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Clinical characteristics of recovered COVID-19 patients with re-detectable positive RNA test

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Background: The characteristics, significance and potential cause of positive SARS-CoV-2 diagnoses in recovered coronavirus disease 2019 (COVID-19) patients post discharge (re-detectable positive, RP) remained elusive.

Methods: A total of 262 COVID-19 patients discharged from January 23 to February 25, 2020 were enrolled into this study. RP and non-RP (NRP) patients were grouped according to disease severity, and the characterization at re-admission was analyzed. SARS-CoV-2 RNA and plasma antibody levels were measured, and all patients were followed up for at least 14 days, with a cutoff date of March 10, 2020.

Results: A total of 14.5% of RP patients were detected. These patients were characterized as young and displayed mild and moderate conditions compared to NRP patients while no severe patients were RP. RP patients displayed fewer symptoms but similar plasma antibody levels during their hospitalization compared to NRP patients. Upon hospital readmission, these patients showed no obvious symptoms or disease progression. All 21 close contacts of RP patients were tested negative for viral RNA and showed no suspicious symptoms. Eighteen out of 24 of RNA-negative samples detected by the commercial kit were tested positive for viral RNA using a hyper-sensitive method, suggesting that these patients were potential carriers of the virus after recovery from COVID-19.

Conclusions: Our results indicated that young patients, with a mild diagnosis of COVID-19 are more likely to display RP status after discharge. These patients show no obvious symptoms or disease progression upon re-admission. More sensitive RNA detection methods are required to monitor these patients. Our findings provide information and evidence for the management of convalescent COVID-19 patients.

Keywords: Coronavirus; SARS-CoV-2; coronavirus disease 2019 (COVID-19)

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Introduction

In December 2019, lots of unexplained pneumonia cases appeared which clinical manifestations suggested viral pneumonia. Deep sequencing of lower respiratory tract samples identified a novel coronavirus named the severe acute respiratory syndrome Coronavirus-2 (SARS-CoV-2), and the disease it caused was named coronavirus disease 2019 (COVID-19) (1). Over 300,000 cases of COVID-19 were reported from December 2019 to March 23, 2020. COVID-19, has resulted in 13,000 deaths, globally (1), with The World Health Organization declaring this virus as a pandemic (2). Generally, COVID-19 is less severe and fatal than the SARS. However, some patients, especially the elderly with co-morbidities are prone to developing more severe symptoms and require urgent medical interventions (3,4). Many literature reports have retrospectively analyzed the clinical characteristics of patients infected with SARS-CoV-2 (3-8). Recently, an increasing number of patients with COVID-19 were discharged from the hospital and received regular follow-up and observation. Re-detectable positive (RP) diagnoses of SARS-CoV-2 RNA test in some recovered patients have been reported (9-13). The first reported four patients with COVID-19 who met criteria for hospital discharge or discontinuation of quarantine in China were reported with positive SARS-CoV-2 RNA test 5 to 13 days later which suggest that at least a proportion of recovered patients still may be virus carriers (9). Parts of pediatric patients remained positive viral RNA in stools for longer than 4 weeks which indicates the potential for the virus to be transmitted through fecal excretion and detection of fecal SARS-CoV-2 RNA would be more sensitive (12). Three readmission patients of COVID-19 with negative IgM and positive IgG was described with no fever symptoms and absorbed lesions on computed tomography (CT) images which suggested serum antibody responsive to SARS-CoV-2 associated the viral shedding (13). The management of RP patients has attracted wide attention. However, the RP population reported in the literature was small, with a short duration of follow-up. In addition, the clinical characteristics, potential impact, and significance of RP patients remain unknown, making it difficult to provide empirical information and evidence-based support for the

management of patients with COVID-19 during recovery.

This study retrospectively analyzed the clinical characteristics of 38 RP patients and 224 non-RP (NRP) patients who recovered from COVID-19. It was revealed that RP patients were characterized by younger age and milder conditions. They also displayed minor symptoms, more sustained remission by CT imaging, earlier RNA negative conversion, and similar plasma antibody levels during their hospitalization period. They showed no obvious disease progression and infectivity when re-admitted to the hospital. The hyper-sensitive detection method which identified SARS-CoV-2 RNA molecules from most samples initially confirmed as RNA-negative by a commercial kit suggests that recovering COVID-19 patients were potential carriers during this period. These findings provide key information for the effective management of COVID-19 patients during their convalescent phase.

We present the following article in accordance with the MDAR reporting checklist (available at http://dx.doi. org/10.21037/atm-20-5602).

Methods

Study design and participants

A total of 262 confirmed COVID-19 patients discharged from Shenzhen Third People's Hospital from January 23, 2020 to February 25, 2020 were enrolled in this study. All discharged patients continued to be isolated and were observed for 14 days. Weekly follow-up in person and SARS-CoV-2 RNA detection were also performed at the same time. Patients were subsequently followed-up for at least an additional two weeks, post isolation. Unlike recovered NRP patients, who were closely followed-up outside the hospital, individuals identified as RP were readmitted to hospital for further medical observation. The close contacts of RP patients also underwent weekly followup in person for 14 day. This study was approved by the Ethics Committee of The Third People's Hospital of Shenzhen (2020-115), which waived the requirement for written patient consent for this retrospective analysis. The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013). All patients provided oral

consent to participate in this retrospective study.

Clinical definition

According to the guideline for the diagnosis and treatment for novel coronavirus pneumonia (the sixth edition), published by National Health Commission of the People's Republic of China (14), all initial diagnoses of COVID-19 were confirmed with positive respiratory RT-PCR tests. The discharge criteria of the recovered patients included: temperature returned to normal for more than 3 days, respiratory symptoms significantly improved, and significant absorption of pulmonary lesions by chest CT imaging, and a minimum of two consecutive negative RNA test results, performed at least 24 hours apart. RP patients were confirmed by digestive (anal swab) and respiratory positive RT-PCR tests. Since February 22, 2020, evaluation of negative anal swab was supplemented for the discharge criteria in Shenzhen Third People's Hospital.

Data collection

The medical records of 262 patients, recovered from COVID 19 were reviewed. These records also comprised of 38 RP patients. The epidemiological, demographic, clinical, laboratory data of the patients were collected, summarized and analyzed. According to the initial chest CT imaging post admission, the severity of pulmonary inflammation was divided into mild, moderate and severe, based on lesions involving unilateral lobe, multiple lobes in both lungs, and all lobes in both lungs, respectively. The remission of the lesions was evaluated by chest CT within 7 days after admission. Temporary progression was indicated by increased lesions, whilst persistent remission was indicated by stable, absorbed or decreased lesions.

qRT-PCR and Sherlock assay for SARS-CoV-2 RNA detection

The quantitative reverse transcription polymerase chain reaction (qRT-PCR) was assessed as described previously (15). Nasopharyngeal and anal specimens collected during hospitalization were sent to the laboratory in a viral transport case. Total nucleic acid extraction from the samples was performed using the QIAamp RNA Viral Kit (Qiagen, Heiden, Germany). Quantitative RT-PCR was undertaken using a China Food and Drug Administration (CFDA) approved commercial kit specific for 2019-nCoV detection (GeneoDX Co., Ltd., Shanghai, China) or Sherlock kit, gifted from Feng Zhang lab according to the manual. Each RT-PCR assay provided a Ct value, representing the number of cycles required for the fluorescent signal to cross the threshold for a positive test. A higher Ct value is correlated with a lower viral load. Specimens were considered positive if the Ct value was \leq 37.0, and negative if the viral load was undetectable. Specimens with a cyclethreshold value higher than 37 were repeated. The specimen was considered positive if the repeat results were equivalent to the initial result and between 37 and 40. If the repeat Ct was undetectable, the specimen was considered negative. All procedures involving clinical specimens and SARS-CoV-2 were performed in a biosafety level 3 laboratory. Meanwhile, next-generation sequencing of samples were performed from samples of three patients.

CMLA assay for anti-SARS-CoV-2 antibodies

The SARS-CoV-2 specific total antibody (Ab), IgG, IgA, and IgM in plasma was tested using a Chemiluminescence Microparticle Immuno Assay (CMIA). Briefly, recombinant SARS-CoV-2-RBD (Receptor binding domain) proteins were immobilized, and HRP-conjugated antigen detected total antibodies, IgM, IgA and IgG. Total antibodies were tested using double-antigens sandwich enzyme-linked immunosorbent assay (Ab-ELISA). IgM was tested by the IgM u-chain capture method (IgM-ELISA). IgA and IgG were tested using indirect ELISA. The testing kits were supplied by the Beijing Wantai Biological Pharmacy Enterprise Co., Ltd., China. The relative fluorescence of sample to control (COI) was used to measure the antibody concentration. The larger the COI value, the higher the concentration. When COI exceeded 1, the result was deemed positive.

Statistical analysis

Statistical analysis was performed using SPSS 26.0 (IBM, Chicago). All tests were two-sided, and a P<0.05 was considered significant. Continuous variables were evaluated using the median and interquartile range (IQR) values. Chi-square or Fisher exact tests were utilized to compare the proportions of the categorical variables.

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Table 1 Baseline characteristi	s of enrolled patier	nts with COVID-19
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		Mild (n=30)		٩	Noderate (n=212	2)
Characteristics	RP (n=11)	NRP (n=19)	P value	RP (n=27)	NRP (n=185)	P value
Age, median (IQR), yr	20 [5–64]	23 [2–63]	0.98	38 [2–60]	48 [1–86]	<0.01
≤60 years old, n (%)	10 (90.9)	18 (94.7)	0.78	26 (96.3)	139 (75.1)	0.11
≤14 years old, n (%)	4 (36.3)	6 (31.6)	0.57	3 (11.1)	6 (3.24)	0.04
>60 years old, n (%)	1 (9.1)	1 (5.3)	0.32	1 (3.7)	46 (24.9)	<0.001
Gender, n (%)						
Male	4 (36.3)	10 (52.6)	0.08	12 (44.4)	90 (48.6)	0.66
Female	7 (63.7)	9 (47.4)	0.12	15 (55.6)	95 (51.4)	0.69
Comorbidities, n (%)	1 (9.1)	0 (0)	NA	1 (3.7)	41 (22.2)	<0.01
History of travel or residence in Hubei, n (%)	10 (90.9)	16 (84.2)	0.61	23 (85.2)	152 (82.2)	0.82
Fever, n (%)	2 (18.2)	7 (36.8)	<0.01	23 (85.2)	133 (71.9)	0.29
Jpper respiratory symptoms, n (%)	5 (45.5)	2 (10.5)	<0.01	4 (14.8)	34 (18.4)	0.53
_ower respiratory symptoms, n (%)	5 (45.5)	7 (36.8)	0.34	14 (51.9)	95 (51.4)	0.96
Digestive tract symptoms, n (%)	0 (0)	2 (10.5)	NA	3 (11.1)	15 (8.11)	0.50
The lesion range of chest CT, n (%)						
Unilateral	0 (0)	0 (0)	NA	6 (22.2)	36 (19.4)	0.66
Multi-lobe of bilateral	0 (0)	0 (0)	NA	15 (55.6)	105 (56.7)	0.92
All-lobe of bilateral	0 (0)	0 (0)	NA	6 (22.2)	44 (23.7)	0.82
Chest CT imaging, n (%)						
Transient progression	0 (0)	0 (0)	NA	4 (14.8)	67 (36.2)	<0.05
Sustained remission	0 (0)	0 (0)	NA	23 (85.2)	118 (63.8)	0.05
Steroids use, n (%)	0 (0)	0 (0)	NA	4 (14.8)	27 (14.6)	0.97

RP, re-detectable positive patients; NRP, non-re-detectable positive patients

Results

Demographic, epidemiological and clinical characteristics

A total of 262 patients were discharged from January 23, 2020 to February 25, 2020 and weekly followedup in person with detection of SARS-CoV-2 RNA in nasopharyngeal and anal specimens for at least 14 days. Mild, moderate and severe patients accounted for 11.4% (n=30), 81.0% (n=212) and 7.6% (n=20), respectively in this study population. Up to March 10, 14.5% of convalescent patients (n=38) were re-detected as SARS-CoV-2 RNA positive during their follow-up period. Severe patients were found to be RNA negative (*Table S1*).

Our findings revealed that the vast majority of RP

patients (94.7%, n=36) were younger than 60 years of age. Among them, patients younger than 14 years old were more prevalent compared to those aged between 14 and 60 years (35.0% vs. 16.0%, P<0.01) (*Table 1*). In addition, the proportion of RP patients displaying mild symptoms (36.7%) was significantly higher than what was observed in NRP patients (12.7%, 19/204, P<0.01, *Table S1*). No significant difference in gender distribution was exhibited between RP and NRP patients. Notably, the incidence of fever as part of the initial symptoms was higher in mild NRP patients, compared to RP patients (P<0.01). Additionally, 45.5% of mild RP patients displayed only upper respiratory symptoms at the first admission, unlike mild NRP patients, who exhibited lower respiratory

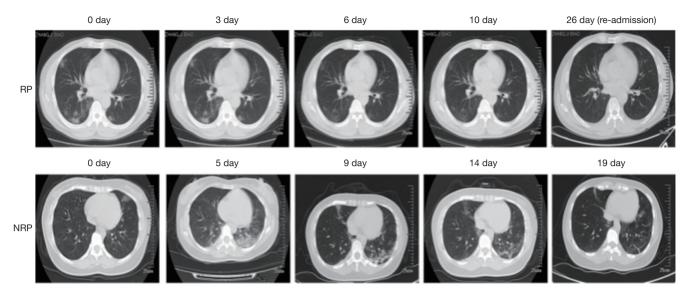


Figure 1 Serial CT imaging of a representative RP and NRP patient. For the RP patient, the first chest CT scan on admission (day 7 since the onset of illness) showed ground-glass opacity in both lungs. At 3, 6 and 10 days after admission (day 10, day 13 and day 17 since the onset of illness), lung lesions in the chest, detected by CT imaging was significantly reduced and accompanied by the disappearance of clinical symptoms. The patient was discharged at day 12 after admission (day 19 since the onset of illness). At day 26 (day 33 since the onset of illness), the patient was re-admitted without fever and cough due to positive RNA detection. The chest CT showed no inflammatory lesions. For the NRP patient, a chest CT scan showed a small ground glass in the upper left lung on admission (day 3 since the onset of illness). On days 2 and 8 after admission (day 5 and day 11 since the onset of illness), the double lower lung lesions increased significantly, as shown by chest CT imaging, although the body temperature and the oxygenation index returned to normal levels. On days 9, 14 and 17 after admission (day 12, day 17 and day 20 since the onset of illness), CT imaging of the chest illustrate the recovery of lesions in both lower lungs. The patient was subsequently discharged without fever and cough at day 18, post admission (day 21 since the onset of illness) when SRAS-CoV-2 RNA was also detected to be negative.

symptoms (*Table 1*). Initial lesion analysis of the chest by CT scan showed no difference in the extent of lesions between moderate RP and NRP patients. However, the incidence of RP (85.2%) was closely related to sustained remission, as indicated by CT imaging of the chest, compared to NRP patients, where 36.2% displayed transient progression during their initial hospitalization (*Table 1* and *Figure 1*). There was no significant difference in the administration of steroid and antiviral therapies between RP and NRP patients during their first hospitalization. In addition, RP patients did not exhibit a higher incidence traveling and living in Hubei province compared to NRP patients.

Differential RNA dynamic in RP and NRP patients

No differences were noted in the duration from illness onset to RNA negative-conversion and hospitalization days between RP and NRP patients. Analysis of the duration between illness onset and RNA negative-conversion was performed. The study revealed that RNA negative conversion occurred mostly within 2–3 weeks from the onset of illness in 63.6% of mild, and 1–2 weeks in 22.2% moderate RP patients. By contrast, there were more NRP patients who displayed RNA negative-conversion after 3 weeks from illness onset, regardless of disease severity (*Table 2*). The data showed that early RNA-negative conversion was characteristic of RP patients, while NRP patients exhibited late viral clearance.

Changes in serum anti-SARS-CoV-2 antibodies

To evaluate the effect of serum-specific antibody levels on the incidence of RP, we analyzed the difference in anti-SARS-CoV-2 antibody levels in RP and NRP patients. A total of 989 plasma samples, including those from 36 RP and 228 NRP patients were examined for total Ab, IgM, IgG, and IgA against SARS-CoV-2. Total antibodies (*Figure 2A*), IgG (*Figure 2B*) and IgA (*Figure 2C*) were detected four

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Table 2 RNA detection in the enrolled patients with COVID-19

Duration of DNA data stick		Mild (n=30)			Moderate (n=212)			
Duration of RNA detection	RP (n=11)	NRP (n=19)	P value	RP (n=27)	NRP (n=185)	P value		
*Days since the onset of illness to								
last RNA negative-conversion	17 [11–22]	15 [8–24]	0.71	18 [9–30]	20 [5–47]	0.17		
Follow-up deadline (March 10)	40 [33–47]	42 [35–49]	0.15	45 [33–54]	46 [30–72]	0.15		
Discharge	15 [14–22]	16 [10–23]	0.72	17 [9–29]	18 [7–35]	0.47		
Days of RNA negative-conversion since	the onset of illness	(n, %)						
Between 7 and 14 days	3 (27.3)	8 (42.1)	0.08	6 (22.2)	18 (9.7)	0.03		
Between 14 and 21 days	7 (63.6)	8 (42.1)	0.04	11 (40.7)	84 (45.4)	0.61		
More than 21days	1 (9.1)	3 (15.8)	0.18	10 (37.3)	83 (44.9)	0.40		

*, median [range]. RP, re-detectable positive patients; NRP, non-re-detectable positive patients

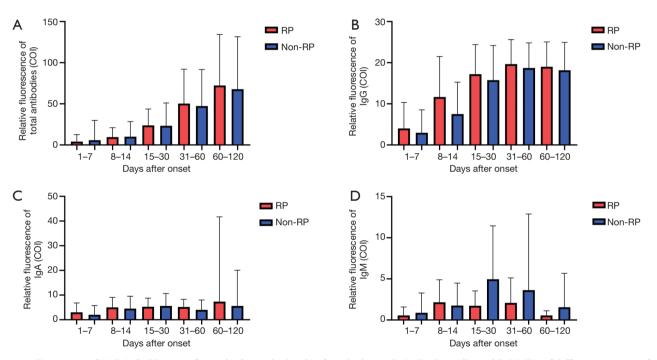


Figure 2 Dynamics of SARS-CoV-2 specific antibodies. The levels of total Ab (A), IgG (B), IgA (C), and IgM (D) of different patients after illness onset. The relative antibody level was estimated using COI, expressed as the mean (denoted by columns) and SD (denoted by error bars). Red and blue columns represent RP and NRP Patients, respectively.

months or longer at sustained levels in both groups. No differences in antibody levels were observed between RP and NRP patients. IgM levels increased slowly before exhibiting a decline in both the groups. Peak IgM levels (*Figure 2D*) in RP patients was slightly higher than NRP patients 15–30 days after illness onset, with a P value (0.056) exceeding 0.05.

Supplementing negative results via anal swab test at discharge failed to reduce RP occurrence of COVID-19 patients

To determine if increasing site tests at discharge affected the incidence of RP, we compared its occurrence before and after the introduction of a supplementary test. The negative

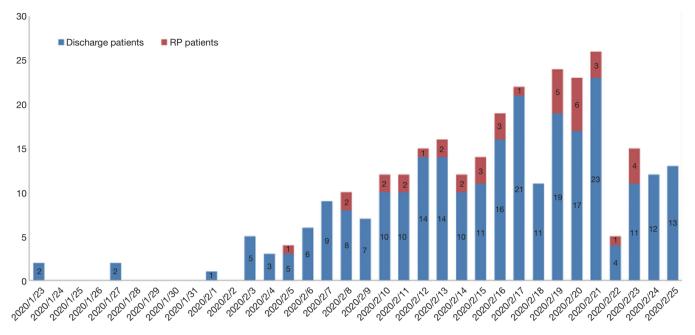


Figure 3 The number of discharged patients and RP patients per day from Jan 23 to March 10, 2020. On Feb 22, 2020, the anal swab negative test was added to discharge criterion. Blue indicates the number of discharge patients. Red represents the number of RP patients.

anal swab detection was an added procedure as part of the discharge criterion on 22^{nd} February, 2020 in COVID-19 patients. Our results showed that there was no statistical difference in the occurrence of RP patients before or after February 22, 2020 (14.5% *vs.* 14.3%, P=0.77, *Figure 3*). The data indicated that supplementary testing at additional detection sites of SARS-CoV-2 RNA failed to reduce the incidence of RP in convalescent patients.

RP patients showed no obvious clinical symptoms and disease progression

All 38 RP patients were re-admitted to hospital for further medical observation. The analysis showed that these patients did not display fever, and a small number of patients reported mild cough and chest tightness, which were previously observed and did not increase in severity (*Table 3*). All patients recovered from mild conditions (n=30). CT imaging revealed 37.0% of moderate patients displayed normal chests with no signs of inflammation. The remaining 63.0% (n=17) of patients in the same group recovered from moderate conditions and displayed stable or reduced chest inflammation, as confirmed by CT imaging (*Figure 1*). All RP patients exhibited lymphocyte, plasma IL-6 and CRP levels within the normal range of the upon admission. Only 1 patient received transient interferonalpha inhalation therapy, and 4 patients received lowflow oxygen inhalation therapy and 11 patients received traditional Chinese medicine after admission (*Table 3*).

All the convalescent patients with COVID-19 in our cohort were required to be isolated at home or undergo intensive isolation, yielding only 21 close contacts. Up to March 10, 2020, all close contacts tested negative for SARS-CoV-2 RNA, and no suspicious clinical symptoms were reported in those close contacts (*Table 3*).

Hyper-sensitive methods potentially improved SARS-CoV-2 RNA detection in RP patients

To investigate the possibility of false negative diagnoses due to the low sensitivity of commercial RNA detection kits, an alternative method with higher detection potential was applied to Anal swab, Blood and Nasal swab from RP and NRP patients displaying similar illness days. Twentyfour samples were obtained from 15 RP patients, 5–7 days since the onset of the re-admission. Our findings revealed that 75% of spike genes and 41.6% ORF1 genes were detected using this hyper-sensitive method on, in contrast to 12.5% N genes and 4.2% ORF1 genes, identified by the commercial detection kit. Eight of 15 RP patients were

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Table 3 Clinical observation of RE	patients at re-admission of hospital
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Clinical conditions	Mild (n=11)	Moderate (n=27)	
Symptoms, n (%)			
Fever	0 (0)	0 (0)	
Cough	1 (9.1)	5 (18.5)	
Chest tightness	0 (0)	2 (7.4)	
Other	0 (0)	3 (11.1)	
Chest CT imaging, n (%)			
Normal	11 (100.0)	10 (37.0)	
Stable or absorb	0 (0)	17 (63.0)	
Progression	0 (0)	0 (0)	
Laboratory examination, n (%)			
Abnormal lymphocyte count	0 (0)	4 (14.8)	
Increasing serum IL-6 level	0 (0)	0 (0)	
Increasing serum CRP level	0 (0)	0 (0)	
Treatment, n (%)			
Low flow oxygen	0 (0)	4 (14.8)	
Traditional Chinese medicine	3 (27.3)	8 (29.6)	
Antiviral therapy	0 (0)	1 (3.7)	
Number of contacts with symptoms	0	0	

confirmed as RNA positive using the hyper-sensitive kit, whereas only 1 person tested positive with the commercial kit. By contrast, 8 samples from NRP patients were detected to be negative by both methods (*Table 4*). The data demonstrates that hyper-sensitive methods potentially improved RNA positive detection in RP patients.

Discussion

Several studies have confirmed the existence of RP patients (9-13), however their clinical characterization was not well defined or with limited clinical samples. This retrospective study analyzed the clinical and follow-up data in a cohort of RP and NRP patients during the same discharge period. Up to March 10, 2020, 38 RP patients were present, accounting for 14.5% of discharged patients during the same follow-up period. These patients displayed several significant features, including younger age and mild and/or moderate symptoms during hospitalization, which is consistent with previous reports (9,12). Mild RP patients were usually below 14 years of age and moderate RP patients were below the age of 60.

By contrast, no severe patients were found to be RP within a similar follow-up period. In addition, more RP patients displayed minor symptoms during hospitalization such as fewer comorbidities, less severe fever, and increased upper respiratory symptoms. CT imaging indicated that RP patients had no lesions or maintained sustained remission in their lungs as those of NRP patients. The data indicated that RP patients were characterized by a younger age and minor symptoms during hospitalization.

Viral load is usually considered to be related to disease outcome (15,16). The present study indicated that RNA negative-conversion commonly occurred 2–3 weeks from the onset of illness in moderate RP patients, compared to over 3 weeks in NRP patients of the same disease severity. The significantly shortened RNA negative-conversion duration may affect the persistence of high adaptive immunity levels (17). Our recent study indicated that a higher antibody titer in the plasma was independently associated with disease severity in patients with COVID-19 (18). However, RP and NRP patients displayed similar plasma IgG and IgM levels. Future studies are required

Table 4	The com	narison	hetween	hvper-sensitivit	v and c	rommon	sensitivity	detection	in RP	and NRP	natients
	I IIC COIII	parison	Detween	nyper-sensitivit	y and c	Johnnon	SCHSILLVILV	ucuculon	III IVI		patients

NI-	Sample	Dates since the	She	erlock	Commercial		
No.	Sample		S gene	ORF1 gene	N gene	ORF1 gene	
1	Anal swab	44	_	_	_	_	
2	Nasal swab	44	+	-	_	-	
3	Anal swab	44	+	-	_	-	
4	Nasal swab	44	+	+	_	_	
5	Anal swab	37	-	+	+	-	
3	Nasal swab	37	+	+	_	_	
7	Anal swab	42	+	-	_	_	
3	Anal swab	43	-	+	-	_	
)	Anal swab	30	+	+	-	_	
10	Blood	42	+	-	-	_	
11	Blood	43	-	-	-	_	
12	Blood	30	+	-	-	_	
13	Nasal swab	42	-	-	+	_	
14	Nasal swab	43	+	-	-	_	
15	Nasal swab	30	+	+	-	-	
16	Anal swab	32	+	+	-	_	
17	Anal swab	37	+	-	-	-	
18	Anal swab	36	+	-	_	_	
19	Anal swab	37	+	+	_	_	
20	Anal swab	41	+	+	-	_	
21	Anal swab	37	-	-	+	+	
22	Anal swab	31	+	-	-	_	
23	Anal swab	43	+	+	-	_	
24	Nasal swab	32	+	-	-	_	
Total, positive (%)			18 (75%)	10 (41.6%)	3 (12.5%)	1 (4.2%)	
25	Nasal swab	47	-	-	_	_	
26	Anal swab	47	-	-	_	_	
27	Nasal swab	45	-	-	_	_	
28	Nasal swab	40	-	-	_	_	
29	Anal swab	54	-	-	_	_	
30	Anal swab	49	-	-	-	_	
31	Nasal swab	44	-	-	_	-	
32	Nasal swab	48	-	-	_	_	
Total, positive (%)			8 (0)	8 (0)	8 (0)	8 (0)	

Note: 1–24, re-detectable positive patients; 25–32: non-re-detectable positive patients. +, represents positive RNA result; –, represents negative RNA result.

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to investigate host immune responses and its effects on determining clinical outcomes in viral infection (19,20).

We also comprehensively characterized the clinical symptoms of RP patients when re-admitted to the hospital. No obvious clinical evidence of disease progression or recurrence was found in these patients from CT imaging and laboratory tests. Additionally, no antibiotics, steroids, antiviral agents and continuous supplemental oxygenation were required. RP patients also displayed significantly reduced inflammatory response during readmission. The data indicated that the diseases of RP patients did not increase in severity despite the positive RNA detection for SARS-CoV-2. More importantly, these patients did not be reinfected after discharge, consistent with a recent longitudinal study in SARS-CoV-2 infected rhesus macaques, where reinfection could not occur in convalescent monkeys (21). Long-term follow-up of close contacts with RP patients will warrant the evaluation of the potential risk of RP.

The mechanisms underlying RP occurrence remain unclear. The possible reasons proposed by a large number of experts are related to several virological, immunological and sampling methodological factors. From a viral standpoint, false negatives (22), viral residual (12), intermittent viral release (12) and viral distribution (23,24) are usually considered as major factors. Our data support the notion that the false negative diagnoses originating from commercial kits may partially account for RPs, as the kits only provide a 30–50% positive rate of detection (24,25). In 24 samples from RP patients, RNA was detected as negative for both N gene and ORF1b gene several days after hospital re-admission using commercial kit, whose lower limit of detection (LOD) was relatively high (500 copies/mL). However, a higher sensitivity Sherlock kit with an LOD of 100 copies/mL (26), confirmed 75% of samples as positive for the S gene and 41.6% for ORF genes. The commercial kit therefore failed to identify half of positive subjects within the RP patient population during hospitalization. By contrast, the Sherlock or commercial kit did not detect any of the 8 samples from NRP patients as positive. However, the Sherlock kit detected a sample previously confirmed by SRAS-CoV-2 sequencing, as positive (data not shown). Therefore, future studies are required with the aim to improve the sensitivity and specificity of detection kits to facilitate the accurate diagnosis of clinical samples. The data indicated that false positive results, detected by the current commercially available kit may account for the occurrence of RP patients, to some extent.

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Another viral factor pertains to long-term virus residual levels in the gut and other tissues, similar to SARS (27). A recent study indicated that SARS-CoV-2 nucleic acid can persist in the digestive tract and feces for nearly 50 days (28). Thus, extending the follow-up time is necessary for the COVID-19 patients when they were discharged. However, our results indicated that additional discharge criterion such as negative results from supplementary RNA testing, which originated from anal swabs, did not significantly reduce the incidence of RP patients. Thus other factors may be associated with the RP patients. We could not exclude sampling methodological factors including differential sampling and operational methods, sample quality, and technician expertise levels. Nor could we exclude immunological factors including low mucosal immune responses such as low IgA levels. These factors may take some uncertain risks leading the occurrence of RP patients (4,28). Future studies focused on the application of the hyper-sensitive detection kit, combined with the detection of multiple samples and more immune markers in should reduce the incidence of RP.

This study has several limitations. Firstly, this study was a single-center retrospective study with a short followup duration. More clinical observations are needed to evaluate the potential risk of SARS-CoV-2 recurrence and infection. Secondly, the dynamics of SARS-CoV-2 RNA in COVID-19 patients need to be monitored and evaluated for RP. Furthermore, additional studies which measure the dynamic changes of serum specific antibody levels in RP patients and evaluate the continuous protective effect of serum specific antibodies on patients with COVID-19 are required. Finally, distinctions are required between RP, relapsed, and convalescent patients, where two distinct prevention and control strategies will be adopted.

Collectively, our findings revealed the clinical features of RP patients who did not exhibit the recurrence of clinical symptoms and abnormal laboratory tests. However, hyper-sensitive detection methods revealed the existence of SARS-CoV-2 RNA in RP patient specimens, initially tested as negative using the commercial kit. Therefore, the development of a more accurate kit is required for quantitative assessments of the RNA dynamics and additional discharge criteria to aid in the guidance of clinical decisions associated with COVID-19. This study provided valuable empirical information and clinical evidence support for effective management of COVID-19 patients during the convalescent period. Further study should evaluate the potential clinical significance and

transmission risk of RP patients.

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Footnote

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Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. This study was approved by the Ethics Committee of The Third People's Hospital of Shenzhen (2020-115), which waived the requirement for written patient consent for this retrospective analysis. The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013). All patients provided oral consent to participate in this retrospective study.

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Supplementary

Table S1 Analysis of disease severity in RP and NRP patients

	Mild (n=30) (%)	Moderate (n=212) (%)	Severe (n=20) (%)	Total (n=262) (%)
RP	11 (36.7)	27 (12.7)	0 (0)	38 (14.5)
NRP	19 (63.3)	185 (87.3)	20 (100.0)	224 (85.5)

RP, re-detectable positive patients; NRP, non-re-detectable positive patients.