Non-Directed and Directed C-H Bond Functionalization*; Kapdi, A. R., Maiti, D., Eds.; Elsevier: Amsterdam, 2017; pp 205–232.

(10) For selected examples of directed acetoxylation, see (a) Dick, A. R.; Hull, K. L.; Sanford, M. S. A Highly Selective Catalytic Method for the Oxidative Functionalization of C–H Bonds. *J. Am. Chem. Soc.* **2004**, *126*, 2300–2301. (b) Giri, R.; Liang, J.; Lei, J.-G.; Li, J.-J.; Wang, D.-H.; Chen, X.; Naggar, I. C.; Guo, C.; Foxman, B. M.; Yu, J.-Q. Pd-Catalyzed Stereoselective Oxidation of Methyl Groups by Inexpensive Oxidants under Mild Conditions: A Dual Role for Carboxylic Anhydrides in Catalytic C–H Bond Oxidation. *Angew. Chem. Int. Ed.* **2005**, *44*, 7420–7424. (c) Wang, G.-W.; Yuan, T.-T.; Wu, X.-L. Direct Ortho-Acetoxylation of Anilides via Palladium-Catalyzed sp² C–H Oxidative Activation. *J. Org. Chem.* **2008**, *73*, 4717–4720. (d) Li, G.; Wan, L.; Zhang, G.; Leow, D.; Spangler, J.; Yu, J.-Q. Pd(II)-Catalyzed C–H Functionalizations Directed by Distal Weakly Coordinating Functional Groups. *J. Am. Chem. Soc.* **2015**, *137*, 4391–4397.

(11) (a) Taniguchi, M.; Yamamoto, I.; Nakagawa, M.; Hino, T. Cyclic Tautomers of Tryptophans and Tryptamines. VIII. Cyclic Tautomers of Cyclo-L-Prolyl-L-tryptophyl and Related Compounds. *Chem. Pharm. Bull.* **1985**, *33*, 4783–4791. (b) Somei, M.; Kawasaki, T.; Ohta, T. A Simple Synthesis of 7-Substituted 1-Acetyl-2-3-Dihydroindoles. *Heterocycles* **1988**, *27*, 2363–2365.

(12) (a) De, P. B.; Banerjee, S.; Pradhan, S.; Punniyamurthy, T. Ru(II)-Catalyzed C7-Acyloxylation of Indolines with Carboxylic Acids. *Org. Biomol. Chem.* **2018**, *16*, 5889–5898. (b) Zhai, W.; Li, B.; Wang, B. Rh^{III}-Catalyzed Directed Selective C7-Hydroxylation and Acetoxylation of Indolines. *ChemistrySelect* **2018**, *3*, 8035–8039. (c) Gao, S.; Xu, X.; Tang, H.; Wu, J.-Q.; Luo, J. Cu-Mediated C7 Acetoxylation of Indolines. *ChemistrySelect* **2019**, *4*, 5835–5838.

(13) See Supplementary Information for details.

(14) (a) Yoneyama, T.; Crabtree, R. H. Pd(II) Catalyzed Acetoxylation of Arenes with Iodosyl Acetate. *J. of Mol. Cat. A.* **1996**, *108*, 35–40. (b) Cook, A. K.; Sanford, M. S. Mechanism of the Palladium-Catalyzed Arene C–H Acetoxylation: A Comparison of Catalysts and Ligand Effects. *J. Am. Chem. Soc.* **2015**, *137*, 3109–3118.

(15) (a) Wang, D.-H.; Mei, T.-S.; Yu, J.-Q. Versatile Pd(II)-Catalyzed C–H Activation/Aryl-Aryl Coupling of Benzoic and Phenyl Acetic Acids. *J. Am. Chem. Soc.* **2008**, *130*, 17676–17677. (b) Engle, K. M.; Wang, D.-H.; Yu, J.-Q. Ligand-Accelerated C–H Activation Reactions: Evidence for a Switch of Mechanism. *J. Am. Chem. Soc.* **2010**, *132*, 14137–14151.

(16) Jiao, L.-Y.; Oestreich, M. Oxidative Palladium(II)-Catalyzed Dehydrogenative C–H/C–H Cross-Coupling of 2,3-Substituted Indolines with Arenes at the C7 Position. *Chem. Eur. J.* **2013**, *19*, 10845–10848.

(17) Griffith, W. P.; Ley, S. V.; Whitcombe, G. P.; White, A. D. Preparation and Use of Tetra-*n*-butylammonium Per-ruthenate (TBAP reagent) and Tetra-*n*-propylammonium Per-ruthenate (TPAP reagent) as New Catalytic Oxidants for Alcohols. *J. Chem. Soc., Chem. Commun.* **1987**, 1625–1627.

(18) (a) Lindgren, B. O.; Nilsson, T. Preparation of Carboxylic Acids from Aldehydes (Including Hydroxylated Benzaldehydes) by Oxidation with Chlorite. *Acta Chem. Scand.* **1973**, *27*, 888–890. (b) Bal, B. S.; Childers Jr., W. E.; Pinnick, H. W.; Oxidation of α,β -Unsaturated Aldehydes. *Tetrahedron* **1981**, *37*, 2091–2096.

(19) Still, W. C.; Kahn, M.; Mitra, A. Rapid Chromatographic Technique for Preparative Separations with Moderate Resolution. *J. Org. Chem.* **1978**, *43*, 2923–2925.

(20) All operations involving trimethylsilyldiazomethane were carried out in a well-ventilated fume hood. This includes but is not limited to: measuring the reagent, execution of the transformation, work-up of the reaction mixture, and concentration of the crude reaction mixture.

(21) This resonance overlaps with CD₂HCN; the exact position of resonance was determined by gHSQC correlation (¹H, ¹³C) in ppm: (2.20, 43.2) and (1.96–1.92, 43.2).

(22) Literature value: 1740 (s), 1490, 1380, 793 (w), 782 (s), 742, 720 (w), and 705 (s), see reference 4c.

(23) Halgren, T. A. Merck molecular force field. I. Basis, form, scope, parameterization, and performance of MMFF94. *J. Comput. Chem.* 1996, 17, 490–519.

(24) Frisch, M. J. T., G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Scalmani, G.; Barone, V.; Mennucci, B.; Petersson, G. A.; Nakatsuji, H.; Caricato, M.; Li, X.; Hratchian, H. P.; Izmaylov, A. F.; Bloino, J.; Zheng, G.; Sonnenberg, J. L.; Hada, M.; Ehara, M.; Toyota, K.; Fukuda, R.; Hasegawa, J.; Ishida, M.; Nakajima, T.; Honda, Y.; Kitao, O.; Nakai, H.; Vreven, T.; Montgomery, J. A., Jr.; Peralta, J. E.; Ogliaro, F.; Bearpark, M.; Heyd, J. J.; Brothers, E.; Kudin, K. N.; Staroverov, V. N.; Kobayashi, R.; Normand, J.; Raghavachari, K.; Rendell, A.; Burant, J. C.; Iyengar, S. S.; Tomasi, J.; Cossi, M.; Rega, N.; Millam, M. J.; Klene, M.; Knox, J. E.; Cross, J. B.; Bakken, V.; Adamo, C.; Jaramillo, J.; Gomperts, R.; Stratmann, R. E.; Yazyev, O.; Austin, A. J.; Cammi, R.; Pomelli, C.; Ochterski, J. W.; Martin, R. L.; Morokuma, K.; Zakrzewski, V. G.; Voth, G. A.; Salvador, P.; Dannenberg, J. J.; Dapprich, S.; Daniels, A. D.; Farkas, Ö.; Foresman, J. B.; Ortiz, J. V.; Cioslowski, J.; Fox, D. J.; Gaussian, Inc.: Wallingford CT, 2009; Vol. Revision D.01.