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SUMMARY

A widely held memory consolidation theory posits that memory of events and space is initially stored in the hippocampus (HPC) in a time-limited manner and is consolidated in the neocortex for permanent storage. Although studies have demonstrated that post-training HPC lesions result in temporally graded amnesia, the precise HPC circuits and mechanisms involved in remote memory storage remain poorly understood. To investigate the role of the trisynaptic pathway, one of the two major excitatory circuits of the HPC, in the consolidation process we employed the CA3-TeTX transgenic mouse, in which CA3 output can be specifically and inducibly controlled. We found that post-training blockade of CA3 output for up to four weeks impairs the consolidation of contextual fear memory. Moreover, *in vivo* hippocampal recordings revealed reductions in the intrinsic frequency of CA1 ripples and a significant decrease in the experience-dependent enhancement of the ripple-associated coordinated reactivation of CA1 cell pairs during post-run slow wave sleep or awake quiescent periods in the mutant mice. Collectively, these results suggest that the post-training integrity of the trisynaptic pathway and the ripple-associated reactivation of hippocampal memory engram is crucial for memory consolidation.

INTRODUCTION

In both human and animals damage to the hippocampus following learning results in temporally graded impairment of long-term memory (Anagnostaras et al., 1999; Kim and Fanselow, 1992; Squire, 1992). A widely held explanation for these behavioral and clinical data is that over time the episodic, contextual or spatial memory initially stored in the HPC (recent memory) is consolidated to the neocortex for permanent storage (remote memory), thus making the HPC dispensable for its recall and expression (Frankland and Bontempi, 2005; McClelland et al., 1995; Squire and Alvarez, 1995; Teyler and Rudy, 2007). Although there remain uncertainties regarding the relative contributions of the HPC and neocortex as the storage sites of recent and remote memories (Buzsaki, 1996; Clark et al., 2007; Lehmann et al., 2007; Martin et al., 2005; Nadel and Moscovitch, 1997), there is agreement among various consolidation models that interaction between the HPC and neocortex after the experience and formation of recent memory is crucial for the consolidation process. However, little is known as to which HPC circuit is important for this interaction.

The neuronal network of HPC and its adjacent cortex, entorhinal cortex (EC), contains two major excitatory circuits, the trisynaptic pathway (EC layer II $\rightarrow$ dentate gyrus $\rightarrow$ CA3 $\rightarrow$ CA1 $\rightarrow$ EC layer V) and the direct pathway (EC layer III $\rightarrow$ CA1 $\rightarrow$ ECV), that converge onto a common HPC output structure, the CA1 region (Amaral and Witter, 1989). Here we investigate whether, after the encoding of a memory in the HPC, continued output from CA3 via the trisynaptic pathway is necessary for systems consolidation of this memory. This issue is particularly important because, despite the fact that the CA3 recurrent network is thought to be a major storage site for recent HPC memory (Marr, 1971; Rolls and Kesner, 2006), recent work has shown that the alternative direct pathway that bypasses CA3 is necessary for the consolidation

process (Remondes and Schuman, 2004). We addressed this issue by taking advantage of the CA3-TeTX inducible transgenic mouse in which blockade of CA3 output could be targeted only to the post-training period following contextual fear conditioning (Nakashiba et al., 2008). We found that the post-training blockade of CA3 output via the trisynaptic pathway impairs consolidation of contextual fear memory.

Further, we employed in vivo multiple tetrode recordings to the CA3-TeTX mice to obtain physiological correlates of the behavioral deficits. Specifically, we investigated whether high frequency field oscillations ("ripples") in CA1 as well as the ripple-associated reactivation of experience-related firing patterns of CA1 pyramidal cells were impaired in the mutants. Previous studies hypothesized that these physiological processes may be involved in the consolidation process, but evidence has been lacking (Buzsaki, 1989, 1996; Ji and Wilson, 2007; Lee and Wilson, 2002; Siapas and Wilson, 1998; Skaggs and McNaughton, 1996; Wilson and McNaughton, 1994). Our data showed that in the CA3-TeTX mice the intrinsic frequency of ripples and the reactivation were both reduced significantly, supporting the hypothesis that these physiological processes are crucial for memory consolidation.

RESULTS AND DISCUSSION

We previously established that in the CA3-TeTX transgenic mice CA3 output is normal while the animals are raised on a diet containing doxycycline (Dox), but becomes blocked following Dox withdrawal without the converging output to CA1 from the temporoammonic (TA) axons being affected (Nakashiba et al., 2008). More specifically, it requires two weeks of Dox withdrawal from the diet before any significant blockade of transmission occurs at the Schaffer

Collateral (SC)-CA1 synapses, while by the end of the third week of Dox withdrawal the field

excitatory postsynaptic potential (fEPSP) at the SC-CA1 synapses is reduced by over 90%

(Nakashiba et al., 2008). Under the latter conditions, no population spikes were induced, even

after applying high frequency stimulation to the SCs, suggesting that the residual transmission at

the mutants' SC-CA1 synapses would not be able to induce action potentials *in vivo* in CA1

pyramidal cells (Nakashiba et al., 2008). Further, under these conditions, the CA3-TeTX mice

were impaired in the acquisition of contextual fear memory (recent memory) when a 3 min

exposure to a novel context was paired with a mild footshock (one CS-US pair protocol,

Nakashiba et al., 2008). For the present study, a stronger conditioning protocol composed of

three CS-US pairs (see Experimental Procedures) was employed in order to study long-lasting

memory (remote memory). We confirmed that normal SC input to CA1 is necessary for the

acquisition of the full level of contextual fear memory in the 3 CS-US pair version of the task

(Figure 1A).

We then targeted the blockade of the SC-CA1 transmission to the period following the training

session in order to investigate its role in systems consolidation. For this purpose, we raised the

CA3-TeTX mice on Dox diet and switched to a Dox-free diet one week prior to the fear

conditioning training session, ensuring the SC input would be intact during memory acquisition

but would cease about one week after training (Figure 1B). Recent and remote contextual

memories were tested one day and six weeks after training in the same context, respectively,

while recent and remote cued memories were tested in a distinct context one day after the

corresponding contextual fear memory test. As expected, mutants did not exhibit a recent

memory deficit in either contextual or cued conditioning (Figure 1B). In contrast, the mutants'

remote fear memory was significantly reduced in the contextual (Figure 1B and D; $P < 0.05$ for both) but not in the cued conditioning task (Figure 1B). The remaining remote memory may be due to a delay in the completion of the post-training blockade of the SC input and/or the residual, off-line reactivation of CA1 memory traces even under the complete blockade of CA3 output (see below).

The above experimental protocol did not allow us to determine if the observed deficit in remote contextual memory was due to a deficit in either or both the consolidation and/or recall process. To distinguish these possibilities, we subjected a second set of mice to fear conditioning, however in this case withdrawing Dox three weeks after training (Figure 1C). Because there is no significant reduction of the SC input up to 2 weeks after Dox withdrawal (Nakashiba et al., 2008), the mutant mice treated this way have had a total of 5 weeks with an intact SC input following the training, a period sufficient for a full consolidation of recent memory (Kim and Fanselow, 1992). On the other hand, the SC input is blocked over 90% at 3 weeks after Dox withdrawal (Nakashiba et al., 2008), which corresponds to 6 weeks after the training, the time point of the remote memory test. Under this protocol, we did not observe a deficit in either contextual or cued remote memories (Figure 1C and D) in the mutants. Together, these experiments suggest that while CA3 output is not required for recall of a remote contextual fear memory, it plays a crucial role in the consolidation of the memory for a period of several weeks following training, but not beyond.

It has been hypothesized that high frequency synchronized field oscillations ("ripples") observed in CA1 during slow wave sleep or awake quiescent periods (hereafter referred to as SQ periods) and the ripple-associated, off-line "reactivation" of experience-dependent coordinated activity patterns of CA1 cells are crucial for memory consolidation (Buzsaki, 1996; Ji and

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Wilson, 2007; Lee and Wilson, 2002; Siapas and Wilson, 1998; Skaggs and McNaughton, 1996; Wilson and McNaughton, 1994). As possible neurophysiological correlates of the observed behavioral consolidation deficit, we examined ripples and ripple-associated reactivation in the CA3-TeTX mice. For this purpose we used multiple-tetrode drives to record both the local field potential (LFP) and single unit activity from mice as they explored a novel linear track (RUN), as well as during SQ periods that bracketed the RUN session (PRE and POST, respectively). In control mice, the dominant high frequency oscillations during SQ periods had an average intrinsic frequency of 148.84 ± 1.57 Hz, resembling classically defined ripples (Buzsaki et al., 1992; O'Keefe and Nadel, 1978) (Figure 2A and B). In the mutant mice, the lack of CA3 output did not alter the number of high frequency oscillations meeting ripple criteria (Control 0.65 ± 0.07 ripples/sec; Mutant 0.48 ± 0.05 ripples/sec; see SOM), but did result in a significant reduction of the average intrinsic frequency (119.50 Hz ± 1.77 Hz; $P<0.001$) of the oscillations, with the majority (~80%) of events ranging between 110-125 Hz (Figure 2A and B). We did not see a reduction in the length of ripple oscillations in mutant animals compared to controls (42.30 ± 5.38 ms vs. 48.62 ± 5.56 ms, respectively); however, there was a significant reduction in the number of wavelets per ripple epoch in the mutants (4.04 ± 0.49 vs. 2.32 ± 0.41, $P<0.01$). Hereafter, for brevity we refer to the control and mutant oscillations as fast and slow ripples, respectively, or sometimes both simply as ripples.

We next examined the spiking properties of individual pyramidal cells and interneurons during SQ periods to assess the cell types participating in the generation of ripples. There was no difference between genotypes in the mean firing rate of pyramidal cells, in the Complex Spike Index (CSI; a measure of bursting (Nakazawa et al., 2002)) or in the mean burst length, although there was a small decrease in the fraction of spikes participating in bursts in the mutant (Table 1).

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Interneurons in the mutants, however, fired significantly less during SQ periods (Table 1), including during ripple episodes (Figure 2D). Similar to past reports (Csicsvari et al., 1999), in both genotypes, pyramidal cells tended to fire on the trough of ripple wavelets while interneurons fired slightly before or after (Figure 2C and D). This suggests that although ripples in the mutants are significantly slower than those in controls on average, the field oscillations observed in CA1 are likely generated via similar mechanisms; namely a strong synchronous depolarization that is coupled with rapid perisomatic inhibition mediated through the local interneuron network (Ylinen et al., 1995). The source of these excitatory inputs in the mutants may be the TA input from the EC layer III and/or the input from the nucleus reuniens of the thalamus (TR input) (Wouterlood et al., 1990). The appearance of slow ripples, coupled with reduced inhibition in the mutants may reflect a change in the numbers or types of interneurons recruited in the absence of SC input. Alternatively, the decrease in inhibitory tone resulting from SC blockade may have revealed the previously hypothesized ability of the CA1 network to generate intrinsic oscillations in the slow ripple frequency range (Buzsaki et al., 1992) alone or in conjunction with TA/TR input.

While ripples reflect the synchronous discharge of CA1 pyramidal cells and interneurons, it is hypothesized that the key to synaptic plasticity and memory consolidation is the specific co-activation of subsets of cells that reflect the neuronal activity present during learning (Foster and Wilson, 2006; Kali and Dayan, 2004). Previous work has demonstrated that pairs of neurons that are spatially correlated in firing during behavior (RUN) exhibited higher temporal correlations following the experience (POST) compared to the SQ periods preceding it (PRE) (reactivation). We have previously shown that CA1 neurons in CA3-TeTX mice have, on average, poorer spatial specificity than control neurons, particularly in a novel space (Nakashiba et al., 2008). To

focus our experiment on the memory consolidation phase, we limited our reactivation analysis to pyramidal cells with robust place fields in both genotypes based on their spatial information value and average firing rate (Table S1; See Experimental Procedures). We classified pairs of simultaneously recorded cells by the degree of spatial overlap of their fields as overlapping (50% or greater spatial overlap; representative examples shown in Figure 3A and B) or non-overlapping (no spatial overlap; representative examples shown in Figure 3C and D). There were no significant differences between genotypes in the average amount of overlap or in the temporal correlations during run of the overlapping pairs. Further, we observed a significant correlation between the spatial overlap measure and the temporal correlation of the cell's firings during behavior in both genotypes (Table S2). When looking for increased correlations of overlapping cell pairs during POST relative to PRE (Δ correlation) outside of ripples, no increase was observed in either control or mutant mice (data not shown). We then calculated the degree of correlation in the firing of cell pairs during all ripple epochs that occurred before (PRE) and after (POST) spatial exploration and determined the difference between them (Δ correlation). Consistent with earlier work (Wilson and McNaughton, 1994), overlapping pairs in control mice showed an increased correlation of firing during POST-run ripples compared to PRE-run ripples (Figure 3A and E), while non-overlapping pairs showed no change (Figure 3C and E). This increase of coordinated firing between overlapping cell pairs was significantly reduced in the mutants, suggesting impairment in the off-line reactivation of neuronal ensembles in a manner reflecting the animals' experience (Figure 3B and E). Thus, while the firing of CA1 pyramidal cells remain phase locked during ripples (Figure 2C) in the absence of the SC input, this activity does not reflect previous experience as well as it does in the presence of SC input.

While experience-dependent co-activation of overlapping pairs was clearly reduced in mutants, it was still higher than that of non-overlapping pairs (Figure 3E). This may in part be due to the remaining ripple events still present in mutants in which the intrinsic oscillation frequency was within the range of control ripples. To test this hypothesis, we calculated the Δ correlation values only for spikes occurring during ripples whose dominant intrinsic frequency was in the top 15% (i.e., those that fell within the Control ripple frequency band, see Experimental Procedures) and found no significant genotype-specific differences (Figure 3F). This indicates that the minority of ripple events with high intrinsic frequency present in mutants are capable of supporting off-line reactivation of experience-related neuronal ensembles, which may have contributed to the remaining remote memory (Figure 1B and D). Further, we observed a significant decrease in spatial information (bits/spike) in the selected mutant cells (Table S1). To ensure this decrease did not bias the deficit in off-line reactivation, we removed cell pairs from either the highest quality control pairs, the lowest quality mutant pairs or both to the point in which the significant decrease was no longer observed (i.e., $P>0.1$). With these limitations, we did not find a relationship between the degree of ripple related reactivation and spatial information in either genotype (Table S3).

Here we demonstrated that blockade of CA3 output targeted to a post-training period of about four weeks led to a specific deficit in the remote memory of contextual fear without affecting its recall or the remote memory of cued fear. These results suggest that the continued CA3 output and the integrity of the trisynaptic pathway after the formation of recent memory in the HPC is important for consolidation of contextual fear memory for a limited period of up to about four weeks, but not beyond. Our results do not exclude the possibility that other hippocampal pathways also contribute to consolidation of hippocampal dependent memory. For

instance, the parallel direct pathway, which bypasses CA3, was shown to be crucial for the consolidation of a HPC-dependent spatial memory in a recent study carried out on rats with the electrolytic lesions of TA axons (Remondes and Schuman, 2004). In that study, however, the area affected by the lesion may have extended to the subiculum, impacting CA1 output and hence blocking, at least partially, the trisynaptic pathway. Nevertheless, it is possible that both trisynaptic and direct pathways contribute to the consolidation process.

Our present work also demonstrated that a highly specific blockade of CA3 output to CA1 resulted in reductions in the intrinsic oscillation frequency of ripples in CA1 as well as a significant decrease in the experience-dependent enhancement of the ripple-associated coordinated reactivation of CA1 cell pairs during post-run SQ periods. When combined with the behavioral deficit observed in the same mutant mice, these results suggest that ripples with high oscillation frequencies and ripple-associated reactivation of experience-related neuronal firing patterns during SQ periods play a crucial role in the consolidation of hippocampal-dependent recent memories into a remote memory that no longer requires CA3 drive. Although the physiological and behavioral deficits observed in mutant mice are correlatory, the association is strengthened by the exquisite specificity of the genetic intervention method employed (Nakashiba et al., 2008). In contrast, the possible role of ripples and ripple-associated reactivation or replay in memory consolidation was previously inferred only on the grounds that such physiological activities would strengthen synaptic plasticity at various sites downstream of CA1, including some cortical regions (Buzsaki, 1989; Foster and Wilson, 2006; Ji and Wilson, 2007).

The intrinsic oscillation frequency of a minor fraction of mutant ripples was in the normal range and a fraction of the experience-dependent increase in the co-activation of CA1 cell pairs

remained in these mice. While it could be argued that these remaining activities reflect "leakiness" in the blockade of transmission at SC-CA1 synapses, we believe this is unlikely because our earlier studies showed that residual synaptic transmission at SC inputs in mutants failed to elicit population spikes in the fEPSP at stimulation intensities far greater than that expected *in vivo* (Nakashiba et al., 2008). We believe that a more likely interpretation of the remaining high frequency ripples in the mutants is that in the absence of SC input, TA and/or TR input can trigger these ripples, albeit less consistently than SC input. Likewise, the remaining experience-associated correlation of CA1 cell pair firings suggest that the TA and/or TR input can provide experience-related information to CA1, albeit less efficiently than SC input. For instance, the multisite recurrent network CA1 → ECV → ECIII → CA1 (Naber et al., 2001) could associate and store a diverse set of information.

In summary, using a mutant mouse in which the CA3 to CA1 synaptic transmission can be specifically inhibited in a temporally controllable manner, we demonstrated the importance of post-recent memory integrity of the HPC trisynaptic pathway for memory consolidation. We also demonstrated that in the mutant mice, CA1 ripples and the ripple-associated reactivation of experience-dependent firing patterns of CA1 neurons are impaired, supporting the hypothesis that these physiological mechanisms underlie the consolidation of HPC-dependent memory.

Table 1. Firing properties of CA1 pyramidal cells and interneurons during SQ periods.

	Control ($N = 14$, $n = 94$)	Mutant ($N = 12$, $n = 91$)
Pyramidal cells		
Mean firing rate (Hz)	0.789 +/- 0.08	0.90 +/- 0.09
Complex spike index (bursting)	29.20 +/- 1.62	28.93 +/- 1.47
Fraction of Spike in Bursts (% total)	37.9+/- 1.23	34.5+/- 1.06*
Mean Burst Length (# of spikes)	2.16 +/- 0.06	2.14 +/- 0.07

	Control ($N = 15$, $n = 56$)	Mutant ($N = 13$, $n = 75$)
Interneurons		
Mean firing rate (Hz)	9.34 +/- 1.00	4.61 +/- 0.68**
Complex spike index (bursting)	1.51 +/- 0.43	1.94 +/- 0.50

* = P =0.042
** = P <0.001

We examined the spiking activity during SQ periods of high spatial information pyramidal cells and interneurons recorded in area CA1. We found a small, yet significant decrease in the fraction of spikes that participated in bursts in mutant mice, however, no differences were observed in CSI, mean rate or mean burst length. Interestingly, interneurons in mutants showed a significant decrease in mean firing rate during SQ periods.

N, number of mice; *n*, number of cells.

FIGURE LEGENDS

Figure 1. Consolidation of contextual, but not cued fear memory, is impaired in CA3-TeTX mice. **(A)** Averaged freezing levels during the recent contextual memory test following 3 weeks Dox withdrawal (Control; blue bar, Mutant; red bar, n = 12 per genotype). Mutants exhibited significantly less freezing (P < 0.05). **(B)** Averaged freezing levels during the contextual and cued memory tests. Dox was withdrawn 1 week before training (-1 week) and recent and remote memories were tested (n = 24 per genotype). Mutant mice displayed significantly less freezing in the remote contextual memory test compared to the control littermates (P < 0.05), but not in the recent contextual memory test. In cued memory tests, mutants and controls showed similar freezing levels in both recent and remote memory tests. **(C)** Averaged freezing levels during the contextual and cued memory tests. Dox was withdrawn three weeks after training (+3 weeks). Mutants exhibited no deficit in either recent or remote contextual or cued memory. Protocols for Dox^+ to Dox^- diet switches relative to the timing of training and memory tests are indicated at the top of each panel (Figure 1A to C). **(D)** Freezing levels of individual mice during remote memory tests were divided by those during the remote memory tests (retention index) for contextual fear conditioning (Control; blue circle, Mutant; red circle, P <0.05 for the -1 week protocol). Data shown as mean ± s.e.m.

Figure 2. Loss of CA3 output to CA1 results in a decrease in the intrinsic frequency of ripple oscillations. (A) LFP samples from Control (top) and Mutant (bottom) mice during SQ periods. Detected ripples are highlighted in either blue (Control) or red (Mutant). Below the LFP traces are individual ripple examples from six randomly selected Controls (blue) and Mutants (red). Note the clear decrease in the Mutant ripple intrinsic frequency. **(B)** The amplitude of ripples is not altered in Mutants, however, there was a significant decrease in the intrinsic frequency of Mutant (N=12) ripples compared to Controls (N=14; P = 4.31e-19). **(C and D)** Both phase preference and probability of firing of pyramidal cells **(C)** were consistent between Mutants and Controls. Although the phase preference of interneurons did not change in the Mutants, the probability of firing was significantly lower in the Mutants **(D)** (P = 0.006). Data shown as mean ± s.e.m.

Figure 3. Blockade of CA3 to CA1 transmission results in a loss of reactivation during post-behavior SQ periods. (A and B) Representative examples of Control (left) and Mutant (right) cell pair correlations during pre-run and post-run SQ periods. Heat maps (right of graphs) illustrate the spatial overlap between the two cells in the novel environment. Note that although the place fields of Mutant cells had significant overlap, little or no increase in their pair-wise correlation was observed during post-SQ periods. **(C and D)** Representative examples of cell pair correlations for cells with non-overlapping place fields. Note the lack of pair-wise correlation increase during post-SQ periodsfor both Controls and Mutants. **(E)** Quantification of all pair-wise correlations during ripple epochs(Control: 91 cells from 14 mice, overlapping pairs=98, non-overlapping pairs=229; Mutant: 94 cells from 12 mice, overlapping pairs=132, non-overlapping pairs=215). Both genotypes show a significant increase in correlation when their place fields overlap and when taking all ripples into account. Note the significant reduction in Δ correlation (post-SQ periodscorrelation – pre-SQ periodscorrelation) in Mutant (red line) animals compared to Controls (blue line two-way analysis of variance, genotype x overlap F(1,670=3.96, P=0.047; Bonferroni post-tests: Control non-overlapping x overlapping, P<0.001; CA3-TeTX non-overlapping x overlapping, P<0.01; Control overlapping x CA3-TeTX overlapping, P<0.01). **(F)** Evaluation of reactivation during ripples only falling into the average Control ripple frequency. When Mutant ripples were in the average Control ripple frequency range, the deficit in Δ correlation was not observed. Data shown as mean ± s.e.m.

EXPERIMENTAL PROCEDURES

Animals
All the experiments were carried out by operators blind to the animal's genotype using male

CA3-TeTX mice (strain C57BL/6) between 14 and 24 weeks of age and their control male

littermates. The specificity, completeness, and time course of blockade of synaptic transmission

in CA3-TeTX mice were reported previously(Nakashiba et al., 2008). Two to four mice were

housed per cage under the conditions of a 12 hr light/dark cycle and ad libitum access to food

and water. All procedures relating to animal care and treatment conformed to the Institutional

and NIH guidelines.

In vivo electrophysiology
All experiments were performed by operators who were blind to the genotypes. Animals were

implanted with a microdrive array consisting of six independently adjustable tetrodes (targeted to

CA1: stereotaxic coordinates from bregma: 1.6 mm lateral; 1.8 mm posterior) as previously

described (McHugh et al., 1996; Nakashiba et al., 2008). In brief, electrodes were slowly

advanced over one week to reach the CA1 pyramidal cell layer, and recordings commenced once

stable cells were present. Putative hippocampal cells and local field potential (LFP) were

recorded during a 20-30 minute SQ session before and after (PRE and POST SQ periods,

respectively) a 10-minute exposure to a novel linear track (~81cm, RUN). Position and

directionality was tracked using a pair of infrared diodes placed 3 cm above the animals' head

and 3 cm front to back. To eliminate periods of immobility, only periods when the animals were

running above 2cm/sec were used for the pair-wise correlation analysis. For SQ, only periods in

which the animals were completely still for greater than 30 sec were used. SQ periods were

further determined using the theta/delta ratio (Csicsvari et al., 1999). After recordings were

completed, mice were given a lethal dose of anesthetic and an electrical current ($50\mu A$) was run

down each tetrode for 8 seconds to create a small lesion at the tip of the probe. Animals were then transcardially perfused with 4% PFA in PB and brains were removed, coronal slices (50µM thick) were prepared using a Vibrotome, mounted and finally counterstained with Nuclear Fast Red to visualize electrode tracks and lesion sites. Recording positions were verified by examination of the lesion sites under standard light microscopy.

Individual neurons were classified offline based on each action potential's relative amplitude across the four recording wires of a tetrode (McHugh et al., 1996). Cells were further classified as pyramidal cells or interneurons based on the average width of their waveforms, bursting properties, and autocorrelation histograms (Csicsvari et al., 1998). Given the previously reported encoding deficits in the CA3-TeTX mice (Nakashiba et al., 2008), we limited the reactivation analysis to cells in both genotypes that best represented the spatial environment. For inclusion, cells were required to have a spatial information index (bits/spike) of >0.5 and an average firing rate of >0.2 Hz on the linear track. Place fields were calculated by counting the number of spikes falling into 2cm bins along the linear track. Bins that had a firing rate below 1 Hz and bins containing less than 10% of the spikes contained in the maximum bin were excluded in the overlap analysis. We then classified all cell pairs as overlapping or non-overlapping based on the average place field location. Overlapping place fields were defined as cells firing on the linearized track with $>50\%$ overlapping bins. Non-overlapping place fields were defined as those with 0% overlap, and all other pairs were discarded. Pair-wise correlations during run were calculated in 10 ms bins +/- 1000 ms. Pair-wise correlations during SQ periods were calculated in 10 ms bins +/- 100 ms from the peak of the ripple or outside of the ripple. Correlation peaks were calculated if the value at time 0 was 3 s.d. above the mean.

To verify that the criterion of spatial information content did not influence the reactivation properties of the control or mutant cell pairs, the data was segregated in three ways: 1) We removed only good control cell pairs to the point in which the spatial information content between controls and mutants was no longer significant (p>0.1), 2) We removed only poor mutant cell pairs to the same point, and 3) We removed both good control cell pairs and poor mutant cell pairs to the same point.

Ripples were detected using modifications to methods previously described (Csicsvari et al., 1999). The LFP was band-pass filtered between 90 and 250 Hz using a Hamming-window based FIR filter. To extract ripple events, the power (root mean square) of the filtered data was quantified and the beginning, middle, and ends of ripple periods were classified for all epochs >5 s.d. The troughs of individual ripple wavelets were then detected by finding the minimum points during each episode that were greater than 20% of the maximum peak amplitude. For cleaner detection of the power and peak frequency for each ripple, the multi-taper method(Buhl et al., 2003) was performed on the filtered ripple epochs. "Fast" (>140Hz) ripples were then classified using a ratio of the power between the 110-125Hz and 140-180Hz frequency bands. Only ripples with significantly higher power in the upper band were classified as "fast".

Behavioral experiments
All experiments were performed by operators who were blind to the genotypes and Dox treatments. For fear conditioning, room and chamber settings were the same as previously described (Nakashiba et al., 2008). Training consisted of a single exposure to the conditioning context for 6 minutes, during which three pairs of tone and electrical foot shock were given. Once placed in the chamber, mice were allowed to freely explore for 3 minutes. We then played a 30 second tone (2KHz; 70db) that co-terminated with a 1.00 mA foot shock (1 s in duration).

The mice received two additional tone and shock pairings at an inter-trial interval of 1 minute. Following the last shock delivery, mice remained in the chamber for 30 seconds. A recent contextual fear memory test was conducted 24 hours later, when the mice were placed in the conditioning chamber for 3 minutes without any tone presentation. A recent tone fear memory was measured in a second distinct chamber 48 hours after the training. The same tone (2 min. in duration) as the one used in the training session was delivered after the mice were placed in the second chamber for 3 minutes. Six weeks after the training session, a remote contextual fear memory test was conducted followed by a remote tone fear memory test on the next day. Procedures for the remote memory tests were the same as the ones used in the recent memory tests.

SUPPLEMENTAL DATA

The Supplemental Data includes three tables.

ACKNOWLEDGEMENTS

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