DECOMPOSITION OF HYDROGEN PEROXIDE AND ORGANIC COMPOUNDS IN THE PRESENCE OF IRON AND IRON OXIDES

by

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Abstract

Most advanced oxidation processes use the hydroxyl radical (•OH) to treat pollutants found in wastewater and contaminated aquifers because •OH reacts with numerous compounds at near diffusion-limited rates. •OH can be made by reacting hydrogen peroxide (H₂O₂) with either Fe(II) (the Fenton reaction), Fe(III), or iron oxide. This dissertation investigated the factors that influence the decomposition rates of H₂O₂ and organic compounds, as well as the generation rate of •OH (VOH), in the presence of dissolved Fe(III) and iron oxide.

The Fe(III)-initiated chain reaction could be the dominant mechanism for the decomposition of H₂O₂ and organic compounds. The degradation rates of H¹⁴COOH, an •OH probe, and H₂O₂ were measured in experiments at pH 4 containing either dissolved Fe(III) or ferrihydrite. Combined with the results from experiments using a radical chain terminator, we concluded that a solution chain reaction was important only in the Fe(III) system. In the ferrihydrite system the amount of dissolved Fe was insufficient to effectively propagate the chain reaction. In addition, a nonradical producing H₂O₂ loss pathway exists at the oxide surface.

The oxidation rate of any dissolved organic compound can be predicted from VOH if the main sinks of •OH in the solution are known. Experiments using H¹⁴COOH and ferrihydrite, goethite, or hematite showed that VOH was proportional to the product of the concentrations of surface area and H₂O₂. Based on these results, a model was created for predicting the pseudo-first-order oxidation rate coefficients of dissolved organic compounds (korg) in systems containing iron oxide and H₂O₂. While our model successfully predicted korg in aquifer sand experiments, it yielded mixed results when compared to measurements from previously published studies.

Some factors that could have caused the disagreements between model predictions and measurements were examined to refine our model. Results from experiments containing goethite, H¹⁴COOH, and 2-Chlorophenol showed that H¹⁴COOH detected more •OH, which is produced at the oxide surface, than did 2-Chlorophenol. This was attributed to electrostatic attraction between the formate anions and the positively charged oxide surface, and could explain why our model, based on H¹⁴COOH, overpredicted the korg values of many neutral compounds.

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Lastly, I thank my parents and my sister for their support.
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Chapter 1: Introduction

or

-OH, you're one radical dude!-
The hydroxyl radical ($\cdot$OH) is a powerful inorganic oxidant that reacts with numerous compounds at or near diffusion-limited rates ($10^{10} \text{ M}^{-1} \text{ s}^{-1}$; I-3). It is of great research interest because it is used in the remediation of wastewater and polluted groundwater (4-9), and is found naturally in surface waters (10-14), clouds (15, 16), and living cells (17-19).

A source of $\cdot$OH common to all of the systems listed above is the oxidation of Fe(II) by hydrogen peroxide ($\text{H}_2\text{O}_2$), also known as the Fenton reaction (20). A wealth of information exists regarding the mechanism and kinetics of $\cdot$OH production during the decomposition of $\text{H}_2\text{O}_2$ by Fe(II) and Fe(III) (21-30). For example, the generally accepted mechanism of the decomposition of $\text{H}_2\text{O}_2$ by Fe(III) consists of a chain reaction where the iron cycles between Fe(III) and Fe(II) as $\text{H}_2\text{O}_2$ is consumed (21, 24, 30).

While the pure solution phase Fenton system has been researched extensively, work on elucidating the mechanism of $\text{H}_2\text{O}_2$ decomposition and $\cdot$OH generation in the presence of iron oxide has only just begun (31-36). The goal of this dissertation work was to further the understanding of the heterogeneous Fenton-like system and to determine the factors that control the decomposition rates of $\text{H}_2\text{O}_2$ and organic compounds, as well as the generation rate of $\cdot$OH ($V_{\text{OH}}$), in the presence of iron and iron oxide. This knowledge can help to increase the efficacy of hydrogen peroxide-based treatment processes of contaminated aquifers.

The contribution of solution phase reactions to the decomposition of $\text{H}_2\text{O}_2$ and organic compounds in the presence of dissolved iron and ferrihydrite at pH 4 was examined in chapter 2. One aspect that was explored in detail was the importance of the chain reaction mechanism in this system because it could determine the lifetimes of $\text{H}_2\text{O}_2$ and organic compounds. $^{14}$C-labeled formic acid was the main $\cdot$OH probe since its reactions with $\cdot$OH and subsequent reactants are well-characterized. It also has a detection limit on the order of nanomolar, meaning that it can be used at concentrations so low that its presence will not drastically alter the kinetics of the system. The experimental results agreed reasonably well with those predicted by a kinetic model of the chain reaction mechanism that is based on published rate constants extrapolated to pH 4. The results from experiments where tert-butyl alcohol was added supported the conclusion that the solution chain reaction mechanism controlled the decomposition kinetics of $\text{H}_2\text{O}_2$ and formic acid when Fe was introduced as dissolved Fe(III). In contrast, the solution chain reaction was not the dominant decomposition pathway of $\text{H}_2\text{O}_2$ and formic acid when ferrihydrite was the iron source. The heterogeneous system was modeled using four different hypothetical mechanisms,
and the one that produced the best fits to the data had a nonradical producing \( \text{H}_2\text{O}_2 \) loss pathway at the iron oxide surface, as well as insufficient amounts of dissolved Fe to effectively propagate the solution phase chain reaction.

In chapter 3, a model was created for predicting the pseudo-first-order oxidation rate coefficient of a dissolved organic compound \( (k_{\text{org}}) \) in mineral-catalyzed Fenton-like systems based on the factors that control \( V_{\text{OH}} \). This was the first time anyone has attempted to formulate a generally applicable approach for predicting \( k_{\text{org}} \) in mineral-catalyzed Fenton-like systems. \( ^{14}\text{C} \)-labeled formic acid was used to measure \( V_{\text{OH}} \) in pH 4 slurries of \( \text{H}_2\text{O}_2 \) and either synthesized ferrihydrite, goethite, or hematite or a natural iron oxide-coated quartzitic aquifer sand. \( V_{\text{OH}} \) was proportional to the product of the concentrations of surface area of the iron oxide and \( \text{H}_2\text{O}_2 \), although different solids produced \( \cdot\text{OH} \) at different rates. Using these results, a model was developed that predicts the degradation rate of an organic compound based on its initial concentration, its second order rate constant with \( \cdot\text{OH} \), the initial concentration of \( \text{H}_2\text{O}_2 \), the concentration and reaction rates of any other important \( \cdot\text{OH} \) sinks in the system, and the surface area, type and quantity of iron oxide. The model successfully predicted \( V_{\text{OH}} \) and \( k_{\text{org}} \) in the aquifer sand experiments and in a number of previously published works, but overpredicted \( V_{\text{OH}} \) and \( k_{\text{org}} \) from other studies.

The work to refine the model discussed in chapter 3 is presented in chapter 4. Three hypotheses that could explain why the model overpredicted \( V_{\text{OH}} \) and \( k_{\text{org}} \) in some instances were tested: (1) laboratory synthesized iron oxide was significantly different from those used in other studies, (2) electrostatic attraction between formate anions and the iron oxide surface must be accounted for, and (3) the compound being degraded or its oxidation products interfered with the generation of \( \cdot\text{OH} \). Experimental results indicated that the discrepancies between model and measured values for \( V_{\text{OH}} \) and \( k_{\text{org}} \) were mainly due to electrostatics. The concentration of formate anions in the double layer was greater than its bulk concentration because it was attracted to the positively charged iron oxide surface. Consequently, the bulk \( ^{14}\text{C} \)-labeled formic acid loss rate was faster than that for neutral compounds because more of the probe reacted with \( \cdot\text{OH} \), which is generated at the oxide surface, than it would have in the absence of electrostatic attraction. As time progressed, the decomposition rate of \( ^{14}\text{C} \)-labeled formic acid decreased, but the oxidation rates of the neutral compounds remained relatively constant. This was a result of the reduction of the positive charge on the iron oxide surface due to sorption of the oxidation products of the
organic compound that was being degraded. This conclusion was supported by the observation that the goethite particles coagulated during the experiment, which had never occurred in all of our previous experiments where the organic compound was not present.

Literature Cited
Chapter 2: Decomposition of Hydrogen Peroxide and Organic Compounds in the Presence of Dissolved Iron and Ferrihydrite

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Abstract

This work examines the contribution of solution phase reactions, especially those involving a chain reaction mechanism, to the decomposition of hydrogen peroxide (H_2O_2) and organic compounds in the presence of dissolved iron and ferrihydrite. In solutions at pH 4, where Fe was introduced as dissolved Fe(III), both H_2O_2 and ^14C-labeled formic acid decomposed at measurable rates that agreed reasonably well with those predicted by a kinetic model of the chain reaction mechanism, using published rate constants extrapolated to pH 4. The ratio of the formic acid and H_2O_2 decomposition rates, as well as the dramatic effect of tert-butyl alcohol on these rates, confirmed that a solution chain reaction mechanism involving •OH controlled the decomposition kinetics of both compounds. In the presence of ferrihydrite as the iron source, the ratio of the rate of formic acid decomposition to that of H_2O_2 decomposition was significantly lower than that observed in the presence of only dissolved Fe. Moreover, neither rate diminished drastically upon addition of tert-butyl alcohol, indicating that the solution phase chain reaction is not a dominant decomposition pathway of H_2O_2 and formic acid. Relative decomposition rates of formic acid and a second •OH probe, benzoic acid, were consistent with oxidation of these compounds by •OH. These observations can be reproduced by a kinetic model including (a) decomposition of H_2O_2 at the iron oxide surface, producing •OH with lower yield than the reaction sequence with dissolved Fe, and (b) low concentrations of dissolved Fe in the presence of ferrihydrite, preventing propagation of the solution phase chain reaction.

Introduction

Fenton’s reagent [Fe(II) + hydrogen peroxide] has been used to oxidize organic pollutants in many applications, from treatment of wastewaters to remediation of contaminated aquifers (1-4). These applications exploit the high reactivity of the hydroxyl radical (•OH) that is generated when hydrogen peroxide (H_2O_2) oxidizes Fe(II) to Fe(III) (the Fenton reaction). A disadvantage of using Fenton’s reagent as a treatment method is that the Fe(III) produced is only sparingly soluble, especially at circumneutral pH, so that high concentrations of ferric oxyhydroxide precipitates are generated when stoichiometric quantities of Fe(II) are used. An alternative is to use catalytic quantities of Fe and regenerate the Fe(II) needed for the Fenton reaction by continuously reducing Fe(III). In batch reactor treatment schemes, UV irradiation can be used to reduce Fe(III) (5-7). For treatment of soils, the slow reduction of dissolved Fe(III) by hydrogen peroxide itself has been found to be sufficient (8, 9).
In hydrogen peroxide solutions in which Fe(II) constitutes a minor fraction of the total dissolved Fe, the overall kinetics of hydrogen peroxide and contaminant decomposition may be controlled by a chain reaction (5, 10-12). In the absence of light, the chain initiation reaction is the reduction of Fe(III) by hydrogen peroxide (Table 2-1, reaction T1.1), producing two chain-propagating species Fe(II) and \( \text{HO}_2/\text{O}_2^- \) (superoxide radical and its conjugate acid) (10, 11). Fe(II) is quickly oxidized by another molecule of \( \text{H}_2\text{O}_2 \), and the chain is then propagated by reactions T1.3 and T1.4. Thus, reactions T1.2, T1.3, and T1.4 form a cycle that sustains itself by regenerating Fe(II) (Figure 2-1). Reactions that will terminate the chain include reactions T1.5 and T1.6. Reactions of \( \bullet \text{OH} \) with organic solutes or other solution constituents may propagate the chain if they produce \( \text{HO}_2/\text{O}_2^- \) or another species capable of reducing Fe(III) (e.g., \( \text{CO}_2^- \) and some organic radical products); otherwise, they act as termination steps (13). For the chain reaction to be significant (with chain lengths much greater than 1), either the concentration of \( \text{H}_2\text{O}_2 \) must be high enough so that it outcompetes other solution constituents for \( \bullet \text{OH} \) or the reactions of \( \bullet \text{OH} \) with the other solutes must be able to propagate the chain. In addition, there must be enough dissolved Fe(III) in solution to ensure that most of the \( \text{HO}_2/\text{O}_2^- \) reacts with it to form Fe(II) [either directly or via an intermediate, e.g., Cu(I) (14)].

Investigators have expanded the range of treatment applications of Fenton chemistry by demonstrating that an iron oxide/\( \text{H}_2\text{O}_2 \) system can effectively oxidize pollutants at pH values ranging from 3 to circumneutral (15-17). However, a consensus on the mechanism of this process does not exist. Proposed mechanisms for the decomposition of hydrogen peroxide generally include reactions analogous to reactions T1.1 and T1.2 taking place on the iron oxide surface (17-20). Many studies suggest that the oxidation of organic compounds takes place via dissolved \( \bullet \text{OH} \) released into solution when the surface analogue of reaction T1.2 occurs. Lin and Gurol (19) postulate that \( \bullet \text{OH} \) formed by a surface reaction would be too reactive to diffuse into the bulk solution. Therefore, they propose that their organic compound may be oxidized either in the sorbed state by near-surface \( \bullet \text{OH} \) or in solution by dissolved \( \bullet \text{OH} \) generated when Fe(II), formed by the surface versions of reactions T1.1 and T1.4, is first released into solution and then oxidized by \( \text{H}_2\text{O}_2 \) (21). Miller and Valentine (22) propose that \( \text{HO}_2/\text{O}_2^- \) diffuses into the bulk solution to generate \( \bullet \text{OH} \) by reacting with \( \text{H}_2\text{O}_2 \). Watts and co-workers invoke an additional surface species capable of oxidizing sorbed compounds at very high (>2%) concentrations of \( \text{H}_2\text{O}_2 \) (18). In summary, the relative contributions of surface and solution reactions to the
decomposition of H\textsubscript{2}O\textsubscript{2} and organic compounds in iron oxide/H\textsubscript{2}O\textsubscript{2} systems remain unclear.

Furthermore, the postulated release of \( \cdot \text{OH}, \text{HO}_2/\text{O}_2^- \) or Fe(II) from surface reactions into the dissolved phase implies that a chain reaction can take place in solution if enough dissolved Fe is present for the chain propagation steps, as is likely in acidic suspensions of iron oxide-containing particles. The chain reaction could even be the dominant mechanism for the decomposition of both hydrogen peroxide and organic compounds. The role of the iron oxide surface would then be restricted to just initiating the solution phase chain reaction. The ability of different solutes present in the solution to participate in chain propagation and termination reactions would become a crucial factor in predicting the overall rates of decomposition of H\textsubscript{2}O\textsubscript{2} and organic compounds. This possibility has not been considered by previous studies.

An important question is whether the chain reaction mechanism is possible at pH values greater than 3. A number of studies have examined the kinetics of H\textsubscript{2}O\textsubscript{2} decomposition in the Fe(III)/H\textsubscript{2}O\textsubscript{2} system in acidic solutions (10, 11, 23-25). More recently, De Laat and Gallard (12, 26) presented a detailed kinetic model of the chain reaction that successfully described the kinetics of both H\textsubscript{2}O\textsubscript{2} and atrazine decomposition under a wide range of conditions at pH values of 3 or less. While H\textsubscript{2}O\textsubscript{2} decomposition rates in the Fe(III)/H\textsubscript{2}O\textsubscript{2} system were observed to decrease dramatically at pH values greater than 3 (5, 12), these studies do not rule out the possibility of a solution phase chain reaction, especially if another initiation reaction besides T1.1 (for example, a surface reaction) can occur.

The objective of this work was to examine the importance of solution phase reactions, especially the Fe(III)-initiated chain reaction, in the decomposition of organic compounds and H\textsubscript{2}O\textsubscript{2}. Experiments at pH 4 with only dissolved Fe(III) added to H\textsubscript{2}O\textsubscript{2} solutions in the presence and absence of the chain terminator tert-butyl alcohol demonstrate that the chain reaction mechanism can still control the kinetics of H\textsubscript{2}O\textsubscript{2} and organic compound decomposition at this pH. Experiments in the presence of ferrihydrite reveal that a surface-initiated chain reaction was not significant in our experimental system. The decomposition kinetics of two organic compounds used as \( \cdot \text{OH} \) probes (formic and benzoic acids) indicate that release of dissolved \( \cdot \text{OH} \) is likely to be the main mechanism of oxidation of these compounds, but the yield of \( \cdot \text{OH} \) was lower than expected.
Materials and Methods

Materials

All reagents were reagent grade and were used without further purification except for 2-nitrophenyl hydrazine (NPH), which was recrystallized once from hot water. H₂O₂ (30%, unstabilized) was from Fluka. All solutions were prepared using 18 MΩ Milli-Q water from a Millipore system unless otherwise noted. NPH and 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide hydrochloride (EDC, protein sequencing grade) were stored frozen until use. Peroxidase (type II from horseradish) and N,N-diethyl-p-phenylenediamine (DPD) solutions were kept in the dark at 4 °C for no more than 2 weeks. Stock solutions of Fe(III) were made fresh daily by dissolving Fe(ClO₄)₂•9H₂O in Milli-Q water that was first acidified to pH < 1 with concentrated HClO₄.

All glassware and other containers used in this work were soaked in 1N HCl for more than 10 h and rinsed with Milli-Q water. The Teflon bottles in which the kinetic experiments were performed were soaked in concentrated nitric acid for more than 10 h and washed with Milli-Q water. Glassware used in the HPLC procedures were soaked in 10% HNO₃, rinsed with Milli-Q water, and then combusted in an oven at 450 °C for a minimum of 8 h. Samples for HPLC analysis were derivatized and stored in borosilicate glass vials with Teflon-lined caps. The pH measurements were made using an Orion model 420A benchtop pH meter calibrated against standard buffers. All spectrophotometric measurements were done on an HP 8453 diode array spectrophotometer. ¹⁴C activity was measured in a Beckman LS 6500 multipurpose scintillation counter. HPLC data were obtained from an HP 1050 series HPLC coupled to a diode array detector. A Spherisorb ODS-2 column (5 μm particle size, 25 cm length x 4.6 mm i.d.) was used for the HPLC separation.

Ferrihydrite was prepared from FeCl₃•6H₂O according to the method described by Wells et al. (27). FeCl₃•6H₂O was dissolved to make a 4 x 10⁻⁴ M solution (5 x 10⁻⁴ M for experiments with high ferrihydrite concentrations). Within 1 h, the solution was heated in a 90 °C water bath for 5 min and then quickly cooled back to room temperature. It was used within 2 days without further purification.

Analytical Techniques

Hydrogen peroxide was measured spectrophotometrically using the DPD method (28) as modified by Voelker and Sulzberger (29) to minimize interference by Fe(II) and Fe(III). Samples
were diluted so that the H$_2$O$_2$ concentration never exceeded 25 μM in the 1-cm path length cuvette.

The ferrozine method (30), as modified by Voelker and Sulzberger (29), was used to measure the concentration of dissolved and total (dissolved plus solid) iron. Dissolved iron was defined as iron that passed through a 0.02-μm filter (Anotop, aluminum oxide membrane, Whatman).

We used $^{14}$C-labeled formic acid as our main •OH probe for various reasons. The reactions of formate and formic acid with •OH are fast (Table 2-2) and are well-characterized (31):

$$\text{•OH} + \text{HCOO}^- \rightarrow \text{H}_2\text{O} \rightarrow \text{CO}_2^- + \text{O}_2^- + \text{HCOO}^- \rightarrow \text{CO}_2 + \text{O}_2^- \quad (1)$$

$$\text{•OH} + \text{HCOOH} \rightarrow \text{HCOO}^- + \text{H}_2\text{O} \rightarrow \text{CO}_2 + \text{O}_2^- + \text{H}^+ \rightarrow \text{CO}_2 + \text{O}_2^- \quad (2)$$

In an oxygenated environment, the carbon radicals create almost no side products besides CO$_2$ and O$_2^-$ because they react with oxygen at nearly diffusion-limited rates (13). This means that any reactions between the iron and the carbon radicals are insignificant. The superoxide produced in reaction 1 propagates the chain reaction (see Figure 2-1). Also, low concentrations (~100 nM) of H$^{14}$COOH can be used since our detection limit (three times the standard deviation of the typical background radiation level) is on the order of 1 nM. These probe properties allow us to investigate •OH formation without any danger of significantly perturbing the system, for example, by adding a chain termination reaction.

To measure the amount of $^{14}$C-labeled HCOOH/HCOO$^-$ in the reactor, a 4-mL aliquot was transferred into a 15-mL polystyrene conical tube. It was subsequently air sparged vigorously with house air using a gas dispersion tube with a fritted cylinder for 30 s or more to drive out the $^{14}$C-labeled carbon dioxide. A total of 1 mL of the aliquot was then mixed with 6 mL of scintillation fluid (ScintiSafe Econo 1, Fisher) in a 7-mL glass scintillation vial. The frit was washed with Milli-Q water and wiped dry with Kimwipes after each sparge. At the end of the day, it was soaked in 1 N HCl overnight to remove any traces of iron. The vials were kept at room temperature, and their $^{14}$C content was measured en masse within 1 day. The $^{14}$C content of an aliquot measured immediately after extraction did not differ from that measured 1 day later. Vigorous air sparging of aliquots for 5 min did not remove $^{14}$C-labeled HCOOH/HCOO$^-$ from an iron-free solution containing 1 mM H$_2$O$_2$ and 50 nM H$^{14}$COOH/H$^{14}$COO$^-$ at pH 3. We also
performed a control experiment to demonstrate that all of the $^{14}$CO$_2$ is removed by sparging: 5 mM each of Fe(NH$_4$)$_2$(SO$_4$)$_2$•6H$_2$O (99+%, Sigma-Aldrich) and H$_2$O$_2$ were added to a pH 3 solution containing 50 nM H$^{14}$COOH/H$^{14}$COO$^-$. Under these conditions, all of the H$^{14}$COOH/H$^{14}$COO$^-$ should be rapidly oxidized to $^{14}$CO$_2$ via the Fenton reaction. The $^{14}$C activity of this solution after sparging was indistinguishable from background radiation. To show that sorption of H$^{14}$COOH/H$^{14}$COO$^-$ to ferrihydrite was insignificant, a pH 4 solution containing no H$_2$O$_2$, 300 µM ferrihydrite, and 70 nM H$^{14}$COOH/H$^{14}$COO$^-$ was stirred for 3 days. There was no statistical difference in $^{14}$C content between samples that were filtered through a 0.02-µm filter and ones that were not. These controls are described in more detail elsewhere (32).

We also performed experiments in which benzoic acid and (nonlabeled) formic acid were used simultaneously as •OH probes. Benzoic acid is a well-characterized •OH-specific probe (33). The two probes were detected using HPLC (34). A total of 0.2 mL of a pyridine (HPLC grade)/HCl (99.999%) buffer (1.25:1.00, v/v), 0.2 mL of 0.3 M EDC, and 0.4 mL of 0.1 M NPH were added to 2 mL of reaction solution. After waiting 90 min or longer, 0.2 mL of 40% (w/v) KOH was added to neutralize the acid. Finally, the vials were put in a 70 °C water bath for 10 min to complete the derivatization. The products were kept frozen until analysis. A total of 0.5 mL of the derivatized sample was injected into a guard cartridge (Dionex IonPac NG1, 4 x 35 mm), which acted as a concentrator, followed by 1 mL of Milli-Q water to flush out the strong KOH. The elution gradient was as follows: water at pH 2.5 (A) and acetonitrile (UV spectrophotometric grade) (B); 0-2 min 10% B, 2-11 min 30% B, and 11-20 min 50% B, at a flow rate of 1.0 mL/min. The column was kept in a 35 °C water bath. The analytes were detected by absorbance at 400 nm. Control experiments showed benzoic acid sorption to ferrihydrite was insignificant.

**Kinetic Experiments**

All experiments were performed in 250-mL Teflon bottles covered completely with black electrical tape to exclude light. Milli-Q water was treated for approximately 45 min with ozone bubbled through a gas dispersion tube with a fritted cylinder. This was done to oxidize any trace impurities that could interfere with the chain reaction and cause, for example, lag periods before the decomposition of H$_2$O$_2$ accelerates (see Results) (32). The water was stirred for a couple of hours prior to adding NaOH to raise the pH above 9, which facilitates the decomposition of dissolved ozone (35). It was left stirring overnight to ensure that all of the ozone was removed.
and acidified to pH ~5 before use. Unless noted otherwise, 200 mL of solution contained 10 mM NaClO₄ (for ionic strength control) and 100 nM H¹⁴COOH/H¹⁴COO⁻. During the course of an experiment, the bottles were loosely capped, and the solutions were kept in a recirculating water bath set at 25 ± 0.5 °C while being stirred by a magnetic stirrer. Experiments lasted from 1 to 4 days. Pseudo-first-order decomposition rate coefficients for H¹⁴COOH/H¹⁴COO⁻ and H₂O₂, i.e., \( k_{HCOOH} \) and \( k_{H₂O₂} \), were obtained by performing nonlinear least squares regression fits to the concentration versus time data from each experiment using the built-in nonlinear curve-fitting routine in SigmaPlot (graphing software from SPSS Inc.).

### Dissolved Iron

The experimental solutions contained ~1 mM H₂O₂ and were adjusted to pH 4 with HClO₄ and/or NaOH. A small volume of an acidified Fe(III) stock solution (typically 20-200 μL to avoid changing the solution pH dramatically) was added to start the experiment. The pH of the solutions remained within ±0.2 unit of their initial pH for the entire experiment without further adjustments.

### Ferrihydrite

The setup is similar to above except that the experimental solutions were adjusted to pH 4 with HClO₄ and/or NaOH after the ferrihydrite was added. Care was taken to avoid making the solution too basic (pH > 6) during the adjustment. H₂O₂ was added at time zero. The solution pH remained steady (±0.2 unit of initial pH) throughout the experiment.

In two of the ferrihydrite experiments, H¹⁴COOH/H¹⁴COO was replaced with 50 μM nonlabeled formic acid; 15 μM benzoic acid was also added. Before collecting the aliquots that would be used for derivatization, the 3-mL syringe and 0.02-μm filter were prerinsed five times with 3 mL of Milli-Q water and once with 3 mL of the experimental solution to remove any residual organic acids. The samples were collected and frozen immediately until they could be derivatized in one batch.

### Kinetic Modeling

All of the modeling results were calculated with the computer program Acuchem (36). At first glance, the kinetic model that we used to investigate the Fenton system (Table 2-1) appears to differ significantly from the recent work of De Laat and Gallard (12), which describes a mechanism for the homogeneous decomposition of hydrogen peroxide by Fe(III) in aqueous solutions at pH < 3. In fact, they are the same with only cosmetic changes. Our model has fewer
reactions than the DeLaat and Gallard model because we have incorporated both equilibrium and rate expressions into composite rate constants instead of using individual rate constants for each species. We chose to use composite rate constants because they are the easiest way to include equilibrium in the Acuchem format and because they make it more apparent which reactions are important to the system at the pH of interest.

The concept of the composite rate constant is illustrated in the following example. HO$_2$ and O$_2^-$ (pK$_a$ = 4.8; 37) react with Fe$^{2+}$ at different rates to form Fe$^{III}$(HO$_2$)$^{2+}$ and are considered as two separate reactions in the De Laat and Gallard model:

$$\text{Fe}^{2+} + \text{HO}_2 \rightarrow \text{Fe}^{III} (\text{HO}_2)^{2+}$$  \hspace{1cm} (3)

$$\text{Fe}^{2+} + \text{O}_2^- + \text{H}^+ \rightarrow \text{Fe}^{III} (\text{HO}_2)^{2+}$$  \hspace{1cm} (4)

In our model (reaction T1.6), they are considered as a single reaction between Fe(II) and HO$_2$/O$_2^-$, whose rate constant (i.e., the composite rate constant) is a weighted sum of the two individual rate constants:

$$k_{T1.6} = \frac{[\text{HO}_2]}{[\text{HO}_2]_T} k_3 + \frac{[\text{O}_2^-]}{[\text{HO}_2]_T} k_4$$  \hspace{1cm} (5)

The weights are determined by the percentage of HO$_2$ and O$_2^-$ in solution, which is a function of pH. Other reactions including HO$_2$ and O$_2^-$ were treated in the same manner. Since Fe$^{III}$(HO$_2$)$^{2+}$ is not a significant species in our system (see below), reaction T1.6 in our model only shows Fe(III) and H$_2$O$_2$ as the products.

Reactions II.1-II.6b in De Laat and Gallard (12) can be expressed as our composite reaction T1.1 if the two Fe(III)-hydroperoxy complexes are not major H$_2$O$_2$ or Fe(III) species. Using the stability constants of the hydrolysis products of Fe$^{3+}$ from Morel and Hering (38) along with eq 15 and the equilibrium constants in Table 2-2 of De Laat and Gallard (12), we calculated that at pH 3 the Fe(III)-hydroperoxy complexes make up less than 5% of the total dissolved Fe(III) if [H$_2$O$_2$] is less than 0.11 M. [The stability constants in the De Laat and Gallard model were calculated for an ionic strength $I$ of 0.1 M. Instead of using those, we referred to literature values and corrected them for $I = 0.01$ M.] If additional hydroperoxy species do not become important at higher pH values (for which the De Laat and Gallard model was not calibrated), Fe(III)-hydroperoxy complexes are insignificant (<5%) to Fe speciation at [H$_2$O$_2$] < 0.088 M at pH 4. The complexes will make up even smaller fractions of the total H$_2$O$_2$ if [H$_2$O$_2$]$_T$ is greater than [Fe(III)]$_T$. Therefore, the assumption that Fe(III)-hydroperoxy complexes are not
significant is always valid under our experimental conditions.

Table 2-1 shows the rate constants, in composite form where appropriate, that were used in our model. All of these were derived from the values used in the De Laat and Gallard model except for the rate constants for two reactions. We used equations from Millero et al. (39) to calculate the rate constants for reaction T1.2 at pH values higher than 3. As for the composite rate constants for reaction T1.1, we computed them based on the literature values of the hydrolysis equilibrium constants of Fe(III) (38), corrected for $I = 0.01$ M, along with the other rate constants in the De Laat and Gallard model. We did not include ionic strength corrections for rate constants because, at our low ionic strength, these should be negligible for reactions of monovalent species with each other or of divalent species with neutral species. These two categories include all of the reactions in Table 2-1, since in reaction T1.4, the main species reacting are $\text{Fe(OH)}_2^+$ with $\text{O}_2^-$, while in reaction T1.6, the main species are $\text{Fe}^{2+}$ and $\text{HO}_2$. As we used formic acid as a hydroxyl radical probe, its oxidation reaction (reaction T1.10) was also included in the model calculations. This rate constant was calculated by averaging the literature values of rate constants obtained at pH 2-5, shown in Table 2-2. Using the recommended values from Buxton of $1.3 \times 10^8$ and $3.2 \times 10^9$ M$^{-1}$ s$^{-1}$ for $\text{HCOOH}$ and $\text{HCOO}^-$ gives a nearly identical rate constant of $2.1 \times 10^9$ M$^{-1}$ s$^{-1}$ at pH 4. The reaction between tert-butyl alcohol and $\cdot \text{OH}$ was added when we explored its effects on the Fenton system. We used $6.0 \times 10^8$ M$^{-1}$ s$^{-1}$, the recommended value from Buxton et al. (40), instead of averaging because there were not many independent measurements of this rate constant.

The direct reaction of $\text{HO}_2/\text{O}_2^-$ with $\text{H}_2\text{O}_2$, suggested by Miller and Valentine (22) to play a role at circumneutral pH, was not included in our models because, at all of the Fe(III) and $\text{H}_2\text{O}_2$ concentrations considered here, the rate of the direct reaction is negligible as compared to the Fe-catalyzed rate (reaction T.1.4 followed by T.1.2). Furthermore, it remains unclear whether this reaction can proceed at all in the absence of metal catalysis (41).

Results

Reactions of Dissolved Fe at pH 4

The purpose of our first set of experiments was to show that chain reaction kinetics can control the rate of hydrogen peroxide and formic acid decomposition and $\cdot \text{OH}$ formation at pH 4 with only dissolved Fe(III) present. The pseudo-first-order decomposition rate coefficients for $\text{H}^{14}\text{COOH}/\text{H}^{14}\text{COO}^-$ and $\text{H}_2\text{O}_2$ ($k_{\text{HCOOH}}$ and $k_{\text{H}_2\text{O}_2}$) were measurable but low (Figure 2-2 and
The total iron concentrations in all of our experiments were slightly above the theoretical saturation limit of amorphous Fe(OH)$_3$(s), which we calculated to be 0.42 $\mu$M at pH 4 (38). In theory, then, solubility should have controlled the amount of dissolved Fe, and it should have been equal to 0.42 $\mu$M in all of the experiments. We extrapolated the De Laat and Gallard model to pH 4 as explained in the Methods section (Table 2-1), then ran it in Acuchem with dissolved Fe equal to 0.42 $\mu$M and particulate Fe assumed to be unreactive. The resulting model calculations were well-described by pseudo-first-order kinetics with $k_{\text{HCOOH}}$ equal to 1.41 day$^{-1}$ and $k_{\text{H}_2\text{O}_2}$ equal to 0.043 day$^{-1}$. Our measured values of $k_{\text{HCOOH}}$ and $k_{\text{H}_2\text{O}_2}$, 0.68 ± 0.13 and 0.030 ± 0.015 day$^{-1}$, respectively (Table 2-3), agree reasonably well with those of the model.

The scatter among the data points in these experiments may be due to two factors. First, if small amounts of chain terminating $\cdot$OH scavengers were present in the Milli-Q water, they could decrease $k_{\text{H}_2\text{O}_2}$ and $k_{\text{HCOOH}}$ by prematurely terminating the chain reaction. The effect of such scavengers was clearly visible in early phases of this work, where we often observed long lag periods before H$_2$O$_2$ decomposition ensued (32). Pretreating the Milli-Q water with ozone decreased the occurrence of these dramatic lag periods, but residual concentrations of $\cdot$OH scavenging contaminants from, for example, the reagent solutions could still be present. Short lag periods (0.5-1 day) remained in three of the seven experiments shown in Table 2-3. In these cases, initial time points were excluded from the data used to obtain the experimental $k_{\text{HCOOH}}$ and $k_{\text{H}_2\text{O}_2}$.

A second possible reason for the scatter in the data is iron speciation. Since we worked at Fe concentrations slightly above saturation, colloids and microscopic-sized solids could have formed throughout each experiment. On the basis of our work with ferrihydrite, colloidal and precipitated iron should not have a large effect on $k_{\text{H}_2\text{O}_2}$ and $k_{\text{HCOOH}}$ at total Fe concentrations of a few micromolar (see Figure 2-3a,b). However, since the dynamics of colloid formation and precipitation could have changed from experiment to experiment, the concentration of dissolved Fe may also have varied, resulting in scatter in the data. (The model calculations predict a strong dependence of $k_{\text{H}_2\text{O}_2}$ and $k_{\text{HCOOH}}$ on the concentration of dissolved Fe.) We did not observe a relationship between observed values of $k_{\text{H}_2\text{O}_2}$ and $k_{\text{HCOOH}}$ and measured concentrations of dissolved Fe (Table 2-3), but this might merely mean that the concentration of Fe that passed
through a 0.02-µm filter did not correspond to the concentration of reactive dissolved Fe.

If most of the $H^{14}COOH/H^{14}COO^- \text{ and } H_2O_2$ were consumed by the chain reaction, then we would expect their decomposition rates to drop drastically when a chain terminator is introduced into the system. In one experiment, we added 100 µM tert-butyl alcohol, which reacts with •OH to form an organic radical that does not participate in the chain reaction (13). The pseudo-first-order decomposition rate coefficients for $H^{14}COOH/H^{14}COO^-$ and $H_2O_2$ that we measured in the presence of 100 µM tert-butyl alcohol were $0.0095 \pm 0.0070$ and $0.0031 \pm 0.0065 \text{ day}^{-1}$, respectively, a factor of 102 and 14 less than the rate coefficients measured in the absence of tert-butyl alcohol. In contrast, this amount of tert-butyl alcohol decreases the fraction of •OH radical reacting with $H_2O_2$ and $H^{14}COOH/H^{14}COO^-$ from 0.99 and 0.0066 to 0.35 and 0.0024, respectively (see eq 7 below). Clearly, the competition for •OH radical by tert-butyl alcohol is not sufficient to explain the observed decrease in $H_2O_2$ and $H^{14}COOH/H^{14}COO^-$ decomposition rates, so the main effect of tert-butyl alcohol must be to decrease the rate of •OH production by acting as an efficient chain terminator. This effect is reproduced by a kinetic model calculation including 100 µM tert-butyl alcohol and, as before, 0.42 µM dissolved Fe, which yields $0.0034 \text{ day}^{-1}$ for $k_{HCOOH}$ and $0.00048 \text{ day}^{-1}$ for $k_{H_2O_2}$, a decrease by a factor of 415 and 90 as compared to the model’s prediction in the absence of tert-butyl alcohol. This calculation neglects the possibility of reactions of Fe with organic radical products of •OH oxidation of tert-butyl alcohol, which could both propagate the chain by reducing Fe(III) or terminate it by oxidizing Fe(II). However, since our experimental result shows that tert-butyl alcohol acts as an efficient chain terminator, we can assume that chain propagation by organic radicals is insignificant.

While the effect of tert-butyl alcohol is a good indicator of a chain reaction mechanism involving •OH, we used another tool to provide evidence that the decomposition of formic acid is caused by reaction with •OH. This tool is based on work describing the oxidation of organic contaminants by hydroxyl radicals produced during ozone decomposition (35). In a well-mixed aqueous solution, the ratio of the decomposition rate of a chemical species (M) versus that of hydrogen peroxide can be defined as the product of the yield of hydroxyl radicals from hydrogen peroxide (Y) and the fraction of hydroxyl radicals that react with the various contaminants (F):
\[
\frac{\text{d}[M]}{\text{d}[H_2O_2]} = \left( \frac{\text{moles of } \cdot \text{OH produced}}{\text{moles of } H_2O_2 \text{ consumed}} \right) \times \left( \frac{\text{moles of } M \text{ consumed}}{\text{moles of } \cdot \text{OH produced}} \right) = YF \quad (6)
\]

where eq 6 is true if \( \cdot \text{OH} \) is the only sink of \( M \). The yield is dependent on the reaction conditions and is equal to 0.5 for the chain reaction described by reactions T1.2-T1.4. If the chain length is sufficiently long, then the initiation and termination steps will not significantly affect this number because they occur once, whereas the propagation steps happen many times. \( F \) can be computed from the concentrations of the chemical species in solution and their second order reaction rate constants of reaction with \( \cdot \text{OH} \) (\( k_{\text{OH,M}} \)):

\[
F = \frac{k_{\text{OH,M}}[M]}{k_{\text{OH,M}}[M] + k_{\text{OH,H_2O_2}}[H_2O_2] + \sum k_{\text{OH},i}[S_i]} \quad (7)
\]

where \( S \) is all other \( \cdot \text{OH} \) sinks besides hydrogen peroxide and \( M \), e.g., tert-butyl alcohol.

In our experiments, \( H^{14}\text{COOH}/H^{14}\text{COO}^- \) is the chemical species of interest (\( M \)), and the largest term in the denominator of eq 7 is \( k_{\text{OH,H_2O_2}}[H_2O_2] \) because \( k_{\text{OH,HCOOH}}[HCOOH]_T \) (where \( [HCOOH]_T = [HCOOH] + [HCOO^-] \), so the rate constant is the composite rate constant) is very small and we have removed all other \( \cdot \text{OH} \) sinks. (This was not true for the experiments that included tert-butyl alcohol, but those were a small subset of this work.) Thus, we can rewrite eq 6 as

\[
\frac{\text{d}[HCOOH]_T}{\text{d}[H_2O_2]} = Y \frac{k_{\text{OH,HCOOH}}[HCOOH]_T}{k_{\text{OH,H_2O_2}}[H_2O_2]}, \quad (8)
\]

which, when integrated, gives

\[
\frac{[HCOOH]_T}{[HCOOH]_0} = \left( \frac{[H_2O_2]_0}{[H_2O_2]} \right)^\gamma \quad (9)
\]

where

\[
\gamma = Y \frac{k_{\text{OH,HCOOH}}}{k_{\text{OH,H_2O_2}}} \quad (10)
\]

If the decomposition of \( HCOOH/HCOO^- \) and \( H_2O_2 \) follow pseudo-first-order kinetics, \( \gamma \) is also equivalent to the ratio of the observed pseudo-first-order decomposition rates of \( HCOOH/HCOO^- \) over that of \( H_2O_2 \).

The \( \gamma \) values from our experiments average to a value of 22.3 ± 9.9 (Table 2-3) as compared to a value of 33 obtained from the De Laat and Gallard model. Both fall within the
uncertainty of the expected value of $\gamma$ of $43 \pm 27$, which was computed using eq 10, the average rate constants from Table 2-2, and a value of 0.5 for $Y$. The discrepancy between the model value and the expected value arises from the fact that our model used DeLaat and Gallard’s value for $k_{OH,H_2O_2}$ instead of the average value in Table 2-2. The large uncertainty in the expected $\gamma$ is mainly attributable to the uncertainty of $k_{OH,H_2O_2}$. The scatter in the measured values of $\gamma$ is much less than the scatter in the measured values of $k_{H_2O_2}$ and $k_{HCOOH}$; this is consistent with the proposed causes of scatter in the rate coefficients because they would affect both decomposition rates equally.

In summary, our experimental results in solutions containing mostly dissolved Fe show that the •OH chain reaction can proceed at pH 4 and dictate the rates at which $H_2O_2$ and $H^{14}COOH/H^{14}COO^-$ decompose in Fe(III)/$H_2O_2$ systems. Furthermore, the model of the chain reaction proposed by De Laat and Gallard, extrapolated to pH 4 with no additional adjustable parameters, produces reasonable approximations of the behavior of this system in the presence and absence of tert-butyl alcohol.

Reactions in the Presence of Ferrihydrite at pH 4

After having established that the solution phase chain reaction occurs at pH 4, we conducted experiments to determine whether the prime effect of iron oxides could be to initiate the solution chain reaction. The difference in this set of experiments from the previous one was that the source of iron was laboratory-synthesized ferrihydrite instead of dissolved Fe(III).

Some initial experiments were performed with 200 $\mu$M ferrihydrite at pH 3. At this pH, some of the ferrihydrite ($\sim 6$ $\mu$M) dissolved rapidly and the observed pseudo-first-order decomposition rate coefficients of both $H_2O_2$ and formic acid were comparable to those expected from the measured levels of dissolved Fe(III), indicating that the effect of ferrihydrite was negligible (except as the source of the dissolved Fe). We therefore worked at pH 4.

The decomposition rates of both $H_2O_2$ and $H^{14}COOH/H^{14}COO^-$ rose as we increased the ferrihydrite concentration (Figure 2-3a,b), with $k_{H_2O_2}$ increasing more steeply than $k_{HCOOH}$ and resulting in a decrease in $\gamma$ with increasing ferrihydrite concentrations (Figure 2-4). Addition of 10 or 100 $\mu$M tert-butyl alcohol did not significantly affect $k_{H_2O_2}$ and only slightly decreased $k_{HCOOH}$. Scatter in the data is most likely due to variations in the reactivity of different batches of ferrihydrite. The amount of dissolved iron released from the ferrihydrite may also have varied.
Our attempts to measure dissolved Fe in these experiments were inconclusive. We observed a lot of variation in measured values of Fe passing through a 0.02-μm filter, but as before, no clear relationship between measured filterable Fe and reaction rate coefficients was observed. About a third of the measurements were less than the detection limit of 0.03 μM; another third were between 0.03 and 1 μM; and the rest were more than 1 μM, often dramatically greater (Appendix A). The abnormally high filterable Fe measurements are likely due to partial breakage of the filter material from high pressures during the filtering process, and it is not clear to what extent the same artifact affected the rest of the measurements. Blank values were consistently low, indicating that contamination was not an issue in these measurements.

To determine the extent to which different simplified mechanisms for the decomposition of H\textsubscript{2}O\textsubscript{2} and HCOOH can explain these experimental results and especially to examine the importance of the solution phase chain reaction in this system, we used a qualitative kinetic model combining both solution phase and surface reactions. Our model of solution phase reactions is identical to the DeLaat and Gallard model discussed in the previous section. The only addition is two or three reactions describing the interactions of H\textsubscript{2}O\textsubscript{2} with the surface of the ferrihydrite. It is important to note that modeling of this kind cannot validate a certain mechanism; our intention is merely to be able to rule out mechanisms inconsistent with the data.

To model the possibility of a surface initiation of the solution chain reaction, we added only two heterogeneous reactions, analogous to reactions T1.1 and T1.2, to the solution phase reactions used for the model of the homogeneous system:

\begin{align}
\equiv\text{Fe}^{III} + \text{H}_2\text{O}_2 &\rightarrow \equiv\text{Fe}^{II} + \text{HO}_2 + \text{H}^+ \quad (11) \\
\equiv\text{Fe}^{II} + \text{H}_2\text{O}_2 &\rightarrow \equiv\text{Fe}^{III} + \cdot\text{OH} + \text{OH}^- \quad (12)
\end{align}

We did not include surface •OH and HO\textsubscript{2} species in this model and instead assumed that the radicals would diffuse rapidly into the bulk solution where they would react primarily with H\textsubscript{2}O\textsubscript{2} and Fe(III), respectively, to initiate the chain reaction (reactions T1.3 and T1.4). Unknown parameters in this model are the rate constants of reactions 11 and 12 and the concentration of total dissolved Fe(III). Reaction 12 was assumed to be fast and an arbitrarily large rate constant; 1 x 10\textsuperscript{5} M\textsuperscript{-1} s\textsuperscript{-1} was chosen. The total dissolved Fe(III) concentration (assumed to be invariant with ferrihydrite concentration) [65 nM Fe(III)] and the rate constant of reaction 11 (1 x 10\textsuperscript{-4} M\textsuperscript{-1} s\textsuperscript{-1}) were chosen to produce \(k_{\text{HCOOH}}\) values that lie within the uncertainty of the data at ferrihydrite concentrations of less than 50 μM and at 190 μM.
The model calculations show that if the decomposition rates of H$_2$O$_2$ and H$^{14}$COOH/H$^{14}$COO$^-$ were both controlled by a surface-initiated solution chain reaction, we would expect to observe three effects (model A in Figures 2-4 and 2-5a,b). First, increasing the amount of ferrihydrite would increase the decomposition rates of both compounds by increasing the rate of the initiation reaction. Second, $\gamma$ would be equal to the value observed in the presence of only dissolved Fe, independent of the ferrihydrite concentration. Third, addition of tert-butyl alcohol as a chain terminator would drastically decrease both decomposition rates. On the basis of the observed decrease in $\gamma$ with increasing ferrihydrite along with the lack of effect of tert-butyl alcohol, we conclude that the chain reaction is not the major mechanism controlling the decomposition of H$_2$O$_2$ and H$^{14}$COOH/H$^{14}$COO$^-$ in this system.

The decrease in $\gamma$ with increasing ferrihydrite concentration probably indicates that some of the H$_2$O$_2$ disproportionates to O$_2$ and H$_2$O at the iron oxide surface without producing dissolved radicals that can initiate a solution chain reaction. This scenario is depicted as “model B” in Figures 2-4 and 2-5a,b, which extends model A by adding a reaction through which the iron oxide surface acts as a catalyst:

$$\text{Fe}^{III} + H_2O_2 \rightarrow \text{Fe}^{III} + \text{non-reactive products}$$  \hspace{1cm} (13)

Model B uses the same parameters as model A, and a rate constant of $7 \times 10^{-3} \text{ M}^{-1} \text{ s}^{-1}$ for reaction 13 was chosen to produce good agreement of modeled and measured $k_{H_2O_2}$ values at 190 $\mu$M ferrihydrite. Model B generated results that behaved qualitatively similar to our observations in the absence of tert-butyl alcohol. However, it was unable to adequately reproduce H$^{14}$COOH/H$^{14}$COO$^-$ decomposition rates when tert-butyl alcohol was included, because in the model, the rate of decomposition of H$^{14}$COOH/H$^{14}$COO$^-$ was still controlled by the chain reaction.

One possible explanation for the lack of effect of tert-butyl alcohol is that there was not enough dissolved Fe(III) present to sustain a chain reaction. This scenario is illustrated by “model C” in Figures 2-4 and 2-5c,d, which is identical to model B except that we chose a lower concentration of total dissolved Fe(III), 15 nM, and adjusted the rate constants of reactions 11 and 13 to values of $7 \times 10^{-4}$ and $8 \times 10^{-3} \text{ M}^{-1} \text{ s}^{-1}$, respectively, to produce values of both $k_{H_2O_2}$ and $k_{HCOOH}$ falling within the range of uncertainty of the observed values at ferrihydrite concentrations of less than 50 and at 190 $\mu$M. A fourth model (model D in Figures 2-4 and 2-
model D is a duplicate of model A except that the amount of total dissolved Fe(III) is 15 nM and the rate constant of reaction 11 is $8 \times 10^{-4}$ M$^{-1}$ s$^{-1}$.

An alternative explanation for the low $\gamma$ values and the small effect of tert-butyl alcohol we observed is that another oxidant besides •OH is responsible for the oxidation of formic acid. We examined this idea by adding 50 µM formic acid and 15 µM benzoic acid simultaneously as probe compounds to solutions containing 190 µM ferrihydrite and 1 mM H$_2$O$_2$. The ratio of pseudo-first-order decomposition rate coefficients of benzoic acid and formic acid that we measured were in good agreement with the ratio calculated from the literature rate constants for •OH with formic and benzoic acids (Table 2-4). This suggests that •OH was the main oxidant in our system.

If the concentration of Fe(III) in the ferrihydrite/H$_2$O$_2$ system is too low to sustain a chain reaction despite release of chain-propagating radicals by surface reactions, then one reason for this may be slow dissolution of ferrihydrite at pH 4. To test whether adding some dissolved Fe(III) to this system would increase the importance of the chain reaction, we compared $k_{H_2O_2}$ and $k_{HCOOH}$ in solutions with 1 mM H$_2$O$_2$ and 40 or 180 µM ferrihydrite with and without an addition of 1.5 µM dissolved Fe(III) at time zero. The additional dissolved Fe(III) either had no effect on the pseudo-first-order rate constants (40 µM ferrihydrite) or increased them by less than a factor of 2 (180 µM ferrihydrite) (data not shown).

In summary, comparing our qualitative model calculations to experimental results shows that the decomposition rates of neither H$_2$O$_2$ nor HCOOH are controlled by a surface initiated chain reaction and that the yield of •OH from H$_2$O$_2$ was much lower than in the presence of dissolved Fe. It should be emphasized that the intention of our model was only to illustrate key characteristics of different proposed mechanisms that could describe the heterogeneous system. While the patterns we discussed did not depend on the values of the rate constants chosen, different sets of fitting parameters (rate constants of reactions 11-13 and total dissolved Fe concentrations) may produce similar or better fits to the data. We did not explore this issue further since our intent was only to constrain the mechanistic possibilities using reasonable values for these parameters.
Discussion

Importance of the Chain Reaction at pH > 3

Our work shows that the chain reaction controls the decomposition rates of both H$_2$O$_2$ and formic acid in an aqueous solution containing only ~0.4 μM dissolved Fe at pH 4. The behavior of this system is described reasonably well by De Laat and Gallard’s kinetic model of the chain reaction mechanism, which is based on experiments conducted at pH 3 or less. The only reaction in Table 2-1 for which the pH dependence of the effective rate constant is not well understood is the initiation reaction, T1.1. At higher pH, other Fe(III)-hydroperoxy complexes [for example, Fe(OH)$_2$(HO$_2$)] could become important contributors to this effective rate constant. Our results show that such additional complexes do not need to be invoked in a kinetic model for pH 4. Extrapolations of the model to higher pH values, still assuming that no new Fe(III)-hydroperoxy complexes become important, predict that if the concentration of dissolved Fe(III) is limited by the solubility of ferrihydrite ($K_{sp} = 10^{-38.8}$; 38), the decomposition of H$_2$O$_2$ and organic compounds will be very slow at pH values higher than 4 (Table 2-5). However, even at pH 5, chain lengths in the hundreds are predicted by the model, where the chain length is equal to the propagation rate, $0.5 \times k_{H_2O_2}[H_2O_2]$, divided by the initiation rate, $k_{T1.1}[Fe(III)][H_2O_2]$ (the factor of 0.5 in the propagation rate accounts for the 2 mol of H$_2$O$_2$ consumed per cycle, see Figure 2-1) (42). The model calculations therefore show that sufficient dissolved Fe can be present at pH 3-5 to effectively propagate the chain reaction, making the chain reaction a potentially important pathway for the decomposition of hydrogen peroxide and organic compounds in an Fe(III)/H$_2$O$_2$ system, especially if it is initiated by a different reaction than T1.1 (for example, a photochemical reaction). However, the modeling result above pH 4 must be treated with caution since the kinetic model does not account for the greater potential of trace species to interfere with the chain reaction as the concentrations of dissolved Fe decrease. The model also does not account for the large variability in the solubility product of ferrihydrite (-log $K_{sp} = 37.0-39.4$; 43), or other potential influences on solubility such as the Kelvin effect (44) (Appendix A).

Mechanisms of Decomposition of Hydrogen Peroxide and Organic Compounds in Iron Oxide/H$_2$O$_2$ Systems

While most previous works assumed that •OH is the intermediate primarily responsible for the decomposition of organic compounds in iron oxide/H$_2$O$_2$ systems, few have performed
experiments that could invalidate this hypothesis. Miller and Valentine (22) showed that the addition of 100 mM butanol dramatically decreased the total decomposition of 0.08 mM quinoline (or 0.11 mM phenol) in a solution containing 58 mM H₂O₂ and 0.2 g/mL of sterilized sand. They attributed this to the competition between butanol and quinoline or phenol for dissolved •OH, but they did not attempt a quantitative comparison of the competition kinetics. A recent study (20) used a highly selective spin-trap technique in conjunction with competition kinetics to show that •OH is produced upon the decomposition of H₂O₂ in the presence of Fe-containing soil samples but did not relate the •OH production rate to the rate of decomposition of organic compounds. Our results show that the decomposition kinetics of two nonsorbing probes having very different chemical structures are consistent with •OH being the only important oxidant.

If dissolved •OH is released from the surface reaction, it is somewhat surprising that no chain reaction occurs in our ferrihydrite/H₂O₂ systems since some Fe must have dissolved from the ferrihydrite. The most plausible explanation for this observation is that dissolved Fe is too low for the chain reaction to take place. We intended to constrain the dissolved Fe value by measuring 0.02-µm filterable Fe in our experiments, but our measurements did not yield reliable results. One-third of our samples were lower than the detection limit of 0.03 µM, suggesting that dissolved Fe was lower during the experiments with ferrihydrite than during those without. This is reasonable since aging the ferrihydrite could have decreased its solubility. The lack of effect of adding dissolved Fe to the ferrihydrite systems could be due to rapid removal of the dissolved Fe by precipitation of additional ferrihydrite.

Despite the lack of dependable dissolved Fe data, we were able to use our modeling results to examine at what range of dissolved Fe concentrations the chain reaction would not be important in the heterogeneous system. This is illustrated by Models C and D; they both have a dissolved Fe(III) concentration of 15 nM, and their predicted k_HCOOH was barely affected by the presence of tert-butyl alcohol. In contrast, Models A and B showed that a dissolved Fe(III) concentration of 65 nM could sustain a chain reaction because their predicted k_HCOOH was greatly decreased by tert-butyl alcohol. As our model has other fitting parameters, it is possible that a different set of values would show the chain reaction to be unimportant at even higher dissolved Fe concentrations.

The decrease in γ values observed with increasing ferrihydrite indicates a sink of H₂O₂ at
the iron oxide surface other than the radical-producing reactions 11 and 12. In other words, assuming that •OH is the oxidizing species, the yield of •OH per H₂O₂ consumed, as defined by eq 6, is lower than the value of 0.5 that is expected if either the solution chain reaction occurs or, in the absence of a chain reaction, if the main reaction sequence is 11 (or T1.1), 12 (or T1.2), T1.3, and T.1.7. In the latter case, the yield of •OH is 0.5 because 3 mol of H₂O₂ are consumed and 1 mol each of H₂O₂ and •OH are produced by this reaction sequence; hence, the predicted γ is the same as when the chain reaction occurs (this is illustrated by the overlap between the lines for models A and D in Figure 2-4). Previous studies [except the direct measurements in Petigara and Blough (20)] have not attempted to calculate a yield of •OH per H₂O₂ consumed, as defined here. Furthermore, most studies have been performed either using natural soil or aquifer solid samples instead of pure mineral phases (e.g., refs 15 and 22) where H₂O₂ may be consumed by reactions with solids other than iron oxide, e.g., manganese oxide (22). Other studies used pure mineral phases but examined systems where not all of the organic compound was dissolved and only the decrease of total concentration over time was measured (e.g., ref 18), making calculation of the observed γ impossible.

Thus, we were able to calculate values of the observed γ from data presented in only two other studies. The experiments of Valentine and Wang (17) examined quinoline loss at pH 7.0 in the presence of three different iron oxide phases. We estimated quinoline and H₂O₂ decomposition rates from Figure 2-4 of their work, using only the data before H₂O₂ was replenished. Since they observed no detectable loss of quinoline in the ferrihydrite solution, its γ was approximately zero. γ for semicrystalline iron oxide was ~0.1, and for goethite it was ~0.3. Using Miller and Valentine’s (22) estimate of the second-order rate constant of •OH with quinoline, 9.28 x 10⁹ M⁻¹ s⁻¹ (no direct measurements of this rate constant have been published), we calculate an expected γ of 140. In the other study, Gurov et al. (45) used goethite to catalyze the decomposition of hydrogen peroxide and n-chlorobutane (BuCl) at pH 7.5. Calculation of γ from their data is complicated by the fact that they observed significant sorptive loss of BuCl (~35%) from their solutions. We attempted to account for this by neglecting the first hour of the data in their Figure 2-3, when sorptive losses were fast, and using the remainder to estimate the rate of loss of BuCl due to oxidation. The decomposition rate of H₂O₂ was computed from their eq 2. The values of γ calculated from their experiments with 44, 75, and 170 mg/L H₂O₂ are ~0.1, ~0.2, and ~2, in contrast to the expected γ of 45, using 3 x 10⁹ M⁻¹ s⁻¹ as the second-order
rate constant of •OH with BuCl (46). In both of these studies, the differences between the expected and the observed γ values were much greater than in ours. H2O2 concentration and the type of iron oxide seem to influence the yield of •OH in these systems, but clearly more studies need to be done to better understand the large variability in this parameter.

While kinetic arguments illustrated by the model calculations show that a sink of H2O2 other than reactions 11 and 12 is needed to explain the observed decrease in γ with increasing ferrihydrite concentration, we cannot distinguish among different mechanisms whose overall effect could be represented by reaction 13. The possibility of a two-electron transfer process has been suggested both for iron oxides and manganese oxides (20). Loss of •OH by reaction with reduced iron sites at the iron oxide surface has also been suggested (22). However, for the latter mechanism to affect γ, this reaction would have to become the major sink reaction of •OH. Since we expect surface Fe(II) to be a short-lived intermediate, and hence present in low concentrations, this appears implausible.

Most of our conclusions still hold if nearly all of the •OH formed at the surface of the iron oxide reacts away in the stagnant surface film before diffusing into the bulk solution, as suggested by Lin and Gurol (19). Since our probe compounds and hydrogen peroxide decompose on time scales that are much too long to be controlled by their diffusion to the surface, their concentration in the surface film must be approximately equal to their concentration in the bulk solution. Therefore, the fraction of •OH that reacts with H2O2 and the probe compounds should still be the same as that of the bulk solution, so the conclusions we drew from the observed values of γ and the ratio of decomposition rates of the two probes remain unchanged. Furthermore, a chain reaction in the bulk solution should still be initiated by the surface processes, since the rate constants in Table 2-1 imply that HO2/O2− and Fe(II) are sufficiently long-lived to diffuse out of the stagnant surface film. However, while reactions of bulk solution radicals with iron oxide surfaces cannot be important due to collision frequency limitations, such reactions are plausible for radicals formed in the surface film. Rapid loss of HO2/O2− to a surface reaction is therefore a possible alternative explanation for the lack of a surface initiated solution chain reaction. Loss of •OH at the surface could explain the low apparent yield of •OH from H2O2 (low γ values), although the reaction would have to be with a more abundant surface species than surface Fe(II), such as sorbed hydrogen peroxide.

Distinguishing among these different possible mechanisms for organic matter and
hydrogen peroxide decomposition is important for optimizing water and soil treatment processes based on these reactions. We show that chain reaction mechanisms can play a role in these processes even at pH values above 3, implying that small concentrations of chain-terminating species could have dramatic effects on the overall decomposition kinetics. The yield of •OH from H₂O₂ appears to vary over several orders of magnitude; to achieve proper dosing of the hydrogen peroxide, we need to understand what controls these variations. Further work is also needed to determine whether organic compounds are oxidized by •OH or other oxidants in the bulk solution or near iron oxide surfaces, all of which are critical for optimizing the effectiveness of hydrogen peroxide-based treatment processes in more complicated systems, such as soils in which contaminants are mostly present as sorbed species.

Acknowledgments

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Table 2-1. Mechanism of the Fe(III)-initiated chain reaction. The rate constants are listed in M$^{-1}$ s$^{-1}$.

<table>
<thead>
<tr>
<th>Reaction</th>
<th>pH 3</th>
<th>pH 4</th>
<th>pH 5</th>
<th>Reaction</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fe(III) + H$_2$O$_2$ $\rightarrow$ Fe(II) + HO$_2$/O$_2^-$ + H$^+$</td>
<td>2.0 x 10$^{-3}$</td>
<td>2.5 x 10$^{-3}$</td>
<td>2.6 x 10$^{-3}$</td>
<td>(T1.1)</td>
</tr>
<tr>
<td>Fe(II) + H$_2$O$_2$ $\rightarrow$ Fe(III) + •OH + OH$^-$</td>
<td>63</td>
<td>1.2 x 10$^2$</td>
<td>5.7 x 10$^2$</td>
<td>(T1.2)</td>
</tr>
<tr>
<td>H$_2$O$_2$ + •OH $\rightarrow$ HO$_2$/O$_2^-$ + H$_2$O</td>
<td>3.3 x 10$^7$</td>
<td>3.3 x 10$^7$</td>
<td>3.3 x 10$^7$</td>
<td>(T1.3)</td>
</tr>
<tr>
<td>Fe(III) + HO$_2$/O$_2^-$ $\rightarrow$ Fe(II) + O$_2$ + H$^+$</td>
<td>7.8 x 10$^5$</td>
<td>6.8 x 10$^6$</td>
<td>3.1 x 10$^7$</td>
<td>(T1.4)</td>
</tr>
<tr>
<td>Fe(II) + •OH $\rightarrow$ Fe(III) + OH$^-$</td>
<td>3.2 x 10$^8$</td>
<td>3.2 x 10$^8$</td>
<td>3.2 x 10$^8$</td>
<td>(T1.5)</td>
</tr>
<tr>
<td>Fe(II) + HO$_2$/O$_2^-$ $\rightarrow$ Fe(III) + HO$_2$</td>
<td>1.3 x 10$^6$</td>
<td>2.4 x 10$^6$</td>
<td>6.6 x 10$^6$</td>
<td>(T1.6)</td>
</tr>
<tr>
<td>HO$_2$/O$_2^-$ + HO$_2$/O$_2^-$ $\rightarrow$ H$_2$O$_2$</td>
<td>2.3 x 10$^6$</td>
<td>1.2 x 10$^7$</td>
<td>2.3 x 10$^7$</td>
<td>(T1.7)</td>
</tr>
<tr>
<td>•OH + HO$_2$/O$_2^-$ $\rightarrow$ H$_2$O + O$_2$</td>
<td>7.1 x 10$^9$</td>
<td>7.5 x 10$^9$</td>
<td>8.9 x 10$^9$</td>
<td>(T1.8)</td>
</tr>
<tr>
<td>•OH + •OH $\rightarrow$ H$_2$O$_2$</td>
<td>5.2 x 10$^9$</td>
<td>5.2 x 10$^9$</td>
<td>5.2 x 10$^9$</td>
<td>(T1.9)</td>
</tr>
<tr>
<td>•OH + HCOOH/HCOO$^-$ $\rightarrow$ CO$_2$ + O$_2^-$</td>
<td>6.5 x 10$^8$</td>
<td>2.2 x 10$^9$</td>
<td>3.2 x 10$^9$</td>
<td>(T1.10)</td>
</tr>
</tbody>
</table>
Table 2-2. Some published rate constants for the reactions of •OH with formic acid, formate, and hydrogen peroxide, in M⁻¹ s⁻¹. We included all rate constants except those where the reaction pH was outside of 2-5.

<table>
<thead>
<tr>
<th></th>
<th>$k_{\text{OH,HCOOH}}$</th>
<th>$k_{\text{OH,HCOO}^-}$</th>
<th>$k_{\text{OH,H}_2\text{O}_2}$</th>
</tr>
</thead>
<tbody>
<tr>
<td>1.3 x 10⁸</td>
<td>4.3 x 10⁹</td>
<td>2.7 x 10⁷</td>
<td></td>
</tr>
<tr>
<td>6.5 x 10⁸</td>
<td>3.2 x 10⁹</td>
<td>4.5 x 10⁷</td>
<td></td>
</tr>
<tr>
<td>1.6 x 10⁸</td>
<td>3.8 x 10⁹</td>
<td>1.2 x 10⁷</td>
<td></td>
</tr>
<tr>
<td></td>
<td>2.2 x 10⁹</td>
<td>1.7 x 10⁷</td>
<td></td>
</tr>
<tr>
<td></td>
<td>3.4 x 10⁹</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Average:</td>
<td>3.1 (±2.9) x 10⁹</td>
<td>3.4 (±0.8) x 10⁹</td>
<td>2.5 (±1.5) x 10⁷</td>
</tr>
</tbody>
</table>

a Buxton et al. (40)
b Ross and Ross (31)
c Elliot et al. (47)
Table 2-3. Experimental pseudo first-order decomposition rates of $^{14}$COOH/$^{14}$COO$^-$ and H$_2$O$_2$, and the corresponding $\gamma$ in pH 4 solutions with dissolved iron, in day$^{-1}$. The uncertainty in the rate coefficients represents one standard deviation calculated from non-linear regressions of the raw data (concentration of $^{14}$COOH/$^{14}$COO$^-$ and H$_2$O$_2$ versus time). The uncertainty in $\gamma$ represents one standard deviation calculated by propagating the errors from the rate coefficients. Measured dissolved Fe (0.02-μm filtered) was 0.41-2.55 μM, but no relationship between reaction rates and dissolved Fe was observed.

<table>
<thead>
<tr>
<th>Experiment #</th>
<th>$k_{\text{HCOOH}}$</th>
<th>$k_{\text{H}_2\text{O}_2}$</th>
<th>$\gamma$</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>1.21 ± 0.14</td>
<td>0.066 ± 0.0019</td>
<td>18.3 ± 2.2</td>
</tr>
<tr>
<td>2</td>
<td>2.23 ± 0.17</td>
<td>0.084 ± 0.012</td>
<td>26.5 ± 4.3</td>
</tr>
<tr>
<td>3</td>
<td>1.73 ± 0.034</td>
<td>0.075 ± 0.0086</td>
<td>23.1 ± 1.8</td>
</tr>
<tr>
<td>4</td>
<td>0.21 ± 0.0094</td>
<td>0.0082 ± 0.0016</td>
<td>25.6 ± 5.1</td>
</tr>
<tr>
<td>5</td>
<td>0.32 ± 0.033</td>
<td>0.017 ± 0.002</td>
<td>18.8 ± 2.9</td>
</tr>
<tr>
<td>6</td>
<td>0.89 ± 0.060</td>
<td>0.046 ± 0.0049</td>
<td>19.3 ± 2.4</td>
</tr>
<tr>
<td>7</td>
<td>0.23 ± 0.0090</td>
<td>0.0090 ± 0.0035</td>
<td>25.6 ± 10.0</td>
</tr>
<tr>
<td>Average$^*$</td>
<td>0.68 ± 0.13</td>
<td>0.030 ± 0.015</td>
<td>22.2 ± 11.7</td>
</tr>
</tbody>
</table>

$^*$Each value is the geometric average of the data set because the data is log-normally distributed.
Table 2-4. Experimental pseudo first-order decomposition rates of benzoic acid and formic acid in pH 4 solutions containing ferrihydrite. The uncertainty in the observed rate coefficients represents one standard deviation calculated from nonlinear regressions of the raw data. The uncertainty in the ratio represents one standard deviation calculated by propagating the errors from the rate coefficients. Also listed are the averages of the literature values of the second-order rate constants with •OH that meet the criterion stated in Table 2-2, with an uncertainty of one standard deviation.

<table>
<thead>
<tr>
<th></th>
<th>$k_{C_6H_5COOH}$ (day$^{-1}$)</th>
<th>$k_{HCOOH}$ (day$^{-1}$)</th>
<th>$\frac{k_{C_6H_5COOH}}{k_{HCOOH}}$</th>
</tr>
</thead>
<tbody>
<tr>
<td>Experiment 1</td>
<td>1.26 ± 0.22</td>
<td>0.59 ± 0.13</td>
<td>2.1 ± 0.6</td>
</tr>
<tr>
<td>Experiment 2</td>
<td>0.66 ± 0.042</td>
<td>0.23 ± 0.034</td>
<td>2.9 ± 0.5</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th></th>
<th>$k_{OH,C_6H_5COOH}$ (M$^{-1}$ s$^{-1}$)</th>
<th>$k_{OH,[HCOOH]}_t$ (M$^{-1}$ s$^{-1}$)</th>
<th>Expected $\frac{k_{OH,C_6H_5COOH}}{k_{OH,[HCOOH]}_t}$</th>
</tr>
</thead>
<tbody>
<tr>
<td>Literature values</td>
<td>4.0 (±1.3) $\times 10^9$</td>
<td>2.2 (±0.5) $\times 10^9$</td>
<td>1.7 ± 0.7</td>
</tr>
</tbody>
</table>
Table 2-5. Estimated solution chain lengths vs. pH, assuming [Fe(III)] is at equilibrium with amorphous ferric hydroxide ($K_{sp} = 10^{-38.8}$).

<table>
<thead>
<tr>
<th>pH</th>
<th>$[\text{Fe(III)}]_{\text{sat}}$</th>
<th>$k_{H_2O_2}$ (day$^{-1}$)</th>
<th>$k_{HCOOH}$ (day$^{-1}$)</th>
<th>Chain length</th>
</tr>
</thead>
<tbody>
<tr>
<td>3</td>
<td>15 μM</td>
<td>~0.47</td>
<td>~4.6</td>
<td>~90</td>
</tr>
<tr>
<td>4</td>
<td>0.42 μM</td>
<td>~0.043</td>
<td>~1.4</td>
<td>~230</td>
</tr>
<tr>
<td>5</td>
<td>33 nM</td>
<td>~0.0063</td>
<td>~0.3</td>
<td>~430</td>
</tr>
</tbody>
</table>
Figure 2-1. Schematic diagram of the Fe(III)-initiated Fenton-like chain reaction.

\[
\begin{align*}
\text{Fe(III)} + \text{H}_2\text{O}_2 & \rightarrow \text{Fe(II)} + \text{O}_2^- \\
\text{Fe(II)} + \text{H}_2\text{O}_2 & \rightarrow \text{Fe(III)} + \text{HO}_2^- \\
\text{HO}_2^- & \rightarrow \bullet\text{OH} \quad \text{or} \quad \text{Fe(II)} + \text{H}_2\text{O}_2 \\
\bullet\text{OH} & \rightarrow \text{HCOOH} + \text{H}_2\text{O}_2
\end{align*}
\]
Figure 2-2. A typical data set of the concentration of hydrogen peroxide (●) and $^{14}$C-labeled formic acid (▽) over time in an experiment with only dissolved Fe at pH 4. Error bars represent one standard deviation. The lines are non-linear regression fits of the data to the function $[C] = [C]_0 \exp(-kt)$, with $[C]_0$ values of 819 μM and 110 nM and $k$ values of 0.046 day$^{-1}$ and 0.89 day$^{-1}$, for H$_2$O$_2$ and HCOOH, respectively.
Figure 2-3a. Experimental (○) pseudo-first-order decomposition rates of $\text{H}_2\text{O}_2$ in the ferrihydrite/$\text{H}_2\text{O}_2$ system. Also included are data from experiments where 10 μM tert-butyl alcohol (■) or 100 μM tert-butyl alcohol (▲) were added. Error bars represent one standard deviation of rate coefficients calculated from non-linear regressions of the raw data.
Figure 2-3b. Experimental (O) pseudo-first-order decomposition rates of H$^{14}$COOH/H$^{14}$COO$^{-}$ in the ferrihydrite/H$_2$O$_2$ system. Also included are data from experiments where 10 µM tert-butyl alcohol (■) or 100 µM tert-butyl alcohol (▲) were added. Error bars represent one standard deviation of rate coefficients calculated from non-linear regressions of the raw data.
Figure 2-4. Comparison of predictions from Models A-D to the experimental γ values (●) of the ferrihydrite/H\textsubscript{2}O\textsubscript{2} system. Error bars represent one standard deviation, calculated by propagating the errors shown in Figures 2-3a,b or from iron measurements. Note that lines for Models A and D are nearly identical.
Figure 2-5a. Pseudo-first-order decomposition rates of H$_2$O$_2$ in an iron oxide/H$_2$O$_2$ system from Models A and B, with and without 100 μM tert-butyl alcohol.
Figure 2-5b. Pseudo-first-order decomposition rates of $\text{H}^{14}\text{COOH}/\text{H}^{14}\text{COO}^-$ in an iron oxide/$\text{H}_2\text{O}_2$ system from Models A and B, with and without 100 µM tert-butyl alcohol.
Figure 2-5c. Pseudo-first-order decomposition rates of H$_2$O$_2$ in an iron oxide/H$_2$O$_2$ system from Models C and D, with and without 100 μM tert-butyl alcohol.
Figure 2-5d. Pseudo-first-order decomposition rates of $\text{H}^{14}\text{COOH}/\text{H}^{14}\text{COO}^-$ in an iron oxide/$\text{H}_2\text{O}_2$ system from Models C and D, with and without 100 $\mu$M tert-butyl alcohol.
Chapter 3: Rates of Hydroxyl Radical Generation and Organic Compound Oxidation in Mineral-Catalyzed Fenton-like Systems

or

-Those iron oxides that I synthesized in my first year could be useful-


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Abstract

The iron oxide-catalyzed production of hydroxyl radical (•OH) from hydrogen peroxide (H₂O₂) has been used to oxidize organic contaminants in soils and groundwater. The goals of this study are to determine which factors control the generation rate of •OH (VOH) and to show that if VOH and the rate constants of the reactions of •OH with the system’s constituents are known, the oxidation rate of a dissolved organic compound can be predicted. Using ¹⁴C-labeled formic acid as a probe, we measured VOH in pH 4 slurries of H₂O₂ and either synthesized ferrihydrite, goethite, or hematite or a natural iron oxide-coated quartzitic aquifer sand. In all of our experiments, VOH was proportional to the product of the concentrations of surface area of the iron oxide and H₂O₂, although different solids produced •OH at different rates. We used these results to develop a model of the decomposition rate of formic acid as a function of the initial formic acid and hydrogen peroxide concentrations and of the type and quantity of iron oxide. Our model successfully predicted the VOH and organic compound oxidation rates observed in our aquifer sand experiment and in a number of other studies but overpredicted VOH and oxidation rates in other cases, possibly indicating that unknown reactants are either interfering with •OH production or consuming •OH in these systems.

Introduction

A powerful method for the remediation of wastewater and contaminated groundwater is chemical oxidation, where a reactive chemical species such as the hydroxyl radical (•OH) is generated in aqueous solution. The hydroxyl radical is often chosen because it is a nonspecific oxidant that reacts with most organic compounds at near diffusion-limited rates (1). The common way to make hydroxyl radicals in situ in contaminated aquifers is to inject acidified solutions of ferrous iron into the groundwater, followed by concentrated hydrogen peroxide (H₂O₂). This is the well-known Fenton’s reagent:

\[
\text{Fe(II)} + \text{H}_2\text{O}_2 \rightarrow \text{Fe(III)} + \text{•OH} + \text{OH}^- \tag{1}
\]

If only Fe(III) is originally present, Fe(II) can be slowly generated by the following reactions:

\[
\text{Fe(III)} + \text{H}_2\text{O}_2 = \text{Fe(HO}_2\text{)}^{2+} + \text{H}^+ \tag{2}
\]

\[
\text{Fe(HO}_2\text{)}^{2+} \rightarrow \text{Fe(II)} + \text{HO}_2 \tag{3}
\]

In addition, the HO₂ formed in reaction 3 may deprotonate (pKₐ = 4.8; 2) and initiate a chain reaction sequence via the reaction

\[
\text{O}_2^- + \text{Fe(III)} \rightarrow \text{Fe(II)} + \text{O}_2 \tag{4}
\]
followed by reaction 1 and

\[ \cdot \text{OH} + \text{H}_2\text{O}_2 \rightarrow \text{HO}_2 + \text{H}_2\text{O} \]  \hspace{1cm} (5)

Thus \( \cdot \text{OH} \) is formed from hydrogen peroxide when either Fe(II) or Fe(III) is present, although generation rates are much slower in the latter case, even when the chain reaction is significant (3, 4).

Researchers have shown that common forms of iron oxide (goethite, hematite, magnetite, and ferrihydrite) can also catalyze the oxidation of organic compounds by \( \text{H}_2\text{O}_2 \) from pH 3 to pH 7 (5-13). Experiments that evaluated different iron oxides under the same conditions exhibited differences in the degradation rates of hydrogen peroxide and contaminants. Magnetite (Fe\(_3\)O\(_4\)) was the most effective catalyst as compared to the other iron oxides (6, 10, 13), possibly because it was the only one that had Fe\(^{2+}\) in its structure to enhance the production rate of \( \cdot \text{OH} \). Valentine and Wang (11) proposed that the surface area of the iron oxide accounts for the difference in reaction activity; they and Huang et al. (14) showed that the decomposition rates of \( \text{H}_2\text{O}_2 \) by goethite, ferrihydrite, and hematite were all relatively similar when normalized to surface area.

However, the consumption rate of \( \text{H}_2\text{O}_2 \) does not equal the generation rate of \( \cdot \text{OH} \) (\( V_{\text{OH}} \)) because \( \text{H}_2\text{O}_2 \) can decompose to water and oxygen via nonradical-producing pathways. Examples of these include reactions involving biological enzymes (e.g., catalase) and inorganic chemical species such as manganese (15) and the iron oxide surface (4). As it is the concentration of \( \cdot \text{OH} \) that determines the oxidation rate of the contaminant, it is therefore crucial to examine and understand the factors that control \( V_{\text{OH}} \) in addition to those that affect the lifetime of \( \text{H}_2\text{O}_2 \).

There have been only four publications (9, 16-18) to our knowledge where the investigators explicitly measured \( V_{\text{OH}} \) from hydrogen peroxide decomposition and looked at the parameters that could affect it. Watts et al. (9, 16) did two studies in systems containing either hematite or a goethite-silica sand and showed that \( V_{\text{OH}} \) remained constant when [\( \text{H}_2\text{O}_2 \)] was varied from 32 mM to 1.6 M and pH was varied from 4 to 7. Huling et al. (17) found that \( V_{\text{OH}} \) in peat-amended suspensions of silica sand mixed with goethite ore at pH ~6 was twice as much as that in unamended suspensions. Petigara et al. (18) measured the \( \cdot \text{OH} \) formation rates of four different soil suspensions and concluded that \( \cdot \text{OH} \) was a major product of \( \text{H}_2\text{O}_2 \) decomposition in soils with low organic matter but was a minor product in soils containing high amounts of organic matter or manganese. Variations in the yield of \( \cdot \text{OH} \) from iron oxide/\( \text{H}_2\text{O}_2 \) systems have
also been noted previously (ref 4 and references therein). These studies illustrate that a more comprehensive study of the factors that control \( V_{OH} \) is needed. Although many other publications have documented the loss of an organic compound in the presence of \( \text{H}_2\text{O}_2 \) and iron oxide, those data cannot be used to study the behavior of \( V_{OH} \) because the compound was not completely dissolved in water, which complicates the data interpretation.

A deeper understanding of the factors that control the concentration of \( \bullet\text{OH} \) in a pure iron oxide/\( \text{H}_2\text{O}_2 \) system would allow us to estimate the disappearance rate of any organic compound that can be oxidized by \( \bullet\text{OH} \) in such systems. This would give us a way to interpret the differences between diverse systems and a basis for predicting the time needed for a contaminant to degrade in soils, where complications arise because of greater heterogeneity, sorption processes, and the presence of multiple sinks of \( \text{H}_2\text{O}_2 \) and \( \bullet\text{OH} \). Although many investigators have presented equations to predict pollutant destruction rates, most of them used empirical fits of experimental data. One exception is the work of Huling et al. (19), who showed that one can use the loss rate of one dissolved organic compound to calculate \( V_{OH} \) of a given \( \text{H}_2\text{O}_2/\text{sand slurry} \) and then use that \( V_{OH} \) to predict the loss rate of another dissolved organic compound under the same conditions. However, they did not attempt to develop a more generally applicable approach for predicting \( V_{OH} \).

This study aims to provide greater comprehension of the parameters that control \( V_{OH} \) and to use that information to predict the degradation rate of any organic compound in an iron oxide/\( \text{H}_2\text{O}_2 \) system. Our experiments at pH 4 with three different iron oxides (ferrihydrite, goethite, and hematite) and \( ^{14}\text{C} \)-labeled formic acid as the \( \bullet\text{OH} \) probe show that \( V_{OH} \) is proportional to the product of the concentrations of surface area of the iron oxide and \( \text{H}_2\text{O}_2 \). Using our results, we formulated a general model that predicts \( V_{OH} \) with four parameters: surface area, type and amount of iron oxide, and concentration of \( \text{H}_2\text{O}_2 \). Our model can also estimate the oxidation rate of any compound if we know its aqueous concentration and its reaction rate with \( \bullet\text{OH} \), along with the concentration and reaction rates of any other important \( \bullet\text{OH} \) sinks in the system. We then compared our model predictions of \( V_{OH} \) and oxidation rates of organic compounds with a number of observations from published studies.

**Materials and Methods**

**Materials**

Detailed descriptions for materials and methods are in a previous publication (4). A brief
summary follows. All reagents were reagent grade and used as received. Solutions were prepared using 18 MΩ-cm Milli-Q water from a Millipore system. Glassware and containers were acid washed and rinsed before use. Colloidal ferrihydrite aggregates, mostly retained by 0.02-μm pore size filters, were prepared from FeCl₃•6H₂O according to the method described by Wells et al. (20). FeCl₃•6H₂O was dissolved to make a 4 x 10⁻⁴ M solution (5 x 10⁻⁴ M for experiments with high ferrihydrite concentrations). Within 1 h, the solution was heated in a 90 °C water bath for 5 min and then quickly cooled back to room temperature. It was used within 2 days as a colloidal suspension without further purification. Goethite was synthesized by aging ferrihydrite in a strong alkaline solution at 70 °C for 60 h (21). Hematite was made in the same manner as goethite except that the solution was aged for 8 days (21). The identities of both solids were verified by X-ray diffraction. The aquifer sand came from Georgetown, SC, and is an iron oxide-coated quartzitic sand that was previously characterized (22, 23). It was sterilized before use by autoclaving, which has been shown to be very effective not only in killing microorganisms but also in deactivating enzymes (24).

**Analytical Techniques**

Hydrogen peroxide was measured spectrophotometrically using the DPD method (25) as modified by Voelker and Sulzberger (26) to minimize interference by Fe(II) and Fe(III). Samples were filtered when necessary to minimize light scattering from iron oxide particles and diluted when needed to ensure that the absorbance through a 1-cm path length cuvette was less than 1.2 au. The concentration of ferrihydrite was measured with ferrozine after reduction with hydroxylamine hydrochloride in strong HCl. H¹⁴COOH was measured using liquid scintillation (ScintiSafe Econo 1 Cocktail, Fisher) after an aliquot of experimental solution was air sparged to remove ¹⁴CO₂ (4). The surface areas of goethite and hematite were measured by single-point N₂ BET analysis (Porous Materials, Inc.) and were respectively 42.5 and 50.1 m²/g, with a standard deviation of less than 0.5 m²/g. BET analysis was not done on the ferrihydrite because we assumed that the drying step would alter its surface area. The bulk surface area of the sand was 15.2 m²/g, calculated from mercury intrusion data (22). Its bulk fraction of organic carbon was 0.109 ± 0.025%, and the total iron content obtained after sediment digestion with HF-HCl-HNO₃ was 267 μmol/g (22). Ti(III)-citrate-EDTA-bicarbonate (TiCEB) and ammonium-oxalate-in-the-dark (AOD) extractions were also performed on the sand. TiCEB extraction measured 189 μmol/g of crystalline and amorphous surface iron, and AOD extraction determined that the
amorphous iron content was 16 µmol/g (22).

*Kinetic Experiments*

All experiments were performed in 250-mL Teflon bottles covered completely with black electrical tape to exclude light. The Milli-Q water was treated with a total organic carbon reduction unit (Aquafine) to oxidize traces of organic contamination. Unless noted otherwise, experimental solutions contained 10 mM NaClO₄ to control ionic strength and 1 µCi of ¹⁴C. The bottles were loosely capped during the experiment, and the solutions were stirred magnetically while being kept in a recirculating water bath. The solutions were adjusted to pH 4 with HClO₄ and/or NaOH before the addition of H₂O₂ at time zero. Care was taken to avoid raising the pH of the solution above 6 during the adjustment. The pH of the solutions remained within ±0.5 unit of the initial pH throughout the experiment except for one instance discussed below. The temperature of the water was 22 ± 5 °C, but the variation was only ±1 °C for each individual set of experiments. A stock solution of sodium formate, kept in the dark at 4 °C and discarded after 2 weeks, was used to augment [HCOOH]ₜ in some of the experiments ([HCOOH]ₜ = [HCOOH] + [HCOO⁻]). In these experiments, the sum of the concentrations of NaClO₄ and sodium formate was 10 mM to maintain a constant ionic strength.

Loss of H¹⁴COOH appeared to follow pseudo-first-order kinetics in all experiments. Significant loss of H₂O₂ was detected only in experiments containing iron oxide and [H₂O₂]₀ of 1 mM or less and in some of the experiments with high [HCOOH]ₜ. H¹⁴COOH loss in H₂O₂-free control reactors containing iron oxide was insignificant. A control experiment consisting of autoclaved sand, Milli-Q water, and H¹⁴COOH showed no loss of H¹⁴COOH for at least 15 h.

For those experiments where [HCOOH]ₜ₀ = 10 mM, the decomposition rate of H¹⁴COOH was obtained using the data points from 0 to 4 h because the pH rose above 5 afterward, which interfered with our sparging technique to separate ¹⁴CO₂ from H¹⁴COOH. We believe the cause of this phenomenon was the high concentration of formic acid (see Appendix B) because in previous experiments at low formic acid concentrations, minimal pH changes were observed.

*Calculating VOH*

A widely used method to measure VOH is to introduce a hydroxyl radical sink and a dilute chemical probe that reacts with •OH in a well-defined manner (27). If the consumption of the •OH probe follows pseudo-first-order kinetics, then its loss rate is
\[
- \frac{d[\text{probe}]}{dt} = k_{\text{probe}}[\text{probe}]
\]

where \( k_{\text{probe}} \) is the observed pseudo-first-order rate constant (s\(^{-1}\)) and is also equal to
\( k_{\text{OH,probe}}[\cdot\text{OH}]_{ss} \), with \( k_{\text{OH,probe}} \) as the second-order rate constant of the probe with \( \cdot\text{OH} \) (M\(^{-1}\) s\(^{-1}\)).

This gives us an equation for \([\cdot\text{OH}]_{ss}\):

\[
[\cdot\text{OH}]_{ss} = \frac{k_{\text{probe}}}{k_{\text{OH,probe}}}
\]

The generation rate of \( \cdot\text{OH} \) must be equal to its scavenging rate at steady state:

\[
V_{\text{OH}} = \text{scavenging rate} = \sum k_{\text{OH,1}}[S_i][\cdot\text{OH}]_{ss}
\]

where \( S_i \) represents \( \cdot\text{OH} \) sink \( i \) in solution. Substituting eq 7 into eq 8 yields

\[
V_{\text{OH}} = \frac{k_{\text{probe}}}{k_{\text{OH,probe}}} \sum k_{\text{OH,1}}[S_i]
\]

\( k_{\text{probe}} \) for our experiments was obtained by using SigmaPlot to do nonlinear least-squares regression fits of the concentration of our probe, H\(^{14}\)COOH, versus time \((k_{\text{HCOOH}})\). (Most of the rate coefficients obtained from linear regression fits to log transformed data differed from those obtained from nonlinear regressions by only a few percent. See Appendix B.) Since H\(_2\)O\(_2\) and HCOOH were the only major sinks of \( \cdot\text{OH} \) in our experiments, just their second-order rate constants with \( \cdot\text{OH} \) were needed for our \( V_{\text{OH}} \) computation. We used \( 2.7 \times 10^7 \) M\(^{-1}\) s\(^{-1}\) \((J)\) for \( k_{\text{OH,H}_2\text{O}_2} \) and \( 2.2 \times 10^9 \) M\(^{-1}\) s\(^{-1}\) \((4)\) for \( k_{\text{OH,\text{HCOOH}}} \). \( k_{\text{OH,\text{HCOOH}}} \) is a composite rate constant and is the weighted sum of the two individual rate constants for formic acid and formate multiplied by the percentage of their respective chemical species at pH 4 \((4)\). The pK\(_a\) of formic acid is 3.745 \((28)\).

Our calculations do not need to distinguish between reactions in the bulk solution and reactions in the diffusion layer surrounding the particles if diffusion is fast as compared to the rate of decomposition of H\(_2\)O\(_2\) and formic acid, so that the concentrations of these species are the same in the diffusion layer and bulk solution \((4, 19)\).

Results

We calculated \( V_{\text{OH}} \) values from the observed pseudo-first-order decay rates of our probe, \( k_{\text{HCOOH}} \), in a variety of iron oxide/H\(_2\)O\(_2\) systems and found that \( V_{\text{OH}} \) is proportional to the product of the concentrations of surface area (a surrogate for reactive sites, see Discussion) and hydrogen
peroxide (Figure 3-1a). However, the proportionality coefficient differed for the different iron oxides (solid lines in Figure 3-1a):

Goethite: \( V_{\text{OH}} = 1 (\pm 0.1) \times 10^{-7} [\text{surface area}] [\text{H}_2\text{O}_2] \)  

Hematite: \( V_{\text{OH}} = 8 (\pm 0.4) \times 10^{-9} [\text{surface area}] [\text{H}_2\text{O}_2] \)  

Ferrihydrite: \( V_{\text{OH}} = 1 (\pm 0.02) \times 10^{-7} [\text{surface area}] [\text{H}_2\text{O}_2] \)

Since we could not measure the surface area of our colloidal ferrihydrite, we plotted those data by assuming that it was 200 m\(^2\)/g. Ferrihydrite has a typical surface area of 100-400 m\(^2\)/g (29). In most of our experiments, \( V_{\text{OH}} \) and \([\text{OH}]_s\) remained constant over time because \([\text{H}_2\text{O}_2]\) did not decrease considerably. In those experiments where \( \text{H}_2\text{O}_2 \) consumption was significant, it is more appropriate to consider our calculated \( V_{\text{OH}} \) values as initial •OH generation rates.

Although \( V_{\text{OH}} \) is directly proportional to \([\text{H}_2\text{O}_2]\) at a constant amount of iron oxide, changing \([\text{H}_2\text{O}_2]\) will have no effect on \([\text{•OH}]_s\) and, consequently, \( k_{\text{HCOOH}} \) if \( \text{H}_2\text{O}_2 \) is the dominant sink of •OH. This is apparent when eq 8 is rewritten:

\[
[\text{•OH}]_s = \frac{V_{\text{OH}}}{k_{\text{OH}, \text{H}_2\text{O}_2} [\text{H}_2\text{O}_2] + k_{\text{OH}, \text{HCOOH}} [\text{HCOOH}]} \quad (13)
\]

We replotted the data in Figure 3-1a to illustrate this point (Figure 3-1b). The model lines were generated by first using eqs 10-12 to determine \( V_{\text{OH}} \) and then using eq 14 to calculate the predicted values of the initial decomposition rate of formic acid:

\[
-\frac{d[\text{HCOOH}]}{dt}|_{t=0} = k_{\text{HCOOH}}[\text{HCOOH}]_T, \quad (14)
\]

(We will use \( d[\text{HCOOH}] /dt \) instead of \( d[\text{HCOOH}] /dt|_{t=0} \) for simplicity.) \( k_{\text{HCOOH}} \) was modeled by multiplying \( k_{\text{OH}, \text{HCOOH}} \) with \([\text{•OH}]_s\), which was computed using eq 13, with \([\text{H}_2\text{O}_2]\) = \([\text{H}_2\text{O}_2]_0\) and \([\text{HCOOH}]_T = [\text{HCOOH}]_{T,0}\). The initial concentration of formic acid in these experiments was low (74-244 nM, with an average value of 131 nM), so hydrogen peroxide was the major sink of •OH. It is obvious from the figure that increasing \([\text{H}_2\text{O}_2]\) by 2-3 orders of magnitude had minimal impact on \( d[\text{HCOOH}] /dt \) (normalized to [Fe]\(_T\)) for all three iron oxides.

Under conditions where \([\text{HCOOH}]_T,0\) is high enough that it is the main sink of •OH, increasing \([\text{H}_2\text{O}_2]\) should cause the oxidation rate of formic acid to increase. To demonstrate this and to ensure that high concentrations of formic acid would not compete with \( \text{H}_2\text{O}_2 \) for the active sites on the iron oxide surface and interfere with the production rate of •OH, we performed additional experiments at 0.6 g/L goethite with HCOOH as the dominant •OH sink. A comparison of the experiments in which HCOOH was the dominant •OH sink with those in...
which H$_2$O$_2$ was the main sink shows that V$_{OH}$ was unaffected by formic acid concentrations as high as 10 mM (Figure 3-2a) and that the expected increase in the rate of oxidation of formic acid with increasing [H$_2$O$_2$]$_0$ occurred (Figure 3-2b). The probe loss was well described by first-order kinetics (eq 6), implying that $k_{HCOOH}$ was approximately constant over the course of the experiments (see Appendix B). As some H$_2$O$_2$ was consumed during the experiments, the V$_{OH}$ values that we calculated using eq 9 with [HCOOH]$_{T,0}$ and [H$_2$O$_2$]$_0$ are more representative of the initial •OH generation rates. Figure 3-2b also illustrates that, at a given [Fe]$_T$ and [HCOOH]$_{T,0}$, there is a theoretical limit to the pseudo-first-order consumption rate of formic acid because there is a maximum value for [•OH]$_{ss}$. This is best shown by the data at [HCOOH]$_{T,0}$ = 50 µM (white squares and black hexagons): adding more H$_2$O$_2$ increased d[HCOOH]$_T$/dt until $k_{OH,H_2O_2}$ [H$_2$O$_2$]$_0$ became much greater than $k_{OH,[HCOOH]}$, [HCOOH]$_{T,0}$, at which point d[HCOOH]$_T$/dt approached its maximum value.

We used the Georgetown sand to examine if our findings from a pure iron oxide system would also apply to natural aquifer material. Goethite was the only type of crystalline iron oxide found in the sand (23). The results of experiments using either 25 or 60 g/L sand and either H$_2$O$_2$ or formic acid as the major •OH sink are shown in Figure 3-3a,b. Although we knew the surface area of the bulk sand, it was unlikely to be equal to the surface area of the goethite in the sand matrix. Instead, we plotted the data assuming that the goethite had a surface area of 40 m$^2$/g, which lies within the typical range of the surface area of goethite (8-200 m$^2$/g) (29), and that there was 173 µmol of goethite/g of sand (the difference between the TiCEB and AOD extractions). The behavior of V$_{OH}$ with respect to the product of the concentrations of surface area and H$_2$O$_2$, with either H$_2$O$_2$ or formic acid as the dominant •OH sink, were analogous to that of the pure iron oxide system. Similar to the results of Figure 3-2b, d[HCOOH]$_T$/dt of the aquifer sand/H$_2$O$_2$ system rose as [H$_2$O$_2$]$_0$ increased when HCOOH was the dominant •OH sink but remained steady when H$_2$O$_2$ was the major •OH sink. The two V$_{OH}$ data points (they lie nearly on top of each other) that are outliers on the far left side of Figure 3-3a,b may have been caused by either of the following phenomena. One, there may have been contamination of the stock formate solution by bacteria whose activity was not suppressed significantly at [H$_2$O$_2$]$_0$ of 100 µM or less. The other explanation is that small amounts of Fe(II) could have been present in the sand, producing a small amount of •OH at a high rate. The effect of this reaction on V$_{OH}$ would only be noticeable when the starting concentration of H$_2$O$_2$ was small, whereas at higher [H$_2$O$_2$]$_0$, its
contribution to $V_{OH}$ would be negligible.

Our predictions for $V_{OH}$ (eq 10) and $d[HCOOH]/dt$ were within a factor of 10 of the measured values (lines in Figure 3-3a,b). The range of predictions for $d[HCOOH]/dt$ was calculated based on the typical range of surface area of goethite. We neglected the contribution of amorphous Fe to $V_{OH}$ and $d[HCOOH]/dt$ because it was only a small percentage (~10%) of the surface iron. Even if we assumed that the surface area of the amorphous iron was 10 times that of crystalline iron, it would only modify $V_{OH}$ and $d[HCOOH]/dt$ by a factor of 2, which is of minor importance considering the uncertainty of other factors involved in the computation.

Discussion

In summary, we showed that $V_{OH}$ is (i) proportional to the product of the concentrations of $H_2O_2$ and iron oxide surface area for goethite, hematite, ferrihydrite, and a natural aquifer sand and (ii) dependent on the type of iron oxide. The $V_{OH}$ measurements from experiments that had similar $[H_2O_2]_0$ but different dominant $OH$ sinks (either formic acid or $H_2O_2$) agreed with each other. Besides being evidence that high concentrations of formic acid did not impact $V_{OH}$, this finding also supports our assumption that $OH$ was the primary oxidant of our system, which agrees with our previous work (4). A different oxidant would react with formic acid and $H_2O_2$ at second-order rates unlike those of $OH$, causing the data sets to not coincide with each other.

Relationship between $[H_2O_2]$, Iron Oxide, and $V_{OH}$

Our observation that $V_{OH}$ is proportional to the product of $[H_2O_2]$ and iron oxide surface area (indicated by the straight lines with a slope of 1 in Figure 3-1a) is consistent with a mechanism whose rate-limiting step involves $H_2O_2$ sorbed on the iron oxide surface. Although a consensus on the reaction mechanism does not exist, it is reasonable to assume that the surface reactions are analogous to those in solution:

$$\equiv Fe^{III} + H_2O_2 = \equiv Fe^{III}H_2O_2$$

(15)

$$\equiv Fe^{III}H_2O_2 \rightarrow \equiv Fe^{II} + HO_2 + H^+$$

(16)

$$\equiv Fe^{II} + H_2O_2 \rightarrow \equiv Fe^{III} + OH + OH^-$$

(17)

We assume that an identical Fe(III) site to that present in reaction 15 is regenerated by reaction 17. If reaction 16 is the rate-limiting step, then the rate of $OH$ production will be proportional to the concentration of $\equiv Fe^{III}H_2O_2$. If reaction 15 is fast enough to reach a pseudoequilibrium, then in accordance with the Langmuir isotherm expression:
\[
[\equiv \text{Fe}^\text{III}]_{\text{H}_2\text{O}_2} = [\equiv \text{Fe}^\text{III}]_{I} \frac{K_{\text{ads}}[\text{H}_2\text{O}_2]}{1 + K_{\text{ads}}[\text{H}_2\text{O}_2]}
\] (18)

where

\[
K_{\text{ads}} = \frac{[\equiv \text{Fe}^\text{III}]_{\text{H}_2\text{O}_2}}{[\equiv \text{Fe}^\text{III}]_{\text{H}_2\text{O}_2}}
\] (19)

and \([\equiv \text{Fe}^\text{III}]_{T} = [\equiv \text{Fe}^\text{III}]_{\text{H}_2\text{O}_2} + [\equiv \text{Fe}^\text{III}].\) We do not expect \(\equiv \text{Fe}^\text{II}\) to make up a significant portion of the surface sites because it is a transient species. If \(K_{\text{ads}}[\text{H}_2\text{O}_2]\) is much less than 1, then eq 18 simplifies to

\[
[\equiv \text{Fe}^\text{III}]_{\text{H}_2\text{O}_2} = K_{\text{ads}}[\equiv \text{Fe}^\text{III}]_{I}[\text{H}_2\text{O}_2]
\] (20)

and \(V_{\text{OH}}\) will be proportional to the product of \([\equiv \text{Fe}^\text{III}]_{T}\) and \([\text{H}_2\text{O}_2]\). This relationship should break down when \([\text{H}_2\text{O}_2]\) is sufficiently high for the surface to become saturated with sorbed \(\text{H}_2\text{O}_2\) (when \(K_{\text{ads}}[\text{H}_2\text{O}_2] \gg 1\)). We saw no evidence of this occurring at \([\text{H}_2\text{O}_2]\) as high as 3.7 M; so if this mechanism is correct, \(K_{\text{ads}}\) must be less than 0.27 M\(^{-1}\).

Previous works have proposed that a chain reaction mechanism contributes to the decomposition of \(\text{H}_2\text{O}_2\) and the generation of •OH in the presence of iron oxide surfaces. The chain reaction could be initiated by reactions 16 and 17 and then propagated by dissolved species (4, 30). Nevertheless, we do not expect propagation of the solution chain reaction to be significant in most of the systems examined here. If the iron oxide/\(\text{H}_2\text{O}_2\) system contains no iron complexing agents and is at pH 4 or greater, then there is very little dissolved iron because of the low solubility of Fe(III) above pH 4. Thus, the amount of hydroxyl radicals generated by reactions in the aqueous phase is likely to be negligible when compared to the amount produced at the oxide surface. We did observe evidence for a contribution of aqueous phase reactions at pH 4 at low concentrations of ferrihydrite (which is more soluble than goethite and hematite; 21), which accounts for most of the deviation of these data points from the general trend (black circles in Figure 3-1a,b; see ref 4 for further discussion of these data points). Models including chain reaction kinetics would not be expected to yield a simple dependence of \(V_{\text{OH}}\) on the concentrations of \(\text{H}_2\text{O}_2\) and iron oxide surface area (e.g., the models discussed in ref 4), so our results suggest that the mechanism of •OH generation does not include a significant contribution by a chain reaction.

We graphed \(V_{\text{OH}}\) versus surface area instead of \([\equiv \text{Fe}^\text{III}]_{T}\) on the assumption that the number of sorption sites per unit surface area was a constant for each form of iron oxide.
However, goethite was better at producing •OH than hematite (Figure 3-1a) even when the same concentrations of surface area were used. (The surface areas of the two iron oxides that were measured with the N\textsubscript{2} BET method on dry surfaces might not be the same as the values in suspension because of, for example, changes due to drying, adsorption of water and/or solutes onto the oxide surface, and molecular size differences between N\textsubscript{2} and H\textsubscript{2}O\textsubscript{2}. ) Petigara et al. (18) also commented on the higher reactivity of goethite, although their measurements were not normalized to surface area. This suggests that factors we did not consider, such as crystalline structure or impurities in the iron oxide, are also important in determining the •OH generation rate by affecting, for example, \( K_{ads} \) or the rate constant of reaction 16. Ferrihydrite has a greater solubility product (-log \( K_{sp} \) = 37.0-39.4) than either goethite (-log \( K_{sp} \) = 43.3-44.0) or hematite (-log \( K_{sp} \) = 42.2-43.3), and ferrihydrite (density = 3.96 g/cm\textsuperscript{3}) has a low crystallinity, whereas goethite (density = 4.26 g/cm\textsuperscript{3}) consists of edge-sharing octahedral crystals, and hematite (density = 5.26 g/cm\textsuperscript{3}) consists of edge- and face-sharing octahedral crystals (21). These variations between the three iron oxides could explain the differences in reactivity that we observed (Figure 3-1A). Interestingly, the decomposition of H\textsubscript{2}O\textsubscript{2} by goethite and hematite has been observed to proceed at comparable rates when normalized to surface area (11, 14), which implies that the yield of •OH from H\textsubscript{2}O\textsubscript{2} is dependent on the type of iron oxide. We had examined this yield in our previous work, but the new data do not provide us with additional information because H\textsubscript{2}O\textsubscript{2} loss was not measurable in most of our experiments.

Predicting \( V_{OH} \) and Organic Compound Degradation Rate

The success of our model prediction of the degradation rate of formic acid in the Georgetown sand at pH 4 shows that the iron oxide in a natural aquifer material with low organic matter content behaves fundamentally similar to pure iron oxide solids. However, the model prediction also illustrates the difficulty of determining the type and quantity of reactive iron oxides present in natural solids. Not all of the “crystalline” surface iron was necessarily goethite, as we assumed. Furthermore, the TiCEB extraction method may have dissolved some of the Fe present in clay minerals (31), so we could have (probably only slightly) overestimated the amount of crystalline surface iron in the aquifer sand. On a similar note, the effect of adsorbed silicate (32) and iron associated with aluminosilicates (approximately 35\% of total, based on the difference between the HF-HCl-HNO\textsubscript{3} and TiCEB extractions, a mole fraction of 0.24 ± 0.08 Al/Fe for goethite aggregates, and a mole fraction of 0.44 ± 0.14 Al/Fe for amorphous iron; 33)
on $V_{OH}$ was unclear. While the surface reactivity of the iron oxides could be influenced by the presence of Al and/or adsorbed silicate, it is more certain that the close association between the two natural oxides and the adsorbed silicate reduced the amount of surface area, i.e., the number of reactive Fe sites, which was exposed to $H_2O_2$. This implies that the data points in Figure 3A should be closer to the predicted $V_{OH}$ values. We chose the approach of using a range of possible goethite surface areas even though it is less satisfactory because it is difficult to obtain the actual value of this parameter.

To test the general applicability of our model, we also compared $V_{OH}$ and organic compound oxidation rates observed in previous studies under a wide variety of conditions to our model’s predictions. Since our model only predicts the oxidation rate of aqueous phase compounds by $•OH$, we considered only studies in which sorption of the organic compound was reported to be minor and in which the aqueous concentrations of the organic compound were measured. In other studies of iron oxide/$H_2O_2$ systems, the loss of hydrophobic compounds over time was measured using extraction techniques that detected the total (aqueous plus nonaqueous) quantity of the compounds remaining in the slurry.

To generate the predictions, we needed to assume that $V_{OH}$ would not be affected by pH. This is not intuitive given that pH would be expected to control the extent of formation of a surface $H_2O_2$ complex, as observed for the solution complex of Fe(III) and $H_2O_2$ (3). Nevertheless, three studies found that $V_{OH}$ was relatively invariant from pH 4 to pH 10, a range that encompasses almost all of the studies that were compared to our model: one in goethite/$H_2O_2$ systems from pH 5 to pH 9 (8), one in lepidocrite/$H_2O_2$ systems at initial pH values of 6.0 and 10.0 (12), and one in goethite-silica sand/$H_2O_2$ systems from pH 4 to pH 7 (16). At low pH values, proton-promoted dissolution of Fe from the oxide surface, as observed by Chou and Huang in their experiment at an initial pH value of 3.2 (12), could increase the importance of the solution phase chain reaction (4). It is unknown if there is a pH effect above pH 10.

When we compare $V_{OH}$ values gathered from the literature (Table 3-1) to our predictions (Figure 3-4), our predictions were within a factor of 10 for slightly more than half of the measurements. All four publications measured $V_{OH}$ with a nonsorbing probe and provided us with information about the type and amount of iron oxide plus the starting $H_2O_2$ concentration. Only in one study, however, was the iron oxide’s surface area measured. For both studies by
Watts et al., the calculations that we did were based on the surface area reported in their other publications (13, 34), which we assumed used the same iron oxides. For modeling the data from Petigara et al. (18), who examined natural soils, we took the approach used for the Georgetown sand, using typical ranges of surface area for goethite and hematite in the model.

Interestingly, our understanding of the relationship between $[\text{H}_2\text{O}_2]$ and $V_{\text{OH}}$ contrasts with that of Watts et al. (9), who observed that the $V_{\text{OH}}$ of hematite at pH 3 remained approximately constant as $[\text{H}_2\text{O}_2]_0$ increased from 0.1 to 5% (black circles in Figure 3-4). There is no obvious difference between the conditions of their experiments and ours, which were performed at similar concentration of $\text{H}_2\text{O}_2$ (0.03-1.6 vs 0.01-2 M) and surface area (9.6 vs 10 m$^2$/L). However, the $V_{\text{OH}}$ values reported by Watts et al. (9), determined with nitrobenzene as the $\bullet\text{OH}$ probe and 2-propanol added as an additional $\bullet\text{OH}$ sink, were in some cases much greater than the decomposition rates of $\text{H}_2\text{O}_2$ measured in separate experiments at the same concentrations of $\text{H}_2\text{O}_2$ and hematite but without nitrobenzene and 2-propanol. The ratio of moles of $\bullet\text{OH}$ created to moles of hydrogen peroxide consumed can, at most, be 1:1. One possible explanation of this discrepancy is that Watts et al. (9) might have measured a fast initial production of $\bullet\text{OH}$ in their probe experiments, while their observations of $\text{H}_2\text{O}_2$ decomposition rates took place on a time scale of 1-3 days (more comparable to those of our experiments). Another possibility is that their probe or sink compounds affected $V_{\text{OH}}$. In contrast, our predictions agreed very well with the $V_{\text{OH}}$ measurements in their other work using lower concentrations of the same probe but 1-octanol as the major $\bullet\text{OH}$ sink (16) (white triangles in Figure 3-4). The other two studies on $V_{\text{OH}}$ included natural materials besides pure iron oxides. While our predictions generally agreed with the observations of Petigara et al. (18), we overestimated those reported by Huling et al. (17).

To further examine our model, we compared the pseudo-first-order loss rates of organic compounds ($k_{\text{org}}$) observed in a variety of systems (Table 3-2) with our predictions (Figure 3-5). (In every study except the one by Huling et al. (19), both the organic compound and $\text{H}_2\text{O}_2$ were initially important $\bullet\text{OH}$ sinks.) Our predictions successfully predicted the values of $k_{\text{org}}$ measured by Lin and Gurol (8) but overpredicted the values of $k_{\text{org}}$ observed in the other studies. We selected these six studies because they satisfied our criteria that the compound that was tracked is highly water-soluble and that the studies report most of the measurements of the parameters that our model needs to estimate $V_{\text{OH}}$ and $k_{\text{org}}$: amount of $\text{H}_2\text{O}_2$, surface area, type and amount of iron.
oxide, concentration of the major $^\bullet$OH sinks and organic compound in the system, and rate constants of their respective reactions with $^\bullet$OH. (We used a composite rate constant for the reaction between the organic compound and $^\bullet$OH if we needed to account for acid-base speciation of the organic compound at the specified pH.) Four of them, however, did not have values for the surface area of the iron oxide because they either used natural aquifer material or did not measure it, so we treated this deficiency in the same way as before.

When other solutes were present, their possible influence on $k_{org}$ was considered. We included carbonate as a hydroxyl radical sink in calculating $k_{org}$ when it was reported to be present (11, 36, 37), but its effect was insignificant. The fast reaction of $Cl^-$ with $^\bullet$OH makes it a candidate as an important $^\bullet$OH scavenger in the studies done by Valentine and Wang (11) and Huling et al. (who included the reaction of $Cl^-$ and $^\bullet$OH in their model of 2-Chlorophenol degradation; 19). However, when $^\bullet$OH combines with chloride, the resulting intermediate can either dissociate back to the reactants or react with a proton to form a chlorine atom:

\[
^\bullet$OH + $Cl^-$ $\rightarrow$ $^\bullet$ClOH$^-$ (21)
\]
\[
^\bullet$ClOH$^-$ $\rightarrow$ $^\bullet$OH + $Cl^-$ (22)
\]
\[
^\bullet$ClOH$^-$ + $H^+$ $\rightarrow$ $Cl^-$ + $H_2O$ (23)
\]

The respective rate constants of reactions 21-23 are $4.3 \times 10^9$ M$^{-1}$ s$^{-1}$, $6.1 \times 10^9$ s$^{-1}$, and $2.1 \times 10^{10}$ M$^{-1}$ s$^{-1}$ (35). A comparison of the rates of reactions 22 and 23 indicates that chloride should not be the major sink of $^\bullet$OH above pH 2 because reaction 22, the reverse of reaction 21, is then the dominant decomposition pathway of $^\bullet$ClOH$^-$. We observed no difference in the decomposition rates of $H^{14}$COOH and $H_2O_2$ in solutions with and without 1 mM NaCl (data not shown; other experimental conditions were pH 4, 1 mM $H_2O_2$, 100 nM $H^{14}$COOH, 10 mM NaClO$_4$, and 1 $\mu$M Fe(III) or 21 mg/L ferrihydrite), confirming chloride’s ineffectiveness as an $^\bullet$OH sink at pH 4.

The possible effects of other system components are more difficult to predict, but they are likely to be unimportant. Sulfate is known to adsorb onto iron oxides, yet we did not find any connection between its presence and the deviations in our predictions. This is consistent with the results of Watts et al. (16), who observed similar $V_{OH}$ values in experiments using unstabilized $H_2O_2$ or $H_2O_2$ containing 27.3 g/L (NH$_4$)$_2$SO$_4$ and 28.6 g/L NaH$_2$PO$_4$•7H$_2$O. Finally, although natural organic matter could inhibit catalytic sites on the iron oxide or act as a radical sink, very little of it ($<0.1\%$) was present in the natural materials that were used in the studies considered here.
There are three principal explanations for the disagreements between our predictions and the measurements. First, we do not have a complete understanding of all of the factors that determine the reactivity of an iron oxide. Hence, our model cannot always make accurate predictions, especially when the iron oxide is not in a pure form (e.g., associated with soil components or contains impurities) or when it is one that we did not examine, as illustrated by the large deviations between our estimates and those measured by Chou and Huang (12; black circles in Figure 3-5). They determined that their brick grain-supported iron oxide was lepidocrocite ($\gamma$-FeOOH), and we used goethite ($\alpha$-FeOOH) in our model as an approximation. If the reactivity of lepidocrocite were instead similar to that of hematite, our predictions would be within 1 order of magnitude of the reported values. This reasoning also applies to the Valentine and Wang experiment (11) that used a semicrystalline iron oxide, which we assumed was equivalent to our ferrihydrite.

The second possible cause is that the compound being degraded or its oxidation products could interfere with the generation of $\mathbf{\cdot}$OH. The oxidation products of many of the aromatic compounds used in these studies are capable of complexing iron and, therefore, could also form complexes on the surfaces of iron oxides. Salicylic acid, for example, is a product of the oxidation of benzoic acid by $\mathbf{\cdot}$OH (38-40). We chose formic acid as our $\mathbf{\cdot}$OH probe to avoid the possibility of Fe reacting with the probe compound or its oxidation product.

Last, it is possible that unknown compounds associated with the solids in some of these systems were the main sinks of radicals. Miller and Valentine proposed that the oxide surface in their aquifer material competed with H$_2$O$_2$ and quinoline (or phenol, in a later study) for $\mathbf{\cdot}$OH and other radicals to explain the lack of increase in the decomposition rate of quinoline (or phenol) with an increase in the amount of solids at constant [H$_2$O$_2$] (36, 37). Reactions representing this phenomenon were included in the mechanism that they used to successfully model the loss of quinoline and phenol in the presence of aquifer material and H$_2$O$_2$. However, we saw no evidence for similar reactions in our aquifer sand experiments.

In summary, we focused our study on determining the factors that affect $V_{\mathbf{\cdot}$OH} in iron oxide/H$_2$O$_2$ systems and showed that if we know $V_{\mathbf{\cdot}$OH} and the reaction rates of the important $\mathbf{\cdot}$OH sinks in the system, the oxidation rate of dissolved organic compounds can be predicted. The simple proportionality we observed between $V_{\mathbf{\cdot}$OH} and the product of H$_2$O$_2$ and iron oxide surface area concentrations in our slurries suggests that it will be possible to predict organic compound
oxidation rates under a wide variety of conditions.

Nevertheless, application of our model to previous studies showed mixed results in its ability to make accurate predictions. The main reason for this may have been that, in most of these cases, the amount and reactivity of the iron oxide present was not precisely known (and in the case of natural solids, difficult to determine). Instead of trying to quantify the surface area and type of iron oxide present in a natural solid, an easier and probably more precise approach would be to measure $V_{\text{OH}}$ at one $[\text{H}_2\text{O}_2]$ and solid concentration to find the proportionality factor $M$ in the general relationship:

$$V_{\text{OH}} = M[\text{H}_2\text{O}_2][\text{solid concentration, e.g., g of solid/L}]$$

(24)

This approach should be effective for a wide range of concentrations of the organic compound, $\text{H}_2\text{O}_2$, and solid and would have accurately predicted the results of our experiments with the Georgetown sand, except at very low concentrations of $\text{H}_2\text{O}_2$.

More work is required to determine if other factors complicate our attempts to formulate a generally applicable model. Some compounds associated with natural solid material could act as the main radical sink. If this phenomenon is generally observed, then the importance of this mechanism will also need to be quantified experimentally for each material. It is conceivable that organic compounds and other solutes can affect $V_{\text{OH}}$ via unknown mechanisms. Unlike most of the other studies we attempted to model, our data were obtained in systems containing no other solutes besides $\text{H}_2\text{O}_2$ and a probe compound, formic acid, that is highly water-soluble, does not complex iron, or sorb onto iron oxides and whose oxidation product, $\text{CO}_2$, also has these properties. If a wide range of solutes, especially the varied and poorly characterized oxidation products of organic contaminants, are found to significantly affect $V_{\text{OH}}$, then our ability to make general predictions will clearly be limited.

Further work is also needed to better understand several other important factors that control the oxidation rate of organic compounds. In cases where sorption is significant, we must consider the possibilities that the desorption rate could control the aqueous concentrations of the organic compound and that it could be oxidized at the surface or in the surface boundary layer at rates different from those in the bulk solution. In cases where $\text{H}_2\text{O}_2$ degradation is not slow as compared to the time needed for decomposition of the organic compound, for example, in soils that have rapid, nonradical-producing $\text{H}_2\text{O}_2$ sinks (18), accurate predictions of $\text{H}_2\text{O}_2$ decomposition rates will be necessary for successful application of our model.
Acknowledgments

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Literature Cited

Table 3-1. Relevant information from the publications used in Figure 3-4.

<table>
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<tr>
<th>Ref</th>
<th>[Probe]₀ (µM)</th>
<th>(k_{OH, \text{compound}}) (10⁻⁹ M⁻¹ s⁻¹)</th>
<th>([H_2O_2]₀) (mM)</th>
<th>pH</th>
<th>Catalyst</th>
<th>Surface area conc. (m²/l)</th>
<th>Solutes</th>
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<td>Nitrobenzene: 3000</td>
<td>3.9</td>
<td>32, 320, 640, 1600</td>
<td>3</td>
<td>Hematite</td>
<td>9.6*</td>
<td>0.3 M 2-propanol, &gt; 1 mM SO₄²⁻</td>
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<tr>
<td>16</td>
<td>Nitrobenzene: 1.5</td>
<td>3.9</td>
<td>2.94</td>
<td>4-7</td>
<td>Goethite-silica sand</td>
<td>24.3**</td>
<td>30 µM 1-octanol, &lt; 1 mM SO₄²⁻</td>
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<td>4POBN†: 665, 490</td>
<td>3.8</td>
<td>140, 130</td>
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<td>‡‡</td>
<td>N/A</td>
<td>0.040</td>
<td>4-5‡</td>
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*Estimated using the surface area value from ref 13.
**Estimated using the surface area value from ref 34.
***Estimated using the typical range of surface area for goethite (8-200 m²/g) and hematite (2-90 m²/g) (29).
†α-(4-pyridyl-1-oxide)-n-tert-butyl nitro barne
‡‡Petigara et al. used excess dimethyl sulfoxide to react with •OH to make a methyl radical that is trapped by 3-amino-2,2,5,5-tetramethyl-1-pyrrolidinoyloxy to form a stable O-methylhydroxylamine.
‡Buffering to pH 7.4 with 0.1 M phosphate had no effect on \(V_{OH}\).
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<th>Ref</th>
<th>Figure(s) examined</th>
<th>[Compound]₀ (µM)</th>
<th>k₌°, compound (10⁻⁹ M⁻¹ s⁻¹)</th>
<th>[H₂O₂]₀ (mM)</th>
<th>pH</th>
<th>Catalyst</th>
<th>Surface area conc. (m²/l)</th>
<th>Solutes</th>
</tr>
</thead>
<tbody>
<tr>
<td>36</td>
<td>7, all points</td>
<td>Quinoline: 81</td>
<td>9.28</td>
<td>54.0</td>
<td>7.8</td>
<td>Sandy Aquifer Material</td>
<td>3.8 - 96**</td>
<td>5 mM NaHCO₃</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>(55.2, 3)</td>
<td>3 mM Na₂SO₄</td>
<td>2 mM CaCl₂</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1 mM NaCl</td>
<td></td>
</tr>
<tr>
<td>8</td>
<td>1, all points†</td>
<td>Butyl chloride: 80</td>
<td>3.4</td>
<td>1.3, 2.2, 5.0, 10</td>
<td>7.5</td>
<td>Goethite</td>
<td>4 – 100**</td>
<td>Not reported</td>
</tr>
<tr>
<td>11</td>
<td>4b and 4c, 0-500 min</td>
<td>Quinoline: 78</td>
<td>9.28</td>
<td>14.7</td>
<td>7.0</td>
<td>Crystalline iron oxide, goethite</td>
<td>55.2, 146.8</td>
<td>5 mM NaHCO₃</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>3 mM Na₂SO₄</td>
<td>2 mM CaCl₂</td>
</tr>
<tr>
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<td></td>
<td></td>
<td></td>
<td></td>
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<td></td>
<td>1 mM NaCl</td>
<td></td>
</tr>
<tr>
<td>12</td>
<td>2a, all points; 3b, all points prior to the turning point</td>
<td>Benzoic acid: 950, 877</td>
<td>pH 3.2: 4.4</td>
<td>pH 6, 10: 5.9, 23.5, 47.06</td>
<td>3.2, 6, 10</td>
<td>Lepidocrocite on a brick grain support</td>
<td>65.3</td>
<td>Not reported</td>
</tr>
<tr>
<td>37</td>
<td>6, all time points</td>
<td>Phenol: 110</td>
<td>6.60</td>
<td>58</td>
<td>7.6</td>
<td>Natural metal oxide-coated materials</td>
<td>2 – 52, 5 – 130, 10 – 260**</td>
<td>5 mM NaHCO₃</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>3 mM Na₂SO₄</td>
<td>2 mM CaCl₂</td>
</tr>
<tr>
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<td></td>
<td></td>
<td></td>
<td></td>
<td>1 mM NaCl</td>
<td></td>
</tr>
<tr>
<td>19</td>
<td>Table 2</td>
<td>4POBN: 1060, 2CP: 320, 350, 510, 530</td>
<td>4POBN: 3.8, 2CP: 12</td>
<td>140</td>
<td>6.0</td>
<td>Goethite-silica sand</td>
<td>110***</td>
<td>0.25 mM SO₄²⁻</td>
</tr>
</tbody>
</table>

*Some figures in some of the references were not evaluated due to complexities involving, for example, phosphate or humic acid.

**Estimated using the typical range of surface area for goethite (8-200 m²/g) (29).

***Estimated using the surface area value from ref 17.
To account for the significant sorptive loss of butyl chloride (~35%), we calculated the decomposition rate of butyl chloride by subtracting the amount lost due to sorption from the concentration that remained at each time point.

*2-Chlorophenol
Figure 3-1a. $V_{OH}$ of three different iron oxides plotted as a function of the amount of surface area and hydrogen peroxide at pH 4. Ferrihydrite concentrations varied from 6-400 µM, with $[H_2O_2]_0 \leq 1$ mM (●) and $\geq 5$ mM (○); goethite concentrations of 0.02 (◇), 0.2 (◆), and 0.6 g/L (◇), and hematite concentrations of 0.2 (▲) and 1 g/L (△) are also shown. The initial concentration of formic acid for all experiments was 74-244 nM. The lines are regression fits of the data to eq 10-12.
Figure 3-1b. Decomposition rate of formic acid in the experiments described in Figure 3-1a (same symbols), normalized to the concentration of total Fe and plotted as a function of the initial amount of hydrogen peroxide. The lines are the expected values based on the regressions.
Figure 3-2a. $V_{OH}$ from experiments containing 0.6 g/L of goethite with either $H_2O_2$ ([HCOOH]$_T, o = 100$ nM (◇) or 50 μM (●)) or formic acid ([HCOOH]$_T, o = 50$ μM (□), 0.3 mM (■) or 10 mM (▼)) as the dominant •OH sink at pH 4. The line is the predicted $V_{OH}$ calculated from eq 10.
Figure 3-2b. Decomposition rate of formic acid in the experiments described in Figure 3-2a (same symbols), normalized to the concentration of total Fe and plotted as a function of the initial amount of hydrogen peroxide. The data for $[\text{HCOOH}]_{T,0} = 100 \text{ nM}$ is the same as Figure 3-1b. The lines are the expected $(d[\text{HCOOH}]_T/dt)/[\text{Fe}]_T$ for each starting amount of formic acid, given the calculated $V_{\text{OH}}$ shown in Figure 3-2a.
Figure 3-3a. $V_{OH}$ from experiments at pH 4 with $H_2O_2$ as the dominant •OH sink and either 25 (●) or 60 g/L (△) of sand and with formic acid as the dominant •OH sink and 60 g/L of sand (■). We assumed that all of the crystalline surface iron in the sand was goethite, and plotted all of the data points using a surface area of 40 m$^2$/g, with a range of 8-200 m$^2$/g depicted by the error bars. The line is the predicted $V_{OH}$ calculated from eq 10.
Figure 3-3b. Decomposition rate of formic acid in the experiments described in Figure 3-3a (same symbols), normalized to the concentration of total Fe and plotted as a function of the initial amount of hydrogen peroxide. The two sets of dashed lines represent the possible range of predicted values for the two systems, assuming that all of the crystalline iron in the sand was goethite, with a surface area ranging from 8 to 200 m$^2$/g.
Figure 3-4. Comparison of measured $V_{\text{OH}}$ from refs 9 (●), 16 (▲), 17 (■), and 18 (◇), versus our predictions. The line represents perfect agreement between predicted and measured $V_{\text{OH}}$ values. Petigara et al. (18) did not have measurements of the surface area of their iron oxide because they used natural soils. Thus, we estimated the $V_{\text{OH}}$ values using typical ranges of surface areas for goethite and hematite and presented the results with error bars.
Figure 3-5. Comparison of the measured pseudo-first-order loss rate of various organic compounds, obtained from refs 36 (▼), 8 (○), 11 (▽), 12 (●), 37 (□), and 19 (■), versus our predictions. The line represents perfect agreement between predicted and measured $k_{org}$ values. The error bars show the range of estimated $k_{org}$ values using typical ranges of surface areas for goethite in studies where the surface area was not measured.
Chapter 4: Influence of Electrostatics on the Oxidation Rates of Organic Compounds in Heterogeneous Fenton Systems

or

- There is electricity in the air... and on the iron oxide-
Abstract

The objective of this work was to refine our model for predicting the pseudo-first-order oxidation rate coefficient of any dissolved organic compound ($k_{org}$) in systems where hydroxyl radicals ($\cdot$OH) are produced by the iron oxide-catalyzed decomposition of hydrogen peroxide ($H_2O_2$). We investigated three hypotheses that could explain why our model overpredicted many of the $k_{org}$ values that other investigators observed in systems containing iron oxide and $H_2O_2$: (1) there is a substantial difference in reactivity between our laboratory synthesized iron oxide and natural ores, (2) the electrostatic attraction between formate anions and the positively charged oxide surface caused $^{14}$C-labeled formic acid, our $\cdot$OH probe, to detect a greater concentration of $\cdot$OH than the amount that reacts with neutral compounds, and (3) the organic compound being degraded or its oxidation products interfered with the $\cdot$OH generation process at the oxide surface. Results from various experiments at pH 4 containing goethite, $H^{14}$COOH, and 2-Chlorophenol (2-CP) showed that our goethite had similar reactivity as natural goethite, and that electrostatic effects were important because $H^{14}$COOH detected more $\cdot$OH than 2-CP did in the same system. We also observed that the concentrations of $\cdot$OH measured by $H^{14}$COOH decreased with time, whereas the amount of $\cdot$OH detected by 2-CP remained constant. This was attributed to the loss of positive charge on the oxide surface as the oxidation products of 2-CP complexed surface Fe species. There was no evidence that the oxidation products of 2-CP interfered with the $\cdot$OH generation process at the concentrations of goethite that we used.

Introduction

The hydroxyl radical ($\cdot$OH) reacts rapidly with numerous compounds (1) and is the key chemical species in many advanced oxidation processes to treat wastewater and polluted aquifers. There are various ways to create $\cdot$OH in solution (2-4), including the use of iron oxide and hydrogen peroxide ($H_2O_2$). Although the iron oxide catalyzed mechanism is not yet fully understood, it most likely contains these reactions,

\[
\begin{align*}
&=Fe^{III} + H_2O_2 = \equivFe^{III}H_2O_2 \quad (1) \\
&\equivFe^{III}H_2O_2 \rightarrow \equivFe^{II} + HO_2 + H^+ \quad (2) \\
&\equivFe^{II} + H_2O_2 \rightarrow \equivFe^{III} + \cdotOH + OH^- \quad (3)
\end{align*}
\]

where $\equivFe$ is a surface iron species. This system has been researched recently (5-14) because it can effectively catalyze the oxidation of different organic contaminants at circumneutral pH, whereas Fenton’s reagent ($Fe(II) + H_2O_2$) is typically applied at low pH to ensure the Fe remains
dissolved throughout the entire remediation process. Therefore, using iron oxide instead of dissolved iron to treat contaminated aquifers would be advantageous when it would be difficult or destructive to lower the pH of the groundwater.

While there has been much research on the mechanism and factors that affect hydrogen peroxide and organic compound decomposition in the presence of iron oxide (4, 5, 7-10, 12, 14-18), none of them adequately answered the need for a general model that can predict the oxidation rate of any organic compound ($k_{\text{org}}$) in this system. We addressed this issue in a previous work (14) by creating a model that predicts the oxidation rate of any dissolved compound in an iron oxide/H$_2$O$_2$ system based on the generation rate of •OH ($V_{\text{OH}}$) and the reaction rate constants of the system’s constituents with •OH. The model uses the following parameters to estimate the pseudo-first-order rate coefficient of oxidation of an organic compound: the surface area, type and amount of iron oxide, the concentrations of H$_2$O$_2$, organic compound and other important •OH sinks in the system, and their respective second-order rates with •OH. It successfully predicted the degradation rate of formic acid that we measured in sand from Georgetown at pH 4, but overpredicted many of the $k_{\text{org}}$ values reported by other investigators who examined contaminant loss in iron oxide/H$_2$O$_2$ systems. We proposed potential explanations for the disagreements between our model predictions and the measured $k_{\text{org}}$ values.

The first hypothesis was that there was a difference in reactivity between our laboratory synthesized iron oxide and those obtained from natural sources. These differences could be due to, for example, the presence of impurities or soil components that were associated with the natural iron oxide. If this were the case, then it could be difficult to create a generally applicable model to estimate the oxidation rate of any organic compound in an iron oxide/H$_2$O$_2$ system since every solid could have unique characteristics. A more practical approach would be to measure $V_{\text{OH}}$ using one concentration of H$_2$O$_2$ and the solid of interest, and then apply our demonstrated linear proportionality between $V_{\text{OH}}$, [H$_2$O$_2$] and solid concentration (14) to extrapolate to conditions of interest.

The second possible explanation was electrostatics. Although we believed that this was not an important issue in our previous studies (14), we now think electrostatics could have affected the decomposition rate of formate. In this case, our predicted $k_{\text{org}}$ values for neutral compounds would have to be adjusted. When oxides or oxyhydroxides are in an aqueous
solution, the surface hydroxyl groups can undergo proton exchange with the surrounding media and possibly acquire a surface charge:

\[ \equiv \text{FeOH}_2^+ = \equiv \text{FeOH} + \text{H}^+ \]  
\[ \equiv \text{FeOH} = \equiv \text{FeO}^- + \text{H}^+ \]  

(4) \hspace{1cm} (5)

The magnitude and sign of the charge can be determined qualitatively based on the pH of the solution and the equilibrium constants of reactions 4 and 5. The goethite surface has a net positive charge at our experimental pH of 4, so formate anions would be attracted toward the oxide surface where \( * \text{OH} \) is being produced. This would cause the actual loss rate of \( \text{H}^{14}\text{COOH} \) to be greater than the value computed from bulk solution properties. On the other hand, neutral compounds are unaffected by the presence of surface charge, so a model based on formate decomposition rates could overpredict \( k_{\text{org}} \) for neutral compounds.

The third hypothesis was that the compound being degraded or its oxidation products interfered with the \( * \text{OH} \) generation process. Surface iron interacts with many functional groups, e.g., deprotonated hydroxy groups \((19, 20)\), and it could become less reactive toward \( \text{H}_2\text{O}_2 \) when it is complexed. We avoided this possibility in our experiments to quantify \( V_{\text{OH}} \) by using \( \text{H}^{14}\text{COOH} \) as our \( * \text{OH} \) probe, but many of the aromatic compounds in the studies that we modeled have oxidation products that could complex surface iron. If this caused \( V_{\text{OH}} \) to decrease with time, then the measured \( k_{\text{org}} \) would be less than the model prediction.

The goal of this work was to improve our model for predicting \( k_{\text{org}} \) by determining the factors that contributed to its overestimation, and we achieved this by testing the above conjectures. We used our goethite to reproduce the experiment done by Huling et al. \((21)\) and measured a similar loss rate for 2-Chlorophenol (2-CP), thus showing that the reactivity of our goethite was not significantly different from that of the natural goethite used by Huling and coworkers. The results from parallel experiments at pH 4 containing 2-CP and/or nitrobenzene (NB) in goethite slurries, with and without \( \text{H}^{14}\text{COOH} \), revealed that electrostatics caused \( \text{H}^{14}\text{COOH} \) to detect a greater concentration of \( * \text{OH} \) than 2-CP and NB. They also indicated that the oxidation products of 2-CP complexed surface iron, but this did not significantly affect the decomposition rate of 2-CP at the amount of iron oxide and \( \text{H}_2\text{O}_2 \) that we used.
Materials and Methods

Materials

Detailed descriptions for materials and methods are in previous publications (4, 14). All reagents were reagent grade and used as received. 2-Chlorophenol (2-CP, 99+%) and nitrobenzene (NB, 99+%) were from Sigma-Aldrich. High load scintillation cocktail (ScintiSafe Plus 50% Cocktail) was from Fisher. The solutions were prepared using 18 MΩ•cm Milli-Q water from a Millipore system. All glassware was soaked in concentrated HCl for 8 hours or more and rinsed before use. Goethite was the same as that described in chapter 3.

Analytical Techniques

2-CP and NB were analyzed by gas chromatography. At each time point a 2-mL aliquot of the experimental solution was removed and centrifuged in a glass container at 5500 rpm for 10 mins. 1-μL of the supernatant was injected into a Perkin Elmer Auto System XL gas chromatograph fitted with a quartz liner at the injector port, 0.32 mm i.d. x 60 m DB-5 1-μm film capillary column, and flame ionization detector. Chromatographic conditions were: helium carrier gas at 25 psi, injector temperature at 125 °C, detector temperature at 300 °C, oven temperature at 120 °C from 0-1 min, rate of 17 °C/min until 190 °C, 5 °C/min until final oven temperature of 205 °C, hold for 0.4 min. 14C activity was measured in a scintillation counter (14). The high load scintillation cocktail was used in those experiments involving goethite. Hydrogen peroxide was measured spectrophotometrically using the DPD method (22) as modified by Voelker and Sulzberger (23) to minimize interference by Fe(II) and Fe(III). Samples were diluted with Milli-Q water to ensure that the absorbance through a 1-cm path length cuvette was less than 1.2 au.

Experimental Setup

Experiments were done in 50-mL glass test tubes with glass stoppers. The Milli-Q water was treated with a total organic carbon reduction unit (Aquafine) to oxidize traces of organic contamination. Parallel experiments were set up so that both test tubes contained the same amounts (within weighing and measuring errors) of goethite, H2O2 and organic compound (2-CP and/or NB) in 40 mL of solution, with the difference being that one was spiked with 14C-labeled formic acid. This allowed us to track the loss rate of H14COOH (kHCOOH) in one tube and the organic compound in the other without contaminating the GC with radioactivity. The ionic strength was set at 10 mM with NaClO4, and the pH was adjusted to 4 (unless noted otherwise)
with HClO₄ and/or NaOH after the goethite and all compounds but H₂O₂ were mixed together. H₂O₂ was introduced at time zero and the contents mixed by a wrist-action shaker. The temperature of the solutions was 23.5 ± 2.5 °C. The pH of the solutions remained within ±0.5 unit of the initial pH throughout the experiment. Control experiments (without H₂O₂) lasting for 43 hours showed no significant loss of 2-CP or NB to air, iron oxide, or glass.

In one set of experiments we examined electrostatic effects by comparing k_{HCOOH} at pH 4 and pH 5. Instead of adding goethite solids directly to the solution, we added equal volumes from the supernatant of a mixture that consisted of 90 mg of goethite in 150 mL of 0.010 M NaClO₄ that stood undisturbed for 2 h. This was done to let the larger goethite particles settle out and decrease the likelihood of coagulation at the higher pH. We can apply Stoke's law to estimate the size of the particles that remained in the supernatant:

\[ \nu_s = \frac{2g(\rho_s - \rho_w)}{9\mu} \cdot r^2 \]  

where \( \nu_s \) is the settling velocity, \( \alpha \) is a nondimensional form factor and is equal to 1 for a sphere (we shall assume the goethite solids are spheres) (24), \( g \) is the acceleration due to gravity (9.81 m/s²), \( \rho_s \) is the density of the solid (4260 kg/m³ for goethite; 25), \( \rho_w \) is the density of water (1000 kg/m³), \( \mu \) is the dynamic viscosity of the fluid (10⁻³ kg/m·s), and \( r \) is the sphere radius. The height of the goethite slurry in the flask was 5 cm, so theoretically only particles with a radius smaller than 1 μm would have stayed in the supernatant.

Results

The aim of our first set of experiments was to determine if our laboratory synthesized goethite crystals were drastically different from the natural goethite that Huling et al. (21) used in their work. The experimental conditions that we used were similar to Huling’s (surface area concentration = 110 m²/L, [H₂O₂]₀ = 150 mM, and [2-CP]₀ = 350 μM) except that we worked at pH 4 instead of pH 6 to facilitate our sparging technique to remove ¹⁴CO₂ from H¹⁴COOH. The change in pH did not significantly affect 2-CP speciation, (pKₐ = 8.49; 26). We also assumed that it did not change how fast •OH was generated since three independent investigations have shown that V_{OH} is relatively invariant from pH 4 to pH 10 (16-18). Huling et al. (21) performed four experiments with [2-CP]₀ ranging from 320-530 μM and reported a 2-CP loss rate of 0.043 ± 0.003 h⁻¹, whereas we measured 0.16 ± 0.02 h⁻¹ (n = 2) in our experiments. The similar 2-CP loss rates, in stark contrast to a difference of over two orders of magnitude between Huling’s results.
and those predicted by our model (14), led us to conclude that the two types of goethite were not significantly different from each other.

If electrostatics were important in our system, then $k_{\text{HCOOH}}$ should decrease when we increase the ionic strength or pH. Raising the ionic strength compresses the thickness of the electric double layer, and increasing the pH reduces the net amount of positive charge on the iron oxide surface. In both cases, the net result is that fewer formate anions are attracted to the oxide surface where the concentration of hydroxyl radicals is the greatest. We tested this hypothesis by comparing the change in $k_{\text{HCOOH}}$ in two different sets of experiments. In the first experiment, $k_{\text{HCOOH}}$ decreased from $-1.0 \text{ h}^{-1}$ to $0.16 \text{ h}^{-1}$ when the ionic strength of a pH 4 goethite slurry was increased from 0.01 to 0.9 M. In the other experiment, $k_{\text{HCOOH}}$ decreased from 0.081 $\text{ h}^{-1}$ to 0.032 $\text{ h}^{-1}$ when the pH of the goethite slurry was increased from 4 to 5 at $I = 10 \text{ mM}$. (The $k_{\text{HCOOH}}$ values at pH 4 and $I = 10 \text{ mM}$ were different because, as mentioned previously, goethite solids were not added directly to the solution used in the second experiment.) The percentage decrease of $k_{\text{HCOOH}}$ in both cases was consistent with theoretical calculations (see Discussion).

To further assess the influence of electrostatics in the heterogeneous system, we compared the steady state concentration of $\cdot \text{OH}$ ($[\cdot \text{OH}]_{\text{ss}}$) that $\text{H}^{14}\text{COOH}$ and 2-CP measured when both were mixed with 0.6 g/L of goethite and 2 M H$_2$O$_2$. If the rate of probe degradation via its reaction with $\cdot \text{OH}$ follows pseudo-first-order kinetics, then $[\cdot \text{OH}]_{\text{ss}}$ is:

$$[\cdot \text{OH}]_{\text{ss}} = \frac{k_{\text{probe}}}{k_{\text{OH,probe}}} \tag{7}$$

where $k_{\text{probe}}$ is the measured pseudo-first-order rate coefficient of $\text{H}^{14}\text{COOH}$ or 2-CP degradation, and $k_{\text{OH,probe}}$ is the second-order rate constant of the probe with $\cdot \text{OH}$. In solutions where electrostatic effects are important, the concentration of a charged probe is a function of its distance from the iron oxide. Since there are high concentrations of both formate anions and $\cdot \text{OH}$ near the oxide surface, $k_{\text{HCOOH}}$ would be higher than expected based on the bulk solution concentrations, and $[\cdot \text{OH}]_{\text{ss}}$ calculated by eq 7 becomes a weighted average of the amount of $\cdot \text{OH}$ that the probe detected in the double layer and in the bulk solution. $[\cdot \text{OH}]_{\text{ss}}$ will be greater for $\text{H}^{14}\text{COOH}$ than for 2-CP if electrostatic effects are indeed important. Figure 4-1 shows the probe loss data from one such experiment, and whereas 2-CP loss was pseudo-first-order, $\text{H}^{14}\text{COOH}$ loss clearly was not. This was surprising as $\text{H}^{14}\text{COOH}$ degradation was always pseudo-first-order in all of our previous experiments where 2-CP was not present (14).
Since adding 2-CP to the goethite/H₂O₂ system should not have introduced a new H¹⁴COOH degradation pathway (supported by experimental results discussed at the end of this section), the logical explanation for the behavior of the H¹⁴COOH data in Figure 4-1 is that as the experiment progressed H¹⁴COOH measured decreasing [•OH]ss. In order to obtain time-resolved information on [•OH] from the H¹⁴COOH data, we calculated the kprobe values of two consecutive time points assuming kprobe remained constant over the time interval:

$$k_{probe, interval} = -\ln \left[ \frac{[probe]_{t}}{[probe]_{t'}} \right] \left( t'-t \right)^{-1}$$  \hspace{1cm} (8)

where t’ was the next time after t at which the amount of probe compound remaining in solution was determined. k_{probe, interval} should approximate kHCOOH for the time interval as long as the difference between t’ and t is small. We then substituted k_{probe, interval} for kprobe in eq 7 to get [•OH]ss, interval values for each interval (Figures 4-2a-c, negative [•OH]ss, interval numbers were excluded). The results showed that (a) early on, [•OH]ss, interval values for H¹⁴COOH were higher than those of 2-CP and NB, (NB is a well-known •OH probe that was used to confirm the [•OH]ss, interval measurements made using 2-CP) and (b) [•OH]ss, interval values for H¹⁴COOH decreased steadily until at approximately 5 hours, they were nearly equivalent to those of 2-CP and NB. As expected from the data in Figure 4-1, 2-CP detected the same amount of •OH in every time interval (within experimental uncertainty).

The non first-order behavior of kHCOOH suggested that we should examine how 2-CP (or NB) or its oxidation products caused the [•OH]ss, interval measurements for H¹⁴COOH to decrease with time. We did parallel experiments where 0.3, 3, 30 or 300 μM of 2-CP was added to 0.6 g/L of goethite and 2 M of H₂O₂, then computed [•OH]ss, interval from the H¹⁴COOH data (Figures 4-3a,b). The results strengthened the idea that 2-CP or its oxidation products affected the degradation kinetics of H¹⁴COOH because the rate at which the [•OH]ss, interval values measured using H¹⁴COOH decreased was a function of [2-CP]₀. There was also noticeable coagulation of the goethite particles in those experiments where [2-CP]₀ ≥ 30 μM (Figures 4-4a,b). We believe these two observations are due to the loss of positive charge on the oxide surface as the intermediates and oxidation products of 2-CP (e.g., chlorocatechol, reaction 9) complexed surface Fe species.
We did not determine the reaction products from our 2-CP experiments. Although there is no direct evidence that 2-Chlorocatechol is formed, 4-Chlorocatechol is a known intermediate of the reaction between •OH and 4-Chlorophenol (27-32), and a study of 2-CP degradation in an irradiated aqueous suspension of TiO$_2$ detected eight intermediates, of which only chlorohydroquinone, hydroxyhydroquinone, and catechol were identified (33). We did not observe any oxidative coupling products, i.e., polyphenols (32, 34), in our suspensions.

Coagulation alone cannot explain the results of Figures 4-2a-c because it leads to the loss of surface area, which should decrease the oxidation rates of both H$^{14}$COOH and 2-CP by the same amount. It may be that not enough surface area was lost to significantly affect the oxidation rates of H$^{14}$COOH and 2-CP.

By the same token, any decrease of Fe reactivity toward H$_2$O$_2$ caused by the oxidation products of 2-CP also cannot explain the results of Figures 4-2a-c because in that case, [•OH]$_{ss, interval}$ measured by both H$^{14}$COOH and 2-CP would be expected to decrease with time. We confirmed this by doing two experiments at pH 3 where 25 and 75 μM of Fe was introduced as dissolved Fe(III) (in the form of Fe(ClO$_4$)$_3$·9H$_2$O) instead of goethite into solutions containing 2 M H$_2$O$_2$ and 200 μM 2-CP (Figures 4-5a,b). Figures 4-6a,b show that both probes measured the same concentration of •OH (within experimental uncertainty), confirming that adding 2-CP to our experiments did not introduce a new H$^{14}$COOH degradation pathway. Moreover, Fe(III) interacted with the oxidation products of 2-CP and became less reactive toward H$_2$O$_2$, as evidenced by the decrease in [•OH]$_{ss, interval}$ for both H$^{14}$COOH and 2-CP with time, in contrast to the data in Figures 4-2a-c. These results suggest that the reactivity of surface Fe toward H$_2$O$_2$ could also decrease when the Fe is complexed by the oxidation products of 2-CP, but as discussed above, we did not observe this in our experiments. Most likely, the number of surface Fe sites that were complexed was too low to adversely affect the production rate of •OH. Nevertheless, the loss of some of the charged surface Fe sites, which make up only a small fraction of the total surface Fe sites, to complexation could still have a substantial effect on the surface charge, causing the observed effects: a decrease in $k_{HCOOH}$ and coagulation.
Discussion

The goal of this work was to improve our model for predicting the oxidation rate of organic compounds in mineral-catalyzed, Fenton-like systems. The model successfully predicted the decomposition rate of H$^{14}$COOH in slurries containing iron oxide and a natural iron oxide-coated quartzitic aquifer sand, but overpredicted many of the $k_{org}$ values that other researchers reported (14). We tested three hypotheses that could explain the disagreements between the model predictions and the measurements, and concluded that electrostatics, which was not considered in our model, played a crucial role by enriching the concentration of formate near the positively charged iron oxide surface where $\cdot$OH is generated.

Lifetime of $\cdot$OH

One method of judging the possible influence of electrostatics on $k_{HCOOH}$ is to compute the lifetime of $\cdot$OH. If $\cdot$OH is relatively long-lived, then it can diffuse far away from the oxide surface. This would imply that $k_{HCOOH}$ is determined by the solute concentrations in the bulk solution, which are unaffected by electrostatics. The lifetime of a chemical species ($\tau$) that is consumed according to first-order kinetics is equal to the inverse of the sum of the product of the concentration of all chemical species that react with it and their respective second-order reaction rate constants:

$$\tau = \frac{1}{\sum k_i [S_i]}$$

$\tau_{OH}$ is estimated to be ~20 ns in a solution containing 2 M H$_2$O$_2$, 200 $\mu$M 2-CP, with $k_{OH,H_2O_2} = 2.7 \times 10^7$ M$^{-1}$ s$^{-1}$ (1) and $k_{OH,2-CP} = 1.2 \times 10^{10}$ M$^{-1}$ s$^{-1}$ (1). The double layer thickness is the reciprocal of the Debye parameter, which at 25 °C is equal to $3.29 \times 10^9$ 1$^{1/2}$ (35), and is ~3 nm at $I = 10$ mM. The typical time for a molecule to diffuse across this boundary is 9 ns, calculated by squaring the length of the double layer and dividing by $10^{-5}$ cm$^2$/s, the diffusivity of a solute (24). As $\tau_{OH}$ is only twice the time needed to travel across the double layer, it agrees with our theory that $k_{HCOOH}$ was significantly affected by electrostatics.

Quantifying Electrostatic Effects

To further determine how electrostatic effects affected our system, we needed a method of predicting how $k_{HCOOH}$ changes as a result of the migration of formate anions toward the positively charged iron oxide surface. It has been shown that the Coulombic correction factor, $\exp(-\Delta ZFP\phi/RT)$, can quantify the electrostatic interactions between solutes and solid surfaces in
solution, i.e., the change in the concentration of charged solutes at the oxide surface caused by the surface charge (35). $\Delta Z$ is the change in the charge of the surface species for the reaction under consideration, $F$ is the Faraday constant (96485 C/mol), $\Psi_0$ is the surface potential, $R$ is the gas constant (8.314 J/K·mol), and $T$ is the temperature. $\Psi_0$ determines the strength of the attraction or repulsion between the ions and the oxide surface and is given by (24):

$$\Psi_0 = \frac{2RT}{zF} \sinh^{-1} \left( \frac{\sqrt{\pi F^2 T^3}}{2eRT} \right)$$  \hspace{1cm} (11)

where $z$ is the valence of the background electrolyte and is equal to 1 in our experiments, $\varepsilon$ is the dielectric constant of water ($7.2 \times 10^{-10}$ C/V-m), and $I$ is the ionic strength. $\sigma$ in eq 11 is the surface charge density, which is the difference between $[\equiv \text{FeOH}^+]$ and $[\equiv \text{FeO}^-]$: 

$$\sigma = \frac{[\equiv \text{FeOH}][H^+]}{K_{a1}^{\text{int}}} e^{-\Psi_0 F \sigma R T} - \frac{[\equiv \text{FeOH}][H^+]}{K_{a2}^{\text{int}}} e^{-\Psi_0 F \sigma R T}$$  \hspace{1cm} (12)

where $[\equiv \text{FeOH}]$ is the density of the reactive surface Fe sites, and $K_{a1}^{\text{int}}$ and $K_{a2}^{\text{int}}$ are the intrinsic surface acid-base equilibrium constants of reactions 4 and 5, respectively. The Coulombic correction factor is required for $H^+$ since we are dealing with surface Fe species and thus need the concentration of protons at the surface. We assumed our goethite had a $pK_{a1}^{\text{int}}$ of 6, a $pK_{a2}^{\text{int}}$ of 9, and a site density of $2 \times 10^{-6}$ mol/m$^2$ (24) because we did not measure these parameters.

Table 4-1 lists $\Psi_0$ of the oxide surface in the experiments that were done to assess the importance of electrostatic effects in the goethite/H$_2$O$_2$ system.

With $\Psi_0$ known, we can predict how $k_{\text{HCOOH}}$ changes with ionic strength or pH. Generally speaking, the pseudo-first-order rate constant that we measure, $k_{\text{HCOOH}}$, is a weighted sum of the rates at which $H^1\text{COOH}$ is oxidized in the bulk solution and near the surface:

$$k_{\text{HCOOH}} = k_{\text{HCOOH,bulk}} \left( \frac{\text{vol}_{\text{bulk}}}{\text{total vol}} \right) + k_{\text{HCOOH,surf}} \left( \frac{\text{vol}_{\text{surf}}}{\text{total vol}} \right)$$  \hspace{1cm} (13)

where "bulk" designates the bulk solution concentration of the solutes, "surf" denotes the surface concentration of the solutes, and the volume fractions weigh the contribution from each region. For electrostatic effects to be important, the contribution from the bulk region must be much less than the surface region, so as a first approximation:

$$k_{\text{HCOOH}} \approx k_{\text{HCOOH,surf}} \left( \frac{\text{vol}_{\text{surf}}}{\text{total vol}} \right)$$  \hspace{1cm} (14)
Using eq 7 to substitute for $k_{HCOOH,\text{surf}}$,

$$k_{\text{HCOOH}} = k_{OH,\text{HCOOH, surf}} [\cdot OH]_{ss, \text{surf}} \left( \frac{\text{vol}}{\text{surf}} \right)$$

which shows that the ratio of $k_{\text{HCOOH}}$ measured from two experiments is the same as the ratio of $k_{OH,\text{HCOOH, surf}}$ for those two experiments as long as the perturbations to $[\cdot OH]_{ss, \text{surf}}$ and the volume fraction are minor. $k_{OH,\text{HCOOH, surf}}$ is equal to:

$$k_{OH,\text{HCOOH, surf}} = k_{OH,\text{HCOOH}} \frac{[\text{HCOOH}]_{\text{bulk}}}{[\text{HCOOH}]_{T}} + k_{OH,\text{HCOOH}, -}\frac{[\text{HCOO}^-]_{\text{bulk}}}{[\text{HCOOH}]_{T}} e^{\frac{\Psi_0}{RT}}$$

where the Coulombic factor corrects for the amount of formate present at the oxide surface and can be computed once $\Psi_0$ is known. Although $pK_{a1}^{\text{int}}$ for goethite ranges from 6.0-7.47 (neglecting one study reporting it as 4.2) (25), $pK_{a2}^{\text{int}}$ ranges from 9.0-11.11 (25), and the site density ranges from 2-4.7 x 10^{-6} mol/m^2 (35), our calculations indicated that the change in $\Psi_0$ at pH 4 due to these variations was less than 0.05 V and more importantly, the ratio of $k_{OH,\text{HCOOH, surf}}$ was unaffected. We predicted $k_{OH,\text{HCOOH, surf}}$ would decrease by a factor of 6.9 when the ionic strength was raised from 10 to 900 mM, and the measured decrease of $k_{\text{HCOOH}}$ was by a factor of 6.3. $k_{\text{HCOOH}}$ decreased by a factor of 2.5 in the experiment where the pH was raised from 4 to 5, which was close to the predicted decrease of a factor of 3.2. The good agreement between the measurements and predictions for both experiments support the hypothesis that electrostatics affected $k_{\text{HCOOH}}$.

Ionic strength dictates the extent of interactions between charged components in solution. The theoretical calculations that we did accounted for the change in the width of the double layer and the influence of surface charge on the distribution of ions in solution. However, they do not consider the change in the activity coefficient within the localized environment of the double layer, or that excess counterions restrict the number of ions that can approach the surface. More experiments would help elucidate the relationship between ionic strength and $k_{\text{HCOOH}}$.

**Implications For Our Model**

In order to improve the ability of our model to predict the $k_{\text{org}}$ of dissolved neutral compounds, we need a method to rectify the higher oxidation rate of H^{14}COOH so that it is the same as that of any dissolved neutral compound. While the previous evaluations were useful for confirming the importance of electrostatics in our system, neither of them met the requirement.
The full analysis involves examining how the consumption rate of $\text{H}^{14}\text{COOH}$ is affected by $\Psi$ and $[\cdot\text{OH}]_{ss}$, both of which decrease with distance away from the iron oxide:

\[
\left( -\frac{d[H\text{COOH}]}{dt} \right)_{el} = \frac{A}{V} \int_0^\infty \left( k_{OH,H\text{COOH}}[\cdot\text{OH}]_{ss}(x)[H\text{COOH}]_{bulk} + k_{OH,H\text{COOH}}[\cdot\text{OH}]_{ss}(x)[H\text{COOH}]_{bulk} e^{-\Delta\Psi(x)/RT} \right) dx \tag{17}
\]

where "el" denotes electrostatic effects, $A$ is the total amount of iron oxide surface area in solution, and $V$ is the volume of the solution. $\Psi(x)$ can be approximated by $\Psi_0 e^{-\kappa x} (35)$, where $\kappa$ is the Debye parameter. We can determine how $\cdot\text{OH}$ changes with distance by applying the law of mole conservation, which says that at steady state the mole inflow rate plus any gains into a control volume must equal the mole outflow rate and any losses within the control volume. The resulting equation for our system is:

\[
D \frac{\partial^2 [\cdot\text{OH}]_{ss}(x)}{\partial x^2} = \sum k_{OH,i}[\cdot\text{OH}]_{ss}(x)[S_i](x) \tag{18}
\]

where $D$ is the diffusion coefficient, and $S_i$ represents $\cdot\text{OH}$ sink $i$ in solution. The boundary conditions are:

\[
- D \frac{\partial [\cdot\text{OH}]_{ss}}{\partial x} \bigg|_{x=0} = \tilde{V}_{OH} \tag{19}
\]

and $[\cdot\text{OH}]_{ss} = 0$ at an infinite distance away from the oxide surface. Eq 19 is Fick's law and describes the influx of $\cdot\text{OH}$, assuming it is generated only at the oxide surface ($x = 0$). $\tilde{V}_{OH}$ is the mole flux per unit area per unit time (equivalent to $V_{OH}$ multiplied by a unit length). If $[S_i]$ is not a function of $x$, then the solution to eq 18 is:

\[
[\cdot\text{OH}]_{ss}(x) = \frac{\tilde{V}_{OH}}{\sqrt{D \sum k_{OH,i}[S_i]}} \exp \left( -x \sqrt{\frac{\sum k_{OH,i}[S_i]}{D}} \right) \tag{20}
\]

We can now refine our model for predicting $k_{org}$ by correcting for the percent increase in the loss rate of formate in the double layer due to electrostatic effects. This was achieved by dividing the increased oxidation rate of $\text{H}^{14}\text{COOH}$ in the double layer due to formate enrichment (integrating eq 17 from 0 to infinity) by the oxidation rate of $\text{H}^{14}\text{COOH}$ in which electrostatic effects are ignored (integrating eq 21 from 0 to infinity).
We assumed the major sink of $\cdot$OH was $\text{H}_2\text{O}_2$, which was true in the experiments that we did to quantify $V_{\cdot\text{OH}}$ using $\text{H}^{14}\text{COOH}$ (Figure 3-1a), and used numerical integration (adaptive Simpson quadrature, adaptive Lobatto quadrature, and Clenshaw-Curtis quadrature) to integrate eq 17 because it does not appear to have an analytical solution. The quotient is approximately proportional to $[\text{H}_2\text{O}_2]^{1/2}$, as expected given the behavior of $[\cdot\text{OH}]_{ss}(x)$. It is $\sim 58$ at pH 4, $I = 10$ mM, $\Psi_0 = 0.16$ V, $[\text{H}_2\text{O}_2]_0 = 2$ M, and a total $\text{H}^{14}\text{COOH}$ concentration of 100 nM, which agrees with the difference in $[\cdot\text{OH}]_{ss,interval}$ observed in Figures 4-2a-c at the beginning of the experiment. Using this information, we divided the predicted $k_{\text{org}}$ values shown in Figure 3-5 by 58 (Figure 4-7). All of the model predictions now are within a factor of 10 of the measurements, even the butyl chloride experiments done by Lin and Gurol (16).

We re-examined the benzoic acid data ($pK_a = 4.19$; 26) for electrostatic effects because those experiments were done at pH 3.2, 6, and 10 (black circles in Figure 4-7). Using the above approach and assuming that lepidocrocite has the same surface charge properties as goethite, we would expect the oxidation rate of benzoic acid, compared to a neutral probe compound under the same conditions, to increase by a factor of 3 at pH 3.2, be approximately the same at pH 6, and decrease by a factor of 2 at pH 10. These correction factors do not have a significant impact on the agreement between the predicted and measured $k_{\text{org}}$.

Most researchers do not consider the effects of electrostatic on the oxidation rates of organic compounds in heterogeneous systems because they deal with pollutants that have $pK_a$ values of 7 or greater. Nevertheless, we showed that electrostatics was the most probable explanation for the discrepancy between our model predictions and the $k_{\text{org}}$ values reported by other researchers because $\text{H}^{14}\text{COOH}$ measured a concentration of $\cdot$OH that was much greater than the amount that reacted with neutral organic compounds. The difference between the two $[\cdot\text{OH}]_{ss}$ was similar to the difference between the predicted and measured $k_{\text{org}}$. Since some organic compounds, for example benzoic acid, have charged species at pH values of 7 or less, it is important to know that electrostatics affects the interactions between organic compounds and iron oxide. Electrostatics could also influence the concentration of any charged $\cdot$OH sinks in the boundary layer, which would impact the oxidation rate of the compound of interest if the charged sink were an important sink of $\cdot$OH.
Acknowledgments

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Literature Cited

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Table 4-1. Effect of ionic strength and pH on $\Psi_0$.

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<th>I (mM)</th>
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<th>$\Psi_0$ (V)</th>
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Figure 4-1. Typical data set of the concentration of $^{14}$C-labeled formic acid (●) and 2-CP (△) in an experiment containing 0.6 g/L goethite, $[\text{H}_2\text{O}_2]_0 = 2$ M, $[2$-CP$]_0 = 300$ µM, and initial $^{14}$C of 10000 DPM at pH 4. The lines are linear regression fits to log transformed data.
Figure 4-2a. [•OH]_{ss, interval} for $^{14}$C-labeled formic acid (●, ○) and 2-CP (▼, ▽) in experiments containing 0.6 g/L goethite, $[\text{H}_2\text{O}_2]_0 = 2 \text{ M}$, initial $^{14}$C of 20000 (●) or 10000 (○)DPM, and $[\text{2-CP}]_0 = 130$ (▼) or 300 (▽) μM at pH 4. The horizontal bars denote the time interval between the two consecutive data points that were used to calculate $[\cdot\text{OH}]_{ss, interval}$. 
Figure 4-2b. $[\cdot\text{OH}]_{\text{ss, interval}} \times 10^{15} (\text{M})$ for $^{14}\text{C}$-labeled formic acid (●, ○) and NB(■, □) in experiments containing 0.6 g/L goethite, $[\text{H}_2\text{O}_2]_0 = 2 \text{ M}$, $[\text{NB}]_0 = 100 \mu\text{M}$, and initial $^{14}\text{C}$ of 40000 (●) or 10000 (○) DPM at pH 4. The horizontal bars denote the time interval between the two consecutive data points that were used to calculate $[\cdot\text{OH}]_{\text{ss, interval}}$. 
Figure 4-2c. $[^14]{\text{OH}}_{ss, \text{interval}}$, interval for $^{14}$C-labeled formic acid (●, ○), 2-CP (▼, ▽), and NB (■, □) in experiments containing 0.6 g/L goethite, $[\text{H}_2\text{O}_2]_o = 2$ M, $[2-\text{CP}]_o = [\text{NB}]_o = 100$ μM, and initial $^{14}$C of 40000 (●) or 20000 (○) DPM at pH 4. The horizontal bars denote the time interval between the two consecutive data points that were used to calculate $[^*]{\text{OH}}_{ss, \text{interval}}$. 
Figure 4-3a. $^{14}$C activity of H$^{14}$COOH in four experiments containing 0.6 g/L goethite, $[H_2O_2]_0 = 2$ M, and $[2$-CP$]_0 = 0.3$ (●), 3 (○), 30 (◆), or 300 (◇) µM at pH 4. The experiments with [2-CP]$_0 = 0.3$ and 3 µM were re-spiked with H$^{14}$COOH after ~20 hours.
Figure 4-3b. \([\cdot \text{OH}]_{\text{ss, interval}}\) for the data shown in Figure 4-3a. The horizontal bars denote the time interval between the two consecutive data points that were used to calculate \([\cdot \text{OH}]_{\text{ss, interval}}\).
Figure 4-4a. Photos of two test tubes containing 0.6 g/L of goethite, $[\text{H}_2\text{O}_2]_0 = 2$ M and $[\text{2-CP}]_0 = 3$ (left) or 300 (right) μM at pH 4. (A) was taken immediately after the test tubes had been shaking for 1.5 hours. They were then left undisturbed for 15 minutes before (B) was taken.
Figure 4-4b. Continuation of the experiment described in Figure 4-4a. (A) was taken immediately after the test tubes had been shaking for 10 hours. They were then left undisturbed for 15 minutes before (B) was taken.
Figure 4-5a. $^{14}$C activity of H$^{14}$COOH (●) and [2-CP] (▽) in an experiment with [Fe(III)] = 25 μM, [H$_2$O$_2$]$_0$ = 2 M, and [2-CP]$_0$ = 170 μM at pH 3.
Figure 4-5b. $^{14}$C activity of H$^{14}$COOH (●) and [2-CP] (▽) in an experiment with [Fe(III)] = 75 μM, [H$_2$O$_2$]$_0$ = 2 M, and [2-CP]$_0$ = 200 μM at pH 3.
Figure 4-6a. \([\cdot \text{OH}]_{\text{ss, interval}}\) for the data shown in Figure 4-5a. The horizontal bars denote the time interval between the two consecutive data points that were used to calculate \([\cdot \text{OH}]_{\text{ss, interval}}\).
Figure 4-6b. $[\cdot \text{OH}]_{\text{ss, interval}}$ for the data shown in Figure 4-5b. The horizontal bars denote the time interval between the two consecutive data points that were used to calculate $[\cdot \text{OH}]_{\text{ss, interval}}$. 
Figure 4-7. Comparison of the measured pseudo-first-order loss rate of various organic compounds, obtained from refs 36 (▼), 16 (○), 10 (▽), 17 (●), 11 (□), and 21 (■), versus our model predictions that account for electrostatic effects. The line represents perfect agreement between predicted and measured $k_{org}$ values. The horizontal bars show the range of estimated $k_{org}$ values using typical ranges of surface areas for goethite in studies where the surface area was not measured.
Chapter 5: Conclusions and Future Research Topics

or

-Just one more experiment…-
Contributions of This Thesis

This dissertation focused on understanding the factors that control the oxidation rates of organic compounds in Fenton and Fenton-like systems. The solution chain reaction was shown to be the dominant decomposition pathway of H₂O₂ and H¹⁴COOH at pH 4 in the presence of dissolved Fe, but this was not the case when the Fe source was ferrihydrite. The probable explanation, based on comparing modeling results from hypothetical mechanisms to experimental data, is that there were insufficient amounts of dissolved Fe to effectively propagate the solution phase chain reaction. Further work in the iron oxide/H₂O₂ system showed that the generation rate of •OH (Vₕₙ) was proportional to the product of the concentrations of surface area of the iron oxide and H₂O₂. A model to estimate the pseudo-first-order rate loss rate coefficients of organic compounds (k₉ₒᵣ₉) based on this relationship and the chemical reactions of the Fenton-like system was created. It successfully predicted Vₕₙ and the loss rate of H¹⁴COOH in aquifer sand experiments, but yielded mixed results when it was used to estimate Vₕₙ and k₉ₒᵣ₉ observed in iron oxide/H₂O₂ systems that other investigators had examined. Subsequent experimental results showed that the most likely explanation was that the attraction between formate anions and the positively charged goethite surface caused H¹⁴COOH to measure a greater [•OH]ₕₙ than the concentration that a neutral •OH probe would detect. Given this analysis, a better model was made that accounted for electrostatic effects, which involves knowing the speciation of the compound of interest, the pH of the solution, and the intrinsic surface acid-base equilibrium constants of the iron oxide.

This work showed that electrostatics affected the oxidation rate of formic acid in the iron oxide/H₂O₂ system. Researchers who study the oxidation rates of organic compounds in heterogeneous systems usually do not need to think about electrostatic effects because many organic pollutants have pKₐ values greater than 7 and therefore are uncharged in neutral and acidic media. Nevertheless, some organic compounds such as benzoic acid have charged species at pH values of 7 or less. It then becomes important to know that electrostatics does affect the interactions between organic compounds and iron oxide. Electrostatics could also influence the concentration of any charged •OH sinks in the boundary layer, which would impact the oxidation rate of the compound of interest if the charged sink were an important sink of •OH.

This work also demonstrated that models could be used to understand key characteristics of the decomposition of hydrogen peroxide and organic compounds in the presence of iron
oxide. Researchers can use models to easily test and constrain mechanistic possibilities, and thus modeling is an efficient approach to investigate the behavior of complicated systems where only some parameters are known. Modeling results should always be compared against experimental data since that is the true test of a model’s accuracy and robustness. When differences occur, as it did in this work, new areas of research become available for those who desire to improve on the current model.

Once some requirements and assumptions are satisfied, our model for predicting $k_{\text{org}}$ from $V_{\text{OH}}$ can be used to determine the proper $H_2O_2$ dosage to oxidize a pollutant in a given period of time. The oxidation process should take place in a well-mixed reactor that is not a continuous flow-through system. The organic pollutant must be primarily in the aqueous phase, and the type and concentration of the contaminant and the iron oxide has to be identified. The pH must be controlled as it dictates chemical speciation and the extent of electrostatic interactions between the various components in the system. The $k_{\text{org}}$ that we are aiming for is then calculated by choosing the time and the factor that the organic compound must be reduced by:

$$k_{\text{org}} = \frac{-\ln([\text{org}]_0/\text{[org]})}{t}$$

(1)

The corresponding steady state concentration of $\bullet OH$ is:

$$[\bullet OH]_{ss} = \frac{k_{\text{org}}}{k_{\text{OH,org}}}$$

(2)

where $k_{\text{org}}$ has been corrected for electrostatic effects (if necessary), and the second-order rate constant of the contaminant with $\bullet OH$ ($k_{\text{OH,org}}$) is known. This is substituted into the expression for $V_{\text{OH}}$ (refer to eq 8 of chapter 3),

$$V_{\text{OH}} = [k_{\text{OH,org}}[\text{org}]_0 + k_{\text{OH,H}_2O_2}[H_2O_2]_0][\bullet OH]_{ss}$$

(3)

where we assume that all other $\bullet OH$ sinks in the groundwater, e.g., dissolved organic carbon and microorganisms, are at negligible concentrations. Lastly, we solve for the initial concentration of $H_2O_2$ by setting eq 3 equal to eq 10, 11, or 12 of chapter 3, depending on the form of the iron oxide in the soil. This assumes that all of the $H_2O_2$ reacts with the iron oxide, i.e., other $H_2O_2$ degradation pathways such as microbial enzymes are unimportant. Once considerable quantities of the pollutant and $H_2O_2$ have been consumed, $V_{\text{OH}}$ will be significantly less than the value that we desire. We would therefore also have to determine when to put in more $H_2O_2$ to the system in order to maintain a steady $V_{\text{OH}}$. For instance, we could decide that the $H_2O_2$ in the reactor should
be replenished when $V_{OH}$ has fallen to 90% of our expected $V_{OH}$, which tells us when more $H_2O_2$ must be added. Then, the contaminant concentration at that time is used in eq 3 to determine how much $H_2O_2$ we need to add to restore $V_{OH}$ back to its original value. This is done iteratively until the pollutant level is reduced to our goal.

For a contaminant that is highly hydrophobic and thus sorbed on soil particles, we would need to know whether or not it is oxidized only in the aqueous phase. This would help us decide if the time that it would require to clean up the polluted soil is dictated by the desorption rate of the compound. If the compound can be oxidized in its sorbed state, then the contribution of the surface oxidation rate to the total oxidation rate has to be evaluated.

If an in situ remediation scheme is used to treat groundwater, then a model for predicting the proper dosage of $H_2O_2$ must also consider issues such as soil porosity, dispersion, and transport of pollutants downstream due to the gradient caused by the well injections. The successful model would likely incorporate our model into a groundwater flow model that is appropriate for the site.

**Future Work**

Many topics related to the Fenton and Fenton-like systems remain to be explored. For instance, the mechanism to describe the iron oxide catalyzed decomposition of $H_2O_2$ is still incomplete. The correct mechanism should show that $V_{OH}$ is proportional to the concentrations of $H_2O_2$ and iron oxide, and explain why $V_{OH}$ is relatively constant from pH 4 to pH 10. Understanding whether $H_2O_2$ is chemisorbed or physisorbed onto the iron oxide could help in formulating this mechanism.

Reactive species such as superoxide ($O_2^-$), perhydroxyl radical (HO$_2^+$), and hydroperoxide anion (HO$_2^-$) are also produced in the Fenton-like system, and they could be involved in reactions that transform organic compounds via reduction instead of oxidation (1-3). A greater understanding of this non-hydroxyl radical degradation pathway could increase the applicability of the Fenton-like system for waste treatment and remediation.

The experiments in this thesis were done in solutions containing only a few solutes, but groundwater typically contains metals, carbon (dissolved and particulate), anions such as phosphate, sulfate, and carbonate, and microorganisms. The impact of these constituents on the kinetics of the reactions of the Fenton system and the oxidation rates of contaminants has not been completely explored (4-9). For example, microbes could be an important nonradical
producing sink of H$_2$O$_2$ because they use catalase and peroxidase to decompose H$_2$O$_2$ to water and oxygen:

\[
2\text{H}_2\text{O}_2 \xrightarrow{\text{catalase}} 2\text{H}_2\text{O} + \text{O}_2 \quad (4)
\]

\[
\text{H}_2\text{O}_2 + \text{NADH} + \text{H}^+ \xrightarrow{\text{peroxidase}} 2\text{H}_2\text{O} + \text{NAD}^+ \quad (5)
\]

[Although this is a disadvantage from the viewpoint of chemical oxidation, H$_2$O$_2$ has been used as a way of delivering oxygen to aerobic microorganisms that are responsible for bioremediation in aquifers (10-12).] Other metal oxides also react with H$_2$O$_2$ (13), but not all make •OH and so they would represent another set of nonradical producing sinks. Experiments will be needed to determine the effect of each component and their interactions as a whole.

Contaminants in the subsurface are often sorbed onto soil particles and may be harder to oxidize in this state (14). Therefore, the rates at which they adsorb and desorb would control their aqueous and sorbed concentrations and determine how quickly they are oxidized by •OH. This subject has not been explored in-depth in the literature (15-17), but understanding it could potentially increase the accuracy of the model predictions of $k_{org}$ for organic compounds that sorb strongly onto solids.

**Literature Cited**


Appendix A

or

- The nitty-gritty, revealed! -
Table A-1. Concentration of dissolved iron (defined as iron that passed through a 0.02-µm filter) in the ferrihydrite experiments. Ferrihydrite was diluted from a freshly prepared $4 \times 10^{-4}$ M stock solution, except where noted, into 200 mL. The detection limit for Fe is 0.03 µM.

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<th>[Ferrihydrite] (µM)</th>
<th>Number of hours in experiment</th>
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<th>$[\text{Fe}]_{\text{diss}}$ at end (µM)</th>
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*a* In this column, the time when the sample was taken was not written down.

*b* Diluted from a $5 \times 10^{-4}$ M solution of ferrihydrite.

*c* Experiment contained 100 µM tert-butyl alcohol.

*d* Experiment contained 10 µM tert-butyl alcohol.
**Kelvin effect**

The Kelvin effect (1) states that colloids and small crystals have a greater solubility than their larger counterparts because they are thermodynamically less stable. It is quantified as follows (2):

\[
\log K_{sp}(r) = \log K_{sp}(r = \infty) + \frac{2\gamma(MW)}{2.303RTpr}
\]

(1)

where \(\log K_{sp}(r)\) is the solubility product of the colloid with radius \(r\), \(r\) is the colloidal radius, \(\log K_{sp}(r = \infty)\) is the solubility product of the large crystals, \(\gamma\) is the interfacial tension of the solid-water interface, \(MW\) is the molecular weight, \(R\) is the gas constant, \(T\) is the absolute temperature, and \(p\) is the mineral density. \(\gamma\) for ferrihydrite is not known, but can be estimated from the values for goethite and hematite because mineral hardness correlates positively with \(\gamma\) (2). Therefore, \(\gamma\) for ferrihydrite is expected to be less than 1600 and 1200 mJ/m\(^2\) (1), the respective values for goethite and hematite. Assuming a value of 250 mJ/m\(^2\) and a colloidal radius of 0.02 µm (this filter size retained most of the ferrihydrite colloids),

\[
\frac{2(2.5 \times 10^{-5} \text{ J cm}^{-2})(107 \text{ g mol}^{-1})}{2.303(8.31 \text{ J mol}^{-1} \text{ deg}^{-1})(298.15 \text{ deg})(3.96 \text{ g cm}^{-2})(2 \times 10^{-6} \text{ cm})} = 0.1
\]

(2)

then the change to the solubility product is small compared to the known range for \(K_{sp}\) (10\(^{-39.4}\) - 10\(^{-37.0}\); 3).

**Literature Cited**


(2) Gschwend, P. M.; Reynolds, M. D. J. *Contam. Hydrol.* 1987, 1, 309-327.

Appendix B

or

\[-0 = 1?\]
pH increase due to oxidation of formic acid

The oxidation of formic acid by •OH is expected to increase the pH of the system in this manner:

\[
\begin{align*}
•OH + HCOO^- + O_2 &\rightarrow CO_2 + O_2^- + H_2O \\
H^+ + O_2^- &\rightarrow HO_2 \\
2HO_2 &\rightarrow H_2O_2 + O_2
\end{align*}
\]

The first two reactions are a significant sink of H\(^+\) in our experiments because the dominant species of formic acid and superoxide at pH 4 are formate and perhydroxyl radical, respectively. The oxygen that reaction 1 consumes is plentiful since the solutions are open to air ([O\(_2\)]\(_{sat}\) = 258 µM at 25 °C; \(I\)) and constantly stirred, which facilitates air-liquid transfer.

Experiments with H\(_2\)O\(_2\) as the dominant •OH sink

Figures B1a-e show typical data sets of loss of \(^{14}\)C-labeled formic acid and H\(_2\)O\(_2\) over time from experiments containing ferrihydrite, goethite, hematite, or Georgetown sand. We compared two methods of determining \(k_{HCOOH}\), non-linear regression fits of the \(^{14}\)C data and linear regression of the log-transformed data, for all experiments except those that used ferrihydrite. The average percent difference in the \(k_{HCOOH}\) obtained was insignificant (4.1, with a standard deviation of 5.2).

Experiments with formic acid as the dominant •OH sink

Figures B2a-f show pseudo-first order and zeroth-order fits to formic acid loss data in experiments containing 0.6 g/L of goethite. Both types of regression produce similarly good fits for most of the data sets.

Given that formic acid is initially the main sink of •OH, if one assumes that [H\(_2\)O\(_2\)], and therefore also \(V_{OH}\), remain constant over time, then the loss of H\(^{14}\)COOH should follow zeroth-order kinetics. However, some H\(_2\)O\(_2\) was consumed during the experiments, thus decreasing \(V_{OH}\), and in the later parts of some experiments H\(_2\)O\(_2\) became a competing •OH sink because [HCOOH]\(_T\) decreased more rapidly than [H\(_2\)O\(_2\)] (see Figures B2a-f and B3). Strictly speaking, then, neither first-order nor zeroth-order kinetics should precisely describe formic acid loss in these experiments. However, since we observed that the loss of formic acid is well described by first-order kinetics, using \(k_{HCOOH}\) from eq 6 to calculate \(V_{OH}\) in eq 9 still provides an excellent estimate of the initial rate of •OH formation.

Literature Cited

Figure B1a. Representative data set of the concentration of $^{14}$C-labeled formic acid (given as disintegrations per minute) and hydrogen peroxide over time in experiments at pH 4, containing 20 μM (○), 200 μM (▽), or 280 μM (■) ferrihydrite. $[\text{HCOOH}]_{T,0}$ was 110 nM.
Figure B1b. Representative data set of the concentration of $^{14}$C-labeled formic acid (given as disintegrations per minute) over time in experiments at pH 4, 190 μM ferrihydrite, $[\text{HCOOH}]_{T,0}$ = 212-219 nM, and $[\text{H}_2\text{O}_2]_0 = 50$ mM (●), 100 mM (□), 200 mM (▲), or 400 mM (▽).
Figure B1c. Representative data set of the concentration of $^{14}$C-labeled formic acid (given as disintegrations per minute) and hydrogen peroxide over time in experiments at pH 4, containing 0.6 g/L of goethite, $[\text{HCOOH}]_{r0} = 92-112$ nM, and $[\text{H}_2\text{O}_2]_0 = 2$ mM (●), 40 mM (□), 0.4 M (▲), or 1 M (▽).
Figure B1d. Representative data set of the concentration of $^{14}$C-labeled formic acid (given as disintegrations per minute) and hydrogen peroxide over time in experiments at pH 4, containing 1 g/L of hematite, $[\text{HCOOH}]_{r,0} = 201-210$ nM, and $[\text{H}_2\text{O}_2]_0 = 10$ mM (●), 100 mM (□), 1 M (▲), or 2 M (▽).
Figure B1e. Representative data set of the concentration of $^{14}$C-labeled formic acid (given as disintegrations per minute) and hydrogen peroxide over time in experiments at pH 4, containing 60 g/L of Georgetown sand, $[\text{HCOOH}]_{T,0} = 162$-166 nM, and $[\text{H}_2\text{O}_2]_0 = 5$ mM (●), 110 mM (○), 675 mM (◆), or 2.7 M (◇).
Figure B2a. Concentration of HCOOH versus time from experiments containing 0.6 g/L of goethite, \([\text{HCOOH}]_T,0 = 10 \text{ mM}\), and \([\text{H}_2\text{O}_2]_0 = 2 \text{ mM} (\bullet) \) or 10 mM (O). (A) shows the nonlinear regression fits to the data, while (B) shows the linear regression fits.
Figure B2b. Concentration of HCOOH versus time from experiments containing 0.6 g/L of goethite, \([\text{HCOOH}]_{T,0} = 10 \text{ mM}\), and \([\text{H}_2\text{O}_2]_{0} = 10 \text{ mM}\) (●), 50 mM (◇), or 100 mM (■). (A) shows the nonlinear regression fits to the data, while (B) shows the linear regression fits.
Figure B2c. Concentration of HCOOH versus time from experiments containing 0.6 g/L of goethite, [HCOOH]_{T,0} = 10 mM, and [H_2O_2]_0 = 50 mM (●) or 100 mM (▲). (A) shows the nonlinear regression fits to the data, while (B) shows the linear regression fits.
Figure B2d. Concentration of HCOOH versus time from experiments containing 0.6 g/L of goethite, $[\text{HCOOH}]_T,0 = 0.3 \text{ mM}$, and $[\text{H}_2\text{O}_2]_0 = 700 \mu\text{M}$ (●), 500 μM (▽), 250 μM (■), or 100 μM (◇). (A) shows the nonlinear regression fits to the data, while (B) shows the linear regression fits.
Figure B2e. Concentration of HCOOH versus time from experiments containing 0.6 g/L of goethite, [HCOOH]₀ = 50 µM, and [H₂O₂]₀ = 600 µM. (A) shows the nonlinear regression fit to the data, while (B) shows the linear regression fit.
Figure B2f. Concentration of HCOOH versus time from experiments containing 0.6 g/L of goethite, [HCOOH]_{T,0} = 50 \mu M, and [H_2O_2]_{0} = 500 \mu M (\bullet), 250 \mu M (\nabla), or 100 \mu M (\blacksquare). (A) shows the nonlinear regression fits to the data, while (B) shows the linear regression fits.
Figure B3. Concentration of H₂O₂ versus time for the experiments shown in Figures B2a-f.
Appendix C

or

-Details, details-
Table C-1. Final pH of experiments using goethite and H₂O₂ to oxidize 2-CP. The initial pH was 4.0.

<table>
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<tr>
<th>[goethite] (g/L)</th>
<th>[H₂O₂]₀ (mM)</th>
<th>[2-CP]₀ (μM)</th>
<th>Number of hours in experiment</th>
<th>Final pH</th>
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<td>6.4</td>
<td>3.5</td>
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</table>

We used eq 17 from chapter 4 (shown below) to determine the impact on electrostatics as a result of a decrease of 0.5-0.7 pH units over the course of an experiment.

\[
\left( - \frac{d[HCOOH]}{dt} \right)_e = \frac{A}{V} \int \left( k_{OH,HCOOH} [OH]_{ss} (x) [HCOOH]_{bulk} + k_{OH,HCOO^-} [OH]_{ss} (x) [HCOO^-]_{bulk} e^{-\Delta zF(x)/RT} \right) dx
\]

The results (Table C-2) indicate that Ψ₀ increases as pH decreases, as expected, but ζ remains relatively constant because the percentage of total formic acid that is formate decreases as pH decreases. Therefore, the changes in pH that we observed have no significant impact on the electrostatics modeling.
Table C-2. \( \int_{0}^{\infty} k_{\text{OH}, \text{HCOO}^-} [\text{OH}](x)[\text{HCOO}^-]_{\text{bulk}} e^{-\Delta G_{\text{OH}}(x)/RT} dx \) (\( \zeta \)) as a function of pH. \( \Psi_0 \) was calculated in the same manner as described in chapter 4, and \( \zeta \) was evaluated at \( I = 10 \) mM and \([\text{H}_2\text{O}_2]_0 = 160 \) mM.

<table>
<thead>
<tr>
<th>pH</th>
<th>( \Psi_0 ) (V)</th>
<th>Percent of total formic acid as formate</th>
<th>( \zeta )</th>
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<td>3.3</td>
<td>0.18</td>
<td>26</td>
<td>5.0 x 10^{-7}</td>
</tr>
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</table>
Appendix D

or

-If a picture is worth a thousand words, then I wrote much more than I thought!-
The graphs in the following pages depict the raw data for the experiments discussed in chapters 2-4. All experiments were done at $I = 10 \text{ mM NaClO}_4$ and pH 4 unless stated otherwise. The rate constants were obtained by fitting the data to the function $[C] = [C]_0 \exp(-kt)$.

Figures D1-14 show data discussed in chapter 2.

Figures D1-7, D11C-D12, and D15-26 show data discussed in chapter 3.

Figures D27-35 show data discussed in chapter 4.
Figure D1. (A) $^{14}$C activity in experiments with [Fe(III)] = 0.79 (●) or 2.11 (▼) μM, and $k = 0.53$ and 1.45 d$^{-1}$, respectively. (B) $[\text{H}_2\text{O}_2]$ in experiments described in (A), with $k = 0.033$ (○) and 0.066 (▼) d$^{-1}$. (C) $^{14}$C activity in experiments with [Fe(III)] = 1.30 (●), 0.58 (▼), or 0.73 (■) μM, and $k = 0.76$, 0.21, and 0.32 d$^{-1}$, respectively. (D) $[\text{H}_2\text{O}_2]$ in experiments described in (C), with $k = 0.072$ (○), 0.0082 (▼), and 0.017 (□) d$^{-1}$. 
Figure D2. (A) $^{14}$C activity in experiments with [ferrihydrite] = 5.59 (●), 37.66 (▼) or 85.69 (■) μM, and $k = 0.36$, 0.49, and 0.41 d$^{-1}$, respectively. (B) [H$_2$O$_2$] in experiments described in (A), with $k = 0.016$ (○), 0.035 (▼), and 0.040 (□) d$^{-1}$. (C) $^{14}$C activity in experiments with [ferrihydrite] = 16.85 (●), 178.56 (▼) or 276.19 (■) μM, and $k = 0.63$, 2.30, and 3.89 d$^{-1}$, respectively. (D) [H$_2$O$_2$] in experiments described in (C), with $k = 0.039$ (○), 0.18 (▼), and 0.35 (□) d$^{-1}$. 
Figure D3. (A) $^{14}$C activity in experiments with [ferrihydrite] = 20.31 (●), 192.93 (▼), or 276.76 (■) µM, and $k = 0.36, 0.49, \text{ and } 0.41 \text{ d}^{-1}$, respectively. (B) $[\text{H}_2\text{O}_2]$ in experiments described in (A), with $k = 0.11$ (Ø), 0.035 (▼), and 0.040 (□) d$^{-1}$. (C) $^{14}$C activity in experiments with [ferrihydrite] = 6.35 (●), 23.74 (▼), 85.34 (■), or 133.91 (◆) µM, and $k = 0.11, 0.31, 0.31, \text{ and } 1.84 \text{ d}^{-1}$, respectively. (D) $[\text{H}_2\text{O}_2]$ in experiments described in (C), with $k = 0.0067$ (Ø), 0.013 (▼), 0.024 d$^{-1}$ (□), and 0.12 (◆) d$^{-1}$. 
Figure D4. (A) $^{14}$C activity in experiments with [ferrihydrite] = 44.86 (●), 84.41 (▼), or 378.99 (■) μM, and $k = 0.53$, 0.56, and 4.00 d$^{-1}$, respectively. (B) [H$_2$O$_2$] in experiments described in (A), with $k = 0.040$ (○), 0.038 (▽), and 0.54 (□) d$^{-1}$. (C) $^{14}$C activity in experiments with [ferrihydrite] = 323.31 (●) or 411.83 (▼) μM, and $k = 1.91$ and 4.97 d$^{-1}$, respectively. (D) [H$_2$O$_2$] in experiments described in (C), with $k = 0.26$ (○) and 0.61 (▽) d$^{-1}$. 
Figure D5. (A) $^{14}$C activity in experiments with [ferrihydrite] = 178.22 μM and $k = 1.77$ d$^{-1}$. (B) [H$_2$O$_2$] in experiments described in (A), with $k = 0.13$ d$^{-1}$. (C) $^{14}$C activity in experiments with [ferrihydrite] = 306.45 μM and $k = 2.20$ d$^{-1}$. (D) [H$_2$O$_2$] in experiments described in (C), with $k = 0.43$ d$^{-1}$.
Figure D6. (A) $^{14}$C activity in experiments with [ferrihydrite] = 166.77 (●) or 183.94 (▼) μM, and $k = 2.63$ and 2.13 d$^{-1}$, respectively. (B) $[\text{H}_2\text{O}_2]$ in experiments described in (A), with $k = 0.14$ (○) and 0.18 (▼) d$^{-1}$. (C) $^{14}$C activity in experiments with [ferrihydrite] = 212.85 (●) or 61.80 (▼) μM, and $k = 2.22$ and 0.83 d$^{-1}$, respectively. (D) $[\text{H}_2\text{O}_2]$ in experiments described in (C), with $k = 0.21$ (○) and 0.036 (▼) d$^{-1}$.
Figure D7. (A) $^{14}$C activity in experiments with [Fe(III)] = 1.30 μM and $k = 0.89$ d$^{-1}$. (B) [H$_2$O$_2$] in experiments described in (A), with $k = 0.046$ d$^{-1}$. (C) $^{14}$C activity in experiments with [ferrihydrite] = 83.68 μM and $k = 1.15$ d$^{-1}$. (D) [H$_2$O$_2$] in experiments described in (C), with $k = 0.071$ d$^{-1}$. 

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Figure D8. (A) $^{14}$C activity in experiments with 0.1 mM tert-butyl alcohol and [ferrihydrite] = 192.63 (●) or 196.87 (▼) µM, and $k = 0.62$ and 1.16 $d^{-1}$, respectively. (B) $[\text{H}_2\text{O}_2]$ in experiments described in (A), with $k = 0.27$ (○) and 0.34 (▼) $d^{-1}$. (C) $^{14}$C activity in experiments with [Fe(III)] = 2.70 (●) or 2.73 and 0.1 mM tert-butyl alcohol (▼) µM, and $k = 0.23$ and 0.0095 $d^{-1}$, respectively. (D) $[\text{H}_2\text{O}_2]$ in experiments described in (C), with $k = 0.0090$ (○) and 0.0031 (▼) $d^{-1}$. 
Figure D9. (A) $^{14}$C activity in experiments with 10 $\mu$M tert-butyl alcohol (except •) and [ferrihydrite] = 85.70 (●), 78.34 (▼), 85.08 (■), or 84.93 and 1 $\mu$M Fe(III) (◆) $\mu$M, and $k =$ 0.75, 0.20, 0.22, and 0.23 d$^{-1}$, respectively. (B) $[\text{H}_2\text{O}_2]$ in experiments described in (A), with $k =$ 0.039 (○), 0.020 (▼), 0.018 (□), and 0.015 (◇) d$^{-1}$. (C) $^{14}$C activity in experiments with [ferrihydrite] = 185.46 (●), 192.95 and 10 $\mu$M tert-butyl alcohol (▼), or 181.93 and 1.5 $\mu$M Fe(III) (■) $\mu$M, and $k =$ 1.86, 1.43, and 2.74 d$^{-1}$, respectively. (D) $[\text{H}_2\text{O}_2]$ in experiments described in (C), with $k =$ 0.088 (○), 0.11 (▼), and 0.15 (□) d$^{-1}$. 
Figure D10. (A) $^{14}$C activity in experiments with [ferrihydrite] = 42.68 (●), 48.93 and 10 μM tert-butyl alcohol (▼), or 42.15 and 1.5 μM Fe(III) (■) μM, and $k = 0.77$, 0.27, and 0.76 d$^{-1}$, respectively. (B) [H$_2$O$_2$] in experiments described in (A), with $k = 0.031$ (O), 0.0037 (▽), and 0.028 (□) d$^{-1}$. (C) $^{14}$C activity in experiments with [ferrihydrite] = 190.11 (●), 187.86 and 10 μM tert-butyl alcohol (▼), or 187.77 and 10 μM tert-butyl alcohol (■) μM, and $k = 1.73$, 0.12, and 0.13 d$^{-1}$, respectively. (D) [H$_2$O$_2$] in experiments described in (C), with $k = 0.44$ (O), 0.036 (▽), and 0.057 (□) d$^{-1}$.
Figure D11. (A) $^{14}$C activity in experiments with $[\text{Fe(III)}] = 0.90$ (●) or 1.18 and 1 mM NaCl (▼) μM, $[\text{ferrihydrite}] = 197.95$ (■) or 197.35 and 1 mM NaCl (◆) μM, and $k = 0.14$, 0.14, 2.85, and 2.63 d$^{-1}$, respectively. (B) $[\text{H}_2\text{O}_2]$ in experiments described in (A), with $k = 0.011$ (○), 0.0087 (▼), 0.25 (□), and 0.19 (◇) d$^{-1}$. (C) $^{14}$C activity in experiments with $[\text{ferrihydrite}] \approx 200$ μM and $k = 2.93$ d$^{-1}$. (D) $[\text{H}_2\text{O}_2]$ in experiments described in (C), with $k = 0.28$ d$^{-1}$.
Figure D12. (A) $^{14}$C activity in experiments with [ferrihydrite] $= 198.75$ μM and $k = 2.40$ d$^{-1}$. (B) [H$_2$O$_2$] in experiments described in (A), with $k = 0.17$ d$^{-1}$. (C) $^{14}$C activity in experiments with [ferrihydrite] $\sim 200$ μM and $k = 1.71$ d$^{-1}$. (D) [H$_2$O$_2$] in experiments described in (C), with $k = 0.19$ d$^{-1}$.
Figure D13. (A) Concentration of benzoic acid (●) and formic acid (▼) in a control experiment containing H₂O₂ and no iron. (B) [H₂O₂] in the experiment described in (A). (C) Concentration of benzoic acid (●) and formic acid (▼) in a control experiment with [ferrihydrite] = 300 μM and no H₂O₂.
Figure D14. (A) Concentration of benzoic acid in duplicate experiments with [ferrihydrite] = 190 μM, and $k = 1.26$ (○) and 0.66 (▽) d$^{-1}$. (B) Concentration of benzoic acid in duplicate experiments with [ferrihydrite] = 190 μM, and $k = 0.59$ (○) and 0.23 (▽) d$^{-1}$. (C) $\left[\text{H}_2\text{O}_2\right]$ in experiments described in (A) and (B), with $k = 0.078$ (○) and 0.090 (▽) d$^{-1}$. 
Figure D15. (A) $^{14}$C activity in experiments with [ferrihydrite] = 190 μM and $[\text{H}_2\text{O}_2]_0 = 5$ (●), 10 (▼) or 30 (■) mM, and $k = 0.13, 0.12, \text{ and } 0.17 \text{ h}^{-1}$, respectively. (B) $^{14}$C activity in experiments with [ferrihydrite] = 190 μM and $[\text{H}_2\text{O}_2]_0 = 50$ (●), 100 (▼), 200 (■) or 400 (◆) mM, and $k = 0.22, 0.26, 0.23, \text{ and } 0.20 \text{ h}^{-1}$, respectively. (C) $^{14}$C activity in experiments with [ferrihydrite] = 90 μM and $[\text{H}_2\text{O}_2]_0 = 5$ (●), 10 (▼) or 30 (■) mM, and $k = 0.11, 0.098, \text{ and } 0.084 \text{ h}^{-1}$, respectively. (D) $^{14}$C activity in experiments with [ferrihydrite] = 90 μM and $[\text{H}_2\text{O}_2]_0 = 50$ (●), 100 (▼), 200 (■) or 400 (◆) mM, and $k = 0.093, 0.080, 0.068, \text{ and } 0.072 \text{ h}^{-1}$, respectively.
Figure D16. (A) $^{14}$C activity in experiments with [ferrihydrite] = 100 μM and $[\text{H}_2\text{O}_2]_0 = 75$ (●) or 300 (▼) mM, or [ferrihydrite] = 200 μM and $[\text{H}_2\text{O}_2]_0 = 75$ (■) or 300 (◆) mM, and $k = 0.061, 0.099, 0.16$ and 0.19 h$^{-1}$, respectively. (B) $^{14}$C activity in experiments with [ferrihydrite] = 160 μM and $[\text{H}_2\text{O}_2]_0 = 0.3$ (●) or 0.4 (▼) M, or [ferrihydrite] = 230 μM and $[\text{H}_2\text{O}_2]_0 = 0.2$ (■) or 0.3 (◆) M, and $k = 0.16, 0.14, 0.18$ and 0.20 h$^{-1}$, respectively. (C) $^{14}$C activity in experiments with 0.2 g/L goethite and $[\text{H}_2\text{O}_2]_0 = 10$ (●) or 100 (▼) mM, or [hematite] = 0.2 g/L and $[\text{H}_2\text{O}_2]_0 = 10$ (■) or 100 (◆) mM, and $k = 0.26, 0.40, 0.024$ and 0.031 h$^{-1}$, respectively.
Figure D17. (A) $^{14}$C activity in experiments with 0.2 g/L goethite and $[\text{H}_2\text{O}_2]_0 = 50$ (●) or 300 (▼) mM, or 0.2 g/L hematite and $[\text{H}_2\text{O}_2]_0 = 50$ (■) or 300 (◆) mM, and $k = 0.53, 0.47, 0.040$ and 0.014 h$^{-1}$, respectively. (B) $^{14}$C activity in experiments with 0.2 g/L hematite with $k = 0.0089$ (●) and 0.013 (▼) h$^{-1}$. (C) $[\text{H}_2\text{O}_2]$ in experiments described in (B), with $[\text{H}_2\text{O}_2]_0 = 0.7$ (○) or 0.9 (▼) M.
Figure D18. (A) $^{14}$C activity in experiments with 0.2 g/L goethite and $k = 0.53$ (●) and 0.27 (▼) h$^{-1}$, or 0.2 g/L hematite and $k = 0.027$ (■) and 0.015 (∆) h$^{-1}$. (B) $[\text{H}_2\text{O}_2]$ in experiments described in (A), with $[\text{H}_2\text{O}_2]_0 = 0.2$ (○), 0.6 (△), 0.8 (□), or 0.9 (◇) M. (C) $^{14}$C activity in experiments with 0.2 g/L goethite and $k = 0.12$ (●) and 0.14 (∆) h$^{-1}$, or 0.6 g/L goethite and $k = 0.76$ (■) and 1.21 (∗) h$^{-1}$. (D) $[\text{H}_2\text{O}_2]$ in experiments described in (C), with $[\text{H}_2\text{O}_2]_0 = 0.002$ (○), 2 (△), 0.010 (□), or 0.1 (◇) M.
Figure D19. (A) $^{14}$C activity in experiments with 0.6 g/L goethite and $k = 0.32$ (●), 1.20 (▼), 1.26 (■) and 1.03 (◆) h$^{-1}$. (B) [H$_2$O$_2$] in experiments described in (A), with [H$_2$O$_2$]$_0$ = 2 (○) or 40 (▼) mM, or 0.4 (□) or 1.0 (◇) M. (C) $^{14}$C activity in experiments with 0.02 g/L goethite and [H$_2$O$_2$]$_0$ = 2 (●), 50 (▼), 140 (■) or 440 (◆) mM, with $k = 0.062$, 0.027, 0.076, and 0.11 h$^{-1}$, respectively. (D) $^{14}$C activity in experiments with 0.2 g/L goethite and [H$_2$O$_2$]$_0$ = 0.9 (●) or 3.7 (▼) M, or 1.0 g/L hematite and [H$_2$O$_2$]$_0$ = 50 (●) or 540 (▼) mM, with $k = 0.12$, 0.062, 0.096, and 0.041 h$^{-1}$, respectively.
Figure D20. (A) $^{14}$C activity in experiments with 1.0 g/L hematite and $k = 0.081 (\bullet)$, 0.11 (\(\nabla\)), 0.039 (\(\blacksquare\)) and 0.050 (\(\blacklozenge\)) $h^{-1}$. (B) $[\text{H}_2\text{O}_2]$ in experiments described in (A), with $[\text{H}_2\text{O}_2]_0 = 10 (\bigcirc)$ or 110 (\(\nabla\)) mM, or 1.0 (\(\square\)) or 2.0 (\(\blacklozenge\)) M. (C) $^{14}$C activity in control experiments with 0.02 g/L goethite (\(\bullet\)) or hematite (\(\nabla\)) and no H$_2$O$_2$. 
Figure D21. (A) $^{14}$C activity in experiments with 0.6 g/L goethite, $I = [\text{NaHCOO}] = 10$ mM, and $k = 0.0020$ (●) and 0.010 (▲) h$^{-1}$. (B) $[\text{H}_2\text{O}_2]$ in experiments described in (A), with $[\text{H}_2\text{O}_2]_0 = 2$ (○) or 10 (▲) mM. (C) $^{14}$C activity in experiments with 0.6 g/L goethite, $I = [\text{NaHCOO}] = 10$ mM, and $k = 0.010$ (●), 0.051 (▲) and 0.079 (■) h$^{-1}$. The second and third $k$ values were obtained using the data points from the first four hours as the pH of the solutions rose above 5 afterwards. (D) $[\text{H}_2\text{O}_2]$ in experiments described in (C), with $[\text{H}_2\text{O}_2]_0 = 10$ (○), 50 (▲), or 100 (□) mM.
Figure D22. (A) $^{14}$C activity in experiments with 0.6 g/L goethite, $I = [\text{NaHCOO}] = 10$ mM, and $k = 0.054$ (●) and 0.099 (▽) h$^{-1}$. The $k$ values were obtained using the data points from the first four hours as the pH of the solutions rose above 5 afterwards. (B) $[\text{H}_2\text{O}_2]$ in experiments described in (A), with $[\text{H}_2\text{O}_2]_0 = 50$ (○) or 110 (▽) mM. (C) $^{14}$C activity in experiments with 0.6 g/L goethite, $[\text{NaHCOO}] = 50$ μM ($I = 10$ mM), and $k = 0.088$ h$^{-1}$. (D) $[\text{H}_2\text{O}_2]$ in experiments described in (C), with $[\text{H}_2\text{O}_2]_0 = 610$ μM and $k = 0.032$ h$^{-1}$.
Figure D23. (A) $^{14}$C activity in experiments with 0.6 g/L goethite, [NaHCOO] = 50 µM (I = 10 mM), and $k = 0.15$ (●), 0.078 (▼), and 0.021 (■) h$^{-1}$. (B) [H$_2$O$_2$] in experiments described in (A), with [H$_2$O$_2$]$_0$ = 530 (○), 260 (▼), or 110 (□) µM, and $k = 0.055$, 0.050, and 0.043 h$^{-1}$, respectively. (C) $^{14}$C activity in experiments with 0.6 g/L goethite, [NaHCOO] = 300 µM (I = 10 mM), and $k = 0.028$ (●), 0.019 (▼), 0.010 (■), and 0.0018 (♦) h$^{-1}$. (D) [H$_2$O$_2$] in experiments described in (C), with [H$_2$O$_2$]$_0$ = 700 (○), 500 (▼), 250 (□), or 100 (♦) µM and $k = 0.037$, 0.037, 0.030, and 0.023 h$^{-1}$, respectively.
Figure D24. (A) $^{14}$C activity in experiments with 0.6 g/L goethite, [NaHCOO] = 50 µM (I = 10 mM), and $k = 1.10$ (●) and 0.74 (▲) h$^{-1}$. (B) [H$_2$O$_2$] in experiments described in (A), with [H$_2$O$_2$]$_0$ = 0.5 (○) or 1.9 (▲) M. (C) $^{14}$C activity in experiments with 25 g/L Georgetown sand, and $k = 0.042$ (●), 0.035 (▲), and 0.052 (■) h$^{-1}$. (●) is the control experiment with $k = 0.0044$ h$^{-1}$. (D) [H$_2$O$_2$] in experiments described in (C), with [H$_2$O$_2$]$_0$ = 4.8 (○) or 100 (▲) mM, or 1.1 (■) M.
Figure D25. (A) $^{14}$C activity in experiments with 60 g/L Georgetown sand and $k = 0.043$ (●), 0.095 (▼), 0.052 (■), and 0.073 (◆) h$^{-1}$. (B) [H$_2$O$_2$] in experiments described in (A), with [H$_2$O$_2$]$_0 = 5.0$ (○), 110 (▼), 675 (□), or 2700 (◆) mM. (C) $^{14}$C activity in experiments with 60 g/L Georgetown sand, and $k = 0.0091$ (●), 0.012 (▼), 0.00034 (■), and 0.0024 (◆) h$^{-1}$. (■) and (◆) contained 10 mM NaHCOO instead of NaClO$_4$. (D) [H$_2$O$_2$] in experiments described in (C), with [H$_2$O$_2$]$_0 = 85$ (○) or 630 (▼) μM, or 1.0 (□) or 10 (◆) mM.
Figure D26. (A) $^{14}$C activity in experiments with 25 g/L Georgetown sand (●), or 60 g/L Georgetown sand and [NaHCOO] = 10 mM (▼) or [NaHCOO] = 300 μM (I = 10 mM) (■), (◆). $k = 0.057, 0.0058, 0.0083,$ and $0.0083 \text{ h}^{-1},$ respectively. (B) $[\text{H}_2\text{O}_2]$ in experiments described in (A), with $[\text{H}_2\text{O}_2]_0 = 30$ (○), 60 (▼), 0.086 (□), or 0.6 (◆) mM.
Figure D27. (A) [2-CP] in an experiment with 2.5 g/L goethite and $k = 0.17 \text{ h}^{-1}$. (B) [H$_2$O$_2$] in the experiment described in (A), with $k = 0.030 \text{ h}^{-1}$. (C) [2-CP] in an experiment with 2.5 g/L goethite and $k = 0.14 \text{ h}^{-1}$. (D) [H$_2$O$_2$] in the experiment described in (C), with $k = 0.050 \text{ h}^{-1}$. 
Figure D28. (A) $^{14}$C activity in an experiment with $[\text{Fe(III)}] = 25$ μM, $[\text{H}_2\text{O}_2]_0 = 1.9$ M, and $[2$-CP]$_0 = 200$ μM, with $k = 0.0038$ h$^{-1}$. (B) [2-CP] in an experiment with $[\text{Fe(III)}] = 25$ μM and $[\text{H}_2\text{O}_2]_0 = 2.0$ M, with $k = 0.039$ h$^{-1}$. (C) $^{14}$C activity in an experiment with $[\text{Fe(III)}] = 75$ μM, $[\text{H}_2\text{O}_2]_0 = 1.7$ M, and $[2$-CP]$_0 = 200$ μM, with $k = 0.032$ h$^{-1}$. (D) [2-CP] in an experiment with $[\text{Fe(III)}] = 75$ μM and $[\text{H}_2\text{O}_2]_0 = 1.8$ M, with $k = 0.97$ h$^{-1}$. 
Figure D29. (A) $^{14}$C activity in an experiment with 0.6 g/L goethite, $[\text{H}_2\text{O}_2]_0 = 2$ M, and $[2\text{-CP}]_0 = 130$ $\mu$M, with $k = 0.28$ h$^{-1}$. (B) $[2\text{-CP}]$ in an experiment with 0.6 g/L goethite, $[\text{H}_2\text{O}_2]_0 = 2$ M, and $k = 0.049$ h$^{-1}$. (C) $[\text{H}_2\text{O}_2]$ in the experiments described in (A, $\bigcirc$) and (B, $\triangle$), with $k = 0.0016$ ($\triangle$) h$^{-1}$. 
Figure D30. (A) $^{14}$C activity in an experiment with 0.6 g/L goethite, $[\text{H}_2\text{O}_2]_0 = 2 \text{ M}$, and $[\text{2-CP}]_0 = 300 \mu\text{M}$, with $k = 0.075 \text{ h}^{-1}$. (B) $[\text{2-CP}]$ in an experiment with 0.6 g/L goethite, $[\text{H}_2\text{O}_2]_0 = 2 \text{ M}$, and $k = 0.052 \text{ h}^{-1}$. (C) $[\text{H}_2\text{O}_2]$ in the experiments described in (A, ○) and (B, △), with $k = 0.0026 \text{ (△) h}^{-1}$. 
Figure D31. (A) $^{14}$C activity in an experiment with 0.6 g/L goethite, $[\text{H}_2\text{O}_2]_0 = 2$ M, and $[\text{NB}]_0 = 100$ µM, with $k = 0.55$ h$^{-1}$. (B) [NB] in an experiment with 0.6 g/L goethite, $[\text{H}_2\text{O}_2]_0 = 2$ M, and $k = 0.058$ h$^{-1}$. (C) $[\text{H}_2\text{O}_2]$ in the experiments described in (A, $\bigcirc$) and (B, $\bigdiamond$), with $k = 0.0042$ ($\bigdiamond$) h$^{-1}$. 
Figure D32. (A) $^{14}$C activity in an experiment with 0.6 g/L goethite, $[\text{H}_2\text{O}_2]_0 = 2$ M, and $[\text{NB}]_0 = 100$ $\mu$M, with $k = 0.73$ h$^{-1}$. (B) [NB] in an experiment with 0.6 g/L goethite, $[\text{H}_2\text{O}_2]_0 = 2$ M, and $k = 0.072$ h$^{-1}$. (C) $[\text{H}_2\text{O}_2]$ in the experiments described in (A, $\bigcirc$) and (B, $\blacklozenge$), with $k = 0.0064$ ($\blacklozenge$) h$^{-1}$. 
Figure D33. (A) $^{14}$C activity in an experiment with 0.6 g/L goethite, $[\text{H}_2\text{O}_2]_0 = 2$ M, and $[2\text{-CP}]_0 = [\text{NB}]_0 = 90$ µM, with $k = 0.20$ h$^{-1}$. (B) $[2\text{-CP}]$ (▲) and $[\text{NB}]$ (◆) in an experiment with 0.6 g/L goethite, $[\text{H}_2\text{O}_2]_0 = 2$ M, with $k = 0.028$ (▲) and 0.012 (◆) h$^{-1}$. (C) $[\text{H}_2\text{O}_2]$ in the experiments described in (A, ○) and (B, △), with $k = 0.0016$ (△) h$^{-1}$. 
Figure D34. (A) $^{14}$C activity in an experiment with 0.6 g/L goethite, $[\text{H}_2\text{O}_2]_0 = 2$ M, and $[\text{2-CP}]_0 = [\text{NB}]_0 = 100$ $\mu$M, with $k = 0.25$ h$^{-1}$. (B) [2-CP] (▲) and [NB] (♦) in an experiment with 0.6 g/L goethite, $[\text{H}_2\text{O}_2]_0 = 2$ M, with $k = 0.046$ (▲) and 0.019 (♦) h$^{-1}$. (C) $[\text{H}_2\text{O}_2]$ in the experiments described in (A, ○) and (B, △), with $k = 0.0014$ (△) h$^{-1}$. 
Figure D35. (A) $^{14}$C activity in an experiment where the iron was from the supernatant of a 0.6 g/L goethite suspension (see chapter 4 for details) and $[\text{H}_2\text{O}_2]_0 = 1$ M at pH 4 ($\bullet$) or 5 (○), with $k = 0.081$ and 0.032 h$^{-1}$, respectively. (B) $^{14}$C activity in an experiment with 0.6 g/L goethite, $[\text{H}_2\text{O}_2]_0 = 1$ M, and $I = 0.9$ M NaClO$_4$, with $k = 0.016$ h$^{-1}$. (C) $^{14}$C activity in four experiment with 0.6 g/L goethite, $[\text{H}_2\text{O}_2]_0 = 2$ M, and $[2\text{-CP}]_0 = 0.3$ (●), 3 (▽), 30 (■), or 300 (◇) μM, with $k = 0.55, 0.52, 0.42$ and 0.043 h$^{-1}$, respectively. The experiments with $[2\text{-CP}]_0 = 0.3$ and 3 μM were re-spiked with H$^{14}$COOH after ~20 hours. (D) [2-CP] in the experiment described in (C) with $[2\text{-CP}]_0 = 300$ μM, with $k = 0.052$ h$^{-1}$.