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Citation: Bochkis, Irina M., Dariusz Przybylski, Jenny Chen, and Aviv Regev. "Changes in Nucleosome Occupancy Associated with Metabolic Alterations in Aged Mammalian Liver." *Cell Reports* 9, no. 3 (November 2014): 996–1006.

As Published: <http://dx.doi.org/10.1016/j.celrep.2014.09.048>

Publisher: Elsevier B.V.

Persistent URL: <http://hdl.handle.net/1721.1/96715>

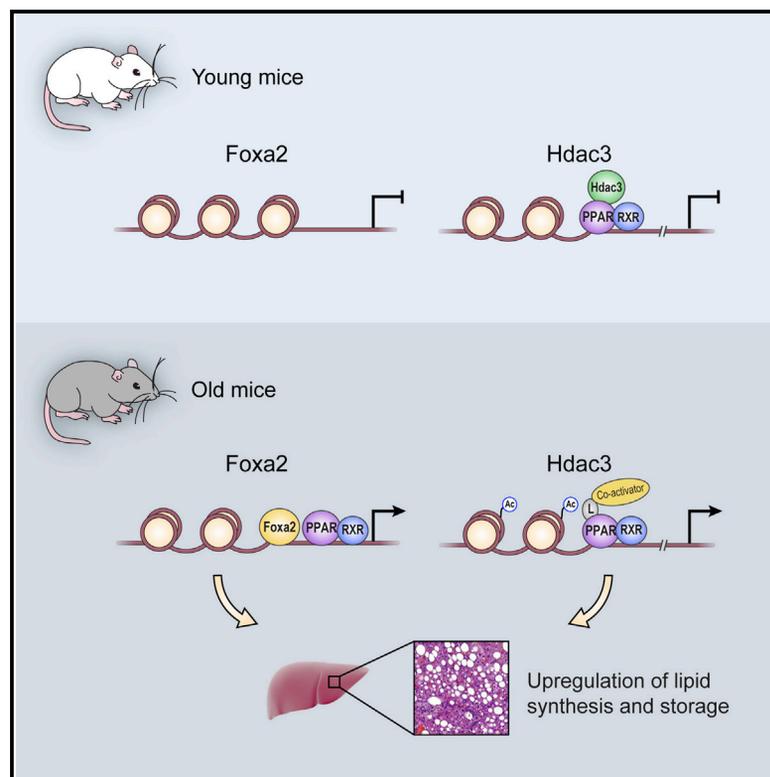
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Changes in Nucleosome Occupancy Associated with Metabolic Alterations in Aged Mammalian Liver

Graphical Abstract



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In Brief

Bochkis et al. use genome-wide profiling in the livers of young and old mice to observe age-dependent changes in PPAR α targets as well as nucleosome occupancy linked to Foxa2 and Hdac3 binding sites. A reciprocal binding pattern of Foxa2 and Hdac3 at PPAR α targets contributes to gene-expression changes that lead to steatosis in the aging liver.

Highlights

An *in vivo* genome-wide nucleosome map in the aging liver is described

Foxa2 binds regions of decreased nucleosome occupancy at PPAR α targets in old livers

Hdac3 and Srf are implicated in age-dependent metabolic dysfunction

Reciprocal binding of Foxa2 and Hdac3 at PPAR α targets contributes to steatosis

Accession Numbers

GSE58006

GSE57809

GSE58005

GSE60393



Changes in Nucleosome Occupancy Associated with Metabolic Alterations in Aged Mammalian Liver

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<http://dx.doi.org/10.1016/j.celrep.2014.09.048>

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SUMMARY

Aging is accompanied by physiological impairments, which, in insulin-responsive tissues, including the liver, predispose individuals to metabolic disease. However, the molecular mechanisms underlying these changes remain largely unknown. Here, we analyze genome-wide profiles of RNA and chromatin organization in the liver of young (3 months) and old (21 months) mice. Transcriptional changes suggest that derepression of the nuclear receptors PPAR α , PPAR γ , and LXR α in aged mouse liver leads to activation of targets regulating lipid synthesis and storage, whereas age-dependent changes in nucleosome occupancy are associated with binding sites for both known regulators (forkhead factors and nuclear receptors) and candidates associated with nuclear lamina (Hdac3 and Srf) implicated to govern metabolic function of aging liver. Winged-helix transcription factor Foxa2 and nuclear receptor corepressor Hdac3 exhibit a reciprocal binding pattern at PPAR α targets contributing to gene expression changes that lead to steatosis in aged liver.

INTRODUCTION

Aging is associated with increased prevalence of metabolic disease and cancer, reduced capacity for tissue regeneration, and physical decline (Rodriguez et al., 2007; Willis-Martinez et al., 2010). In particular, triglyceride accumulation is the common metabolic phenotype of aging liver and of metabolic syndrome, an age-related disorder that increases the risk of developing diabetes and cardiovascular disease. While mechanisms of age-dependent defects in tissue regeneration have been studied extensively (Jin et al., 2010; Jin et al., 2009), causes of age-onset metabolic impairments remain largely unknown.

Previous studies suggest a role for chromatin and nuclear organization in aging-associated lipid dystrophies. First, changes in chromatin organization mediate age-dependent impairments in several tissues (Chambers et al., 2007; Jin et al., 2010).

Furthermore, mutations in lamin A/C (*LMNA*), a nuclear envelope protein, cause progeria, a premature aging syndrome, and lamin-dependent defects have been connected to physiological human aging (Scaffidi and Misteli, 2006). *LMNA* is also mutated in partial lipodystrophy (Shackleton et al., 2000), a condition associated with insulin-resistant diabetes and hepatic steatosis. Clinical features of lipodystrophy due to mutations in *LMNA* closely resemble those in individuals with mutations in *PPARG*, a nuclear receptor involved in pathogenesis of fatty liver (Gavrilova et al., 2003). Progeria attributed to defects in DNA repair (Schumacher et al., 2009) is modeled in mice by deletion of *Ercc1*, an enzyme crucial to nucleotide excision repair (Niedernhofer et al., 2006). *Ercc1* mutants exhibit upregulation of targets of the hepatic nuclear receptors (PPAR α and PPAR γ) and hepatic steatosis. Finally, liver-specific loss of *Foxa2*, a pioneer transcription factor regulating nucleosome dynamics, leads to premature aging, increased hepatic lipogenesis, and age-onset obesity (Bochkis et al., 2013). While the progeroid models described above propose a role for epigenetic conformation in modulating age-dependent metabolic impairments, this hypothesis has not yet been tested in a model of physiological aging.

To study the relation between chromatin organization and age-dependent metabolic impairments in the liver, we examined gene expression and genome-wide nucleosome occupancy in livers isolated from young (3 months) and old (21 months) mice (Figure 1A). Age-dependent induction of expression in lipid synthesis and storage genes, similar to metabolic changes seen in progeroid syndromes, is consistent with derepression of the nuclear receptors PPAR α , PPAR γ , and LXR α . Analysis of transcription factor binding sites that are overrepresented in regions where nucleosome occupancy changes with age identified established regulators of age-dependent metabolic dysfunction and lamina-associated candidates. Winged-helix transcription factor Foxa2 that regulates nucleosome dynamics binds regions of decreased nucleosome occupancy at PPAR α targets in old livers. Conversely, binding of nuclear receptor corepressor Hdac3, detected from motifs found in regions of increased nucleosome occupancy, exhibits a reciprocal pattern. Together, altered Foxa2 and Hdac3 occupancy at PPAR α targets contributes to gene expression changes that lead to steatosis in aged liver.

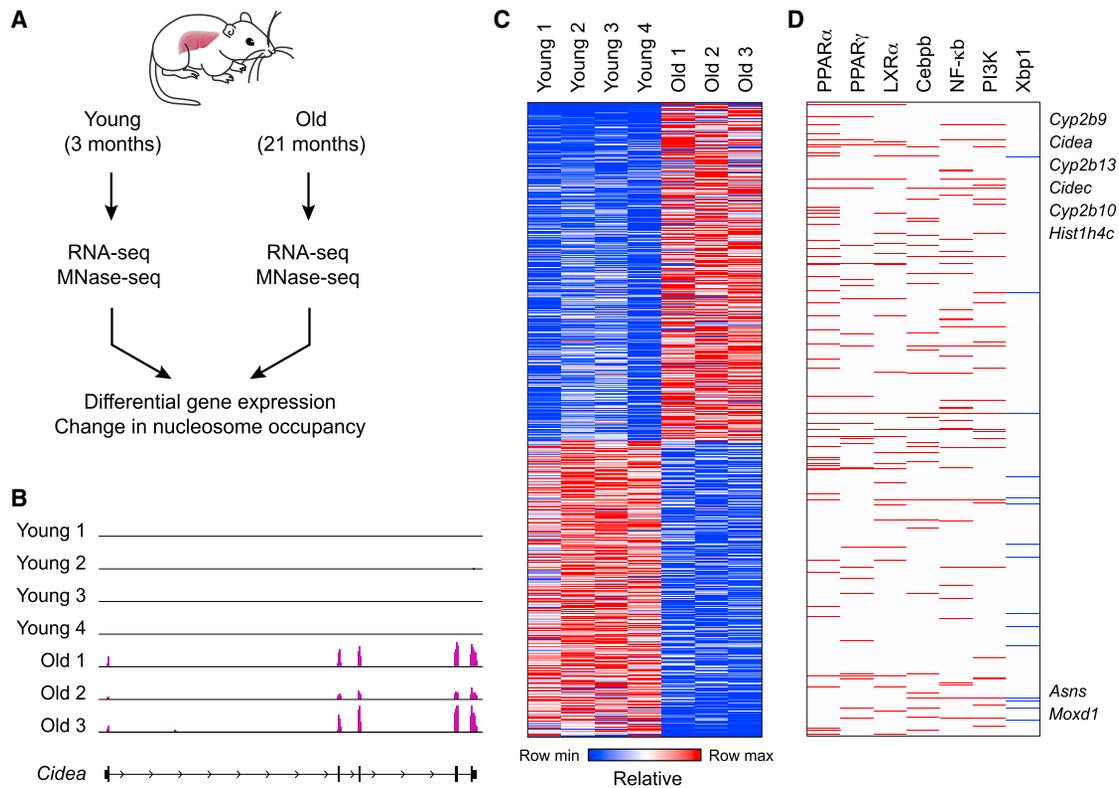


Figure 1. Nuclear-Receptor-Dependent and Inflammatory Targets Are Induced during Physiological Aging in the Liver

(A) Experimental design includes comparing gene expression (RNA-seq) from livers of young (3 months), and old (21 months) and nucleosome occupancy (MNase-seq) of young and old livers.

(B) RNA-seq track view in Integrative Genome Viewer (IGV) of expression of *Cidea*, a target of nuclear receptors *Ppar α* and *Ppar γ* , not expressed in young healthy livers and highly induced in steatotic older hepatocytes.

(C) Heatmap of differentially expressed genes in aged livers (FDR 5%, 727 upregulated and 525 downregulated).

(D) Regulators of gene expression in heat map in (C). Factors regulating lipid synthesis and storage are nuclear receptors *Ppar α* (p value 1.1×10^{-20}), *Ppar γ* (p value 2.7×10^{-8}), *LXR α* (p value 4.2×10^{-13}), and *Cebpb* (p value 2.1×10^{-9}). Additional regulators include *NF- κ B* (p value 1.8×10^{-3}), *PI3K* (p value 6.4×10^{-6}), and *Xbp1* (p value 3.6×10^{-4}). Networks with upregulated and downregulated targets are shown in red and blue, respectively. p values were determined by the Fisher's exact test (using IPA).

RESULTS

Inflammatory and Nuclear Receptor Target Genes Are Induced in Aged Liver

To characterize transcriptional changes in aging, we profiled gene expression by RNA sequencing (RNA-seq) in young (3 months) and old (21 months) male mice (three to four animals per age group; Figure 1A) and identified 1,252 genes differentially expressed between young and old mice (727 induced with age; 525 repressed; false discovery rate [FDR] 5% edgeR; Robinson et al., 2010; Experimental Procedures; Table S2). Consistent with previous studies of genetically induced aging (Niedernhofer et al., 2006), genes induced with age were enriched for "Lipid & Fatty Acid Metabolism" functions (88 of 1,157 pathway genes, p value = 4.1×10^{-9} , Fisher's exact test). Among the highly induced genes were *Cidea*, a target of nuclear receptors *PPAR α* and *PPAR γ* , encoding a lipid-associated protein only detected in fatty and diabetic livers (Gong et al., 2009); related family members *Cideb* and *Cidec*, cytochrome p450 detoxification enzymes (*Cyp2b9*, *Cyp2b10*, and

Cyp2b13) involved in stress response; and histone H4 transcript (*Hist1h4c*; Figures 1B–1D). In contrast, mRNA levels of *Moxd1*, an enzyme in the endoplasmic reticulum (ER) and *Asns*, an enzyme upregulated by ER stress response, are downregulated in aged hepatocytes.

Genes induced with aging were also enriched for known targets of key transcriptional regulators of lipid homeostasis, including peroxisome proliferator activated receptors (*PPAR α* and *PPAR γ* ; p values 1.1×10^{-20} and 2.7×10^{-8} , respectively, Ingenuity Pathway Analysis [IPA], Fisher's exact test), liver X receptor (*LXR α* , *Nr1h3*, p value 4.2×10^{-13}), *PGC1 α* (p value 1.8×10^{-4}), a coactivator of PPARs, and *CEBPB* (p value 2.1×10^{-9}), known to coregulate *PPAR γ* targets (Lefterova et al., 2008) (Figures 1D, S1A, and S1B; Table S3). IPA also identifies gene networks regulated by agonists for *PPAR α* (pirinixic acid/WY-14643 and clofibrate, p values 9.6×10^{-20} and 1.8×10^{-7} , respectively) and *LXR α* (T0901317, p value 4.2×10^{-13}), suggesting that *PPAR α* and *LXR α* are ligand activated in aged liver. In addition, genes activated by inflammatory regulators (*NF- κ B/RELA*, *IRF3*, and

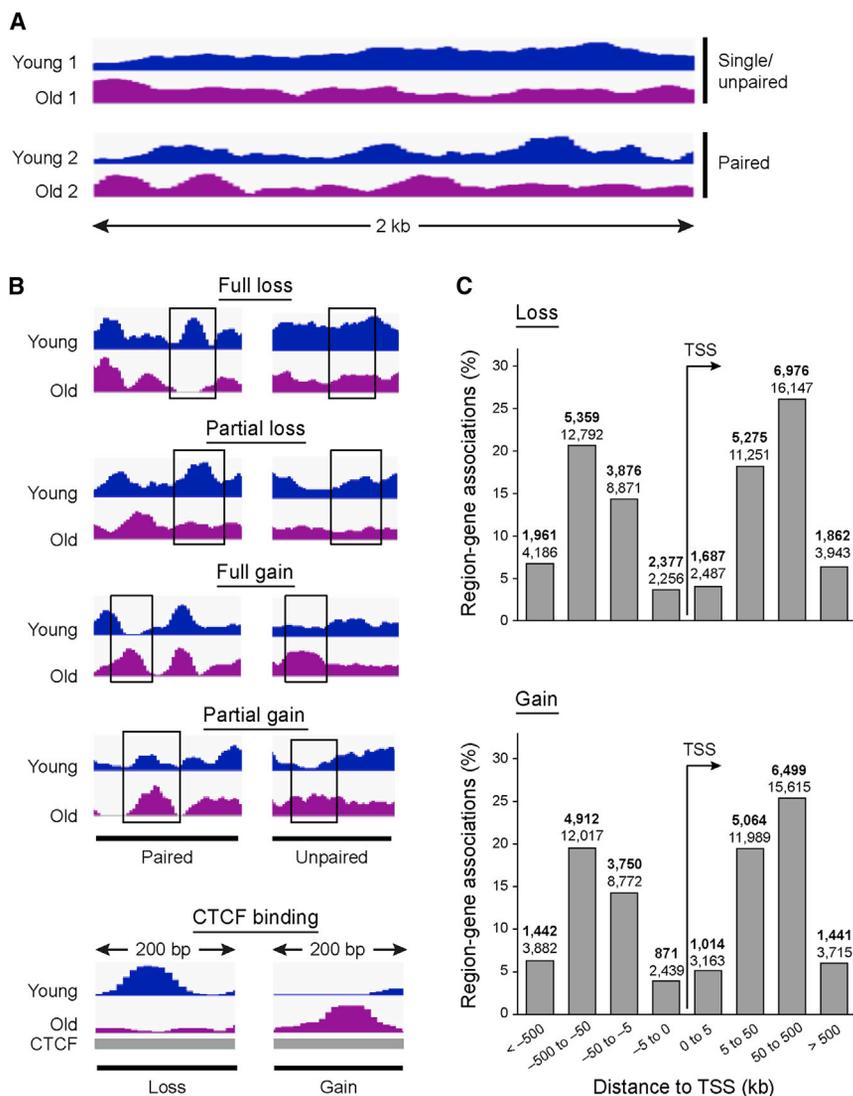


Figure 2. Characterization of Regions of Age-Dependent Change in Nucleosome Occupancy

(A) MNase-seq tracks show nucleosome occupancy of young (3M, blue) and old (21M, purple) biological replicates in a 2 kb region (top, unpaired; bottom, paired).

(B) Examples of loss (full and partial, two top panels) and gain (full and partial, two middle panels) in nucleosome occupancy for paired (left) and unpaired (right) replicates. Examples of loss and gain in nucleosome occupancy overlapping a CTCF binding site (bottom panel).

(C) The distribution of age-dependent nucleosome occupancy change regions (both losses and gains) around the TSS. Values for replicate 1 (in bold) and replicate 2 are shown above each bar. The majority of changes occur distally, 50 to 500 kb from the TSS.

TLR4; p values 1.8×10^{-3} , 2.9×10^{-2} , and 2.1×10^{-3} respectively) are also upregulated in aged mice (Figures 1D, S1A, and S1B; Table S3).

These results are consistent with a model where one or more of these regulators is activated during aging, leading to increased transcription of key lipid metabolism genes. Notably, PPARs are upregulated in progeroid syndromes, LXR α is activated in prematurely aged *Foxa2* mutants (Bochkis et al., 2013), and inflammatory signaling is known to be activated by dietary fatty acids and contributes to insulin resistance (Fessler et al., 2009).

Age-Dependent Changes in Nucleosome Occupancy Are Enriched in Distal Elements

To study the role of chromatin organization in these age-dependent changes in transcription, we next measured genome-wide nucleosome occupancy profiles in livers from young (3 months) and old (21 months) mice. We used MNase digestion of chromatin followed by sequencing (MNase-seq; Experimental Proce-

occupancy changes of well-positioned nucleosomes (29,355 and 61,869 regions of loss and 24,979 and 61,540 regions of gain for replicates 1 and 2, respectively; Figure 2B). Finally, similar pathways are enriched for genes with changes in nucleosome occupancy in either replicate (Figure S3B).

Most age-dependent changes in nucleosome occupancy occur distally, 50 to 500 kb from the transcription start site (TSS; Figure 2C). Furthermore, there is an overlap between regions with changes in nucleosome occupancy and those bound the insulator CTCF or marked with H3K4me1 (10% and 9% of all regions with occupancy change for replicate 1 and 2, respectively) or with H3K9ac (9% and 3% of all regions with occupancy change for replicate 1 and 2, respectively) in young livers, modifications associated with enhancers and active euchromatin, respectively. This suggests that distal enhancers, known to be bound by Foxa factors and nuclear receptors (Bochkis et al., 2012; Lefterova et al., 2008), may be the genomic features most associated with nucleosome occupancy changes.

Foxa2 Binds Regions of Reduced Nucleosome Occupancy at PPAR α Targets in Aged Liver

To identify transcriptional regulators whose binding may be affected by changes in nucleosome positions, we performed both de novo motif discovery (using MEME; Bailey et al., 2009; Experimental Procedures) and positional weight matrix (PWM) scan analysis (using PscanChIP; Zambelli et al., 2013; Experimental Procedures) of 150 bp windows spanning all nucleosomes gained or lost with age for either replicate.

Motifs associated with forkhead transcription factors are the most overrepresented in regions of changed occupancy (both gain and loss) for both replicates. FOXO factors, transducers of insulin signaling, can decompact chromatin (Hatta and Cirillo, 2007), and FOXA2 is known to bind nucleosomal DNA in vivo (Li et al., 2011). Indeed, FOXA2 targets (defined as differentially expressed in 1-year-old *Foxa2* mutants versus wild-type [WT] liver; Bochkis et al., 2013) are particularly differentially expressed between our WT old versus young livers (Figure S3C; Kolmogorov-Smirnov [K-S] p value 2.4×10^{-9}).

Motifs for nuclear receptors (ROR α p value 3.1×10^{-56} , LXR p value 7.6×10^{-85} , PPAR p value 9.7×10^{-76}) and interferon regulatory factors (IRF p value 8.1×10^{-80}) were highly enriched in regions of nucleosome occupancy loss (Figure 3A). Protein expression of the lipid metabolism regulators FOXA2 and PPAR α is not altered in aged livers, while that of PPAR γ is induced, consistent with previous reports of PPAR γ induction in fatty liver (Panasyuk et al., 2012) (Figure 3B). IPA of genes with nucleosome occupancy loss near TSSs identified networks regulated by ligand-activated nuclear receptors PPAR α , PPAR γ , and LXR α . Expression was induced for one-quarter of these targets (fold change ≥ 1.3 ; Figure 3C).

Since *Foxa2* plays an important role in lipid metabolism in aged liver (Bochkis et al., 2013), we next examined *Foxa2* binding in young and old livers using chromatin immunoprecipitation sequencing (ChIP-seq). Strikingly, *Foxa2* occupies substantially more sites in old hepatocytes (12,834) than in young ones (6,605; Figure 3D). Additional bound regions in the old livers are found at PPAR target genes as well as at the *Ppara* promoter (Figure 3E). The increase in binding is not due to change in FOXA2 expression (Figure 3B).

A subset of 734 *Foxa2* sites that are “gained” in old livers correspond to regions of decreased nucleosome occupancy. Interestingly, while the majority of changes in nucleosome occupancy occur distally, these sites are found mostly at promoters (Figure 3F). Genes associated with the binding sites are enriched in functional categories including “hepatic steatosis” (p value 1.5×10^{-4}), “nuclear import” (p value 3.8×10^{-4}), and “increased circulating VLDL” (p value 3.5×10^{-5}). In addition, the PPAR/DR-1 element is enriched at these sites, suggesting that age-dependent *Foxa2* binding could enable PPAR binding at these regions. Alternatively, newly bound FOXA2 at the promoter may interact with PPAR proteins that are bound to existing distal enhancer elements to activate PPAR targets (Figure 6).

Increased Nucleosome Occupancy Suggests a Role for cKrox-Hdac3 in Age-Dependent Dysfunction

Regions associated with increased nucleosome occupancy were enriched, among other elements, with a motif bound by

Srf (p value 3.3×10^{-45}), a factor that interacts with the nuclear lamina (Swift et al., 2013) (Figure 3A) and with motifs identified as a GAGA repeat (short motif p value 1.8×10^{-111} , long motif p value 2.3×10^{-8} ; Figure 4A). The short GAGA motif is frequent along the genome (background frequency 0.416), but the prevalence of this sequence in the regions of age-dependent occupancy gain was higher (frequency 0.527). The GAGA repeat is associated with lamina-associated domains (LADs) (Lund et al., 2013) and is bound by the transcriptional repressor cKrox (*Zbtb7b*) in a complex with Hdac3. While SRF protein levels decrease, expression of HDAC3 is not altered in older livers (Figure 4B). Hdac3 can act as a corepressor of nuclear receptors regulating hepatic lipid metabolism genes (Knutson et al., 2008; Sun et al., 2012). Furthermore, the Ncor complex, an activating cofactor for Hdac3 (Guenther et al., 2001), is predicted to be repressed in our aged livers based on the expression of its known targets (IPA; Experimental Procedures). Taken together, this suggests that Hdac3 activity may also be reduced in older animals, resulting in induction of its targets by nuclear receptors.

To test this hypothesis, we first compared the genes differentially expressed between (young) *Hdac3* liver-specific mutants versus WT (Knutson et al., 2008) with the genes differentially expressed between aged and young livers. We found that the Hdac3-dependent genes are particularly differentially expressed in old versus young livers (compared to the background of all old versus young differentially expressed genes; K-S p value 2.2×10^{-16}). In addition, genes from the same biological functions and pathways were affected in both young *Hdac3* mutant mice and differentially expressed genes in aged livers (Figures 4C and 4D).

Next, we used ChIP-seq to measure Hdac3 binding in young and old livers and found markedly less occupied regions in old livers (5,828 in young, 2,930 in old; Figure 4E), especially at PPAR-dependent genes. PPAR response element (DR-1) was significantly overrepresented in the sequences bound by Hdac3 in young, but not old, livers (p value 1.4×10^{-31}). While the overlap between Hdac3 sites bound in young livers and regions of increased nucleosome occupancy is small (152 regions), the GAGA motif is also enriched at sites bound by Hdac3 in the young livers (short motif p value 7.0×10^{-44} , long motif p value 3.6×10^{-13} ; Figure 4F), suggesting the overlap could be greater with deeper coverage of MNase-seq data and more statistically significant nucleosome occupancy changes calls.

Sites for serum response factor (SRF) were also enriched in regions with age-dependent gain of nucleosome occupancy (p value $< 6 \times 10^{-74}$, PScan ChIP analysis for overrepresented PWMs). Furthermore, SRF-dependent genes were differentially expressed in older livers (IPA analysis; Experimental Procedures; Figure 4G), similar gene functions are enriched in genes differentially expressed in liver-specific *Srf* mutants (versus WT) or in old liver (versus young; Figure 4H), and most (36 out of 48) *Srf* target genes with age-dependent gain in nucleosome occupancy near the TSS are induced in aged livers (Figure 4I). Altogether, these data suggest a downregulation of *Srf* activity with aging, consistent with reduced SRF protein levels (Figure 4B), leading to derepression of its gene targets. *Srf* activity has also been shown to be repressed during senescence by nuclear exclusion (Ding et al., 2001).

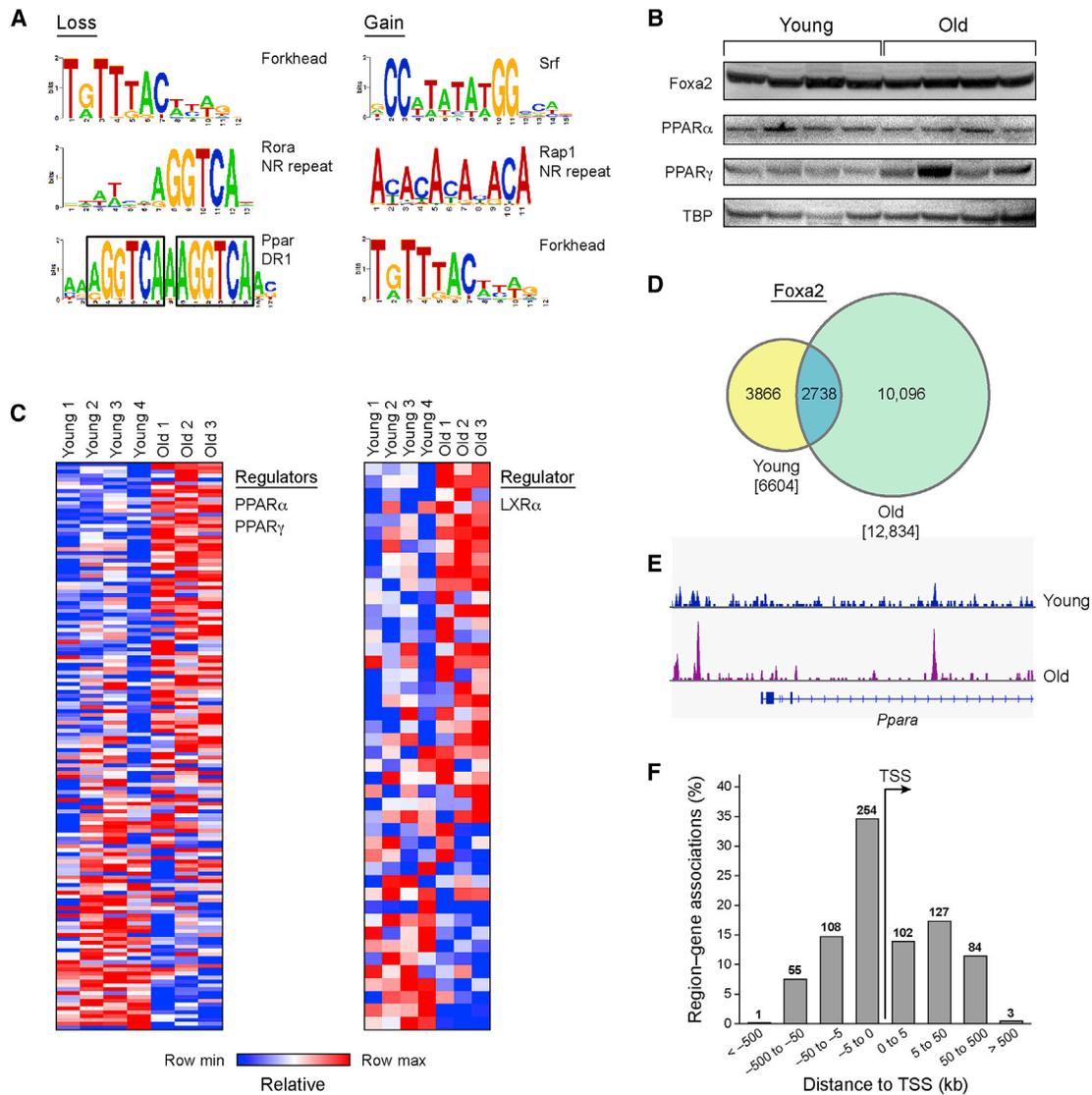


Figure 3. Foxx2 Binds PPAR α Targets in Old Liver

(A) PWM scan analysis identifies numerous forkhead PWMs and nuclear receptor motifs (Rora p value 3.1×10^{-56} , LXR p value 7.6×10^{-85} , PPAR p value 9.7×10^{-76}) in regions of age-dependent loss of nucleosome occupancy. PPAR factors bind a direct repeat (DR-1 element). The repeats are enclosed by black rectangles. Forkhead PWMs, as well as matrices for Srf (p value 3.3×10^{-45}) and Rreb1 (p value 5.4×10^{-98}), and a sequence resembling a telomeric repeat bound by Rap1 (p value 5.9×10^{-64}), are significantly overrepresented in regions of gain of nucleosome occupancy.

(B) Western blot analysis of protein nuclear extracts from four young (3 months) and four old (21 months) mouse livers with antibodies to FOXA2, PPAR α , PPAR γ , and TATA box binding protein (TBP, loading control).

(C) Heatmaps showing expression of Ppar α and Ppar γ targets (left) and LXR α targets (right) with age-dependent nucleosome occupancy loss near the TSS.

(D) Venn diagram showing the results of genome-wide location analysis for Foxx2 in young and old liver, identifying 6,605 binding sites in young and 12,834 in old, of which 2,738 were called bound by both factors by PeakSeq.

(E) ChIP-seq track view in IGV of increased Foxx2 binding in old liver at the *Ppara* locus.

(F) The distribution of regions of additional Foxx2 binding in old livers that correspond to loss of nucleosome occupancy around the TSS. While the majority of changes in nucleosome occupancy occur distally, these sites are found mostly at promoters.

Foxx2 and Hdac3 Exhibit Reciprocal Binding Pattern at PPAR α Targets

Strikingly, we found that Foxx2, a pioneer factor that binds the forkhead motif, occupies substantially more regions in old livers (6,605 in young, 12,834 in old), whereas lamina-associated nuclear receptor corepressor Hdac3 binds markedly less regions

(5,828 in young, 2,930 in old). These differential binding sites for both factors are found at functional PPAR α target genes (Rakhshandehroo et al., 2010) (Figure 5) and contain the PPAR response element. This is consistent with a model where Foxx2 binding likely leads to nucleosome eviction in older livers. Foxx2 cooperates with PPAR receptors, either enabling PPAR

binding at the promoter or interacting with existing PPAR proteins bound to enhancer elements, leading to upregulation of targets involved in lipid synthesis and storage (Figure 6A). Hdac3 may regulate hepatic lipid targets in either of two ways: (1) through GAGA sites bound by cKrox/Hdac3 or (2) by repressing PPAR sites in young, but not old, livers (Figure 6B). Together, the reciprocal binding pattern of Foxa2 and Hdac3 contributes to gene expression changes leading to steatosis in aged liver.

DISCUSSION

Here, we used an unbiased approach to find candidate regulators that affect age-dependent metabolic dysfunction. Since nucleosomes and transcription factors compete for DNA binding (Workman and Kingston, 1992), mapping genome-wide nucleosome composition and tracking changes in nucleosome occupancy in aged mice *in vivo* allowed us to test for differences in transcription factor binding that are responsible for downstream gene regulation governing age-dependent phenotypes.

Motifs bound by forkhead transcription factors and nuclear receptors are significantly overrepresented in regions of age-dependent loss of nucleosome occupancy. We have examined binding of Foxa2 in young and old livers, and it is likely that other Fox factors, especially Foxa1 and Foxa3 and members of the Foxo subfamily, could play a role in this process, and that possibility should be explored further. While nucleosome occupancy dynamics observed in aged livers associates with distal enhancers, elements bound by forkhead transcription factors and nuclear receptors in young livers (Bochkis et al., 2012; Lefterova et al., 2008), we find that most Foxa2 sites that are bound only in old livers and correspond to regions of decreased nucleosome occupancy are found near the promoters. These sites are also enriched for the PPAR/DR-1 element, suggesting that additional Foxa2 binding might enhance accessibility and enable recruitment of PPAR factors to these elements (Figure 6A). We also observe upregulation of PPAR-dependent gene expression for genes with a nucleosome loss at the promoter. A recent study has challenged the classical model of nuclear-receptor-dependent gene regulation, reporting that LXR α and PPAR α binding to their target loci in the liver is largely ligand dependent, with the agonists enabling the receptors to occupy less accessible sites (Boergesen et al., 2012). Two additional reports involving progesterone receptor and estrogen receptor showed that nucleosome occupancy observed in unstimulated cells is significantly depleted upon hormone activation (Ballaré et al., 2013; Tropberger et al., 2013), allowing for nuclear receptor binding. Our findings are consistent with this revised model and suggest that nucleosome dynamics may mediate ligand-dependent activation of “metabolic” nuclear receptors.

While Foxa2 binding sites are also enriched for the PPAR/DR-1 element, we cannot pinpoint which PPAR receptor (PPAR α , PPAR γ , or PPAR δ) binds these sites and in which physiological condition. PPAR α mediates the hepatic fasting response, and binding of this factor should also be examined in the fasted state. Hence, binding of PPAR receptors should be explored in young and old livers to determine the relationship between the factors and their roles in aged livers.

We find that shifts in hepatic gene expression in physiological aging mirror differences observed in progeroid conditions. Changes in nucleosome occupancy are associated with our inferred derepression of nuclear receptors regulating hepatic lipid metabolism, leading to fatty liver (Figure 6). Examining changes in nucleosome occupancy *in vivo* highlighted lamina-associated regulators, Hdac3 and Srf, whose role in age-dependent metabolic dysfunction should be explored further.

Histone deacetylases related to Hdac3, Hdac1, and Sirt1 are known to play important roles in aging liver (Jin et al., 2011; Willis-Martinez et al., 2010). Liver-specific deletion of *Hdac3* leads to fatty liver, a phenotype associated with aging, due to derepression of nuclear hormone receptor-dependent gene expression (Sun et al., 2012; Knutson et al., 2008). *Hdac3* mutant livers also exhibit upregulation of mTOR signaling similar to a model of premature aging due to hepatocyte-specific ablation of *Foxa2* (Bochkis et al., 2013). Deletion of *Hdac3* also impacts DNA repair and reduces heterochromatin content, as observed in aging nuclei (Bhaskara et al., 2010). Loss of Hdac3 binding and transcriptional derepression of targets is observed in adipocytes in a mouse model of progeria (Karakasilioti et al., 2013). Hence, it is likely that Hdac3 is a pivotal regulator of epigenetic and metabolic changes during chronological aging.

The second candidate, Srf, regulates liver proliferation, hepatic lipid metabolism, and growth hormone/Igf-1 signaling crucial to longevity (Sun et al., 2009). Transcription factors, including Hif1a, Hsf1, and Xbp1, that govern different stress responses, similar to Srf, affect gene expression during aging (Henis-Korenblit et al., 2010; Hsu et al., 2003; Kang et al., 2005). Loss of Srf in the liver also alters mRNA levels of histone proteins and chromatin regulators, similar to changes seen in aged livers. A recent study reported that lamin A regulates Srf mRNA levels and Srf-dependent gene transcription (Swift et al., 2013), providing another link to aging.

Notably, “nuclear lumen” genes, including a number of histone transcripts, were highly overrepresented in targets changed in older livers. Histone expression has been reported to decline in a number of aging paradigms (Feser et al., 2010; Celona et al., 2011; Liu et al., 2013). In contrast, we found that whereas some histone transcripts are downregulated with age, others are upregulated (Figures S2A–S2C). Downregulated histone H2 transcripts included replication-dependent (Hist2h2aa and Hist1h2b) and replication-independent genes (H2afx). H2afx is the principal chromatin component involved in DNA repair, and reduced levels of this histone could explain defects in DNA repair in aged livers. Histone variants differ in stability and DNA binding and play distinct functions in the nucleus (Talbert and Henikoff, 2010). Changing composition of histone variants in aged tissues *in vivo* could impact gene regulation and should be investigated further.

Premature aging, due to either mutation in lamin A or defects in DNA repair, is associated with dysregulation of lipid homeostasis and upregulation of PPAR-dependent gene expression (Niederhofer et al., 2006; Savage, 2009). We find that similar pathways, also implicated in metabolic syndrome, are perturbed in chronologically aged livers. We suggest a relationship between lamina-associated factors and age-dependent dysregulation of hepatic lipid metabolism. Whether lamina-dependent

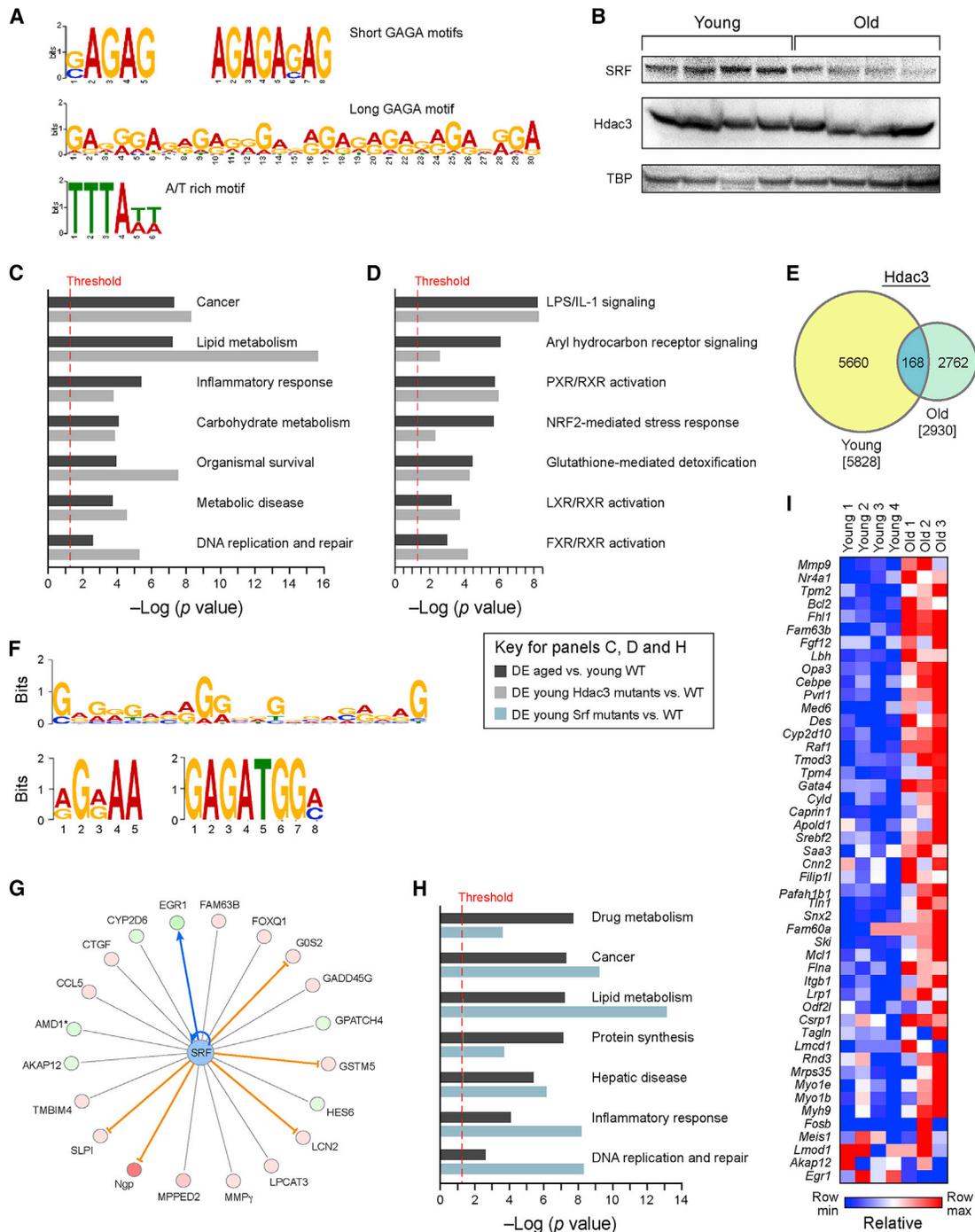


Figure 4. Regulators Associated with Nuclear Lamina Are Implicated in Mediating Age-Dependent Dysfunction in the Liver

(A) GAGA repeat motif (short motif p value 1.8×10^{-11} , long motif p value 2.3×10^{-8}), bound by a transcriptional repressor cKrox (*Zbtb7b*) at the nuclear lamina, and an A/T rich sequence associated with lamina-associated domains (LADs), are enriched in regions of gain of nucleosome occupancy.

(B) Western blot analysis of protein nuclear extracts from four young (3 months) and four old (21 months) mouse livers with antibodies to SRF, HDAC3, and TATA box binding protein (TBP, loading control).

(C and D) Comparison of overrepresented biological functions (C) and pathways (D) between differentially expressed genes in aged WT livers and young liver-specific Hdac3 mutants (K-S p value 2.2×10^{-16}).

(E) Venn diagram showing the results of genome-wide location analysis for Hdac3 in young and old liver, identifying 5,828 binding sites in young and 2,930 in old, of which 168 were called bound by both factors by PeakSeq.

(F) GAGA repeat motif (short motif p value 7.0×10^{-44} , long motif p value 3.6×10^{-13}) is also enriched in regions bound by Hdac3 in young liver.

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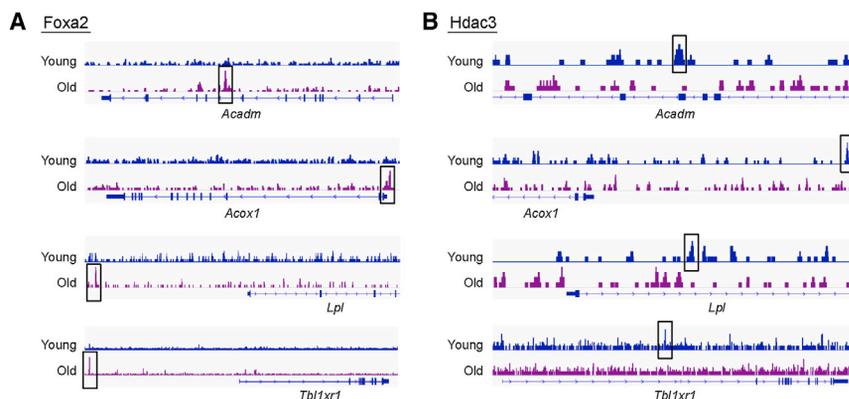


Figure 5. Foxa2 and Hdac3 Exhibit Reciprocal Binding Pattern at PPAR α Targets
(A and B) Chip-seq track view of (A) Foxa2 and (B) Hdac3 binding in young and old livers at the loci encoding *Ppara* targets.

mechanisms could mediate age-onset degeneration in other tissues remains to be explored.

EXPERIMENTAL PROCEDURES

Young and Old Mice

Male mice (C57BL6) were purchased from the National Institute of Aging aged rodent colony (Charles River Laboratories). Four young (3 months), and three old (21 months) mice were used for RNA studies. Two biological replicates of young and old mice were used for micrococcal nuclease digestion and sequencing. Two biological replicates of young and old mice were used for chromatin immunoprecipitation and sequencing. Animals were housed for a week after arrival to acclimate to the light-dark cycle at the Massachusetts Institute of Technology (MIT) facility before tissue harvest. All animal work was approved by MIT's Committee on Animal Care (CAC protocol number Regv-0612-058-15).

RNA Isolation and Profiling

Liver RNA isolation and quantitative real-time PCR was performed as previously described (Bochkis et al., 2013). Total RNA (2 μ g) was enriched for mRNA (Dynabeads mRNA direct kit, Life Technologies) used for strand-specific library preparation. mRNA was fragmented (Ambion fragmentation buffer, 70°C for 3 min), dephosphorylated, and concentrated (RNA Clean & Concentrator columns, Zymo Research). Ligation with RNA adapters was followed by first-strand cDNA synthesis (AffinityScript reverse transcriptase enzyme). RNA was degraded (EDTA and NaOH) while adapters containing barcodes were ligated to cDNA, which subsequently was amplified for ten cycles of PCR (KAPA HiFi DNA Polymerase, Kapa Biosystems). Libraries were sequenced on an Illumina HiSeq 2500 (31 bp paired-end reads; Table S1).

Micrococcal Nuclease Digestion and Sequencing

Digestion was performed on snap-frozen liver, as previously described (Umlauf et al., 2004) with a few modifications. Briefly, 100 mg of tissue was pulverized (Covaris cryoPrep impactor), and nuclei were purified using a sucrose gradient. The nuclei were digested with MNase (Roche catalog number 10107921001) at 37°C for 12 min. Several titrations of MNase enzyme were tested. Samples that were well digested, with most of the input as a mononucleosome (150 bp) and a slight band of a dinucleosome (300 bp) visible on a BioAnalyzer trace, to prevent overdigestion, were used for subsequent library preparation. Libraries (starting material 10 ng) were made as previously described (Bochkis et al., 2012) and sequenced on an Illumina HiSeq 2000 instrument (36 bp single-end and 25 bp paired-end read; summary in Table S4).

(Seven Hills Bioreagents, WRAB-1200) and rabbit antibody to HDAC3 (Abcam, ab7030) were used for immunoprecipitation. Libraries were sequenced on an Illumina HiSeq 2500 with 60 bp single-end reads (Table S6).

Western Blot Analysis

Nuclear extracts preparation and protein immunoblot analysis were performed as reported previously (Bochkis et al., 2008). The primary antibodies used were rabbit antibody to FOXA2 (Seven Hills Bioreagents, WRAB-1200, 1:5,000), rabbit antibody to HDAC3 (Abcam, ab7030, 1:5,000), rabbit antibody to PPAR α (Santa Cruz, sc-9000, 1:100), rabbit antibody to PPAR γ (Santa Cruz, sc-7196, 1:100), rabbit antibody to SRF (Santa Cruz, sc-335, 1:200), and rabbit antibody to TBP (Santa Cruz, sc-273, 1:100).

RNA-Seq Data Analysis

RNA-seq reads were aligned with TopHat (Trapnell et al., 2009) to mouse genome build mm9 (library and alignment statistics in Table S1). Expression levels were calculated using RSEM (Li and Dewey, 2011). Differential expression analysis of RNA-seq (p value < 0.05) was performed in R using EdgeR package (Robinson et al., 2010), with a Benjamini-Hochberg FDR of 5%.

Functional Analysis

Ingenuity curates information gathered from the literature, as well as genomic experiments (microarray, RNA-seq, and ChIP-seq) to determine sets of targets controlled by a regulator (a transcription factor, chromatin remodeler, kinase, small molecule, etc.) and the logic of this regulation (activation or repression). IPA's analysis method compares the overlap of each such set of targets with the subset of genes that are controlled by each regulator in an experimental gene list and reports the p value of the overlap (Fisher's exact test). If the overlap is significant, and if the expression changes agree with those expected from the regulatory connection (genes activated by the regulator are activated in the analysis data set and repressed genes are repressed, for example), IPA analysis predicts whether the regulator associated with the gene targets is itself activated or inhibited. Analysis of overrepresented functional categories and upstream regulators in IPA and heatmap generation was performed as described previously (Bochkis et al., 2013).

ChIP-Seq Analysis

Reads were aligned to the mouse genome (mm9; NCBI Build 37; Table S6) using BWA (Li and Durbin, 2009). Duplicate reads were removed. Reads (phred score > 20) that aligned uniquely were used for subsequent analysis. Data from two biological replicates were merged for each condition (Foxa2 young, Foxa2 old, Hdac3 young, and Hdac3 old). PeakSeq (Rozowsky et al., 2009) was used to identify bound peaks against input controls (Foxa2: FDR 10%, q-value = 0.12; Hdac3: FDR 5%, q-value = 0.2).

(G) IPA Analysis of Srf-dependent network of genes differentially expressed in older livers suggests that Srf activity is downregulated with aging (blue line represents activation, orange line repression, and gray line the association with expression change).

(H) Comparison of overrepresented biological functions in aged WT-type livers and young liver-specific *Srf* mutants.

(I) Heatmap of *Srf* target genes with age-dependent gain in nucleosome occupancy (replicate 2; pattern similar for replicate 1).

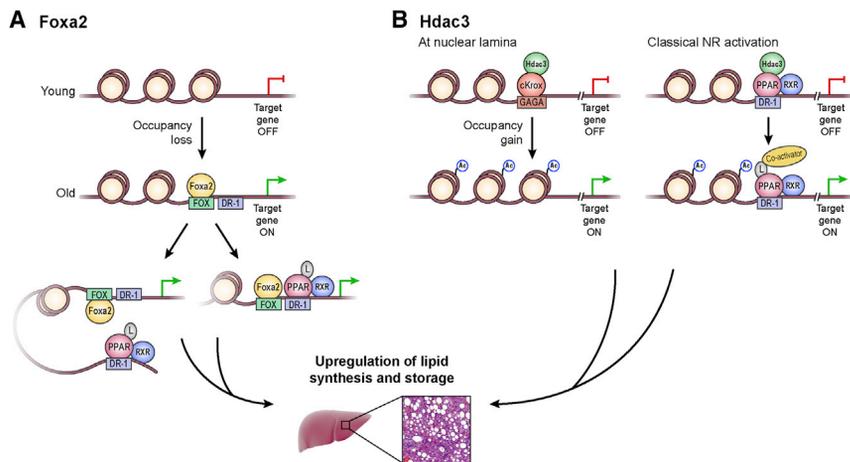


Figure 6. A Model Relating Chromatin Changes to Development of Fatty Liver during Aging

Change in nucleosome occupancy in aged liver is associated with upregulation of nuclear receptor targets and development of steatosis.

(A) Foxa2 binding leads to nucleosome eviction in older livers. Foxa2 cooperates with ligand-activated PPAR receptors (L, ligand), either interacting with existing PPAR proteins bound to enhancer elements (left) or enabling additional PPAR binding at the promoter (right), leading to upregulation of targets regulating lipid synthesis and storage.

(B) Hdac3 regulates hepatic lipid targets in two ways: (1) at the nuclear lamina through GAGA sites bound by cKrox/Hdac3 (left), and (2) by repressing PPAR sites in young, but not old, livers in a classical mechanism of nuclear receptor (NR) action. Regions with a GAGA motif, bound by cKrox (*Zbtb7b*)

in complex with Hdac3, inhibit expression of lipogenic targets in young livers. Age-dependent gain in nucleosome occupancy at these locations leads to eviction of histone deacetylase Hdac3. Nucleosomes can now be acetylated, leading to active transcription of nuclear receptor targets. In addition, in a classical model of nuclear receptor activation, Hdac3 binds unliganded PPAR α in young livers and is evicted upon agonist stimulation in old livers. A coactivator with histone acetyltransferase activity is recruited to the PPAR complex. Nearby nucleosomes are acetylated, and gene expression of the targets is turned on. The reciprocal binding pattern of Foxa2 and Hdac3 at loci encoding PPAR α targets contributes to dysregulation of hepatic lipid homeostasis during aging.

MNase-Seq Analysis

Reads were aligned using BWA (Li and Durbin, 2009) to mm9. Duplicate reads (replicate 1) and fragments (replicate 2) were removed from the analysis. Reads (replicate 1) and fragments (replicate 2; phred score > 30) that aligned uniquely were used for subsequent analysis. Reads for each sample were down sampled to the same number (150 million). Nucleosome calls (~8.5 million nucleosome peaks) and changes in nucleosome occupancy were calculated by the DANPOS software (p value < 1×10^{-7} in Poisson test, parameters: w = 150, d = 150, extend = 148 for single end, 74 for paired end).

Regions Annotation

Regions of changed nucleosome occupancy were associated to closest genes with the GREAT analysis tool (McLean et al., 2010). Overlap with histone modifications and CTCF binding (ENCODE data, 8-week mouse liver (Karolchik et al., 2014) was computed using Galaxy genome analysis tools (Hillman-Jackson et al., 2012). Sequencing reads were visualized with the Integrative Genome Viewer (IGV) (Robinson et al., 2011). To compute correlations between the biological replicates, we trimmed reads from replicate one (single end) to 25 bp and disregarded pairing information in reads from replicate two (paired end) and used single reads (read 1) of 25 bp. Reads were aligned using BWA to *Mus musculus* genome build mm9, and duplicate reads were removed from the analysis. Each read alignment was extended to 150 bp (size of a nucleosome), and resulting data were used to compute genome coverage and Spearman correlation.

Motif Analysis

MEME was used for de novo motif finding (default parameters: zero or one motif per sequence, minimum width = 6, maximum width = 50, maximum number of motifs = 3) (Bailey et al., 2009). PscanChIP (Zambelli et al., 2013) was utilized to identify overrepresented PWMs from Jaspar Version 5.0_Alpha (Vlieghe et al., 2006) and Transfac 7.0 Public (Matys et al., 2006) databases (parameters: assembly = mm9, background = mixed) among sequences corresponding to age-dependent change in nucleosome occupancy (150 bp nucleosome-wide regions with significant change in occupancy as calculated by DANPOS).

Statistical Analysis

The Kolmogorov-Smirnov test was performed to test the distributions of fold changes of targets differentially expressed in both mutant mice (Foxa2 [Bockkiss et al., 2013] and Hdac3 [Knutson et al., 2008]) and in aged WT livers and fold changes of all genes differentially expressed in old versus young livers. The

chi-square test was used to assess whether a given chromatin mark is over-represented in the set of age-dependent nucleosome occupancy changes (as compared to a background set).

ACCESSION NUMBERS

Genomic data from this study have been deposited to the NCBI Gene Expression Omnibus under accession number GSE58006 (GSE57809 for RNA-seq, GSE58005 for MNase-seq, and GSE60393 for ChIP-seq).

SUPPLEMENTAL INFORMATION

Supplemental Information includes three figures and six tables and can be found with this article online at <http://dx.doi.org/10.1016/j.celrep.2014.09.048>.

AUTHOR CONTRIBUTIONS

I.M.B. developed the project, performed experiments and data analysis, and wrote the manuscript. D.P. and J.C. analyzed the data. A.R. edited the manuscript and provided funding for the study.

ACKNOWLEDGMENTS

We thank D. Thompson and R. Majovsky for critical reading of the manuscript and L. Gaffney, R. Raychowdhury, H. Whitton, and I. Wortman for technical assistance. This work was supported by NHGRI CEGS P50 HG006193 and HHMI (A.R.). I.M.B. was supported by National Diabetes and Digestive and Kidney Diseases Institute K01 award DK-101633.

Received: May 16, 2014
Revised: August 15, 2014
Accepted: September 24, 2014
Published: October 23, 2014

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